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724 **Supplementary figures:**

725 **Figure S1A. E-score distribution of top four 6mers recognized by FUS3 and ABI3.**

726 The elements are shown in descending order in relation to their E-score. All top scoring elements
727 recognized by both proteins share the sequence GCATG (underlined), which could be considered as a
728 RY-core. The motif ACATGT corresponds to a “mutant” version of RY-core in which the nucleotide G
729 at the 5' end has been replaced by an A, reflecting a decrease in E-score. Both proteins recognize
730 similar elements, although DNA-binding affinity was slightly lower for ABI3 compared to FUS3.

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732 **Figure S1B.** Score plots of the *proOLE1* sequence scanned with DNA-binding matrices described for
733 LEC2, FUS3 and ABI3.

734 Since matrices correspond to pseudo-palindromic sequences, each binding site is systematically
735 identified on both DNA strands with slightly different scores. This explains the presence of double bars
736 in the plots.

737

738 **Figure S2.** Schematic representation of the mutagenized versions of the *OLE1* promoter used in this
739 study.

740 Regulatory elements (RYL and GBL domains) present in the minimal *OLE1* promoter were
741 mutagenized using site-directed mutagenesis. Mutagenized RYL (blue box) and GBL (red box) are
742 represented. Figures indicate the position of these elements relative to the translational start site of
743 *OLE1*.

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745 **Figure S3.** Activation of the native or mutated *OLE1* promoter versions by LEC2 in moss protoplasts.

746 Means of at least 3 replicates +/- SD

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748 **Figure S4.** Complementary analyses related to activation studies in moss protoplasts.

749 Transient activation of the *OLE1* promoter by LEC2 (A) or ABI3 (B) fused to a strong activator domain
750 (VP16) were assayed in moss protoplasts. Means of at least 3 replicates +/- SD are presented.

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752 **Figure S5. Test of synergistic effect of LEC2, LEC1, and ABI3 on the activation of several**
753 **oleosin promoters in moss protoplasts.**

754 (A) Transient expression assays were carried out in moss protoplasts. GFP activity was measured in
755 protoplasts transfected with *ProOLE1:GFP*, *ProOLE2:GFP*, or *ProOLE3:GFP* reporter and constructs
756 allowing the expression of L-AFLs (LEC2, ABI3 or LEC1), alone or in combination. Means of at least 3
757 replicates +/- SD are presented. AU, arbitrary units.

758 (B) Schematic representation of *proOLE1* (At4g25140), *proOLE2* (AT5G40420), and *proOLE3*
759 (At5g51210) genomic regions. The RYL and GBL elements are highlighted in blue and red,
760 respectively.

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762 **Figure S6. *In vitro* binding of the L-AFLs to the *OLE1* promoter.**

763 Electrophoretic mobility shift assays (EMSA) with LEC1, LEC2, and ABI3 alone or in combination were
764 carried out using a labelled 100-bp long probe corresponding region -228 to -121 (relative to the
765 translational start site) of the *OLE1* promoter. Competition experiments were performed by adding
766 non-labelled competitor DNA added in 25-fold molar excess. Shifted bands are indicated with stars,
767 arrows indicate supershifts.

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769 **Figure S7. Complementary results to pull-down experiments (Figure 6).**

770 (A-B) NF-YC2 interacts with LEC1 or its mutated version (LEC1m, D55K).

771 The HA- or Myc-tagged proteins have been co-expressed in *Arabidopsis* protoplasts. INPUT, 2 % of
772 the full protein extract; IP, immunoprecipitated sample; *, heavy and light chains of the antibody used
773 for the immunoprecipitation.

774 (C-D) LEC1 and NF-YC2 interact with each other but do not interact with LEC2

775 The HA- or Myc-tagged proteins have been co-expressed in *Arabidopsis* protoplasts. INPUT, 2 % of
776 the full protein extract; IP, immunoprecipitated sample; *, heavy and light chains of the antibody used
777 for the immunoprecipitation.

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780 **Figure S8. Study of protein-protein interactions in yeast.**

781 (A) Summary of interaction studies carried out using a yeast two-hybrid system in strain AH109.
782 Reciprocal interactions between LEC1, LEC2, ABI3, and FUS3 were assayed.
783 AD, fusion to activation domain; BD, fusion to DNA-binding domain; W, tryptophan; L, leucine; H,
784 histidine; A, alanine.
785 (B) Study of homodimer formation between native or mutant LEC1 proteins. Yeast two hybrid
786 experiments were performed using native (LEC1) or mutant (LEC1m D55K) proteins. W, tryptophan;
787 L, leucine; H, Histidine; A, alanine; 3-AT, 3-amino -1,2,4-triazole.

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794 **Supplementary table 1. Primers and genes used in this study.**

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Primer name	5'→3' sequence	Gene	AGI
oSB110R	TATGCTCGAGCTAGTAGAAGTCATCGA	FUS3	At3g26790
oETH1132	GGCCATGGTTGATGAAAATGTGGAA	FUS3	At3g26790
oSB109F	TGCCATATGGCAGAGACTCAGTTACCT	ABI3	At3g24650
oSB109R	TGCGGCCGCTTGCCGCTCGGTTGTCT	ABI3	At3g24650
73 mer selex	TGGAGAAGAGGAGAGATCTAGC(N) ₃₀ CTTGTTCTTCTTCGATTCCGG	selex	
Selex F	TGGAGAAGAGGAGAG ATCTAG	selex	
Selex R	CCGG AATCGAAGAAGAACA	selex	
LEC2 F GW	GGGGACAAGTTTGTACAAAAAGCAGGCTCGATGGATAACTTCTTACC	LEC2	At1g28300
LEC2 R GW	GGGGACCACTTTGTACAAGAAAGCTGGGTTCCACCACCACTCAAAG	LEC2	At1g28300
ABI3 F GW	GGGGACAAGTTTGTACAAAAAGCAGGCTCGATGAAAAGCTTGCATGTGGCG G	ABI3	At3g24650
ABI3 R GW	GGGGACCACTTTGTAAAGAAAGCTGGGT GGAAATGGTGTGGTTATAT	ABI3	At3g24650
FUS3 F GW	GGGGACAAGTTTGTACAAAAAGCAGGCTCGATGATGGTTGATGAAAATGTG G	FUS3	At3g26790
FUS3 R GW	GGGGACCACTTTGTAAAGAAAGCTGGGTCTAGTAGAAGTCATCGAGAG	FUS3	At3g26790
LEC1 F GW	GGGGACAAGTTTGTACAAAAAGCAGGCTCG ATGGAACGTGGAGCTCCCTTC	LEC1	At1g21970
LEC1 R GW	GGGGACCACTTTGTAAAGAAAGCTGGGTTCACTTATACTGACCATAATGG	LEC1	At1g21970
NF-YC2 F GW	GGGGACAAGTTTGTACAAAAAGCAGGCTTCATGGAGCAGTCAGAAGAGGGT C	NF-YC2	AT1G56170
NF-YC2 R GW	GGGGACCACTTTGTACAAGAAAGCTGGGTCTTAAGACTCATCAGGGTGTG	NF-YC2	AT1G56170
LEC2-stop R GW	GGGGACCACTTTGTAAAGAAAGCTGGGT CCCACTCAAAGTCGTTAAA	LEC2	At1g28300
LEC1-stop R GW	GGGGACCACTTTGTAAAGAAAGCTGGGTCTTATACTGACCATAATGG	LEC1	At1g21970
ABI3-stop R GW	GGGGACCACTTTGTAAAGAAAGCTGGGTCAATGGTGTGGTTATAT	ABI3	At3g24650
NF-YB 7 F GW	GGGGACAAGTTTGTACAAAAAGCAGGCTATGACTGAGGAGAGCCAGAAGA AG	NF-YB7	At2g13570

NF-YB 7 R GW	GGGGACCACTTTGTACAAGAAAGCTGGGTCTCACCAGTGAATTGAATCAATGT TATTATC	NF-YB7	At2g13570
L1L F GW	GGGGACAAGTTTGTACAAAAAAGCAGGCTATGGAACGTGGAGGCTTCCATGG C	L1L	At5g47670
L1L R GW	GGGGACCACTTTGTACAAGAAAGCTGGGTCTCAGTACTTATGTTGTTGAGTCG G	L1L	At5g47670
pOleo R GW	GGGGACCACTTTGTAAAGAAAGCTGGGTCTTTTTTGTCTTGTTTACTAG	pOleo	At4g25140
pOleo Δ526 F GW	GGGGACAAGTTTGTACAAAAAAGCAGGCTCGGAAGAGAGACGACG	pOleo	At4g25140
pOleo Δ257 F GW	GGGGACAAGTTTGTACAAAAAAGCAGGCTCAACGCCACATTCTGAG	pOleo	At4g25140
pOleo Δ220 F GW	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCTTTGAATAGACTCCTCTCGTTA AC	pOleo	At4g25140
pOleo Δ187 F GW	GGGGACAAGTTTGTACAAAAAAGCAGGCTGCGGCTGCATGGTGACGCC	pOleo	At4g25140
pOleo Δ140 F GW	GGGGACAAGTTTGTACAAAAAAGCAGGCTCTCTCCATTGACACGTGACTTC	pOleo	At4g25140

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808 Supplementary table 2. List of primers used for site-directed mutagenesis of
 809 *ProOLE1*.
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Name of the construct	Mutations (*)	Construct used as a DNA template	Primers	5'→3' Sequence
mutA	RY3,2,1/G*,2,1	ProS3-257 (wild type) in pDONR207	mutA-F mutA-R	AGGCAAAAAACAATTGTGTCTTTGAA TTCAAAGACACAATTGTTTTTGCCT
mutB	RY3,2,1/G3,2,*	ProS3-257 (wild type) in pDONR207	mutB-F mutB-R	CATGATGTCTCCATTGACAGTTGACTTCTCGTCTCCTTTC GAAAGGAGACGAGAAGTCAACTGTCAATGGAGACATCATG
mutC	RY3,2,1/G*,*,1	mut3 (RY3,2,1/G3,*,1)	mutC-F mutC-R	AGGCAAAAAACAATTGTGTCTTTGAA TTCAAAGACACAATTGTTTTTGCCT
mutD	RY3,2,1/G3,*,*	mut3 (RY3,2,1/G3,*,1)	mutD-F mutD-R	CATGATGTCTCCATTGACAGTTGACTTCTCGTCTCCTTTC GAAAGGAGACGAGAAGTCAACTGTCAATGGAGACATCATG
mutE	RY3,2,1/G*,2,*	mutA (RY3,2,1/G*,2,1)	mutE-F mutE-R	CATGATGTCTCCATTGACAGTTGACTTCTCGTCTCCTTTC GAAAGGAGACGAGAAGTCAACTGTCAATGGAGACATCATG
mutF	RY3,*,1/G3,2,1	ProS3-257 (wild type) in pDONR207	mutF-F mutF-R	CGTTAACACATGCAGCGGCTACAACGTGACGCCATTAACACGTG CACGTGTTAATGGCGTCACGTTGTAGCCGCTGCATGTGTTAACG
mutG	RY3,2,*/G3,2,1	ProS3-257 (wild type) in pDONR207	mutG-F mutG-R	GGCCTACAATTACAACATGTCTCCATTG CAATGGAGACATGTTGTAATTGTAGGCC
mutH	RY*,2,1/G3,2,1	ProS3-257 (wild type) in pDONR207	mutH-F mutH-R	AGACTCCTCTCGTTAACAGTTGTAGCGGCTGCATGGTGACG CGTCACCATGCAGCCGCTACAACGTGTTAACGAGAGGAGTCT
mutI	RY*,*,1/G3,2,1	mutH (RY*,2,1/G3,2,1)	mutI-F mutI-R	CGTTAACAGTTGTAGCGGCTACAACGTGACGCCATTAACACGTG CACGTGTTAATGGCGTCACGTTGTAGCCGCTACAACGTGTTAACG
mutJ	RY3,*,*/G3,2,1	mutG (RY3,2,*/G3,2,1)	mutJ-F mutJ-R	CGTTAACACATGCAGCGGCTACAACGTGACGCCATTAACACGTG CACGTGTTAATGGCGTCACGTTGTAGCCGCTGCATGTGTTAACG
mutK	RY*,2,*/G3,2,1	mutH (RY*,2,1/G3,2,1)	mutK-F mutK-R	GGCCTACAATTACAACATGTCTCCATTG CAATGGAGACATGTTGTAATTGTAGGCC
mutL	RY*,*,*/G3,2,1	mutK (RY*,2,*/G3,2,1)	mutL-F mutL-R	CGTTAACAGTTGTAGCGGCTACAACGTGACGCCATTAACACGTG CACGTGTTAATGGCGTCACGTTGTAGCCGCTACAACGTGTTAACG
mutM	RY*,*,1/G*,2,1	mutI (RY*,*,1/G3,2,1)	mutM-F mutM-R	AGGCAAAAAACAATTGTGTCTTTGAA TTCAAAGACACAATTGTTTTTGCCT
mutN	RY*,*,1/G3,2,*	mutI (RY*,*,1/G3,2,1)	mutN -F mutN -R	CATGATGTCTCCATTGACAGTTGACTTCTCGTCTCCTTTC GAAAGGAGACGAGAAGTCAACTGTCAATGGAGACATCATG
mutP	RY*,*,1/G3,*,1	mutI (RY*,*,1/G3,2,1)	mutP-F mutP-R	CGTGACGCCATTAACAGTTGGCCTACAATTGCATG CATGCAATTGTAGGCCAAGTCAACTGTTAATGGCGTCACG
mutQ	RY3,*,*/G*,2,1	mutJ (RY3,*,*/G3,2,1)	mutQ-F mutQ-R	AGGCAAAAAACAATTGTGTCTTTGAA TTCAAAGACACAATTGTTTTTGCCT
mutR	RY3,*,*/G3,2,*	mutJ (RY3,*,*/G3,2,1)	mutR-F mutR-R	CAACATGTCTCCATTGACAGTTGACTTCTCGTCTCCTTTC GAAAGGAGACGAGAAGTCAACTGTCAATGGAGACATGTTG
mutS	RY3,*,*/G3,*,1	mutJ (RY3,*,*/G3,2,1)	mutS-F mutS-R	CGTGACGCCATTAACAGTTGGCCTACAATTACAAC GTTGTAATTGTAGGCCAAGTCAACTGTTAATGGCGTCACG
mutT	RY*,2,*/G*,2,1	mutK (RY*,2,*/G3,2,1)	mutT-F mutT-R	AGGCAAAAAACAATTGTGTCTTTGAA TTCAAAGACACAATTGTTTTTGCCT
mutU	RY*,2,*/G3,2,*	mutK (RY*,2,*/G3,2,1)	mutU-F mutU-R	CAACATGTCTCCATTGACAGTTGACTTCTCGTCTCCTTTC GAAAGGAGACGAGAAGTCAACTGTCAATGGAGACATGTTG
mutW	RY*,2,*/G3,*,1	mutK (RY*,2,*/G3,2,1)	mutW-F	GGTGACGCCATTAACAGTTGGCCTACAATTACAAC

			mutW-R	GTTGTAATTGTAGGCCAACTGTTAATGGCGTCACC
mut2	RY3,2,1/G*,**	mutD (RY3,2,1/G3,*,*)	mut2-F mut2-R	AGGCAAAAAACAATTGTGTCTTTGAA TTCAAAGACACAATTGTTTTTGCCT
mut3	RY3,2,1/G3,*,1	ProS3-257 (wild type) in pDONR207	mut3-F mut3-R	GGTGACGCCATTAACAGTTGGCCTACAATTGCATG CATGCAATTGTAGGCCAACTGTTAATGGCGTCACC

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	proOLE1:GFP	proOLE1:GFP + LEC1	proOLE1:GF P + LEC2	proOLE1:GFP + ABI3	proOLE1:GFP + FUS3	proOLE1:GFP + LEC1 + LEC2	proOLE1:GFP + ABI3 + LEC1	proOLE1:GFP + FUS3 + LEC1	proOLE1:GFP + ABI3 + LEC2	proOLE1:GFP + FUS3 + ABI3	proOLE1:GFP + LEC2 + ABI3 + LEC1
proOLE1:GFP + LEC1	0.2406										
proOLE1:GFP + LEC2	5.2e-13	1.9e-07									
proOLE1:GFP + ABI3	6.6e-12	4.2e-07	1.33e-4								
proOLE1:GFP + FUS3	4.8e-07	6.4e-05	3.819e-4	4.741e-08							
proOLE1:GFP + LEC1 + LEC2	3.21-11	1.2e-06	4.066e-3	0.6098	NR						
proOLE1:GFP + ABI3 + LEC1	6.6e-12	4.2e-07	1.515e-09	8.839e-06	NR	8.83e-05					
proOLE1:GFP + FU3 + LEC1	3.06e-08	1.0e-05	7.082e-05	3.647e-09	0.2636	5.41e-07	1.802e-07				
proOLE1:GFP + ABI3 + LEC2	1.3e-11	5.3e-07	8.532e-08	6.117e-3	NR	NR	0.0151	NR			
proOLE1:GFP + ABI3 + LEC1 + LEC2	8.2e-15	6.0e-08	2.299e-11	1.643e-07	NR	6.331e-07	0.02368	NR	6.243e-05		
proOLE1:GFP + FUS3 +ABI3	8.742e-07	5.017e-05	5.891 ^e -3	0.472	2.955e-4	NR	0.00783	NR	0.1375	NR	
proOLE1:GFP + FUS3 + ABI3 + LEC1	2.997e-06	9.486e-05	8.544e-05	6.115e-3	6e-4426	NR	0.9628	6.426e-4	NR	0.09447	8.03e-3

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	pS3 ⁻²⁵⁷ :GFP	pS3 ⁻²⁵⁷ :GFP + LEC2	pS3 ⁻²⁵⁷ :GFP + LEC2 + LEC1	pS3 ⁻²⁵⁷ :GFP + ABI3	pS3 ⁻²⁵⁷ :GFP + ABI3 + LEC1	pS3 ⁻²⁵⁷ :GFP + ABI3 + LEC2	pS3 ⁻²⁵⁷ :GFP + LEC2 + LEC1 + ABI3	pS3 ⁻²⁵⁷ :GFP + LEC2 + ABI3 + NFYB7
pS3 ⁻²⁵⁷ :GFP + LEC2	5.222e-13							
pS3 ⁻²⁵⁷ :GFP + LEC2 + LEC1	3.214e-11	4.066e-3						
pS3 ⁻²⁵⁷ :GFP + LEC2 + NFYB7	4.209e-3	0.195	0.3667					
pS3 ⁻²⁵⁷ :GFP + LEC2 + LEC1mut	0.004209	0.1003	0.01807					
pS3 ⁻²⁵⁷ :GFP + ABI3	6.654e-12	1.332e-4	0.6098					
pS3 ⁻²⁵⁷ :GFP + ABI3 + LEC1	6.654e-12	1.515e-09	8.83e-05	8.839e-06				
pS3 ⁻²⁵⁷ :GFP + ABI3 + NFYB7	4.209e-3	NR	NR	0.4407	0.02917			
pS3 ⁻²⁵⁷ :GFP + ABI3 + LEC1mut	0.004209	NR	NR	0.04274	5.092e-3			
pS3 ⁻²⁵⁷ :GFP + ABI3 + LEC2	1.336e-11	8.532e-08	NR	0.006117	0.01515			
pS3 ⁻²⁵⁷ :GFP + LEC2 + LEC1 + ABI3	8.232e-15	2.299e-11	6.331e-07	1.643e-07	0.02368	6.243e-05		
pS3 ⁻²⁵⁷ :GFP + LEC2 + ABI3 + NFYB7	1.821e-4	NR	NR	NR	NR	0.2421	8.974e-3	
pS3 ⁻²⁵⁷ :GFP + ABI3 + LEC2 + LEC1mut	4.209e-3	NR	NR	NR	NR	0.1337	0.01367	0.8607

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C-1

		pWt +LEC2	pWt +ABI3	pWt +LEC1 LEC2 ABI3	pmutA +LEC2	pmutA +ABI3	pmutA +LEC1 LEC2 ABI3	pmut3 +LEC2	pmut3 +ABI3	pmut3 +LEC1 LEC2 ABI3	pmutB +LEC2	pmutB +ABI3	pmutB +LEC1 LEC2 ABI3	pmut2 +LEC2	pmut2 +ABI3	pmut2 +LEC1 LEC2 ABI3	pmutH +LEC2	pmutH +ABI3	pmutH +LEC1 LEC2 ABI3	
mut A	gain LEC2	0.12																		
mut A	gain ABI3		0.28																	
mut A	gain LEC2 LEC1 ABI3			0.014	0.005	0.002														
mut 3	gain LEC2	0.25			0.15															
mut 3	gain ABI3		0.11			0.25														
mut 3	gain LEC2 LEC1 ABI3			0.001			0.008	0.97	0.012											
mutB	gain LEC2	0.84			0.40			0.34												
mutB	gain ABI3		0.16			0.09			0.039											
mutB	gain LEC2 LEC1 ABI3			0.057			0.52			0.019	0.007	0.016								
mut2	gain LEC2	0.003			0.004			0.05			0.079									
mut2	gain ABI3		0.001			0.008			0.13			0.023								
mut2	gain LEC2 LEC1 ABI3			0.002			0.004			0.014			0.010	0.010	0.007					
mutH	gain LEC2	0.009			0.003			0.051			0.072			0.34						
mutH	gain ABI3		0.13			0.26			0.89			0.037			0.21					
mutH	gain LEC2 LEC1 ABI3			0.02			0.20			0.42			0.13			0.086	0.047	0.056		
mutF	gain LEC2	0.007			0.002			0.052			0.077			0.55			0.35			
mutF	gain ABI3		0.18			0.37			0.88			0.047			0.14					0.79
mutF	gain LEC2 LEC1 ABI3			0.002			0.024			0.16			0.040			0.002				0.81
mutG	gain LEC2	0.013			0.006			0.06			0.11			0.26			0.004			
mutG	gain ABI3		0.75			0.45			0.18			0.31		0.06					0.17	
mutG	gain LEC2 LEC1 ABI3			0.040			0.76			0.07			0.45			0.026				0.31
mutI	gain LEC2	0.006			0.0014			0.049			0.070			0.30			0.82			
mutI	gain ABI3		0.011			0.03			0.34			0.026			0.27				0.47	
mutI	gain LEC2 LEC1 ABI3			0.005			0.014			0.13			0.022			0.010				0.20
mutJ	gain LEC2	0.009			0.005			0.049			0.070			0.26			0.41			
mutJ	gain ABI3		0.018			0.044			0.42			0.026			0.23				0.55	
mutJ	gain LEC2 LEC1 ABI3			0.003			0.006			0.017			0.012			0.42				0.081
mut K	gain LEC2	0.009			0.004			0.050			0.071			0.28			0.57			
mut K	gain ABI3		0.08			0.29			0.70			0.052			0.035				0.63	
mut K	gain LEC2 LEC1 ABI3			0.006			0.08			0.31			0.057			0.042				0.93
mutL	gain LEC2	0.009			0.005			0.045			0.062			0.13			0.03			
mutL	gain ABI3		0.002			0.022			0.32			0.033			0.16				0.45	
mutL	gain LEC2 LEC1 ABI3			0.001			0.007			0.30			0.008		0.12					0.21

C-2

	pmutF +LEC2	pmutF +ABI3	pmutF +LEC1 LEC2 ABI3	pmutG +LEC2	pmutG +ABI3	pmutG +LEC1 LEC2 ABI3	pmutI +LEC2	pmutI +ABI3	pmutI +LEC1 LEC2 ABI3	pmutJ +LEC2	pmutJ +ABI3	pmutJ +LEC1 LEC2 ABI3	pmutK +LEC2	pmutK +ABI3	pmutK +LEC1 LEC2 ABI3	pmutK +LEC2	pmutK +ABI3
pmut A gain LEC2																	
pmut A gain ABI3																	
pmut A gain LEC2 LEC1 ABI3																	
pmut 3 gain LEC2																	
pmut 3 gain ABI3																	
pmut 3 gain LEC2 LEC1 ABI3																	
pmutB gain LEC2																	
pmutB gain ABI3																	
pmutB gain LEC2 LEC1 ABI3																	
pmut2 gain LEC2																	
pmut2 gain ABI3																	
pmut2 gain LEC2 LEC1 ABI3																	
pmutH gain LEC2																	
pmutH gain ABI3																	
pmutH gain LEC2 LEC1 ABI3																	
pmutF gain LEC2																	
pmutF gain ABI3																	
pmutF gain LEC2 LEC1 ABI3	0.002	0.009															
pmutG gain LEC2	0.013																
pmutG gain ABI3		0.22															
pmutG gain LEC2 LEC1 ABI3			0.14	0.020	0.026												
pmutI gain LEC2	0.34			0.09													
pmutI gain ABI3		0.32			0.052												
pmutI gain LEC2 LEC1 ABI3			0.027				0.0003	0.015									
pmutJ gain LEC2	0.18			0.005			0.76										
pmutJ gain ABI3		0.38			0.10			0.84									
pmutJ gain LEC2 LEC1 ABI3			0.005			0.026			0.001	0.0014	0.19						
pmut K gain LEC2	0.21			0.003			0.87			0.79							
pmut K gain ABI3		0.86			0.23			0.13			0.18						
pmut K gain LEC2 LEC1 ABI3			0.84			0.20			0.12			0.041	0.023	0.031			
pmutL gain LEC2	0.048			0.007			0.11			0.021			0.036				
pmutL gain ABI3		0.31			0.10			1			0.82			0.10			
pmutL gain LEC2 LEC1 ABI3			0.08			0.033			0.92			0.11			0.12	0.034	0.052

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- 3 (A) Statistical analyses corresponding to the results presented in Figure 3.
- 4 (B) Statistical analyses corresponding to the results presented in Figure 4
- 5 (C) Statistical analyses corresponding to the results presented in Figure 5 (C1, to be continued with C2)

6 Red: significant differences at 5%

7 NR: not relevant.

8 For A and B, the normality of the data was checked with a Shapiro-Wilk test. As the samples were not all following a Normal law, a Kolmogorov-Smirnov test
 9 was then performed on the relevant comparisons. All the tests are included in the R package

2 C: The normality of the data was checked with a Shapiro-Wilk test. Student t-test was performed on the relevant comparisons using the R stats Package
3 (<https://www.r-project.org/>) version 3.2.