

## **Supplementary Information**

### **Sesterterpene ophiobolin biosynthesis involving multiple gene clusters in *Aspergillus ustus***

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**Table S1.** *Aspergillus ustus* 094102 genome assembly and main feature

Statistic	Value
Coverage	>68×
Genome assembly size (Mb)	40.09
Number of scaffolds	174
Scaffold N50(Kb)	1,814
Number of contigs	301
Contig N50(Kb)	675
CDS total length (Mb)	20.44 (50.98% of genome)
CDS average size (bp)	1461.62
Predicted protein-coding genes (Number)	13,982
Predicted protein-coding genes, total length (Mb)	23.19
Predicted protein-coding genes, mean length (kb)	1658.33
Exon total length (Mb)	20.44
Exon total number	43,400
Exon average length (bp)	470.88
Intron total length (Mb)	2.75
Intron total number	29,418
Intron average length (bp)	93.50
GC content (%)	51.22

**Note:** The gene density is 349 genes/ Mb scaffold. On average, the predicted genes contain 3.10 introns that are 471 bp long. This is in the range seen for other fungi, where the average intron densities range from just over 1.0 intron/ kb coding sequence (cds) in *Schizosaccharomyces pombe* to approximately 5.0 introns/ kb cds in *Cryptococcus neoformans*. For all of the predicted proteins, 8,882 had hits in the SwissProt database, 7,929 genes were mapped in the Kyoto Encyclopaedia of Genes and Genomes (KEGG) database, and 5,061 were classified in the Clusters of Orthologous Groups (COG) database, 8,336 were classified in the Gene Ontology (GO) database, 12,785 in the non-redundant database of NCBI (NR), and 10,928 in the Tremble database, 13,982 were hit in all the above databases.

**Table S2 Blastnp of the presumed 27 terpene synthases in *A. ustus* 094102 genome (done in Oct, 2012)**

Presumed Aupts type	AUPTS ID (Au)	AA No.	Intron No.	RTS	Max score	Query coverage	Max identity
Sesqui AUPTS	226	382	8	EEA21257.1  pentalenene synthase, putative [ <i>Penicillium marneffei</i> ATCC 18224]	125	95%	24%
	12847	408	8		116	83%	24%
	13884	363	8		141	93%	29%
	6852	390	8	XP001390370.2  pentalenene synthase [ <i>Aspergillus niger</i> CBS 513.88]	457	91%	62%
Di AUPTS	11092	340	0	ACZ56398.1  trichodiene synthase [ <i>Fusarium incarnatum</i> ]	126	69%	31%
	6061	343	0	BAF45925.1  fusicoccadiene synthase [ <i>Phomopsis amygdali</i> ]	207	97%	37%
	6241	744	7		288	89%	29%
	7240	762	8		258	94%	28%
	8003	725	3		517	100%	39%
	11189	710	5		292	94%	30%
	13192	708	8		425	98%	36%
	13606	755	7		307	85%	31%
	13676	717	9		294	85%	30%
Tri AUPTS	1332	430	2	EIT75610.1  squalene synthetase [ <i>Aspergillus oryzae</i> 3.042]	501	91%	59%
	7332	470	3		816	97%	83%
	1334	642	2	XP001388569.2  squalene-hopene-cyclase [ <i>Aspergillus niger</i> CBS 513.88]	655	95%	51%
	1561	382	1	GAA92416.1  squalene/phytoene synthase [ <i>Aspergillus kawachii</i> IFO 4308]	560	100%	71%
	5911	735	3	EAW24229.1  oxidosqualene:lanosterol cyclase [ <i>Neosartorya fischeri</i> NRRL 181]	1407	100%	89%
	11571	731	2	EAL89589.1  lanosterol synthase, putative [ <i>Aspergillus fumigatus</i> Af293]	1153	96%	76%
Unknown AUPTS	606	387	1	EAA65430.1  hypothetical protein AN0654.2 [ <i>Aspergillus nidulans</i> FGSC A4]	696	98%	86%
	7669	362	2	gb EAA59165.1  hypothetical protein AN8143.2 [ <i>Aspergillus nidulans</i> FGSC A4]	593	100%	83%
	11565	692	7	EAA64518.1  hypothetical protein AN2407.2 [ <i>Aspergillus nidulans</i> FGSC A4]	759	99%	53%
	13624	689	7	EAU32979.1  conserved hypothetical protein [ <i>Aspergillus terreus</i> NIH2624]	1036	100%	72%
	329	336	2	CAK48316.1  unnamed protein product [ <i>Aspergillus niger</i> ]	529	99%	74%
	3446	456	1	EAA63201.1  hypothetical protein AN2767.2 [ <i>Aspergillus nidulans</i> FGSC A4]	832	98%	90%
	6064	404	3	CCD54927.1  BcSTC5, similar to sesquiterpene cyclase [ <i>Botryotinia fuckeliana</i> ]	38.1	18%	31%
	6298	345	1	EAA59634.1  hypothetical protein AN8012.2 [ <i>Aspergillus nidulans</i> FGSC A4]	643	99%	88%

Note: AUPTS: *Aspergillus ustus* presumed terpene synthases; Prefix “Au” for all the ID of the AUPTS

**Table S3 Primers used for gene deletion and complementation of *Au3446***

Type	Full name	Sequence: 5'—3' (restriction endonuclease)	Product size (bp)
pUC18	M13-47	CGCCAGGGTTTCCCAGTCACGAC	156
	RV-M	GAGCGGATAACAATTACACAGG	
Selective marker	<i>Sh ble</i> -s	CCCACACACCATAGCTTCAAAT	1172
	<i>Sh ble</i> -a	AGCTTGCAAATTAAAGCCTTCG	
	<i>Hpt</i> II-s	AAATTGACGCTTAGACAACTTAA	2146
	<i>Hpt</i> II -a	GCAGCTTGCCAACATGGTG	
<i>Au3446</i> deletion	<i>Au3446L</i> '-s	CGGAATTCATCTTGACGGATT (EcoRI)	1224
	<i>Au3446L</i> '-a	GGGGTACCTTGCCTGTTGG (KpnI)	
	<i>Au3446R</i> '-s	AACTGCAGCTAATCGAGATGTGC (PstI)	1657
	<i>Au3446R</i> '-a	CCCAAGCTCCTTCAAATGG (HindIII)	
	<i>Au3446</i> -s	AAAGATGTTCTCCCTTGGCG	1167
<i>Au3446</i> complementation	<i>Au3446</i> -a	GAGGTCGCGGGCCTGATA	
	<i>Au3446L</i> '-	GGGGTACCATACACTCTCATCCTAC (KpnI)	2502
	<i>Au3446</i> -s		
	<i>Au3446L</i> '-	CTGCAGAACCAATGCATTGCCGGTTCATGCC (BstXI)	
	<i>Au_3446</i> -a		
	<i>Au_3446R</i> '-s	CTGCAGAAAAAAAAGCTCGCTTTGCAC (PstI)	1012
	<i>Au_3446R</i> '-a	AAGCTTGAACCTTTCTAGCCCCGCC (HindIII)	

**Table S4** Plasmids used for gene deletion and complementation of *Au3446*

Type	Serial number / Full name	Characteristics
Expression vectors	pGAPZαA	$P_{GAP}$ , α-factor signal, myc epitope tag, C-terminal polyhistidine tag, $T_{Aox1}$ , $P_{Tefl}$ , $P_{Em7}$ , $Sh ble$ (Zeocin <sup>r</sup> ), $T_{Cycl}$ , pUC ori (ColE1 ori)
	pMD18-T	Derived from pUC18, pUC ori (ColE1 ori), $O_{LacZ}$ , $bla$ (Amp <sup>r</sup> )
	pMD18-T-Simple	Derived from pMD18-T, without multiple cloning site
Cloning vectors	pCAMBIA1301	$P_{CAMV35S}$ , $Gus$ , $T_{Nos polyA}$ , $P_{CAMV35S}$ , $HptII$ (HmB <sup>r</sup> ), $T_{CAMV35S polyA}$ , pVS1 rep, pVS1 sta, pBR322 ori, pBR322 born, Kan <sup>r</sup>
	pTFCM	Derived from pCAMBIA1301, $P_{CAMV35S}$ and $HptII$ (HmB <sup>r</sup> ) were substituted with $P_{TrpC}$ , $HptII$ (HmB <sup>r</sup> ), and $T_{TrpC}$
	JS	$P_{TrpC}$ , $eGFP$ , $T_{TrpC}$ , $HptII$ (HmB <sup>r</sup> ), pVS1 rep, pVS1 sta, Kan <sup>r</sup>
Random insertion vectors	pWHU2201 / pUC18 / $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{Cycl}$	Derived from pMD18-T and pGAPZαA, pUC ori (ColE1 ori), $O_{LacZ}$ , $bla$ (Amp <sup>r</sup> ), $P_{Tefl}$ , $P_{Em7}$ , $Sh ble$ (Zeocin <sup>r</sup> ), $T_{Cycl}$
	pWHU2202 / pUC18 / $P_{CAMV35S}$ - $HptII$ - $T_{CAMV35S polyA}$	Derived from pMD18-T and pCAMBIA1301, pUC ori (ColE1 ori), $O_{LacZ}$ , $bla$ (Amp <sup>r</sup> ), $P_{CAMV35S}$ , $HptII$ (HmB <sup>r</sup> ), $T_{CAMV35S polyA}$
	pUC18 / <i>Au_3446L</i>	Derived from pMD18-T-Simple
Deletion and complementation vectors	pUC18 / <i>Au_3446R</i>	Derived from pMD18-T-Simple
	pUC18 / <i>Au_3446L'</i>	Derived from pMD18-T-Simple
	pUC18 / <i>Au_3446R'</i>	Derived from pMD18-T-Simple
Genetic deletion and complementation system (gene <i>Au_3446</i> )	pUC18 / <i>Au_3446L'-P<sub>Tefl</sub>-P<sub>Em7</sub>-Sh ble-T<sub>Cycl</sub></i>	Derived from pUC18 / $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{Cycl}$
	pWHU2203 / pUC18 / <i>Au_3446L'-P<sub>Tefl</sub>-P<sub>Em7</sub>-Sh ble-T<sub>Cycl</sub>-Au_3446R'</i>	Derived from pUC18 / <i>Au_3446L'-P<sub>Tefl</sub>-P<sub>Em7</sub>-Sh ble-T<sub>Cycl</sub></i>
	pWHU2204 / JS / <i>Au_3446L'-P<sub>Tefl</sub>-P<sub>Em7</sub>-Sh ble-T<sub>Cycl</sub>-Au_3446R'</i>	Derived from JS and pUC18 / <i>Au_3446L'-P<sub>Tefl</sub>-P<sub>Em7</sub>-Sh ble-T<sub>Cycl</sub>-Au_3446R'</i> , $P_{TrpC}$ , $eGFP$ , and $T_{TrpC}$ were substituted with <i>Au_3446L'-P<sub>Tefl</sub>-P<sub>Em7</sub>-Sh ble-T<sub>Cycl</sub>-Au_3446R'</i>
Genetic deletion and complementation system (gene <i>Au_3446</i> )	pUC18 / <i>Au_3446-Au_3446L'-P<sub>CAMV35S</sub>-HptII-T<sub>CAMV35S polyA</sub></i>	Derive from pUC18 / $P_{CAMV35S}$ - $HptII$ - $T_{CAMV35S polyA}$
	pWHU2205 / pUC18 / <i>Au_3446-Au_3446L'-P<sub>CAMV35S</sub>-HptII-T<sub>CAMV35S polyA</sub>-Au_3446R'</i>	Derive from pUC18 / <i>Au_3446-Au_3446L'-P<sub>CAMV35S</sub>-HptII-T<sub>CAMV35S polyA</sub></i> - <i>Au_3446R'</i>

**Table S5** Primers used for gene cluster inactivation

Type	Full name	Sequence: 5'—3' (restriction endonuclease)	Product size (bp)
pUC18	M13-47	CGCCAGGGTTTCCCAGTCACGAC	
	RV-M	GAGCGGATAACAATTACACAGG	156
Selective marker	<i>Sh ble</i> -s	CCCACACACCATACTTCAAAAT	
	<i>Sh ble</i> -a	AGCTTGCAAATTAAAGCCTTCG	1172
	<i>HptII</i> -s	AAATTGACGCTTAGACAACTTAA	
	<i>HptII</i> -a	GCAGCTTGCCAACATGGTG	2146
POC3446 deletion	POC3446L-s	GAATTCCCACAGATAGTATGCGAGTC ( <i>EcoRI</i> )	
	POC3446L-a	TCTAGACAAACGCCAGAACATCAGC ( <i>XbaI</i> )	1419
	POC3446R-s	GTCGACTAGGGTGGTATTGCGT ( <i>Sall</i> )	
	POC3446R-a	AAGCTTGGGCATCCGTATGATGTCG ( <i>HindIII</i> )	1128
POC8003 deletion	POC8003L-s	GAATTCTCGTACAGTCAGGTAGGGA ( <i>EcoRI</i> )	
	POC8003L-a	GGTACCAAGGAGGAGTGCAAGGAA ( <i>KpnI</i> )	1085
	POC8003R-s	CCTGCAGGTTGCCGAGGACAAGG ( <i>SbfI</i> )	
	POC8003R-a	AAGCTTCATCTGTGGACGGAGTAAAG ( <i>HindIII</i> )	1061
	<i>Au_8003</i> -s	CGCCCACCACTCATCTTTAG	
POC13192 deletion	<i>Au_8003</i> -a	CGACCTGATCCGATGCTGTC	1063
	POC13192L-s	GAATTCTAACTCAAATCAGCGAGGA ( <i>EcoRI</i> )	
	POC13192L-a	TCTAGAATACCCACGGACCCAAC ( <i>XbaI</i> )	1224
	POC13192R-s	CCTGCAGGAGCAGCATCTCGATTGG ( <i>SbfI</i> )	
	POC13192R-a	AAGCTTATTCTGGACAGGAACTAG ( <i>HindIII</i> )	1077
POC11565 deletion	<i>Au_13192</i> -s	AAGAACAAAGCACGGACTGCC	
	<i>Au_13192</i> -a	CCTGCCCTCGTCCAATACAGAT	1096
	POC11565L-s	GAATTCTTGTGAGGCCTTGTG ( <i>EcoRI</i> )	
	POC11565L-a	TCTAGAGTCATCGTGCAGTAGG ( <i>XbaI</i> )	1058
	POC11565R-s	CCTGCAGGCATCCGCAGTCATAGTAGT ( <i>SbfI</i> )	
POC6298 deletion	POC11565R-a	AAGCTTCGCCTCCACGTTG ( <i>HindIII</i> )	1018
	<i>Au_11565</i> -s	CTCGTATTGCGCTTGGTTGA	
	<i>Au_11565</i> -a	CCAGCATTGTCGGGTTCG	1106
	POC6298L-s	GAATTCGCAGCCAGTCCCTCCCTA ( <i>EcoRI</i> )	
	POC6298L-a	GGATCCTTTTACCCAAGCCCATC ( <i>BamHI</i> )	1508
POC6298 deletion	POC6298R-s	CCTGCAGGAGGGAAATACCGCCTCAAAG ( <i>SbfI</i> )	
	POC6298R-a	AAGCTTAGCCTCCTCGTTC ( <i>HindIII</i> )	1208
	<i>Au_6298</i> -s	TTTGAGGCGGTATTCCC	
	<i>Au_6298</i> -a	ACCTCCTTCTCAGTCCCT	1038

**Table S6 Plasmids used for gene cluster inactivation**

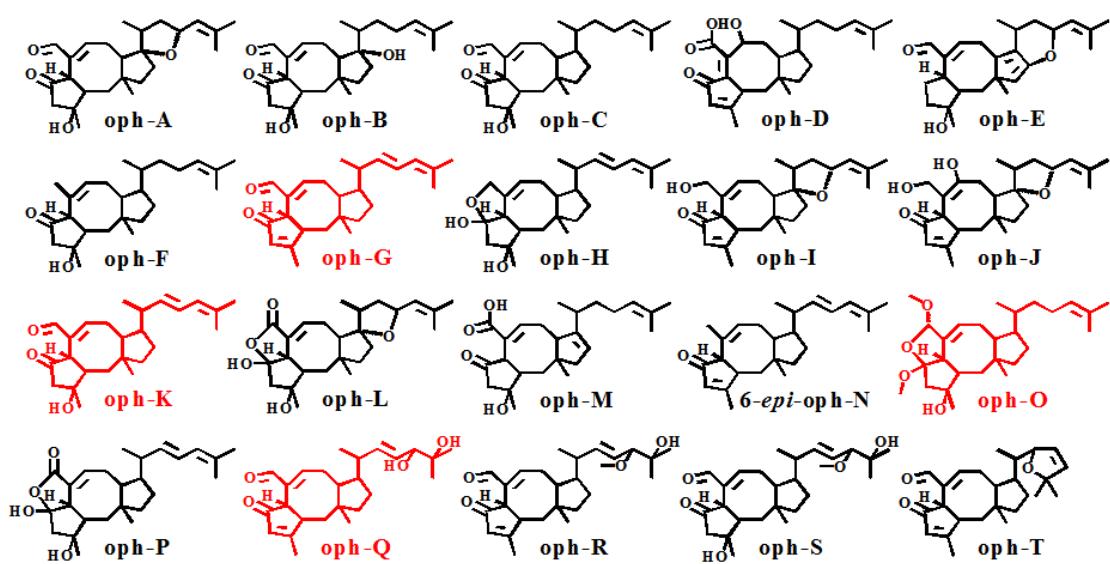
Type	Serial number / Full name	Characteristics
Expression vectors	pGAPZαA	$P_{GAP}$ , α-factor signal, myc epitope tag, C-terminal polyhistidine tag, $T_{AoxI}$ , $P_{Tefl}$ , $P_{Em7}$ , $Sh ble$ (Zeocin <sup>r</sup> ), $T_{CycI}$ , pUC ori (ColE1 ori)
	pMD18-T	Derived from pUC18, pUC ori (ColE1 ori), $O_{LacZ}$ , $bla$ (Amp <sup>r</sup> )
Cloning vectors	pMD18-T-Simple	Derived from pMD18-T, without multiple cloning site
	pCAMBIA1301 (CAMBIA, Canberra, Australia)	$P_{CAMV35S}$ , $Gus$ , $T_{Nos polyA}$ , $P_{CAMV35S}$ , $HptII$ (HmB <sup>r</sup> ), $T_{CAMV35S}$ polyA, pVS1 rep, pVS1 sta, pBR322 ori, pBR322 born, Kan <sup>r</sup>
	pTFCM	Derived from pCAMBIA1301, $P_{CAMV35S}$ and $HptII$ (HmB <sup>r</sup> ) were substituted with $P_{TrpC}$ , $HptII$ (HmB <sup>r</sup> ), and $T_{TrpC}$
POC3446 deletion	pUC18 / POC3446L	Derived from pMD18-T-Simple
	pUC18 / POC3446R	Derived from pMD18-T-Simple
	pUC18 / POC3446L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$	Derived from pUC18 / $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
	pWHU2206 / pUC18 / POC3446L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC3446R	Derived from pUC18 / POC3446L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
	pWHU2207 / JS / POC3446L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC3446R	Derived from JS and pUC18 / POC3446L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC3446R, $P_{TrpC}$ , eGFP, and $T_{TrpC}$ were substituted with POC3446L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC3446R
	pUC18 / POC8003L	Derived from pMD18-T-Simple
	pUC18 / POC8003R	Derived from pMD18-T-Simple
	pUC18 / POC8003L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$	Derived from pUC18 / $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
POC13192 deletion	pWHU2212 / pUC18 / POC8003L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC8003R	Derived from pUC18 / POC8003L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
	pUC18 / POC13192L	Derived from pMD18-T-Simple
	pUC18 / POC13192R	Derived from pMD18-T-Simple
	pUC18 / POC13192L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$	Derived from pUC18 / $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
POC11565 deletion	pWHU2213 / pUC18 / POC13192L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC13192R	Derived from pUC18 / POC13192L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
	pUC18 / POC11565L	Derived from pMD18-T-Simple
	pUC18 / POC11565R	Derived from pMD18-T-Simple
	pUC18 / POC11565L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$	Derived from pUC18 / $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
	pWHU2214 / pUC18 / POC11565L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC11565R	Derived from pUC18 / POC11565L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
POC6298 deletion	pUC18 / POC6298L	Derive from pMD18-T-Simple
	pUC18 / POC6298R	Derive from pMD18-T-Simple
	pUC18 / POC6298L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$	Derived from pUC18 / $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$
	pWHU2223 / pUC18 / POC6298L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$ -POC6298R	Derived from pUC18 / POC6298L- $P_{Tefl}$ - $P_{Em7}$ - $Sh ble$ - $T_{CycI}$

**Table S7 BlastnP result in GenBank database of 5 TS/TC proteins with reference sequences. Underlined sequences were added in 2015; shaded were unidentified sequences**

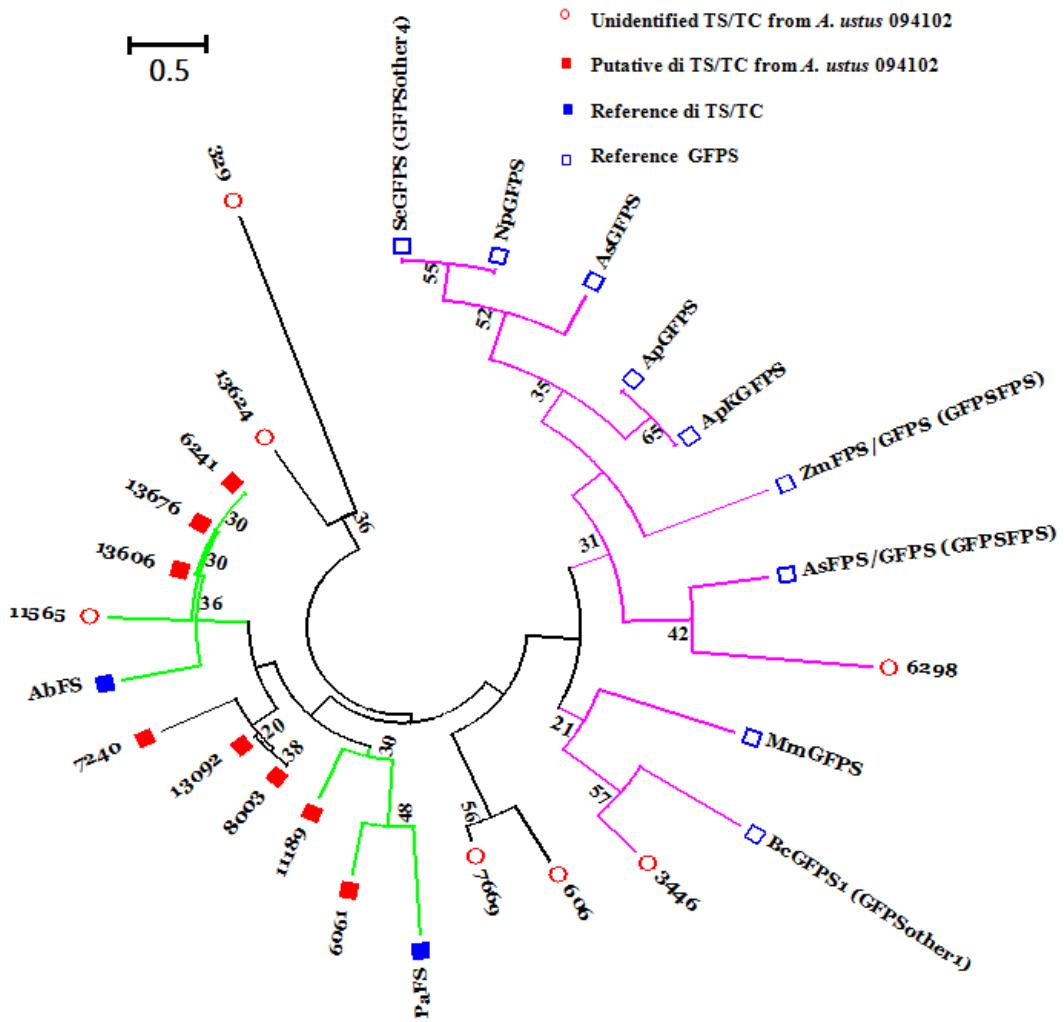
	sequence	sources	description		
			Max score	Query cover	Ident
Au8003	AcOS	<i>Aspergillus clavatus</i>	979	100%	65%
	<u>EvVS</u>	<u><i>Emericella variecolor</i></u>	<u>273</u>	<u>85%</u>	<u>45%</u>
	<u>EvSS</u>	<u><i>Emericella variecolor</i></u>	<u>552</u>	<u>100%</u>	<u>41%</u>
	PaPS	<i>Phomopsis amygdali</i>	516	100%	40%
	PaFS	<i>Phomopsis amygdali</i>	517	100%	39%
	AbFS	<i>Alternaria brassicicola</i>	466	98%	36%
	<u>NfSS</u>	<u><i>Neosartorya fischeri</i></u>	<u>290</u>	<u>96%</u>	<u>30%</u>
	<u>EvVS</u>	<u><i>Emericella variecolor</i></u>	<u>1115</u>	<u>99%</u>	<u>78%</u>
Au13192	<u>NfSS</u>	<u><i>Neosartorya fischeri</i></u>	<u>218</u>	<u>83%</u>	<u>46%</u>
	PaFS	<i>Phomopsis amygdali</i>	425	98%	36%
	PaPS	<i>Phomopsis amygdali</i>	394	97%	35%
	<u>EvSS</u>	<u><i>Emericella variecolor</i></u>	<u>412</u>	<u>98%</u>	<u>34%</u>
	AbFS	<i>Alternaria brassicicola</i>	359	96%	34%
Au11565	AcOS	<i>Aspergillus clavatus</i>	402	98%	33%
	<u>NfSS</u>	<u><i>Neosartorya fischeri</i></u>	<u>191</u>	<u>87%</u>	<u>40%</u>
	AcOS	<i>Aspergillus clavatus</i>	263	96%	32%
	<u>EvSS</u>	<u><i>Emericella variecolor</i></u>	<u>274</u>	<u>96%</u>	<u>30%</u>
	<u>EvVS</u>	<u><i>Emericella variecolor</i></u>	<u>237</u>	<u>75%</u>	<u>29%</u>
	PaFS	<i>Phomopsis amygdali</i>	209	73%	29%
	AbFS	<i>Alternaria brassicicola</i>	206	71%	28%
	PaPS	<i>Phomopsis amygdali</i>	259	98%	28%
	WcFPPS	<i>Wolfiporia cocos</i>	397	98%	58%
	CpFPPS	<i>Chimonanthus praecox</i>	326	100%	50%
Au6298	EpFPPS	<i>Euphorbia pekinensis</i>	318	100%	49%
	OsaFPPS	<i>Ornithogalum longebracteatum</i>	315	98%	48%
	RpFPPS1	<i>Ornithogalum longebracteatum</i>	310	97%	48%
	RpFPPS2	<i>Ornithogalum longebracteatum</i>	315	97%	47%
	RsFPPS	<i>Rhizosolenia setigera</i>	319	99%	46%
Au3446	Nf	<i>Neosartorya fischeri</i>	786	100%	86%
	Ao	<i>Aspergillus oryzae</i>	776	100%	85%
	An	<i>Aspergillus niger</i>	745	100%	79%
	Ak	<i>Aspergillus kawachii</i>	742	100%	79%
	Ml	<i>Micrococcus luteus</i>	125	60%	32%
	Ss	<i>Sulfolobus solfataricus</i>	49.3%	34%	26%

**Table S8 Primers used for gene expression**

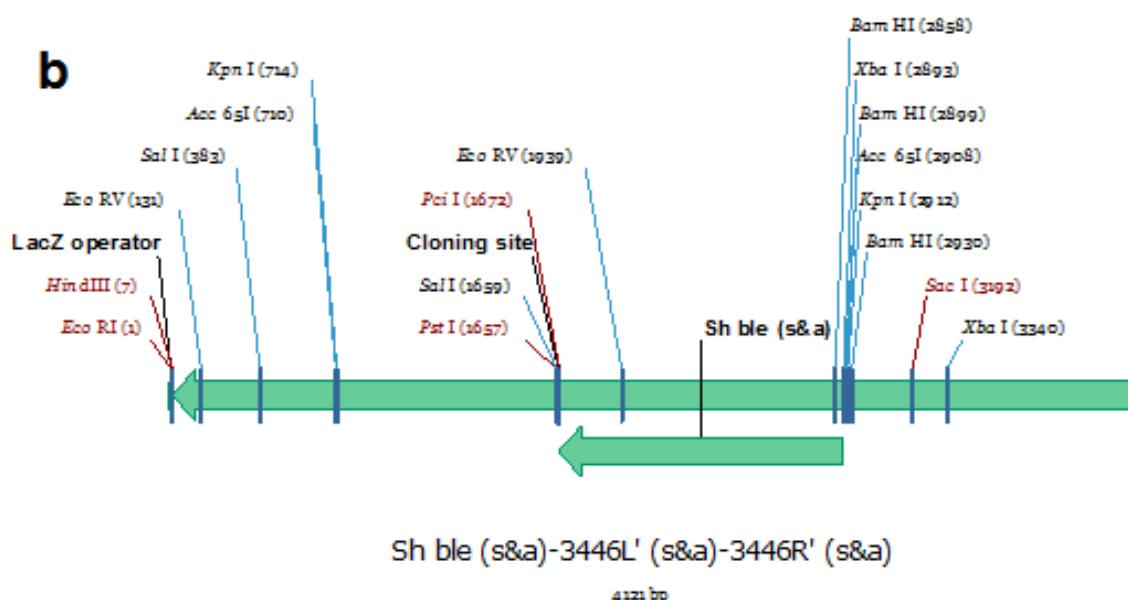
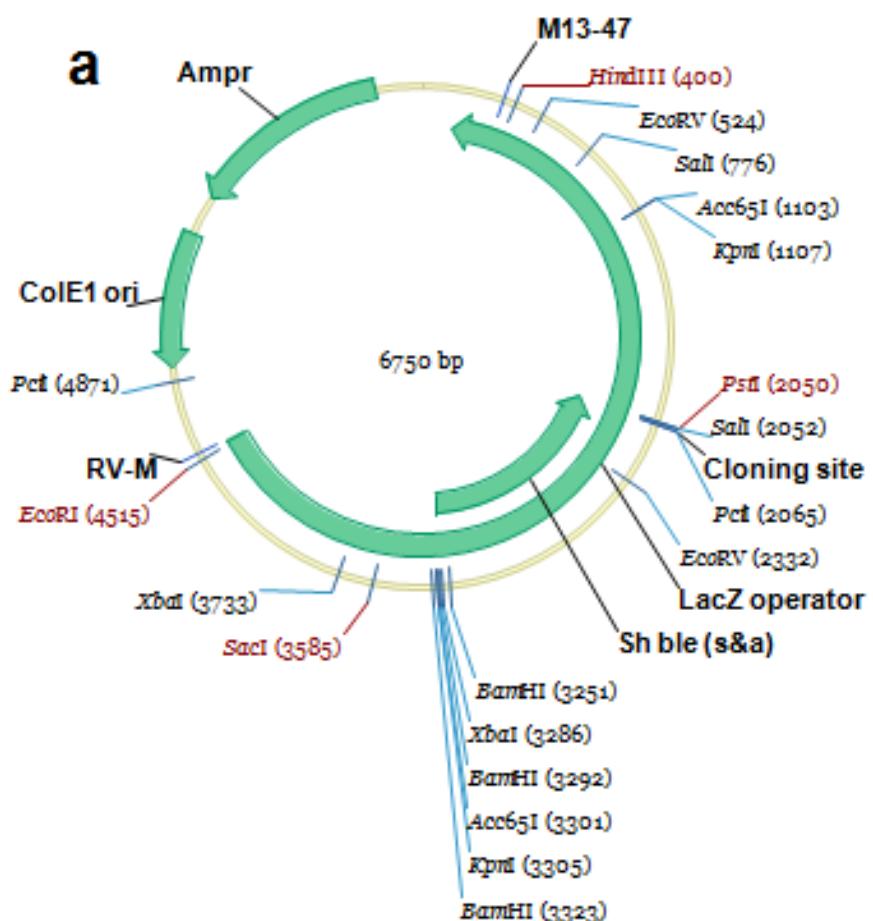
Type	Full name	Sequence: 5'—3' (restriction endonuclease)	Product size (bp)
Au8003 expression	Au8003-F	CATATGATGGAGTATAAGTACTCGACC( <i>Nde</i> I)	2178
	Au8003-R	AAGCTT TCAAACCTTCAGCAGCTCCA( <i>Hind</i> III)	
Au6298 expression	Au6298-F	CCCATATGATGTCTTCACCGCGTGCC ( <i>Nde</i> I)	1038
	Au6298-R	CCAAGCTTTATTGGTGCCTGTAAA ( <i>Hind</i> III)	
Au11565 expression	Au11565	GGGGTACCGCGATGTACACCCCTCGA ( <i>Kpn</i> I)	2405
	Au11565	GCTCTAGATACTTCAGCAAAAGCA ( <i>Xba</i> I)	
Au13192 exon fusion and expression	Au13192E1-s	AAGCTTATGCCACAAA CACAATCC( <i>Hind</i> III)	326
	Au13192E1-a	GCCGACTCGACAACATTATCATAACAGAAAC	
		GCATACTCAAAGATATAACAAATAA	
	Au13192E3-s	TGATAATGTTGTCGAGTCGGCGGCCAATTC	301
		TACGTTGAACATGGACACGGACAA	
	Au13192E3-a	CATGAGCATGTCGACGAAGGGGGCGCCGGTATCGAT	
	Au13192E4-s	CTTCGTCGACATGCTCATG	766
	Au13192E4-a	GCCGGGGCGAGGGAGGATCTCGTCTCCTAGCTGCAC	
	Au13192E5-s	ATCCTCCTCGCCCCGGC	175
	Au13192E5-a	AGTCTTCGATGTCGTCGAGCATGAGCGAGGCATTGTGTA	
	Au13192E6-s	CTCGACGACATCGAAGACT	309
	Au13192E6-a	CTGCACCATCACCTCGTAAGCAACCGGAAC	
		AACCCCACCGGTCTTCTGCCTAACCATCTCCAAAT	
	Au13192E8-s	TTACGAGGTTGATGGTGCAGATTGCGCCGGT	126
		GC GGCGGAAAGATCTCGACGGCATTTATCAT	
	Au13192E8-a	CCCTTTGGCCGGTGTACTCTTCAGTTAGGTTCTGTAGT	
	Au13192E9-s	AGTACACCGGCCAAAGGG	333
	Au13192E9-a	GC GGCGCTACACCTCAACCTCTGCAC( <i>Not</i> I)	

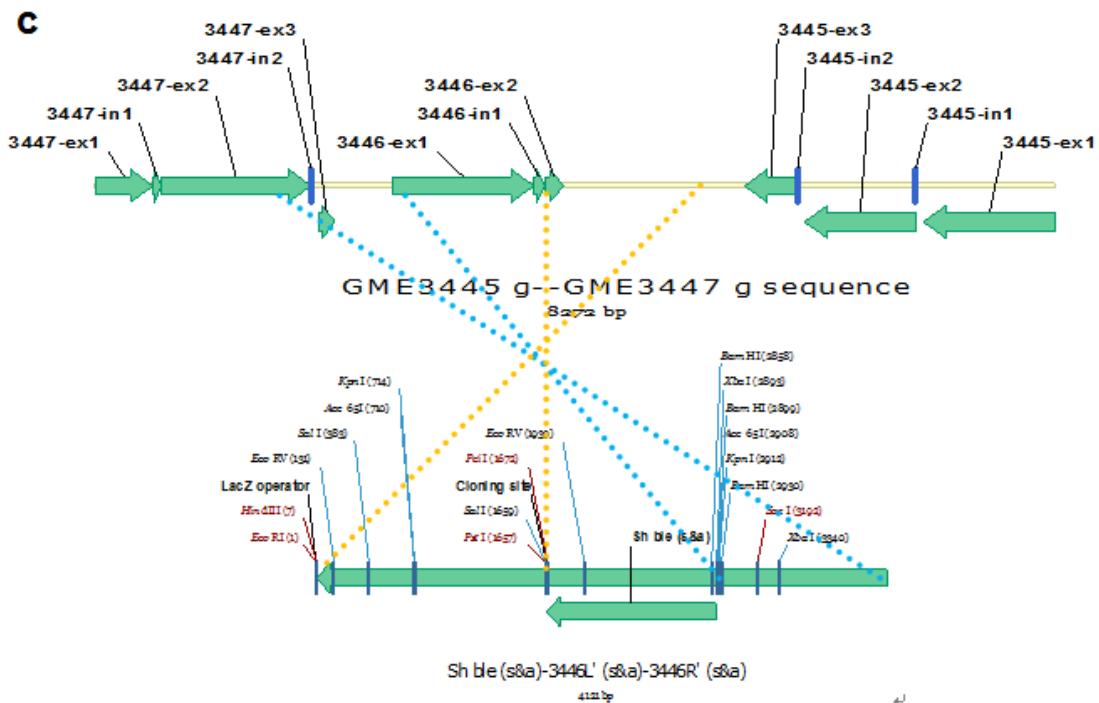


**Figure S1 Twenty reported subgroups (1957–2013) of ophiobolins with 5-8-5 ring scaffolds.**  
These structures were from references 12,14,15 and 16; red color shows ophiobolins identified from *Aspergillus ustus* 094102

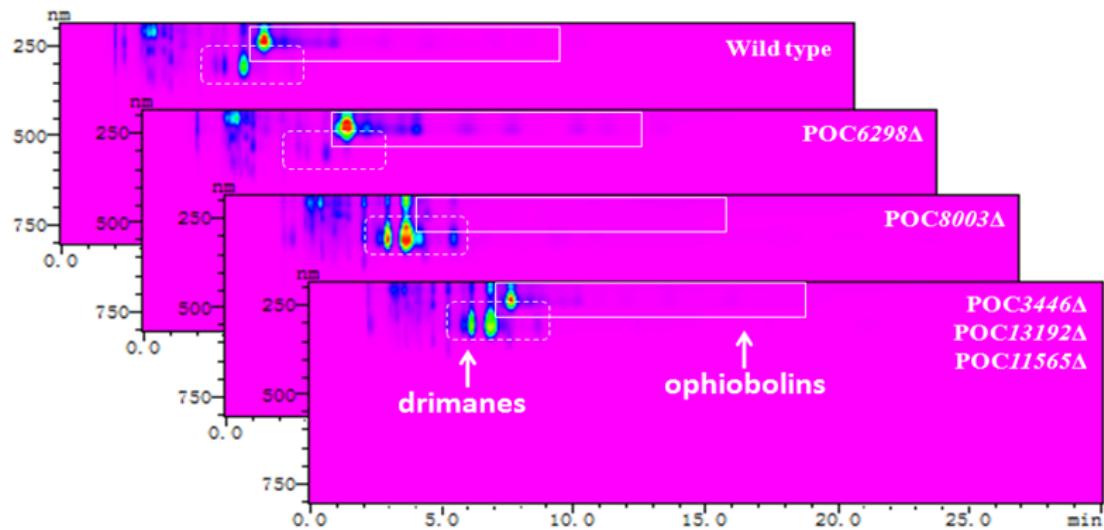


**Figure S2 Neighbour-joining phylogenetic tree based on amino acid sequences of chain length determination motif (Hisashi et al<sup>25</sup>) of the 15 trans-IPP terpene synthetase proteins.** The tree was generated based on Kimura 2-parameter matrix in MEGA software (version 5.0). Numbers at nodes indicate bootstrap values with 1000 iterations. Bar, 0.5, substitutions per nucleotide position. PaPS (*Phomopsis amygdali* phomopsene synthase); PaFS (*Phomopsis amygdali* fusicoccadiene synthase); NpGFPPS (GFPPS from *Natronobacterium pharaonis*); ApGFPPS (GFPPS from *Aeropyrum pernix*); MmGFPPS (GFPPS from *Methanoscarcina mazei*)





**Figure S3 Gene deletion of Au3446.** (a) Plasmid pMD18-T-3446L'-Sh ble- 3446R' constructed with the selective marker and the left and right homologous sequences for recombination; (b) The detailed targeting fragment; (c) Homologous recombinant of the cassette with the genomic DNA to delete the target gene.

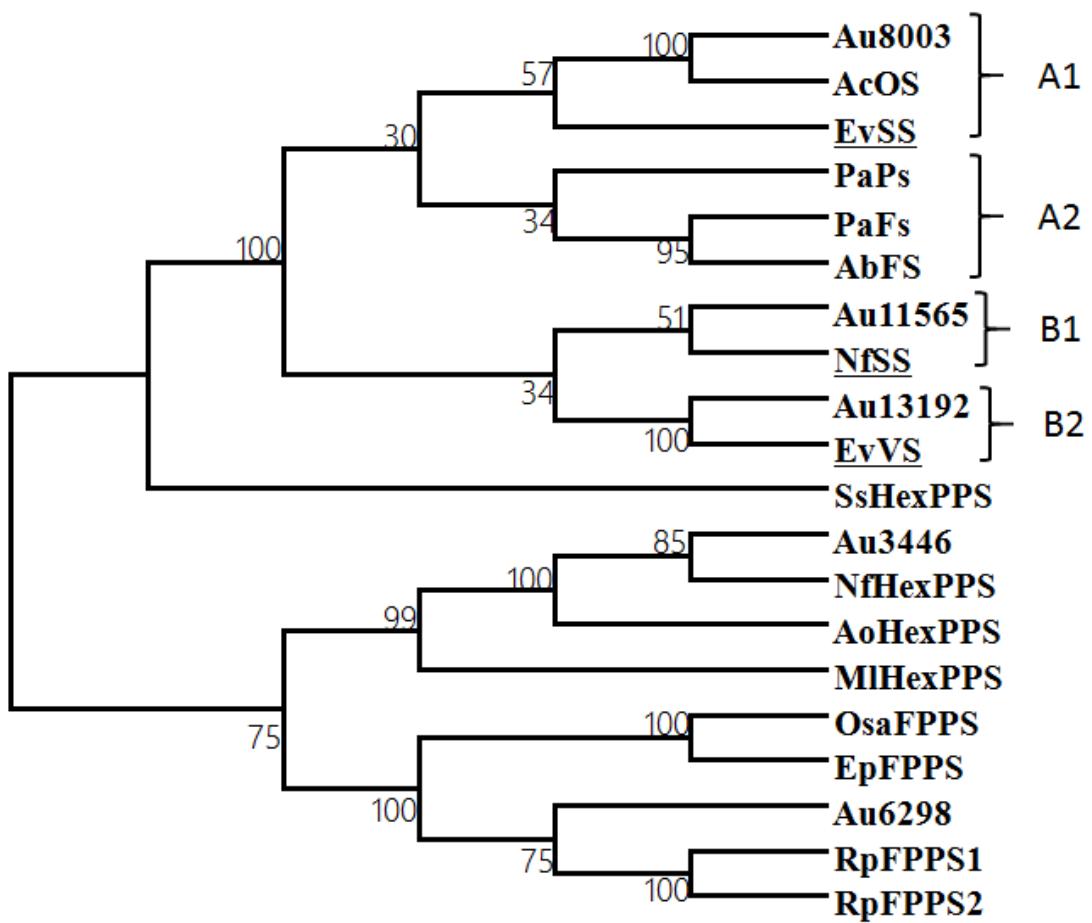


**Figure S4** Full wavelength range from 210–400 nm analysis collected for ophiobolin and drimane on the fermentation products of wild type strain and the five gene cluster inactivated mutants. Ophiobolin did not appear for  $\Delta$ POC8003, increased for  $\Delta$ POC6298, partially decreased for  $\Delta$ POC13192,  $\Delta$ POC3446 and  $\Delta$ POC11565; drimane was increased for  $\Delta$ POC8003,  $\Delta$ POC13192,  $\Delta$ POC3446 and  $\Delta$ POC11565, but decreased for  $\Delta$ POC6298.

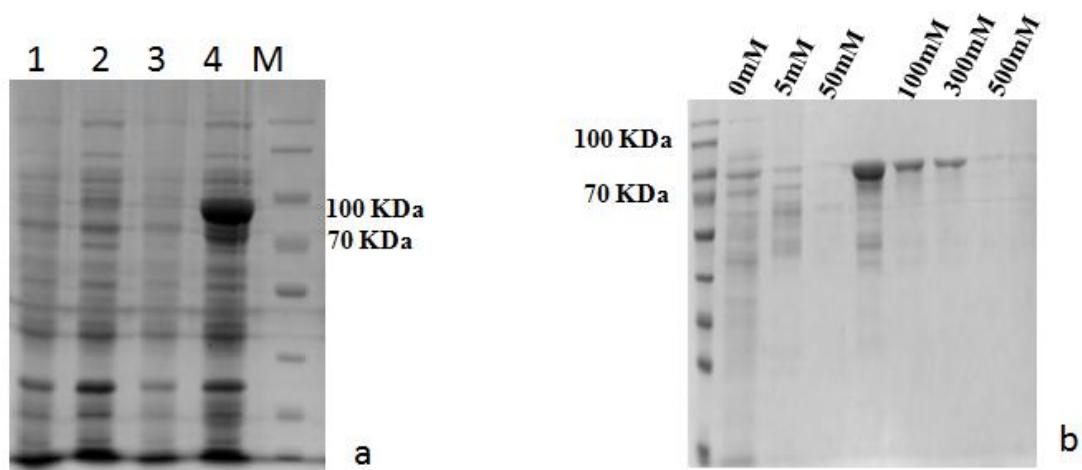
Au8003 1 - - - - - MEYKYSITIVDSSKKWDPEGLIEGIPLRKHAEQDLEEVGSFRVQEDWRRLVGPVEN - PFRGSL  
 AcOS 1 - - - - - MACKYSTLIDSSLYDREGLCPGIDLRRHVAGELEEVGAFRAQEDWRRLVGPLPK - PYAGLL  
 EvSS 1 - - - - - MEYKYSTVVDPGTYETHGLCEGYEVRYHKNAELEDIGCLRCEHWRSQVGPL - AFKGTL  
 PaPS 1 - - - - - MEFKYSEVVEPSTYYTEGLCEGIDVRKSFKFTLEDRGAIRAHEDWNKHIGPCG - EYRGTL  
 PaFS 1 - - - - - MKYQFSIIVDPATYDNEGLNSGIDLRLKNNFTLEDRGAIRAHEDWNKHIGPCG - QFKGTL  
 Au13192 1 MPQTQSESYLNPSLSTPVRSTPDIAGFCAGYELRHHHEHLANEGLSLRCRTDWEQFIGPIER - WGSCN  
 EvS 1 MSQS-SDFILNSTLSSVERSTPDIAGFCAGYELRHHHEHLANEGLSLRCRTDWEQFIGPIER - WGSCN  
 Au11565 1 - - - - - MYTLDDEPFKYSLPNPEAERPAKFFNSLPYRVTRIEDDCGSTIDRVVAEWKAITGKVP - DAVG  
 NSS 1 - - - - - MEVWEHSRPIADDTIKKTPSFTTLPRINKQNDVADAATRALRDWDYLYHDGLAERALISI  
 Au6298 1 -  
 RpFPSS1 1 -  
 RpFPSS2 1 -  
 EpFPSS 1 -  
 OsaFPSS 1 -  
 Au3446 1 -  
 SsHexPPS 1 -  
 MiHexPPS 1 -  
  
 DD/NNX/D/E motif  
 Au8003 61 GPEISFIYTVPPECLPERILEATSYGLDYGFHDQEITTKIEE - - AEELDDVGAALAAQGGSTG - - KIQE  
 AcOS 61 GPDFSFIFTGAVPECHPDRMEIYVAYALEFGFMHDVDIDTDVNH - - ASLDEVGHTLDQSRG - RTG - - KIED  
 EvSS 60 GNPFNLLSLVIPPECLPDRLSIVGFANELAFIHDDVTDIVQYG - - DAHNNDKEAFNSMATTG - - SM  
 PaPS 62 GPKRFSFTAVTLPCECRPDRMELVSYIMEFAFLHDVDIDTDVNH - - ASLDEVGHTLDQSRG - RTG - - SA  
 PaFS 60 GPRFSFISVAVPECIPERLEIISYANEFAFLHDVDVTDVHGHDGTGEVENDEMVTPLLEAHNTGAIDTSNKV  
 AbFS 60 GHDFSMFTVCVPCPECIPERLEIISYANEFAFLHDVDIDTDVNH - - DAHNNDKEAFNSMATTG - - WSPPQ  
 Au13192 69 PWEHGFAGAVVLPLCPKPERLAVICYIFEYAFYLVDNVVESAANSTLNMDTDNIALDETEYRTVR - -  
 EvS 68 PWEHGFAGAVVLPLCPKPERLAVICYIFEYAFYLVDNVVESAANSTLNMDTDNIALDETEYRTVR - -  
 Au11565 65 GHASDLIRYVLPCEPVEFVAPVTRLIMYILLWIDDATDPLVSHENLMLDFRVRGVVSQMKG - - YCQ  
 NSS 63 SELGNLGAFAYPEVPPERLAITYLTLDGLIHDGYEAMMDMQARATEHREFGALDFPHEQLP - - SRR  
 Au6298 1 -  
 RpFPSS1 1 -  
 RpFPSS2 1 -  
 EpFPSS 1 -  
 OsaFPSS 1 -  
 Au3446 1 -  
 SsHexPPS 1 -  
 MiHexPPS 1 -  
  
 Au8003 124 GTKSSGKRKMAAQILREMMALDPERAMTLAKSWAQ - GVQHSARRVEEKDWKSLDEYIPFRCMDLGYMHWH  
 AcOS 122 - KGSDGDKRMVPTQJIREMMAIDPERAMTVKSWAS - GVRHSSRKEDTNFKALEQYIPYRALDVGYMLWH  
 EvSS 122 ENAASKGRALQAYIAREMVRIDKERAIAKAWAK - FVDYGG - RQETTRKEYTEYRQDIGHWFY  
 PaPS 126 KGKKSGEGLILKEVTLIDKVEVTPDVRRAELMKFWK - LDLDVSRDRKHF - FRDFDDDMYERIVDCASYFLI  
 PaFS 130 DIRRAGKCKRILQSKQFLPEMLAIDPCEAKTTMKSWAR - FVEVGSRRQHETRVELAKYIIPYRIMDVGEMFWF  
 AbFS 126 DQSSSGKTRILQSKQFLSEMMECIAKATMKAWAE - FLRVGGSSRQHGTVFTRLKDYLIPYRICKDVGEMFWF  
 Au13192 130 - SILDGKTQIQSkmLLELLAIDARCAEVVINSWKE-MISTTAKKDKTTRAFFNNLEEYVDYRILDGAPFVD  
 EvS 129 - SILDGKTQIQSkmLLELLSIDAPRAEVVINSWKE-MISTTAKKDKTTRAFFNNLEEYVDYRILDGAPFVD  
 Au11565 130 CELEMNRRIYIQSVLALVVQQAGRNLDQMGDFKRGLEVFDFSTARQGAPPQTITWEEMYKRHRTTSIGGRЛИ  
 NSS 126 GTRAALKLKLVSQILLEAIRIDRDMGMYMFDMYNKGWLVSAGGEGKVPQFKSVEEYQAYRRDDFGIRAFW  
 Au6298 1 -  
 RpFPSS1 1 -  
 RpFPSS2 1 -  
 EpFPSS 1 -  
 OsaFPSS 1 -  
 Au3446 1 - MKARTVPAARRLWASPIKPQTTLICWQCLRNDLFLERIQCQTRKYHPTRRKDVSPPGAASAAQTIFK  
 SsHexPPS 1 -  
 MiHexPPS 1 -  
  
 Au8003 193 GLVTFGCATITPEEEEEERRTLLEPAVIACLMTNDLFSYEKEKNDNNPQ - - - - NAAVIMKIHKCSE  
 AcOS 191 GLVTFGCATITPNEEEEAKRLIIPALVQASLNDLFSYEKEKNDNNPQ - - - - NAVLIVMNEHGCSE  
 EvSS 190 GLLSFAMALDVPEHEREMCHEVCRTAYVQIMVHDLASWEKEKLNAALG-KDVTI - - IVFLMEEHGI  
 PaPS 192 ALSTFAMALTIPAEODKEVFTLTRPVMAAAAATNDVPSWEKEDKLKFQKDNTADMTNGWWMLMKQYSIGV  
 PaFS 199 GLVTFLGLHLIPDHEELLCRELMANAWIAVGLQNDLWSWPKFRDAATLHG-KDHVVNAIWVLMQEHQTDV  
 AbFS 195 GVVTFGMALHIDPHEMDACHKLMPEAVIAVGLANDVFSWPKFRDASQRLG-RTHVVNAWVLMQEHEGFSQ  
 Au13192 198 MLMRFGMGIILTPEEQARTPLVKPCYAAGLANDYFSFIDWEEFQREGAKTTMTNAWVLFMQWEGLSV  
 EvS 197 MLMRFGMGIILTPEEQOKRIEPIVKPCYAAGLANDYFSFIDWEEFQAESDKTTMTNAWVLFMQWEGLSV  
 Au11565 197 ALIPIPSMRKLRLDQDALDSVSHLAEICLPLVFSWPKFRDASQRLG - - - - FEEHARAGSLDGIHNGMAVLMSRYGYEE  
 NSS 198 PMVEFGMAMRLSDEDKKLIELPVMEPIDKAIWTNDYWSFDRHYESHTING - - - - SRLTNVMEVVRQIENKSI  
 Au6298 1 -  
 RpFPSS1 1 - MNKML-TFTRLSRRSAYFLS-DAAAVRENLF - - PMSTVRAAPPVP - - PA  
 RpFPSS2 1 - - - - MNKMLSTFTRTLNRPTAFLFLRRTATANRDWHRFSMTMSVVHSPPV - - - - PV  
 EpFPSS 1 -  
 OsaFPSS 1 -  
 Au3446 69 GLPKAPPGISVDPPLRIVGKELKFLSKNIRQLLGSGHPLLDKVAKYYTRSE - - - - GKHMRPLVLLMSQ  
 SsHexPPS 1 -  
 MiHexPPS 1 -  
  
 Au8003 256 EEARDICKQIRLECRKYARIVKETTLARTDISDLKRYIEIMQYTVSGNWAWSTQCPRYHADAKFNEQ  
 AcOS 254 EEAIDLKKRIRLECRKYARIVKETTLARTDISDLKRYIEIMQYTVSGNWAWSTQCPRYHADAKFNEQ  
 EvSS 259 EEAARKERCERAKTAAADYLKIVEYKARD - DISLSDSRKVIEESWLYTISGNTVWSTFCPRYHNGPTKFNELQ  
 PaPS 261 EEAARKRILGKAREHAEVFKLTSQIHNRLLDLSLSDSRKVIEAMQYTMGNSLWMGISTPRYHSDQSLDEMM  
 PaFS 268 DGAAMQICRKLIVEYVAKYLEVIEATKNDE-SISPLDLRKYLDAMLYSISGNVWVWSLECPRYHNPDVSFNKTQ  
 AbFS 264 EQARQYCRLAAQFVAQYDNIRNIKNEE-SISPDLRTVYEAQMYSIGNVIWSKFCPRYHPEKRFNQTQ  
 Au13192 268 EDAAKRRVREVTNMYERQYLKNVMEFCASEGKDEPKLQYERQAQGHQVPGNVVAWSLRCPRYHPWLCREAGE  
 EvS 267 EEAQAKRVEVTKQMEQQYLRNIAFDAAGEGENIKLDTKLQYKAGQVPGNVVAWSLRCPRYHPWLCKEAS  
 Au11565 270 SEAAEEIMKKEILSAERTLMEKYEIWKSSSAPKSDELRHFLVLIISLAVGAGSYWQSFSPRYQRANLSTTVE  
 NSS 266 DEAKAAVQRQLLNVNLQQYLERKRAIYAQNPSIPSHLRKWLVEVGITVAGTHFWASCSPRTHAWRNNSRNG  
 Au6298 1 - MSS - - PRAAFEAVPFLPSQADILDHAKKYNLPQ - - - - NALEWLEKNLNTN - -  
 RpFPSS1 45 ITGTAWSKDETDRDFMAVFDPDVRLTDTGRNLNDIP - - - - DVTWKWLSKVLOY - -  
 RpFPSS2 49 LIGTSISKDELGDMAVPFDIMKOLTDNGLNDLDP - - - - DVNKWLKLEIILHYN - -  
 EpFPSS 1 - MAD - - - - LKSTELDLYSQLKTELLNDT-AFEWSP - - - - DSROWILERMLDYN - -  
 OsaFPSS 1 - - - - MADSNGLDTKSRFQIYGRKLSLELLDP-AFDFTD - - - - DARQWIDRMLDYN - -  
 Au3446 133 ATALTTPQRQNRPVSEADSTATMDPITSP - - - -  
 SsHexPPS 4 IEFWLEAKATIDRLIEOFINNSNRDWDLVD - - - -  
 MiHexPPS 1 - MIALSYKAFLNPMIIEVEKRYECIQS - - - -  
  
 Au8003 325 MLRAEHGVAKYPARYSLENRKNGANGVNGVNGINGVNGVNGVNGKRKRSGEETA - - - -  
 AcOS 323 LRLSEHGLAKYPSRWQSENRTSLG - - - - VEGDCHESKPN - - ELKRKRGNSVSD - -  
 EvSS 328 LELMKNGVPKDPASGSTNTGTSNGT - - - - SNGTSHVAVNG - - NGHVTNDDLSAN - -  
 PaPS 330 VARMKYGPWNHREVTKLTSLENRGTTKRTHQDTEGVQSVKRFNGASTKNGINGTNGI - - - -  
 PaFS 337 LEWWRQGLPSLECPVLAERSPEID - - - - SDESAVSPTAD - - ESDSTEDLSGS - -  
 AbFS 333 LDWMQNGLPS - - - - TVE - - - - LDGASNTSSSF - - LSTSTHGSPAS - -  
 Au13192 338 LLRDAAESREEKPYIYETETTRWGSSPSESESDASEGSPFWSGSSRSSVRSSVS - - - -  
 EvS 337 LLHQDTIQLQEAEAG - - - - RLPNYPVVPVAMTKSTNGYSES LTNR - - - -  
 Au11565 340 DRAQLVGRSYTAG - - - - LRLPNYPVVPVAMTKSTNGYSES LTNR - - - -  
 NSS 336 LK PANHVAAPTLITPSNNLNSSKGSEEQMQQSDSDNGTRTOMCPANDHEVMQLNAKLSLGKQDGHHAMRAAL  
 Au6298 43 - - - -  
 RpFPSS1 92 - - - -  
 RpFPSS2 96 - - - -  
 EpFPSS 42 - - - -  
 OsaFPSS 47 - - - -  
 Au3446 161 - - - -  
 SsHexPPS 32 - - - -  
 MiHexPPS 28 - - - -



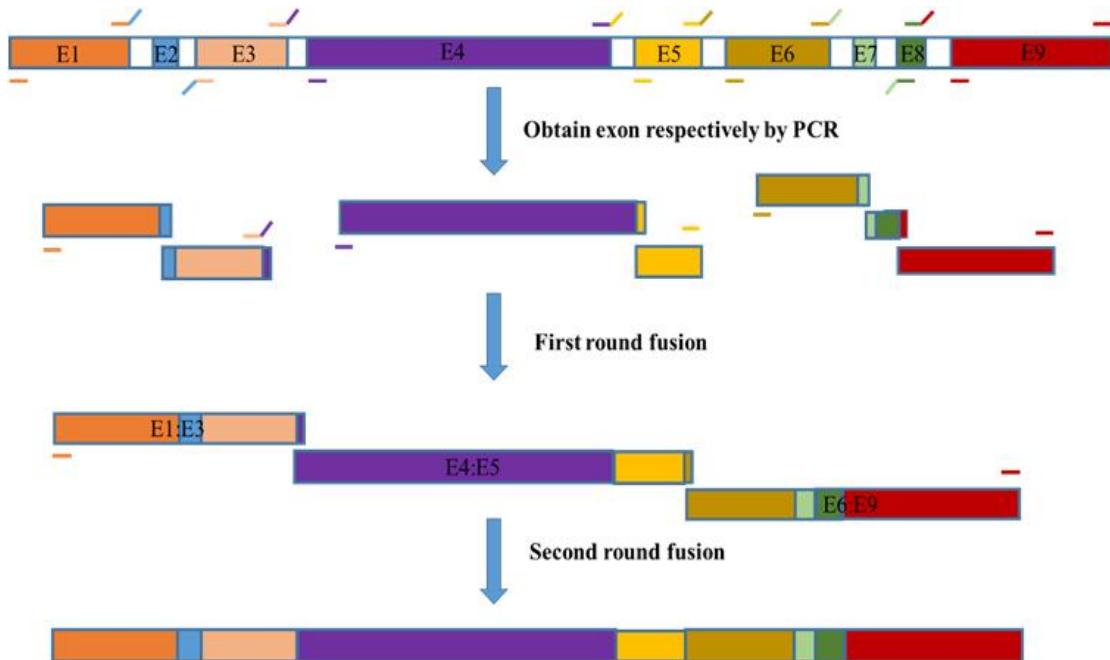
**Figure S5. Alignment of amino acid sequences of the five terpene synthesis related proteins in ophiobolin biosynthesis with identified functions.** Shown here are DDXXD/E (Terpene cyclase (TC) domain), DDXXD (Prenyltransferases (PT) domain), GQ(PT), DDXXN (bifunction PT only) DDXXD (single function PT only), *Aspergillus clavatus* ophiobolin F synthase (AcOS A1C8C3), *Emericella variecolor* stellata-2,6,19-triene Synthase (EvSS LC073704), *Phomopsis amygdali* phomopsene synthase (PaPS AB254159), *Phomopsis amygdali* fusicoccadiene synthase (PaFS AB267396), *Alternaria brassicicola* fusicoccadiene synthase (AbFS C9K2Q3), *Emericella variecolor* Variediene Synthase (EvVS LC063849), *Neosartorya fischeri* sesterfisherol synthase (NfSS EAW16201), *Rhopalosiphum padi* isoprenyl diphosphate synthase (RpFPPS1 and RpFPPS2 HQ850372 and HQ850373), *Ornithogalum saundersiae* farnesyl pyrophosphate synthase (OsaFPPS KF509889), *Euphorbia pekinensis* farnesyl diphosphate synthase (EpFPPS FJ755465), *Sulfolobus solfataricus* Hexaprenyl Pyrophosphate Synthase (SsHexPPS 2AZK\_B), and *Micrococcus luteus* heterodimeric hexaprenyl diphosphate synthase (MIHexPPS 3AQB\_B).



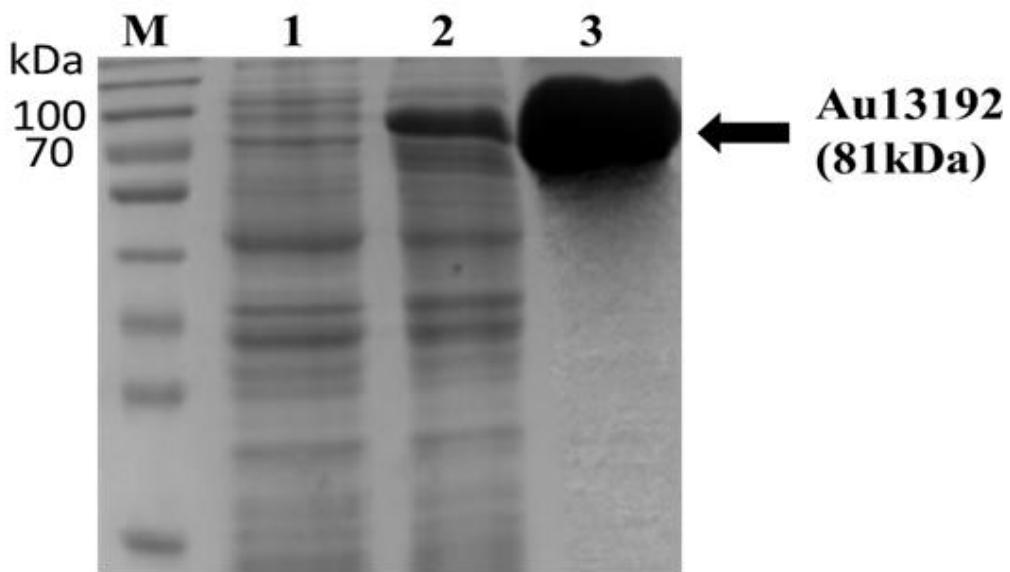
**Figure S6 Neighbour-joining phylogenetic tree based on amino acid sequences of bifunctional terpene synthases, farnesyl pyrophosphate synthases and hexprenyl pyrophosphate synthases.** The tree was generated based on Kimura 2-parameter matrix in MEGA software (version 6.0). Numbers at nodes are bootstrap values obtained using 1000 iterations. The scale bar indicates 0.2 substitutions per nucleotide position. Reference sequences were the same as in Figure S5.



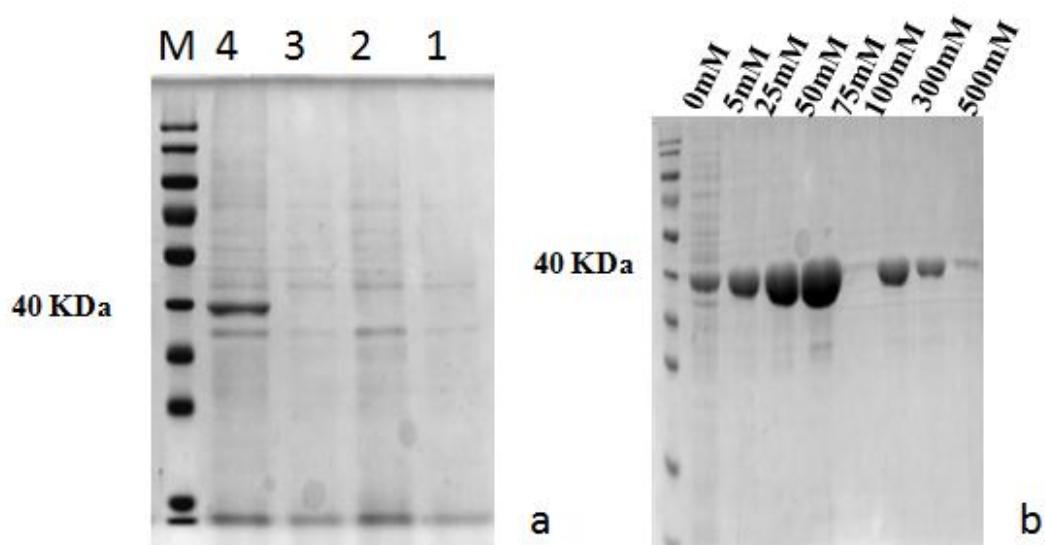
**Figure S7. SDS-PAGE analysis of expression and purification of Au8003.** (a) Overexpression of Au8003 gene in BL21(DE3). Lane M: Marker; Lane 1: uninduced (without isopropyl (3-D-thiogalactopyranoside) IPTG) bacterial protein carrying plasmid pET28a; Lane 2: induced (with IPTG) bacterial protein carrying plasmid pET28a, Lane 3: uninduced bacterial protein carrying the Au8003 cDNA; Lane 4: induced bacterial protein carrying the Au8003 cDNA; (b) purification with different concentrations of imidazole.



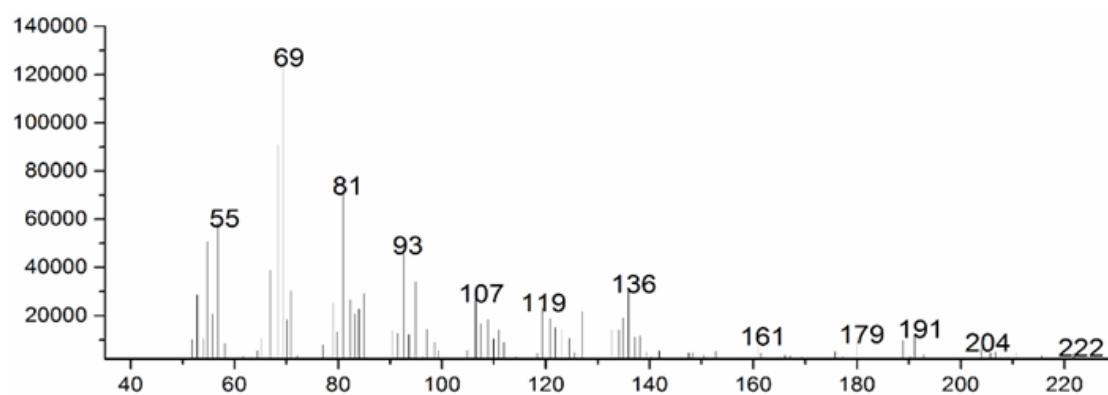
**Figure S8. Strategy of Au13192 multiple exon fusion by overlap extension PCR.** Stage 1: designing primers and amplifying exon 1 (E1) to exon 9 (E9) with overlap sequences through normal PCR (because the E2 and E7 segments are too short, the two gene sequences were designed according to the adjacent two exons of the reverse primer and forward primer, respectively); Stage 2: first round fusion obtained E1:E3/E4:E5/E6:E9, using adjacent exon as template with forward primers of front end exon and reverse primers of tail end exon; Stage3: second fusion obtained Au13192 cDNA following stage 2.



**Figure S9. SDS page gel detection of overexpression and purification of Au13192.** Lane M: Marker; Lane 1: uninduced (without isopropyl (3-D-thiogalactopyranoside) bacterial protein carrying the 2.1-kb Au13192 cDNA; Lane 2: induced (with isopropyl p-D-thiogalactopyranoside) bacterial protein carrying the 2.1-kb Au13192 cDNA; Lane3: purified Au13192



**Figure S10. SDS-PAGE analysis of expression and purification of Au 6298.** (a) Overexpression of Au6298 genes in BL21(DE3). Lane M: Marker; Lane 1: uninduced (without isopropyl (3-D-thiogalactopyranoside) IPTG) bacterial protein carrying plasmid pET28a; Lane 2: induced (with IPTG) bacterial protein carrying plasmid pET28a, Lane 3: uninduced bacterial protein carrying the Au6298 cDNA; Lane 4: induced bacterial protein carrying the Au6298 cDNA. (b) purification with different concentrations of imidazole.



**Figure S11 Mass spectrum of authentic farnesol.**