

Supplementary Table S1. Gene-specific primer pairs used in RT-PCR experiments.

Gene	Primer (5' to 3')
<u>For RT-PCR</u>	
At3g04160 (U11-48K)	Forward: ATGGATCGACCACCGTCGTT Reverse: TGGGCAACGAAGTGAATGAAGAA
At1g56280 (Di19-1)	Forward: CTTTAGAGTCGAAGAACGCGGTATGC Reverse: GAGAACTCTGTGATCGTGTAGGTGAG
At1g02750 (Di19-2)	Forward: CGATATAGCAGTGGAGTATCCATGC Reverse: GGCAACTCATCAAGATGAGGTTGAAG
At3g05700 (Di19-3)	Forward: GGGAAAGAGTTTGCTTGCCCTTTCTG Reverse: CCTCCAAAGAGGCTCTGAAAGTTTCC
At3g06760 (Di19-4)	Forward: CAGAGTTTATATGCCCTTTTTGCGCTG Reverse: GAAGATCCACCAAGAAGAGACTGTAG
At2g30470 (HSI2)	Forward: GGTAAGCTAATGAGTCCAGAAAGTAC Reverse: CATGGACTGTATGCATGGAGTGAC
At5g22370 (QQT1)	Forward: GAAGGTTGCTATTGTTAATCTGGATCC Reverse: CTTGCATCAATGCCGGCAAATATG
At5g44200 (CBP20)	Forward: GAGAGGATACTGAAGATGCAGTCAAG Reverse: CTTCTTGAGTTATCGCGGCCATAG
At1g54370 (NHX5)	Forward: CGTGGTTTAATTTCCACGAAGAGTTC Reverse: GAAGAGTATAGAAACAATGCCGGAG
At1g79610 (NHX6)	Forward: GTTGGTGGTTTAGCGAACATCTCGAAC Reverse: CGATACGATGCCAGATAGACTGAGAC
At2g36010 (E2FA/E2F3)	Forward: CATGCTTGCCTATTGCTTCTAGCTC Reverse: CAACAGCTGTTGGTGCTATTTTCGC
At5g22220 (E2FB/E2F1)	Forward: GTTCTGATGAACCCTCTAATGTCC Reverse: GTGATCTCGTAGCAGTGGATTCTTC
At5g03740 (HDT3)	Forward: GAATGGCAAACCACTTCACCTAG Reverse: GTCATCATCTTGCTTGGCTTTGAC
At2g21880 (RabG2)	Forward: GGTCATTGTTCTTGGAGATAGCGG Reverse: GAGGAACTCTGTGTGCCAGTTATTG
At1g16540 (ABA3/los5)	Forward: GATTATTATGGATACCCAGATGGTCCC Reverse: GCATATTCCCTAATAACCAAGTACACTG
At2g41740 (VLN2)	Forward: GATTTGACTGTTATAACAATGTCAAC Reverse: GCAAGTATATAACCGGGTCTC
At3g57410 (VLN3)	Forward: TAGCCACTAATAACAAAATGTCTGG Reverse: TCCTCCTCCGGTTTTTTAAATC
At3g04630 (WDL1)	Forward: TGCTTATGGACCGAAACGC Reverse: TTCAAGCTCGTTCCTCTCAG
At5g12210 (RGTB1)	Forward: GCTGACAAGCATGTACGTTAC Reverse: CAACGCAGCAGAAAATTTGTC
At3g12070 (RGTB2)	Forward: GAAGATGAAGTTGTGTCATGGG Reverse: ATGAGACTCCGCTCCAGGTG
At3g51460 (RHD4)	Forward: AAGCTCTTAAGTGTAGCAGAAAAG Reverse: CTTGTTAACGAGATCAACTGCC
At4g02560 (LD)	Forward: CGCAAATTGATTCGCAGAGTCAATTGC Reverse: GAATATCATCAGCAGACATTGGGCG
At5g27380 (GSH2)	Forward: GATCAGATTGGCATAGACTCTGAACG Reverse: GACGGATGATCATTAGGAGTATAGCC
Actin	Forward: CAGCAGAGCGGGAAATTGTAAGAG Reverse: TTCCTTTCAGGTGGTGCAACGAC

For real-time RT-PCR

GA20ox	Forward: ACATGGGTTTCAGCCATTTGGGAAGGTGTA Reverse: AGATGGGTTTGGTGAGCCAATCTGAAAAGG
GA3ox	Forward: CCAACATCACCTCAACTACTG Reverse: CTCTTTCCATGTCACCGATTG
GA2ox	Forward: GTGTCGTACAAGGTGTTGGAGA Reverse: CCAAGTCTATAATCACCAAGC
Actin	Forward: TGCGATAATGGAAGTGGTATGG Reverse: AAGACAGCCCTGGGCGCATCA

Supplementary Table S2. Gene-specific primer pairs used in quantitative analysis of splicing efficiency by real-time RT-PCR.

Gene		Primer (5' to 3')
HSI2	exon1	Forward: CCA ACA TGG TCC ATT GCT CT
	intron1	Reverse: CTG TCC AGA ATA AAA TAT TTG GC
	exon2	Reverse: CAG GCT TTT GGA AGA ACT AG
QQT1	exon1	Forward: GCC AAA TAT TTT ATT CTG GAC AG
	intron1	Reverse: CAA CAT ACA CCT GCA ATT TTT AAC
	exon2	Reverse: AAC ATC TTC TAA CTT GAT CAA TTC
NHX5	exon1	Forward: CTC CGC CGT CAT CGA TTC C
	intron1	Reverse: TAG CAG CAG CGA ATT TGA AG
	exon2	Reverse: GTA ACA ATG GCT CCA AAG TTA G
NHX6	exon1	Forward: CGG CGG CTC TGC GTT CTT T
	intron1	Reverse: CAA AAT TAG TAA GAG AAG TGC TC
	exon2	Reverse: AGT GAA CCT GTT AGT CCT CG
E2FA	exon1	Forward: CTC AGG ATC TAC AGG ACA CC
	intron1	Reverse: CTG CAC AAC TTC CTA TTT CTA TG
	exon2	Reverse: GTT TTC CAA ATA TCC GTC ATG C
E2FB	exon1	Forward: AAG CAC CGA AAG AAA CAT GG
	intron1	Reverse: AAG AGA GAT TAG ATT AAA TCA CAG
	exon2	Reverse: GTG GTC TTG ATC AAA TGT AAT C
HDT3	exon1	Forward: GTT TTG AGT GTG ACT GTT GG
	intron1	Reverse: TAT CTT CCA GTC TTA AAT ATA GAG
	exon2	Reverse: CAG GAG CAG GAA CAG CTT C
GSH2	exon1	Forward: GAT TGG CAT AGA CTC TGA AC
	intron1	Reverse: CAT AAG AAG TCA AGA TCG ACC
	exon2	Reverse: TCC TCT TGT ACA CTC CCT TC
LD	exon1	Forward: GTA TTG CTG AAA TCA TGG GTG
	intron1	Reverse: GTT AAC TTA TCC AAT GCA TTG AG
	exon2	Reverse: TGC GGA CTC TTT CCA GCA G
ABA3	exon1	Forward: AGA TGG TCC CAA GAA CAT TC
	intron1	Reverse: CAT TAA AAT CTG AGA TTG AAA GCC
	exon2	Reverse: GAT GCA TTA AAG TAT TCA AGC AC
Di19-1	exon1	Forward: CTT TAG AGT CGA AGA ACG CG
	intron1	Reverse: GAA TAT CTG TAA GTT ATA AAC GTA G
	exon2	Reverse: CAA CAG AAC CAT TAC GAG AAG
Di19-2	exon1	Forward: TAT GAT TTA GTT GAA TTG TGT CAC
	intron1	Reverse: AAT CAC ATT CAT TAT AGC GAC GG
	exon2	Reverse: GCT GAT GAT GAT TCA TAG ATG G
CBP20	exon1	Forward: TCC TGT TCT ACT CTA GAG AG
	intron1	Reverse: TTA ATT GTA AAG AAA CGG TCA GG
	exon2	Reverse: GCT GCC TTT GTG CTT CGA G

Supplementary Table S2. (Continued)

Gene		Primer (5' to 3')
RGTB1	exon1	Forward: TGG CTT TGC CGG TAA CAC TG
	intron1	Reverse: GAT TAA TTA ACA TAT GAA AGT TCC AC
	exon2	Reverse: ACA TTT CAA TAT TGA GAG ACA GC
RGTB2	exon1	Forward: GTG GCT TTG CCG GTA ATA CT
	intron1	Reverse: ACA ACA AAC GAA TTA CCG AG
	exon2	Reverse: ATA TTG AGA GGC AGC ATA TAG C
RHD4	exon1	Forward: GAG CTC ACA ACG CTT GCA TG
	intron1	Reverse: ACG TAA CCA TGA AGC GAA AAT AG
	exon2	Reverse: TGT ACC GTT TCT CCT AGT GC
VLN2	exon1	Forward: GTT CAT TGG CGT TTG TAG TAC
	intron1	Reverse: AAC AGT CCA AAC ATT CAG GAA G
	exon2	Reverse: CAC CCC CTT TAT TCT GTG TTG
VLN3	exon1	Forward: CTC TTG AAG GAA GAG ATT AGC
	intron1	Reverse: AAC ATT ACC AGC AAA CTG GTG
	exon2	Reverse: CAA ATA GAT AAG CGC CAC CC
WDL1	exon1	Forward: ATG CAC CAA ATG TTG GAT CTG
	intron1	Reverse: TAA ACA TCC ATA CAT AAG TAT CAA G
	exon2	Reverse: CAC TCC TAA ACG TTG GAG C

Supplementary Table S3. Comparison of U11-48K proteins and other U11/U12-proteins found in diverse plant species and humans.

48K	Species	Accession No.	aa*	Sequence homology (%)**									
				1	2	3	4	5	6	7	8	9	10
Dicots	<i>A. thaliana</i>	At3g04160	714	75	77	40	33	27	33	29	28	24	
	<i>E. salsugineum</i>	XP_006408216	733	78	39	33	27	32	28	31	23		
	<i>C. rubella</i>	XP_006297086	703	41	31	27	32	30	31	24			
	<i>R. communis</i>	XP_002525479	722	35	29	32	30	31	20				
	<i>G. max</i>	XP_003535384	687	30	32	30	32	19					
Monocots	<i>H. vulgare</i>	BAJ90436	656	58	57	91	10						
	<i>S. bicolor</i>	XP_002466198	481	85	56	13							
	<i>Z. mays</i>	NP_001145685	527	55	12								
	<i>T. urartu</i>	EMS45165	388	3									
Humans	<i>H. sapiens</i>	NP_689764	339										
	Number of amino acids			Sequence homology (%)									
	Arabidopsis	Other dicot and monocot plants											
65K	442	441~467		55~92									
59K	382	384~420		55~93									
35K	333	314~403		60~95									
31K	261	261~275		60~95									
25K	165	155~187		60~95									

*Number of amino acid (aa)

**The sequence homology was calculated using the ClustalW2 program.

Supplementary Fig. S1 Amino acid sequence comparison of U11-48K proteins in diverse plant species. The alignment was made using the ClustalW2 program. Gaps in the sequences are indicated by dashes. The position of CHHC-type zinc finger and arginine-rich region is indicated by a box and a thick line, respectively. Accession numbers of 48K-related proteins are as follow; *A. thaliana* (At3g04160), *R. communis* (XP_002525479), *C. clementina* (XP_006443313), *E. salsugineum* (XP_006408216), *C. rubella* (XP_006297086), *G. max* (XP_003535384), and *M. truncatula* (XP_003590902).

Supplementary Fig. S2 Development-defect phenotypes of the *u11-48k* mutant plants. (A) Schematic presentation and the sequences of artificial miRNA2 (amiR2) along with its target U11-48K (48K). Exons and introns are represented as gray boxes and thick lines, respectively. Downregulation of *U11-48K* in the transgenic plants was confirmed by RT-PCR and real-time RT-PCR analysis. (B) Development-defect phenotypes of the *u11-48k* knockdown plants. The wild-type (WT) and artificial miRNA-mediated knockdown mutant (amiR2) plants were grown in soil for the indicated times, and the abnormal leaf shape, arrested stem growth, and delayed senescence of the *u11-48K* mutant plants are shown. Identical results were obtained from three independent experiments, for which a representative example is shown.

Supplementary Fig. S3 Effect of exogenously applied hormones on the stem length of the wild-type plants. Twenty four-day-old Col-0 plants were treated with 100 μ M GA, 50 μ M kinetin, 5 μ M BR, or 0.5 μ g/ml NAA, and stem lengths of the plants were measured 2 weeks after the application of each hormone. Values are means \pm SE obtained from three independent experiments (n=5).

Supplementary Fig. S4 Splicing of *GA20ox*, *GA3ox*, and *GA2ox* genes in the mutant plants. The splicing patterns of *GA20ox*, *GA3ox*, and *GA2ox* transcripts were analyzed by RT-PCR in wild type (WT), *48k*, and *65k* mutant plants. Exons and introns are represented as gray boxes and thin lines, respectively, and the positions of primers used are indicated by thick lines. The sequences of primer sets are shown. Identical results were obtained from three independent experiments, one of which is shown.

Supplementary Fig. S5 Abnormal splicing of U12 introns in the *u11-48k* mutant plants. The splicing patterns of selected U12 intron-containing transcripts were analyzed by RT-PCR in wild type (WT) and three different knockdown plants (amiR1-1, amiR1-2, and amiR1-3). Identical results were obtained from three independent experiments, one of which is shown.

Supplementary Fig. S6 Comparison of abnormal splicing of U12 introns in the *u11/u12-31k*, *u11-48k*, and *u11/u12-65k* mutant plants. The splicing patterns of selected U12 intron-containing transcripts were analyzed by RT-PCR in wild type (WT), *31k*, *48k*, and *65k* mutant plants. Identical results were obtained from three independent experiments, one of which is shown.

Supplementary Fig. S7 Splicing of selected U2 introns in the mutant plants. The splicing patterns of selected U2 intron-containing transcripts were analyzed by RT-PCR in wild type (WT), *31k*, *48k*, and *65k* mutant plants. Exons and introns are represented as gray boxes and thin lines, respectively, and the positions of primers used are indicated by thick lines. The sequences of primer sets are shown. Identical results were obtained from three independent experiments, one of which is shown.

A. thaliana 1 MDRPPSLPHYQNPENLVEYHPPFNENFNFFRPPPPPLQNPENNYIVSPPPIRELSCTLSSLSKSLSECOITD---SLSQNLALDH
R. communis 1 -----MNPSSAEPDYEQNSNYLIPNFVHSLQPPPPHITITPTTP---LIDLSTLSSLANLLSLSQOTRN---SLS-----
C. clementina 1 -----MSSSYAAPPPTSFQQNPNFNSSSIPGQS-----DLSTLSSLALSLFCHQTLQ---NYSF-----
E. salsugineum 1 MDRRRSFPNYQNPYIFHHQPPFNHNPNFFRPPP--LQNPNTYSIAPSPPPIRELSCTLSSLSKSLSECOITLA---SLSENLALDH
C. rubella 1 MDRPSPFPHNQENPNFHHVPPFNPNFNFFRPPPPLQNPNTYSIAPSPPPIRELSCTLSSLSKSLSECOITLD---SLSQNLALDH
G. max 1 -----MHAGGLPLVDWILEELVRRQSRFQGVRSMMNPSFSSSSLSLSTLSSLNLLTSLNHVLSLTPS-----
M. truncatula 1 -----MNPPEAQLPSPSQLPPLSPPPQCHPPPLLPPLPPLNLLTSSLTNLLFSTQILSTTPQ-----

A. thaliana 87 SLLQKQDENGCFVFCPEFDSNHMPPEALFLHSLRCPNPLDITHLLES---SSYRNTLELPCPEQLNNGDGDLCISLDDLAD--FGSNF
R. communis 71 --LTKPNKNVKEFCFNPNPNHLMPPESLFLHSLRCPSEFQDPPLSLVNSLHMPKTLNQNPNPNFKNSDNALCLISLGGFYN--EFSNF
C. clementina 56 --LTPKQNDNLFCFNPNPQHLMPPELFLHSLRCPPLDLDPPNYRNTLHSSLLNQON--APLTIQDHIQELCFSLDDYLSNVRVSF
E. salsugineum 85 SLLQKQDENGCFVFCPEFDPNHLMPPPEALFLHSLRCPNPLDITHLLES---SSYRNTLELPCPEQLNNGDGDLCISLDDLAD--FGSNF
C. rubella 87 SYLQKQGGNGCFVFCPEFDSNHMPPEALFLHSLRCPNPLDITHLLES---SSYRNTLELPSQVQLNSDAGDLCVSLDELAD--FGTNE
G. max 63 -----PPTLNSNLQCFPNFPHLPLPPELFLHSLRCPSPRPLPLNPN-----SPS-LTYPKTLQSSQQLFSLDSLNSN-----F
M. truncatula 62 ----PQPTTNTLFCLEFNPNHLPPELFLHSLRCPSPRPLPLNPN-----LLSLSYPKTLQSPSIHLSYSLDFTN-----F

A. thaliana 171 FYRDCPGAVKFSFLDGGK---RLLTLFHLVLSVECSDFVGSDF---EKVKKIVLDKCLGVLPDLICAMKNEIDQWRDFP--SSYSSSVLS
R. communis 158 FYKDCPGAVQFSLDSSS---KTEFLPAVLSVECANFVAR---IEDIKGFDINEFRILPSDLVVKREVESWADV--SMYSMAVFC
C. clementina 142 FYQDCPFAVAISFPHASTSISKTLALFGLCMECANVVCSDGAEKNAEAGFGEVGRVLSGLDFTREVESWRDHEHMSYSRNVFC
E. salsugineum 169 FYRDCPGAVNFSFLDGGK---RLLTLFHLVLSVECSDFVGSDF---EKVKKIVLDKCLGVLPDLICAMKNEIDQWRDFP--SSYSSSVLS
C. rubella 171 FYRDCPGAVNFSFLDGGK---RLLTLFHLVLSVECSDFVGSDF---EKVKKIVLDKCLGVLPDLICAMKNEIDQWRDFP--SSYSSSVLS
G. max 134 FYRDCPFAVAISFPHASTSISKTLALFGLCMECANVVCSDGAEKNAEAGFGEVGRVLSGLDFTREVESWRDHEHMSYSRNVFC
M. truncatula 133 FYRDCPFAVAISFPHASTSISKTLALFGLCMECANVVCSDGAEKNAEAGFGEVGRVLSGLDFTREVESWRDHEHMSYSRNVFC

A. thaliana 251 STVCSKVEISALRKKWLVNSTRYGVITDTYMRDHFLLFRLCLSAVKEAGCFERMESDATDVGEQKIMSCKSSSTFPCVFTQVLSWLAS
R. communis 238 AILRLNVIKGSDLRRWIIFNSPRYGVVIVYMRDHISVLFRLCLNAIRREAFSFMGHQMN-----VKISSNCPVLSQVFMWVP
C. clementina 232 AILGLRTVNVISLQKWLNVNSTRYGVITDTYMRDHFLLFRLCLSAVKEAGCFERMESDATDVGEQKIMSCKSSSTFPCVFTQVLSWLAS
E. salsugineum 249 SILGSEANETSLSWLVNSTRYGVITDTYMRDHFLLFRLCLSAVKEAGCFERMESDATDVGEQKIMSCKSSSTFPCVFTQVLSWLAS
C. rubella 247 AILGSKALETSELNSWLVNSTRYGVITDTYMRDHFLLFRLCLSAVKEAGCFERMESDATDVGEQKIMSCKSSSTFPCVFTQVLSWLAS
G. max 213 AILGLGANDRDLTDMWIANSPRYGVITDTYMRDHFLLFRLCLSAVKEAGCFERMESDATDVGEQKIMSCKSSSTFPCVFTQVLSWLAS
M. truncatula 209 AILGIGAKSEMVNWLNSSTRYGVITDTYMRDHFLLFRLCLSAVKEAGCFERMESDATDVGEQKIMSCKSSSTFPCVFTQVLSWLAS

A. thaliana 341 QLVVLYGEGNGKFFALDMFKQCIYESASQVMLFRLEGRRSKSCGVVE---LDDARLRNKDVIEMKPEFNSSGGCGKTLDS---PQVIS
R. communis 318 QLVVLYGERNAKCFATLHFCQICILDVSNMIFP-----LEANKEISTEINGNGSDVRDLKIQEPLBGSIKCETDABVEHVDKEVIF
C. clementina 318 QLVVLYGQVSGKIFALEIFKQCILEASGILLFPLEQSLTESLDLKEGDLTLHASSSGARDVVRQEPFERNANSGLDETVEGTVHVKVIF
E. salsugineum 339 QLVVLYGEGNGKFFALDMFKQCIYESASQVMLFRLEGRRSKSCGVVE---LDDARLRNKDVIEMKPEFNSSGGCGKTLDS---PQVIS
C. rubella 337 QLVVLYGEGNGKFFALDMFKQCIYESASQVMLFRLEGRRSKSCGVVE---LDDARLRNKDVIEMKPEFNSSGGCGKTLDS---PQVIS
G. max 289 QVSLYLYGAANGKAVLVNFVKKICILVGSVILLFPLGDNAAKQESQNLGTESGDPKAKPGAQCCEKKNWIL-----NRKIS
M. truncatula 272 QVSLYLYGVS-GKLEVLNFVKKICILVGSVILLFPLGDNAAKQESQNLGTESGDPKAKPGAQCCEKKNWIL-----NRKIS

A. thaliana 425 VSRVAASVAALHERSLEEKIRAVR---YAQPLRYQRIISLHLSLIPHVSENRNRCSYRPIIDHDLGRPRORSINQDMDRKMKTREELL
R. communis 401 VSRVAASVAALHERSLEEKIRAVR---YAQPLRYQRIISLHLSLIPHVSENRNRCSYRPIIDHDLGRPRORSINQDMDRKMKTREELL
C. clementina 408 VSRVAASVAALHERSLEEKIRAVR---YAQPLRYQRIISLHLSLIPHVSENRNRCSYRPIIDHDLGRPRORSINQDMDRKMKTREELL
E. salsugineum 423 VSRVAASVAALHERSLEEKIRAVR---YAQPLRYQRIISLHLSLIPHVSENRNRCSYRPIIDHDLGRPRORSINQDMDRKMKTREELL
C. rubella 414 VSRVAASVAALHERSLEEKIRAVR---YAQPLRYQRIISLHLSLIPHVSENRNRCSYRPIIDHDLGRPRORSINQDMDRKMKTREELL
G. max 366 VSRVAASVAALHERSLEEKIRAVR---YAQPLRYQRIISLHLSLIPHVSENRNRCSYRPIIDHDLGRPRORSINQDMDRKMKTREELL
M. truncatula 332 VSRVAASVAALHERSLEEKIRAVR---YAQPLRYQRIISLHLSLIPHVSENRNRCSYRPIIDHDLGRPRORSINQDMDRKMKTREELL

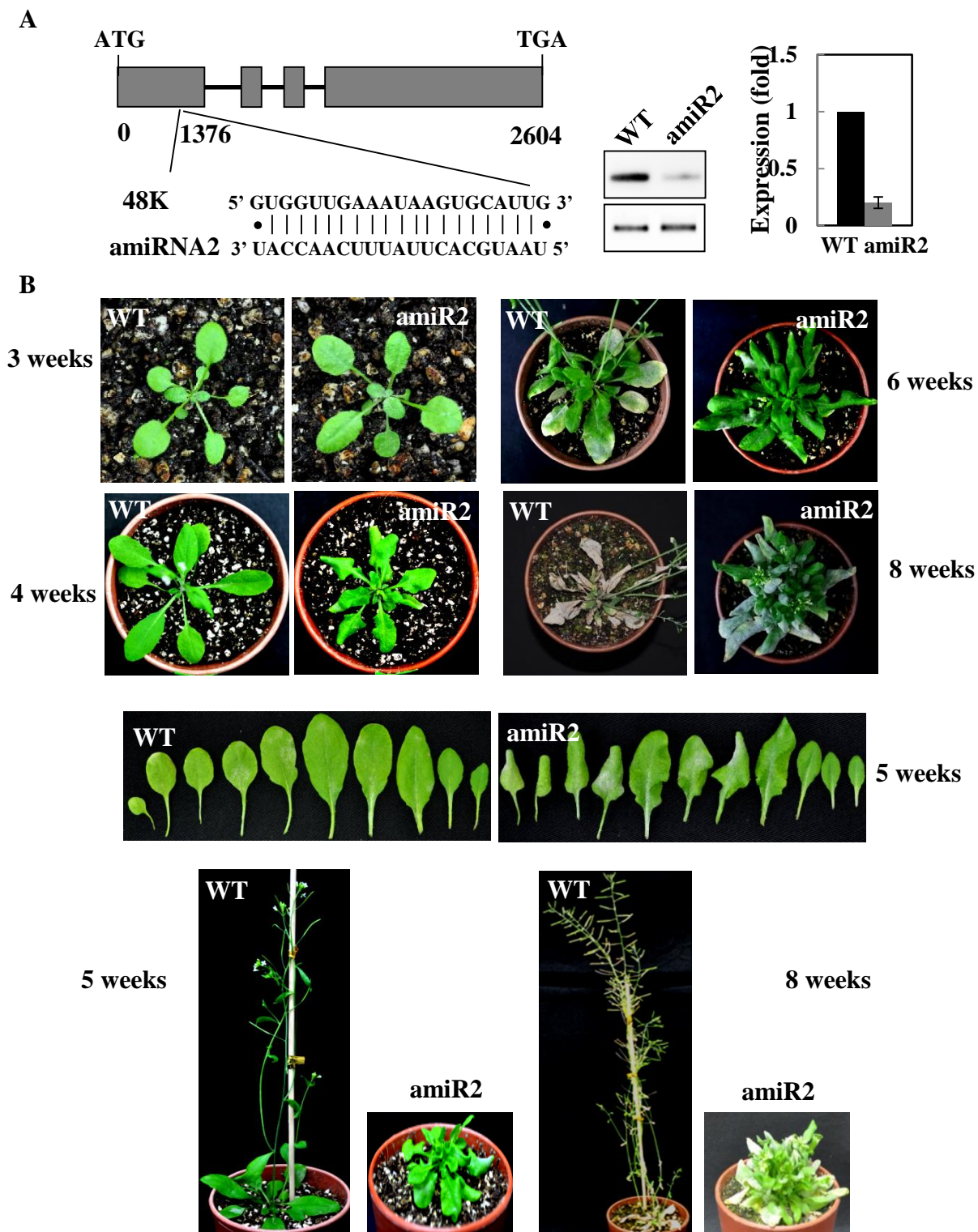
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C. clementina 496 ABERDYKRRRMSYRGKVKRTPROVLEHPIEYIEEIKLAGGIGCFEKGMPLOSSEPIGN---DQKESDFGYS-----IPSTDKW
E. salsugineum 508 ABERDYKRRRMSYRGKVKRTPROVLEHPIEYIEEIKLAGGIGCFEKGMPLOSSEPIGN---DQKESDFGYS-----IPSTDKW
C. rubella 499 ABERDYKRRRMSYRGKVKRTPROVLEHPIEYIEEIKLAGGIGCFEKGMPLOSSEPIGN---DQKESDFGYS-----IPSTDKW
G. max 451 ABERDYKRRRMSYRGKVKRTPROVLEHPIEYIEEIKLAGGIGCFEKGMPLOSSEPIGN---DQKESDFGYS-----IPSTDKW
M. truncatula 417 ABERDYKRRRMSYRGKVKRTPROVLEHPIEYIEEIKLAGGIGCFEKGMPLOSSEPIGN---DQKESDFGYS-----IPSTDKW

A. thaliana 590 -KGENRADIEYPIDNQNQKVKRHDYDSSGSSORQSHRSYKSDRRD-----KLRDRRDR-----NDRRD
R. communis 573 NHYQKQSHIDNNNRSAACKNASQDYERWRKVNHRHSHVYEQKDSDRHGRDYSSASPERHKGHGPLHERDEAFNISKRHDVRSNGK
C. clementina 583 NHYQKQSHIDNNNRSAACKNASQDYERWRKVNHRHSHVYEQKDSDRHGRDYSSASPERHKGHGPLHERDEAFNISKRHDVRSNGK
E. salsugineum 595 -KGENRADIEYPMDRHTDKKRYEYDSSGSSORRSHRSYKQHSDEEYDSSSSORQSHRSYKSDRRD-----NDRRD
C. rubella 586 -KGENRADIEYKDNFNNDKVNREHEYDSSGSSORRSHRSYKSDRRD-----NDRRD
G. max 538 RCSECCSDSNCCDQSKLEDAFSDRYQORHEHRSYCRBEDQONADQGYHRDRHSISPERYSSY-----SDRRD
M. truncatula 507 DHEEQSHITNSDKSKVVDATSRDYQORQGHQGSHEHGGDERGTDQ-ENHRSHASTSPERHRSR-----SDRRD

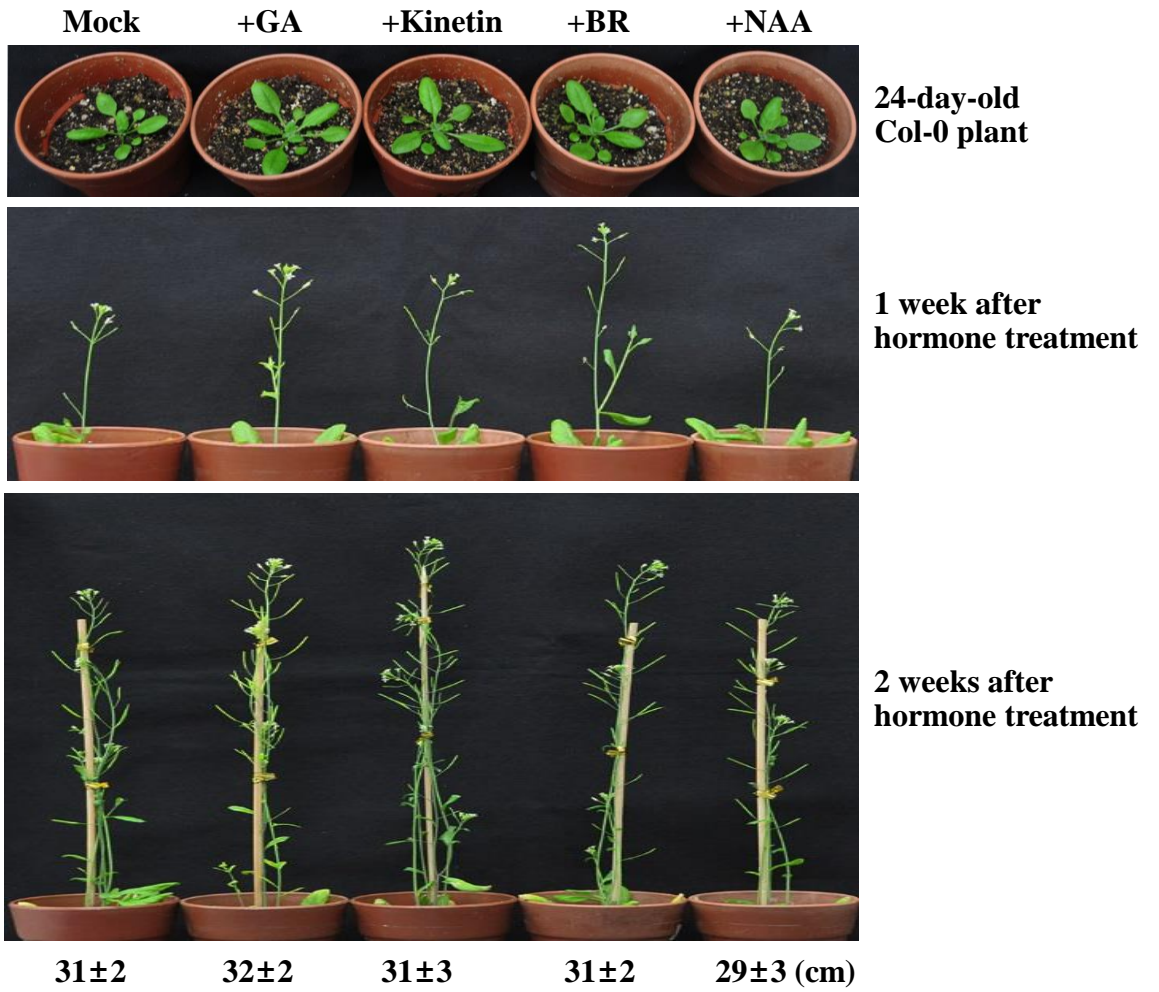
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R. communis 663 SNYQNYSSCFGSDSANDPGVQKDGDKLDVRDWLRLNSYGTSSSTFLVKNAFEDRYDPAE-----
C. clementina 660 MELTANGRIGVERQSLGSSKYCDYRSYSTNSHRRRRRNDHSDTLVNRNAFEDRYDPEESHNRDENVSDGGYIKPE-----
E. salsugineum 666 DEFTRNRKHSLEGS-YQNYSSSEKSSSDYKTKEDDQYDRRSQOPRNLNLFEDRYLPTKE-----
C. rubella 642 DEFTRNRKHSLEGS-YQNYSSSEKSSSDYKTKEDDQYDRRSQOPRNLNLFEDRYLPTKE-----
G. max 604 ---SRSEHSSHNKQDYYPNRKHNSSRIKDGWQKDHRSHSISDFNNAFSDRYDPEESLVICEDDISSDSKYIKSKDFYDKEGY---
M. truncatula 572 ---SRSEHHRVHEEKQDYSGRKKYNNSSRTKDRWQNDHKNHISDSFS-----RSDPESRSGVGEHDISSDDKYIKFLCLLEALVRLSC

A. thaliana -----
R. communis -----
C. clementina -----
E. salsugineum -----
C. rubella -----
G. max -----
M. truncatula 653 DEVTSMGLQMLVTTEEVWAQHETVFNFFLLQEKSSQFFLCTRAEQSYVVRWIVTCLCPFFRT

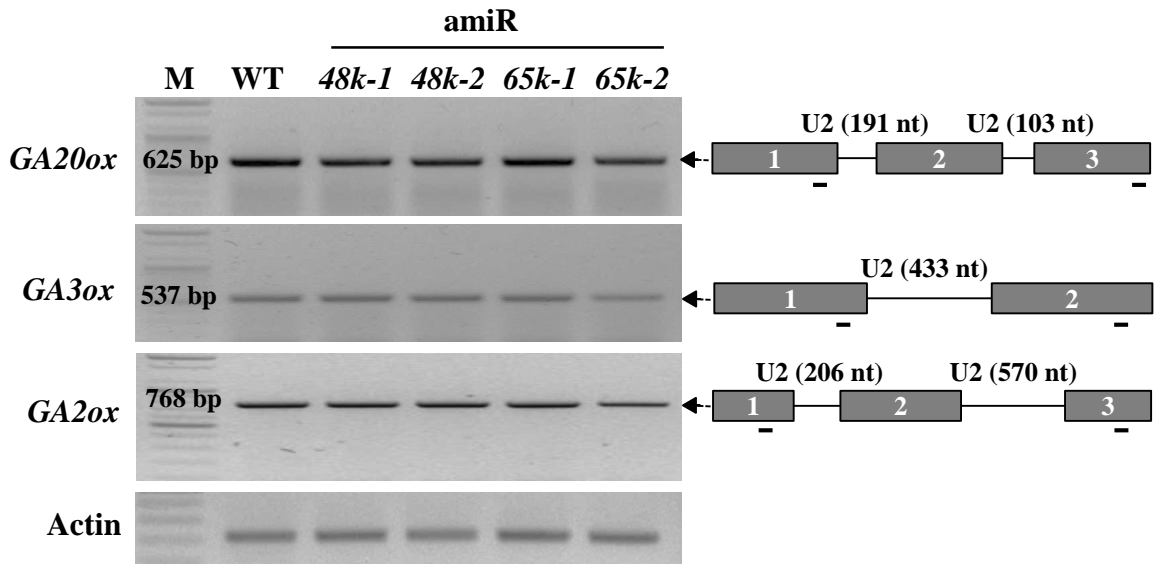
Supplementary Fig. S1



Supplementary Fig. S2



Supplementary Fig. S3



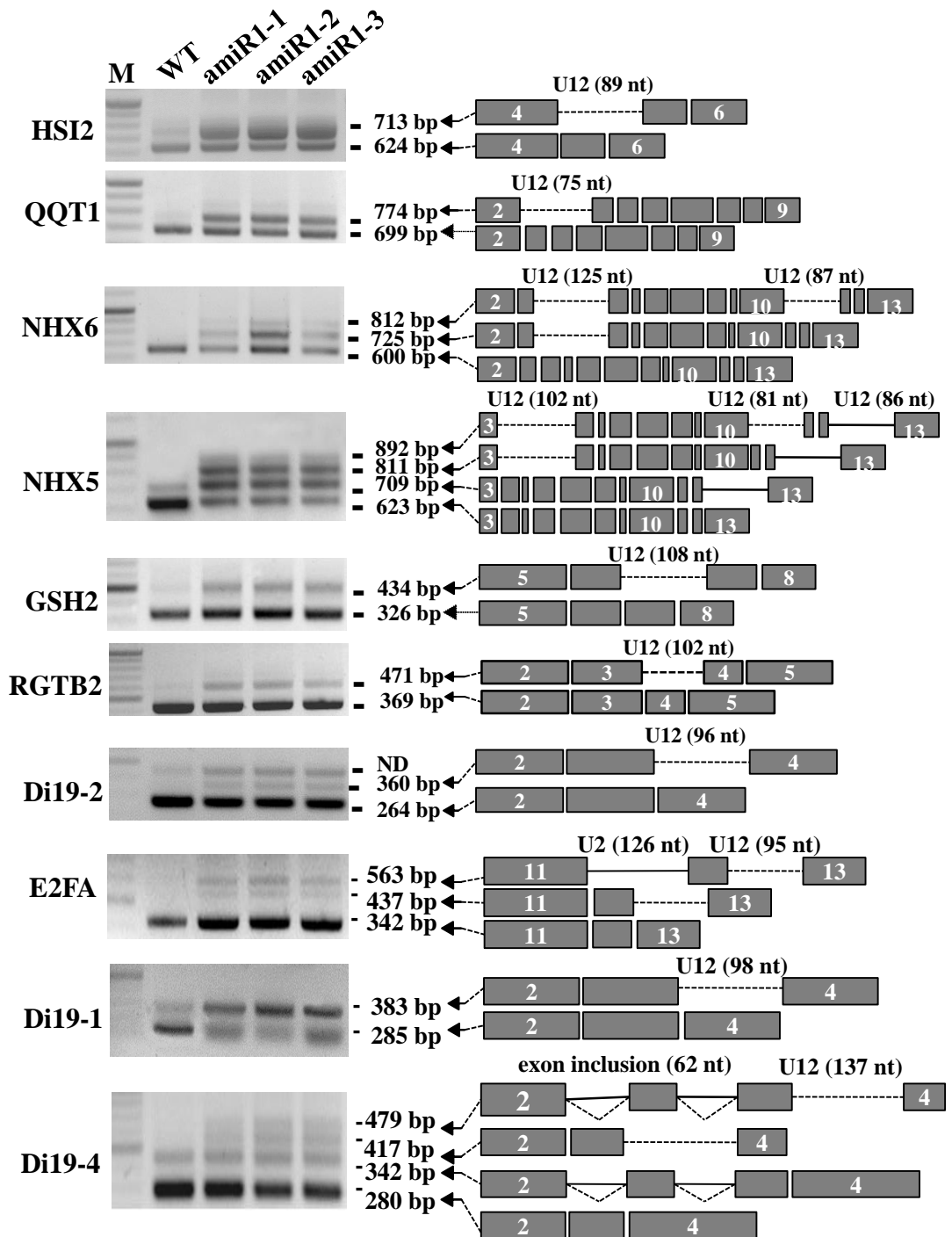
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Reverse: AGATGGGTTTGGTGAGCCAATCTGAAAAGG

GA3ox Forward: CCAACATCACCTCAACTACTG
Reverse: CTCTTTCATGTCACCGATTG

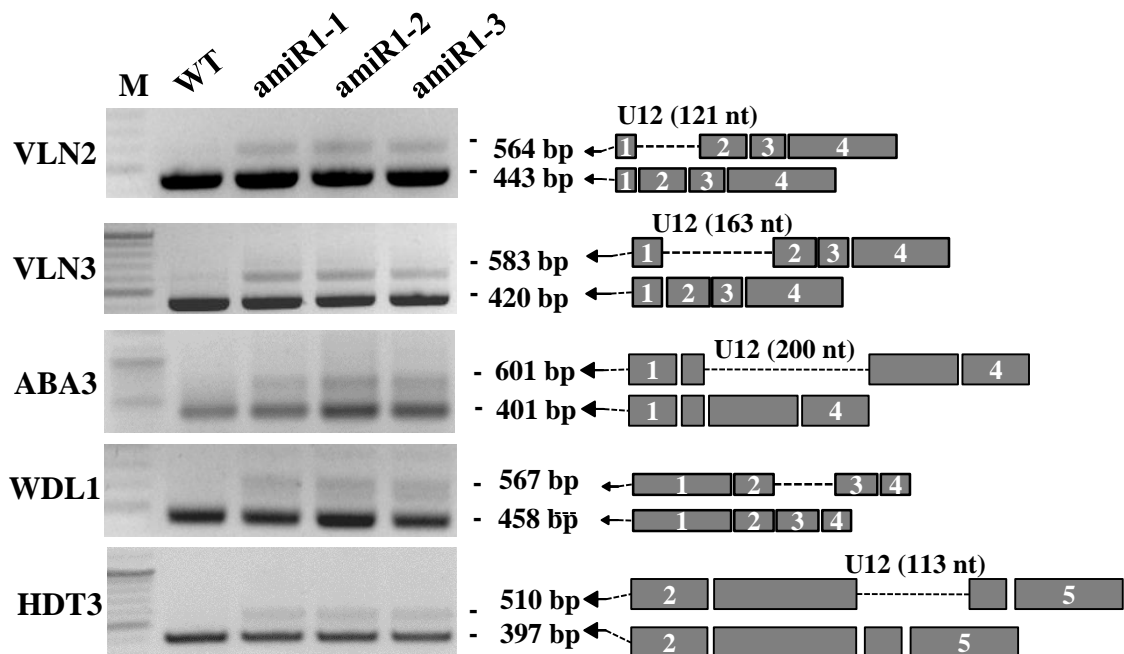
GA2ox Forward: GCAGGAGGCTATTGGCTTCTTC
Reverse: CCAAGTCTATAATCACCAAGC

Actin Forward: CAGCAGAGCGGGAAATTGTAAGAG
Reverse: TTCCTTTCAGGTGGTGCAACGAC

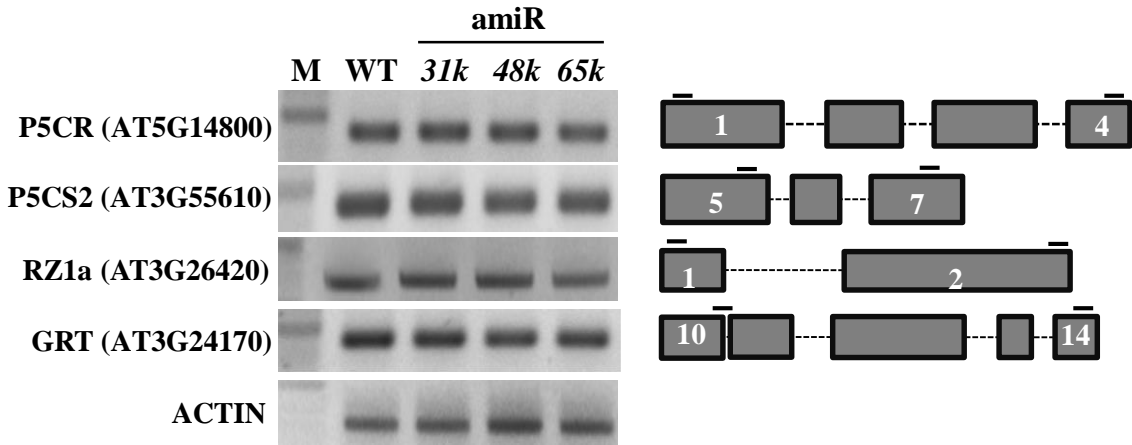
Supplementary Fig. S4



Supplementary Fig. S5



Supplementary Fig. S5 (Continued)



P5CR Forward: 5' TCCGGCGGAGAGTTTCAAGGTAG 3'

P5CR Reverse: 5' CAGCGGCAGGTGTATTAGGCATC 3'

P5CS2 Forward: 5' TGATGCTATAAGCACTCGCAGAGCC 3'

P5CS2 Reverse: 5' TTGATGGAACAGGGTACCAACACG 3'

RZ1a Forward: 5' ATGTCTGAAGATCCGGAG 3'

RZ1a Reverse: 5' GTGGAAACCACCAGCTCTG 3'

GRT Forward: 5' AGGCTGGAGCTGTGAAGGTTGAC 3'

GRT Reverse: 5' CATGATCTCAGCTGCATCAGGACC 3'

Supplementary Fig. S7