

High-Throughput Single-Cell Derived Sphere Formation for Cancer Stem-Like Cell Identification and Analysis

Yu-Chih Chen^{1,3*}, Patrick N. Ingram^{2*}, Shamileh Fouladdel³, Sean P. McDermott³, Ebrahim Azizi³, Max S. Wicha³ and Euisik Yoon^{1,2}

¹Department of Electrical Engineering and Computer Engineering, University of Michigan, 1301 Beal Avenue, Ann Arbor, MI 48109-2122;

²Dept. of Biomedical Engineering, University of Michigan, Ann Arbor, MI, USA, 2200 Bonisteel, Blvd. Ann Arbor, MI 48109-2099

³University of Michigan Comprehensive Cancer Center, 1500 East Medical Center Drive, Ann Arbor, MI 48109-5940

*Equal contribution as first authors

Corresponding author:

Euisik Yoon, 1301 Beal Avenue, Ann Arbor, MI 48109-2122, 734-615-4469, esyoon@umich.edu.

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SUPPLEMENTARY INFORMATION

Cell Lines	Cell Capture Rate (%)
SUM159	74.1±8.6
MDA-231	83.2±11.3
MCF-7	67.8±18.5
SKOV3	81.7±7.5
C2C12	70.1±3.2

Supplementary Table 1: Single cell capture rates of 5 different cell lines utilizing passive hydrodynamic capture in the single cell devices. Data are shown as the mean ± SD (N = 4).

	Evaporation Process	Proposed Process
Thickness	30.5±2.71 μm	2.04±0.26 μm
Roughness RMS***	3.15±0.50 μm	0.19±0.04 μm
Peak Height	6.55±0.62 μm	0.56±0.07 μm
Valley Depth	9.24±1.65 μm	0.65±0.14 μm

Supplementary Table 2: The comparison between the surface roughness of conventional polyHEMA coating techniques and the presented improved methodology. Data are shown as the mean ± SD (N = 5), *** P < 0.001.

Cell Lines	Sphere Formation Rate (%)
A549	0.6±0.3
SKOV3	2.3±2.1
C6	2.6±2.1

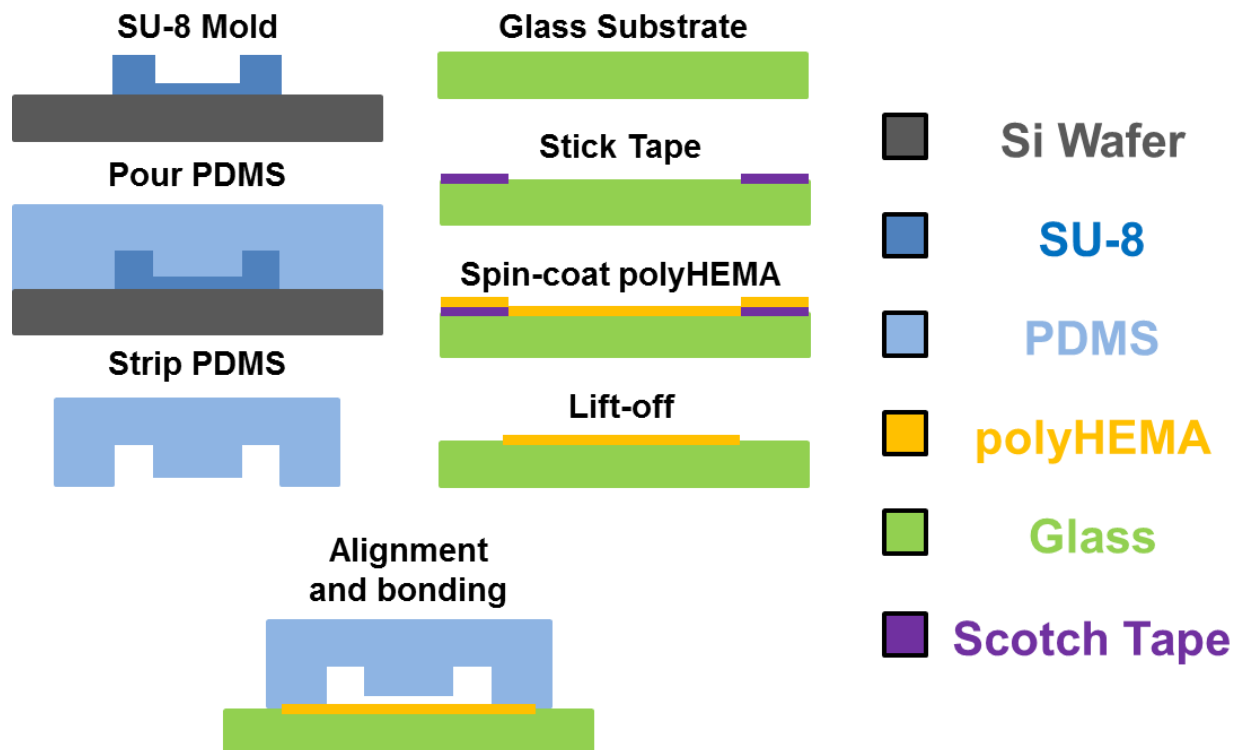
Supplementary Table 3: Single cell-derived sphere formation rates of non-breast cancer cell lines. A549 is lung cancer cell line; SKOV3 is ovarian cancer cell line; and C6 is brain tumor cell line.

Gene	P-Value
EpCAM	1.9E-23
STAP2	9.5E-17
TAZ	3.9E-12
CD14	7.2E-09
ID1	8.8E-09
YAP1	4.1E-07
CCND1	7.8E-06
TM4SF1	8.0E-06
TSPAN6	1.5E-05
ITGA6	9.1E-05
IL6R	1.3E-04
ID2	2.6E-04
CD24	7.5E-04
CXCR1	3.2E-03
CDH1	3.7E-03
TNKS1BP1	7.5E-03
CD44	1.4E-02
GAPDH	1.6E-02
PTEN	3.0E-02
HES1	3.6E-02

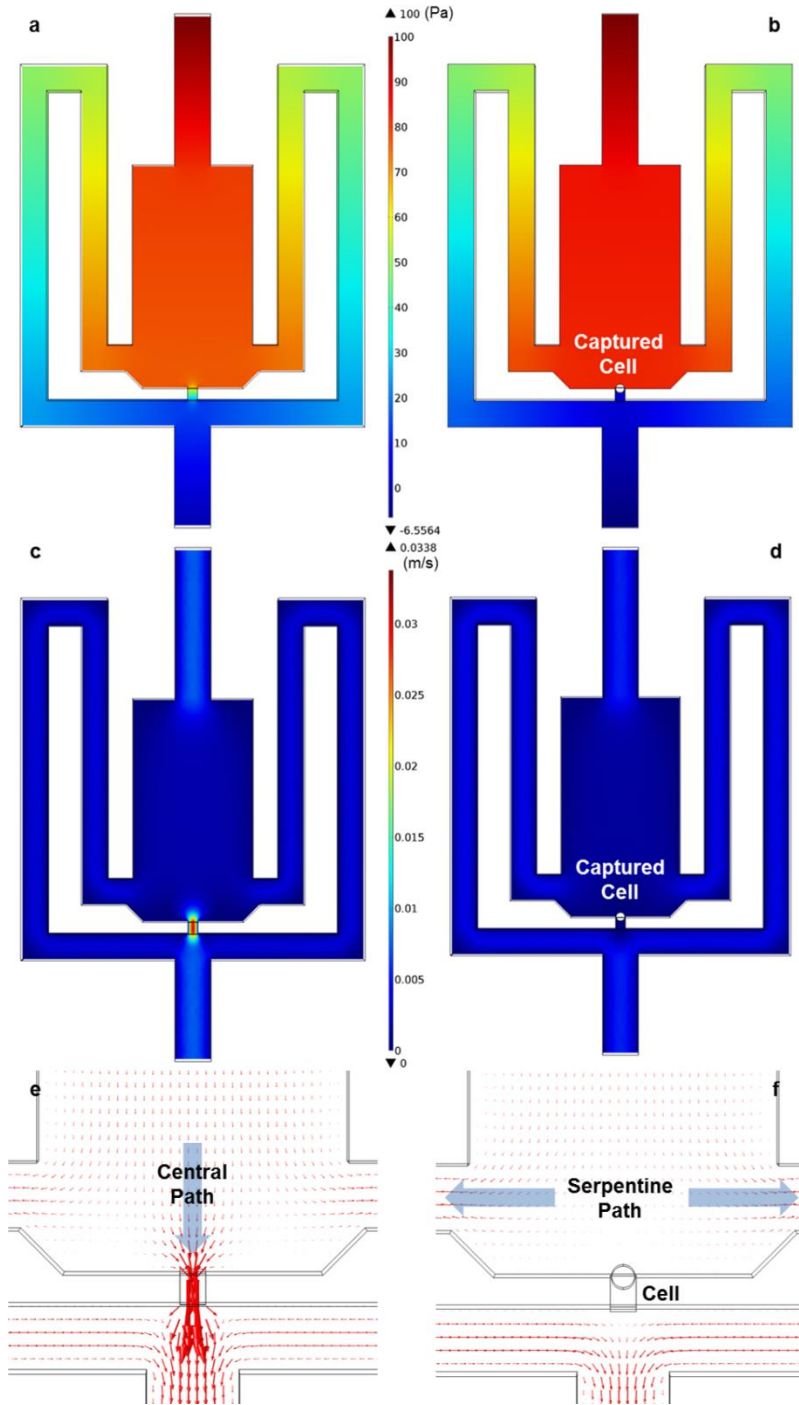
Supplementary Table 4: The p-values of genes that are significantly different between Notch+ sphere derived cells and Notch- sphere derived cells for Figure 5.

Gene	P-Value
ALDH1a3	3.4E-12
bCatenin	1.7E-04
AKT1	8.1E-04
CXCR1	8.3E-04
CXCR4	1.5E-03
TCEA3	3.0E-03
GAPDH	3.5E-03
PPL	5.7E-03
p53	1.0E-02
NUMB	1.1E-02
DLL3	1.6E-02
NESTIN	1.6E-02
TGFb	1.6E-02
DLL4	1.6E-02
TNKS1BP1	2.0E-02
Notch1	2.3E-02
TAZ	2.5E-02
EGFR	2.6E-02
STAP2	3.2E-02
JAG2	5.2E-02
NYNRIN	5.6E-02
TM4SF1	5.7E-02

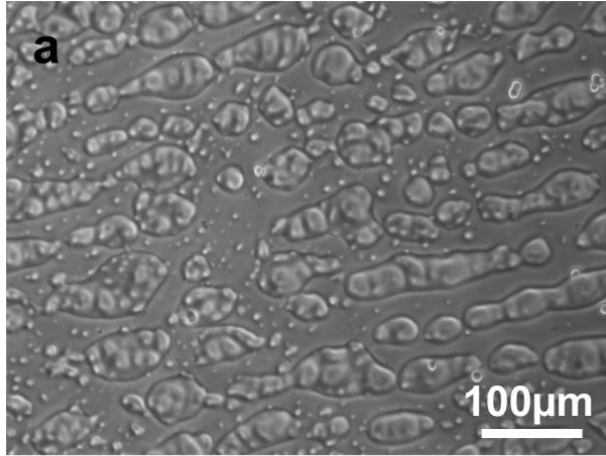
Supplementary Table 5: The p-values of genes that are significantly different in Notch+/ALDH1a3+ cells as compared to Notch+/ALDH1a3- cells in Figure 5c,d.



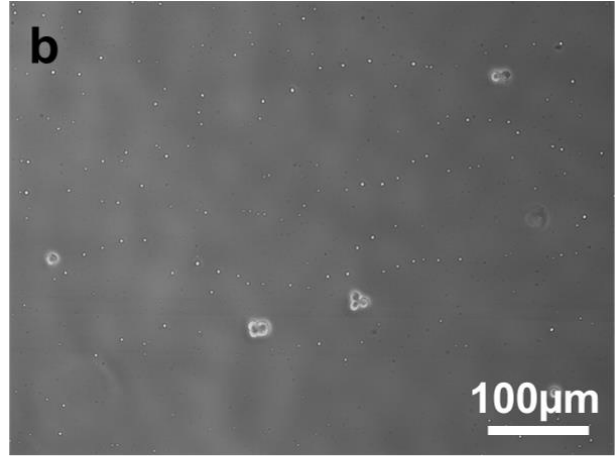
Supplementary Figure 1: Fabrication process of high throughput single cell suspension culture device for mammosphere assays. SU-8 mold is fabricated in a 2-step photolithographic process. PDMS is poured over the mold and cured. After overnight curing, the PDMS is removed from the mold for bonding to the substrate. The glass substrate (microscope slide) is cleaned and masked with scotch tape to restrict the polyHEMA pattern to the area of the microchambers. A polyHEMA ethanol solution (60mg/mL) is spin-coated (1000 rpm for 60 second) over the substrate and reflowed at 150°C after the tape mask is removed. After oxygen plasma oxygenation (100 Watt, 60 second), the substrate and PDMS layer are bonded.



Supplementary Figure 2: Simulations of flow velocity and pressure to evaluate cell capture scheme. (a, b) Simulations of pressure distribution before and after cell capture. (c, d) Simulations of flow velocity before and after cell capture. (e) The simulated flow pattern before cell capture. The red arrows indicating flow direction and velocity suggest that the cells are likely to be guided to the capture site and get captured. (f) After cell capture, the captured cell blocks the flow, so the next cell will be guided into the serpentine path.

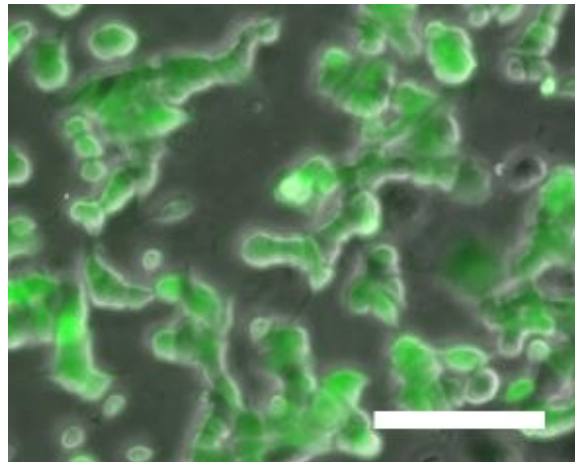


Spin Coating

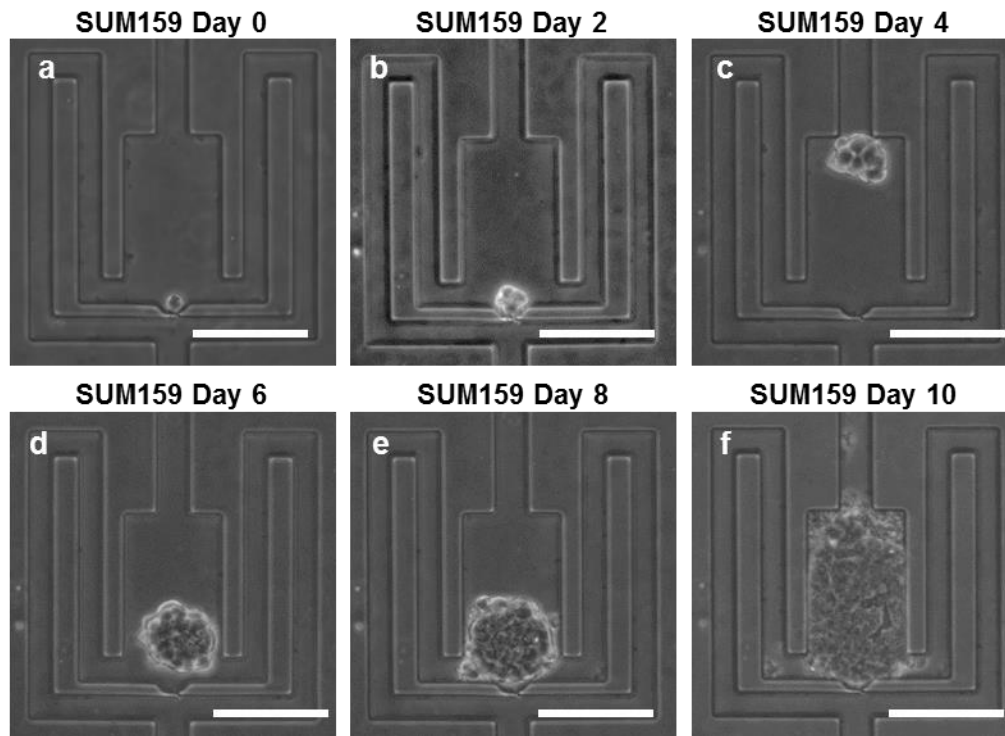


Spin Coating + Reflow

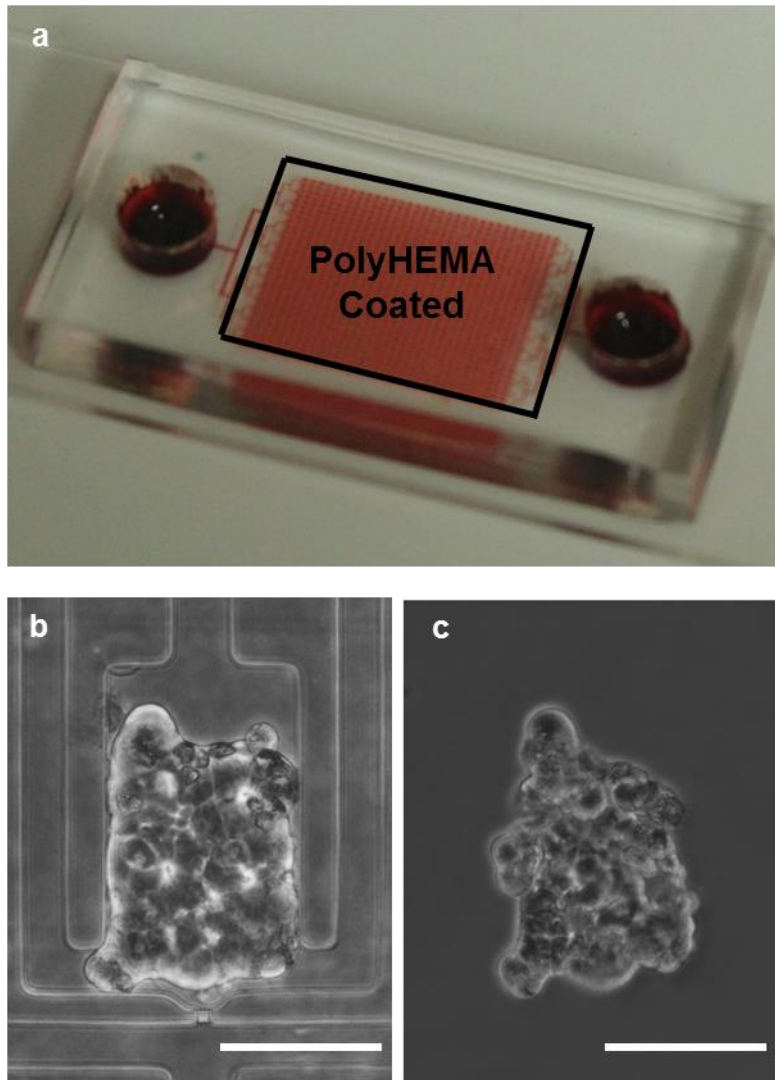
Supplementary Figure 3: PolyHEMA spin-coated surface with and without reflow process: (a) the polyHEMA spin-coated surface without reflow. As the ethanol evaporates fast during the spin-coating, the radial trenches are formed on the substrate. Also, some evaporation bubbles were trapped in the polyHEMA film, making the surface coarse. (b) After reflow at 200 °C overnight, the trench profile was flattened, and the trapped bubbles were released, making the coated surface smoother.



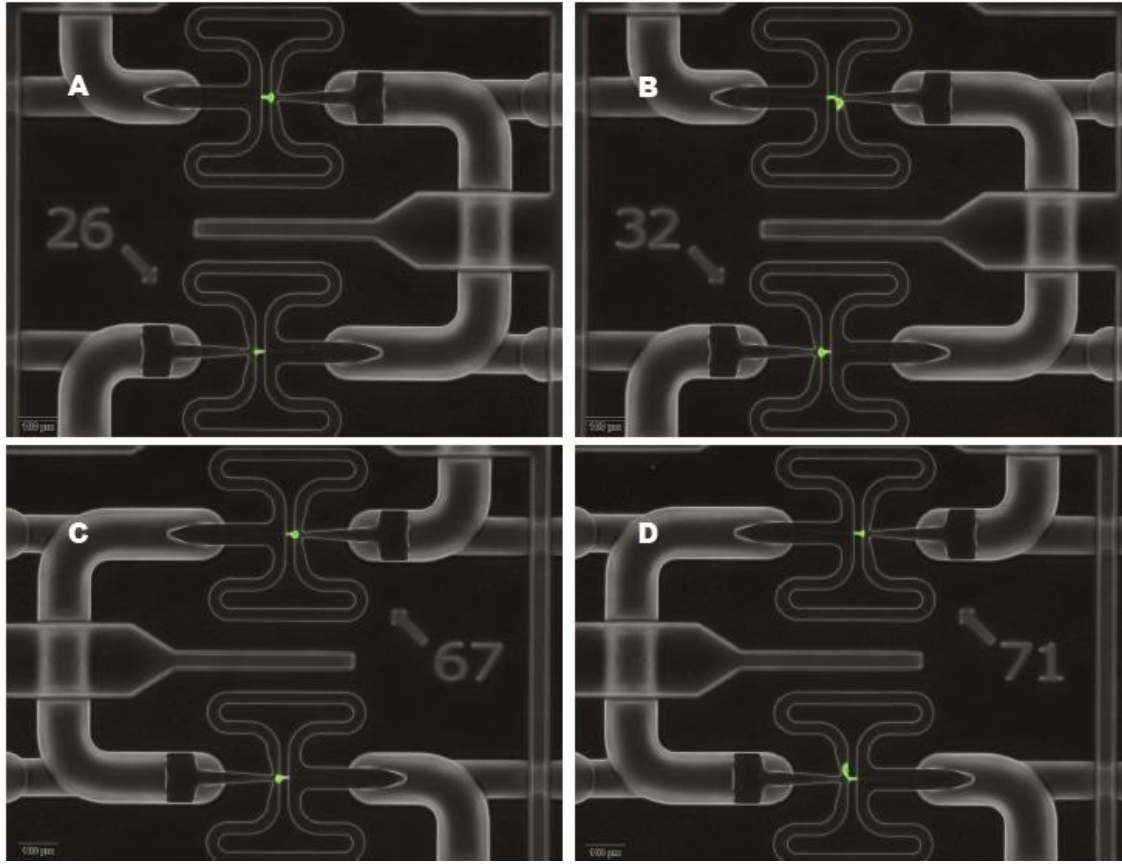
Supplementary Figure 4. T47D culture on blank PDMS (scale bar: 100 μm)



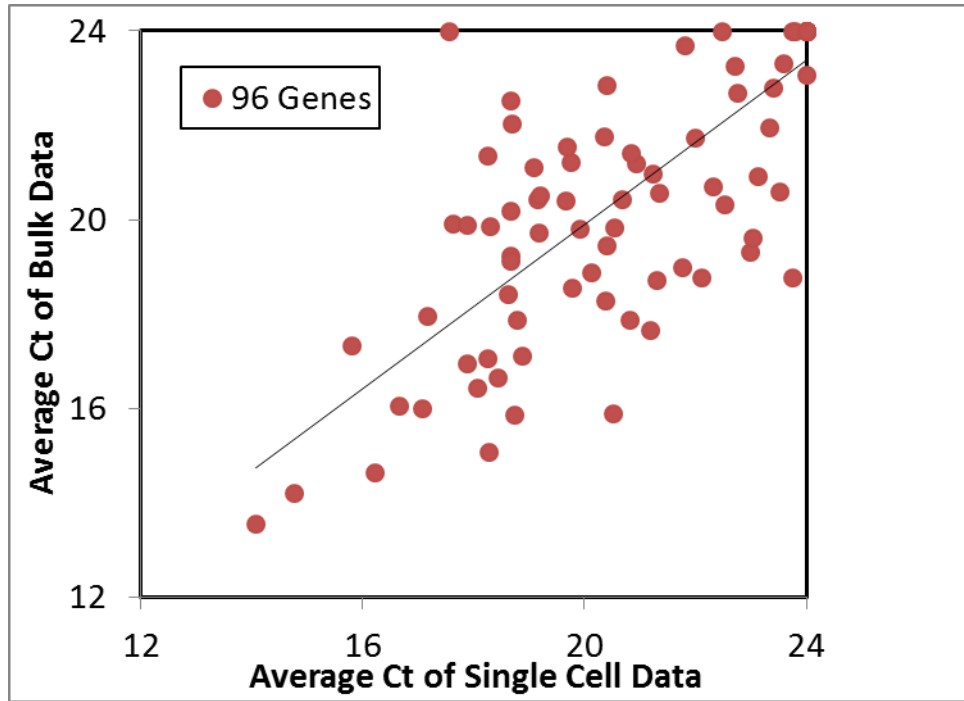
Supplementary Figure 5: Formation and morphology of cancer spheres derived from a single SUM159 breast cancer cell: (a) day 0, (b) day 2, (c) day 4, (d) day 6, (e) day 8, and (f) day 10.



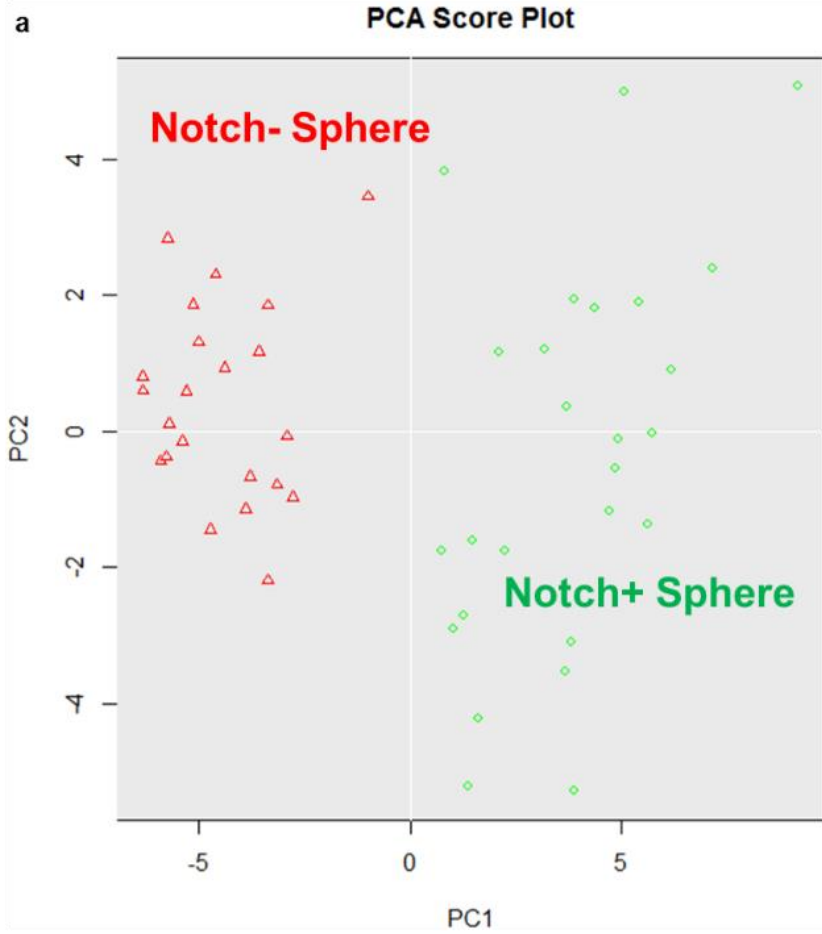
Supplementary Figure 6: Single cell derived sphere retrieval process: (a) Photograph of fabricated 1,024 well suspension culture device with polyHEMA coated area shown. (b) Example of single cell derived sphere in microchamber. After 14 days of culture, the microchamber might restrict further sphere growth. (c) Example sphere of interest in multiwell plate after release and retrieval from microfluidic platform.



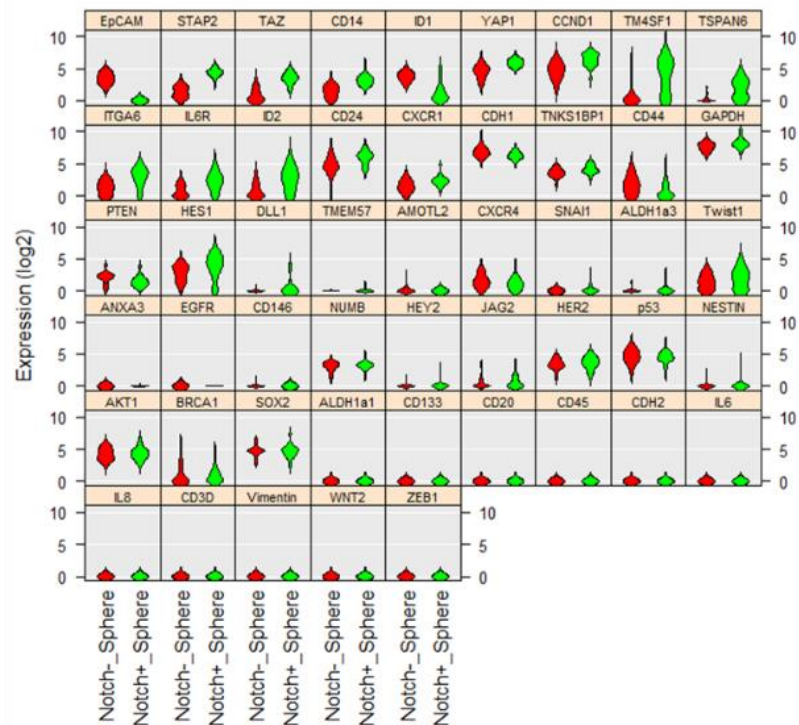
Supplementary Figure 7. Representative images of single breast cancer cells isolated within the C1 chip. (a) Large versus small size round shape single cells, (b) elongated versus round shape single cells, (c) similar size round shape single cells (d) and small round shape versus large elongated single cells.



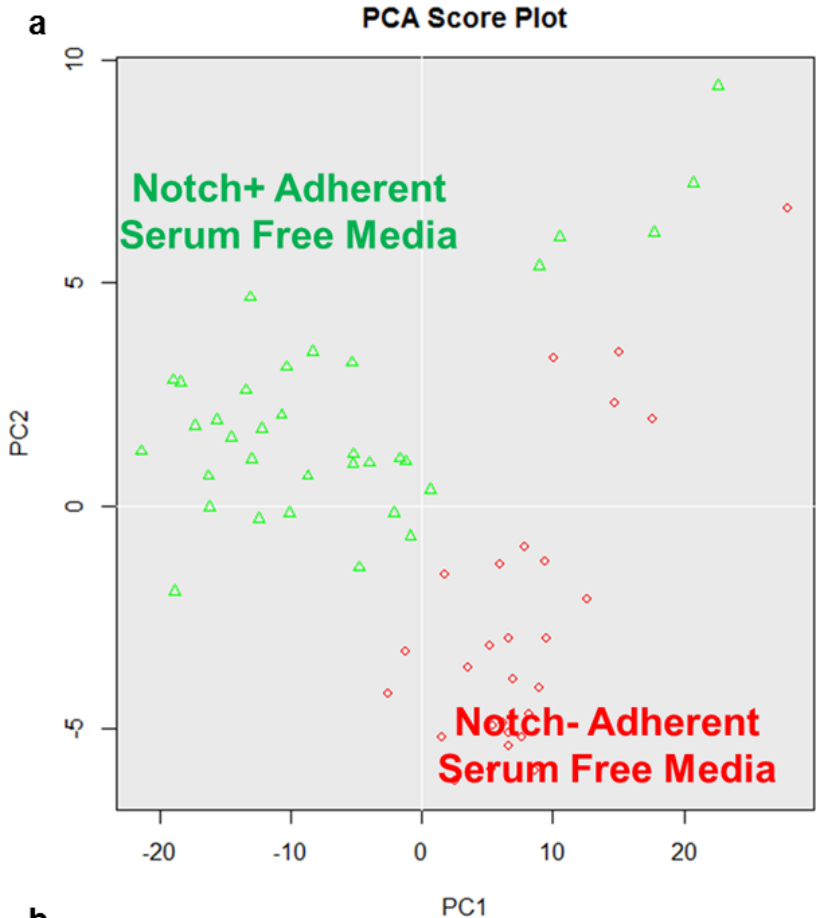
Supplementary Figure 8. Correlation between single T47D cell (n = 28) expression data and bulk T47D (5,000 cells each sample, n = 3) expression data (both normalized to the geometric mean of GAPDH and RAB7A). The average Ct of 96 genes are highly correlated (correlation coefficient $r = 0.79$) between single cell and bulk data.



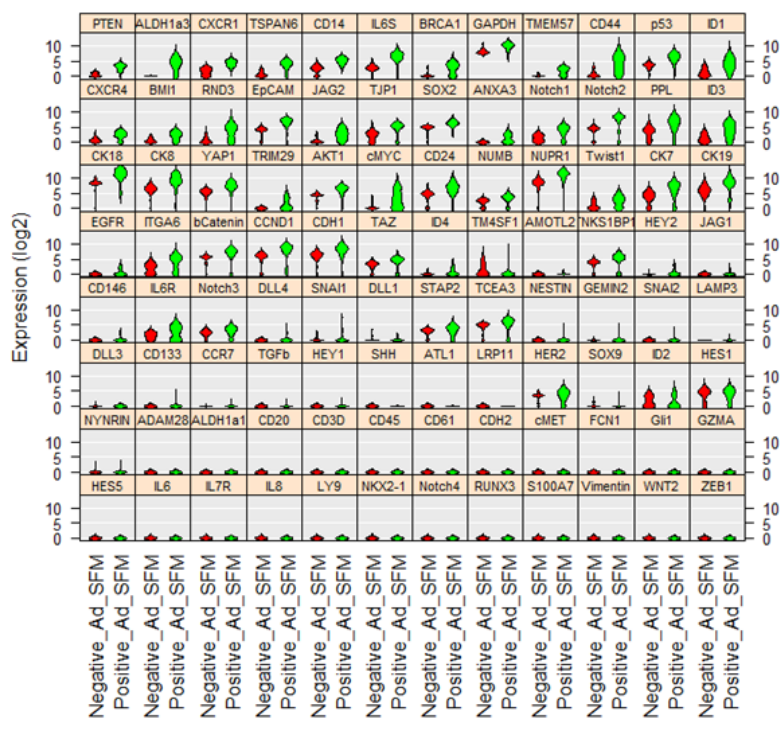
b **Violin Plot of Gene Expression By the Order of PCA Gene Scores**



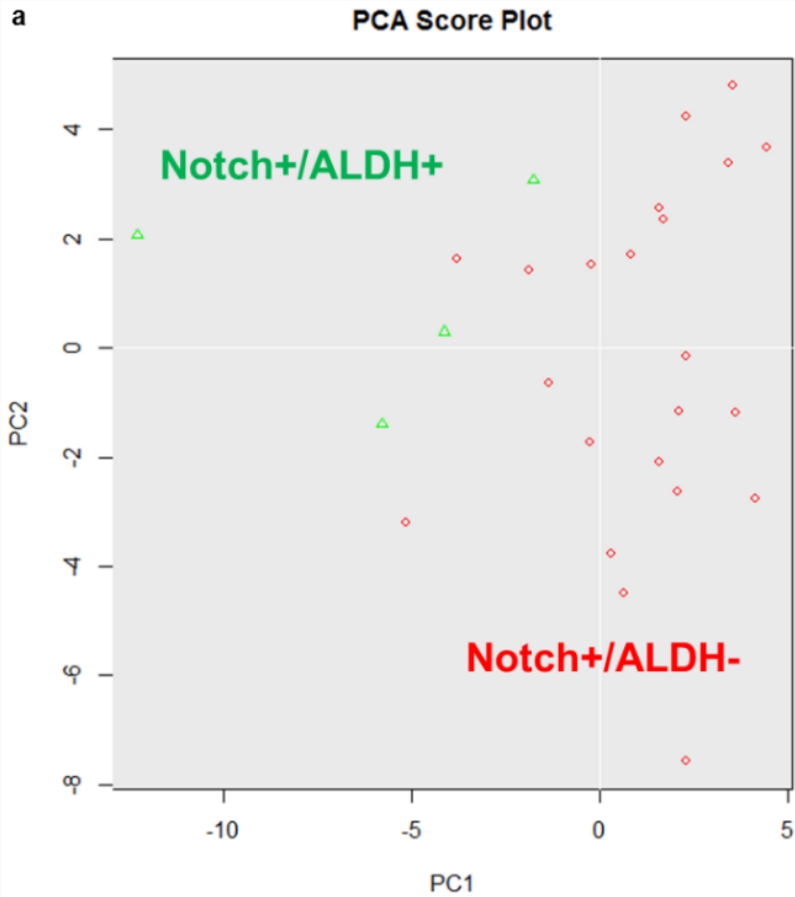
Supplementary Figure 9: Expression data of single cells from retrieved spheres utilizing Fluidigm C1/Biomark HD for multiplexed gene expression analysis comparing cells derived from T47D Notch+ cells and those derived from T47D Notch- cells. (a) PCA clustering of single cell expression analysis for Notch+ cells compared to cells derived from Notch- cells. (b) Violin plots of 50 genes for differences between Notch+ derived sphere cells and Notch- derived sphere cells.



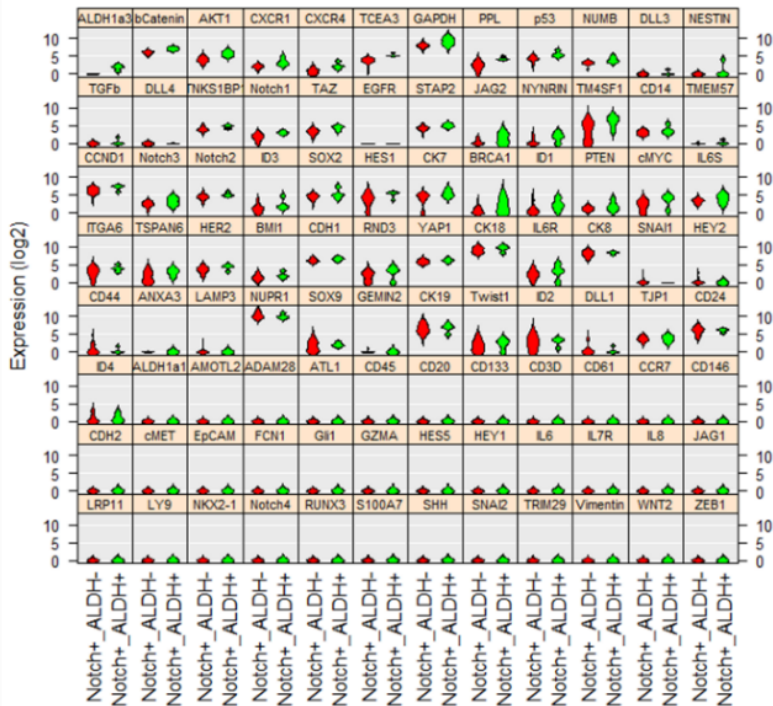
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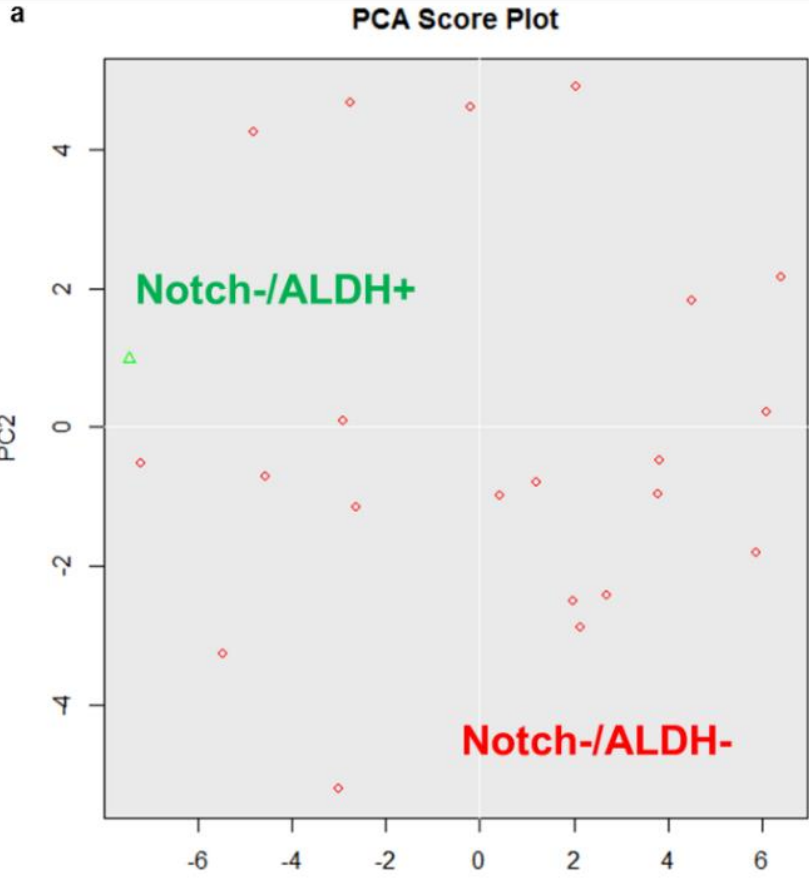
Supplementary Figure 10: Single cell expression data from Notch+ and Notch- cells in adherent culture in serum-free media utilizing Fluidigm C1/Biomark HD for multiplexed gene expression analysis. (a) PCA clustering of single cell expression analysis for Notch+ and Notch- adherent cells. (b) Violin plots of the entire 96-gene panel for differences between Notch+ and Notch- cells from adherent culture.



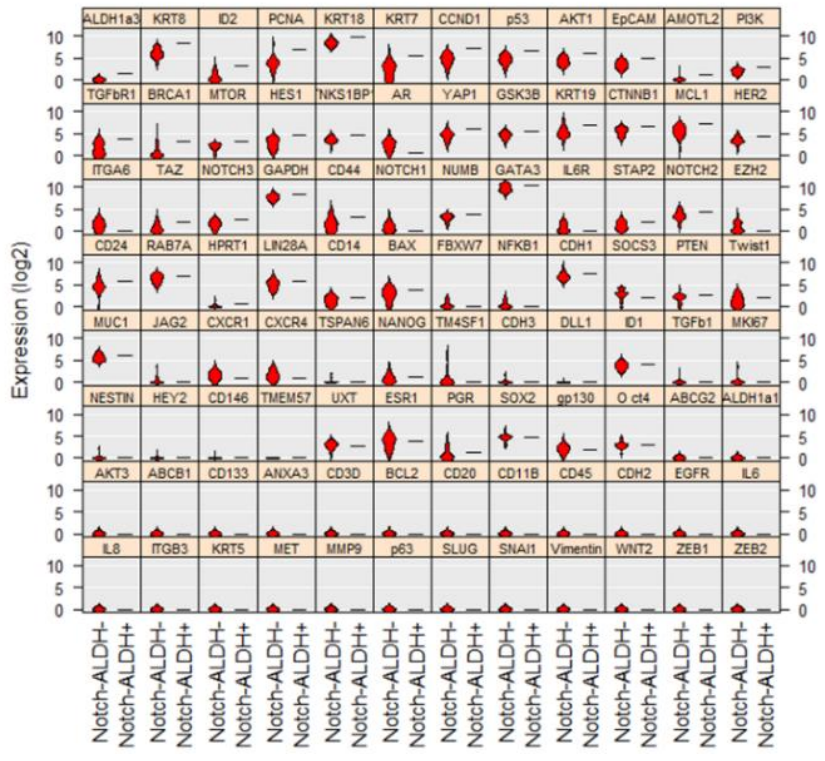
b **Violin Plot of Gene Expression By the Order of PCA Gene Scores**



Supplementary Figure 11: Single cell expression data of retrieved spheres utilizing Fluidigm C1/Biomark HD for multiplexed gene expression analysis comparing Notch+ALDH1a3 high (N=4) and Notch+ALDH1a3 low (N=22) cells. (a) PCA clustering of single cell expression analysis for Notch+ derived ALDH high cells versus ALDH low cells. (b) Violin plots of the entire 96 gene panel for differences between Notch+ derived ALDH high cells versus ALDH low cells

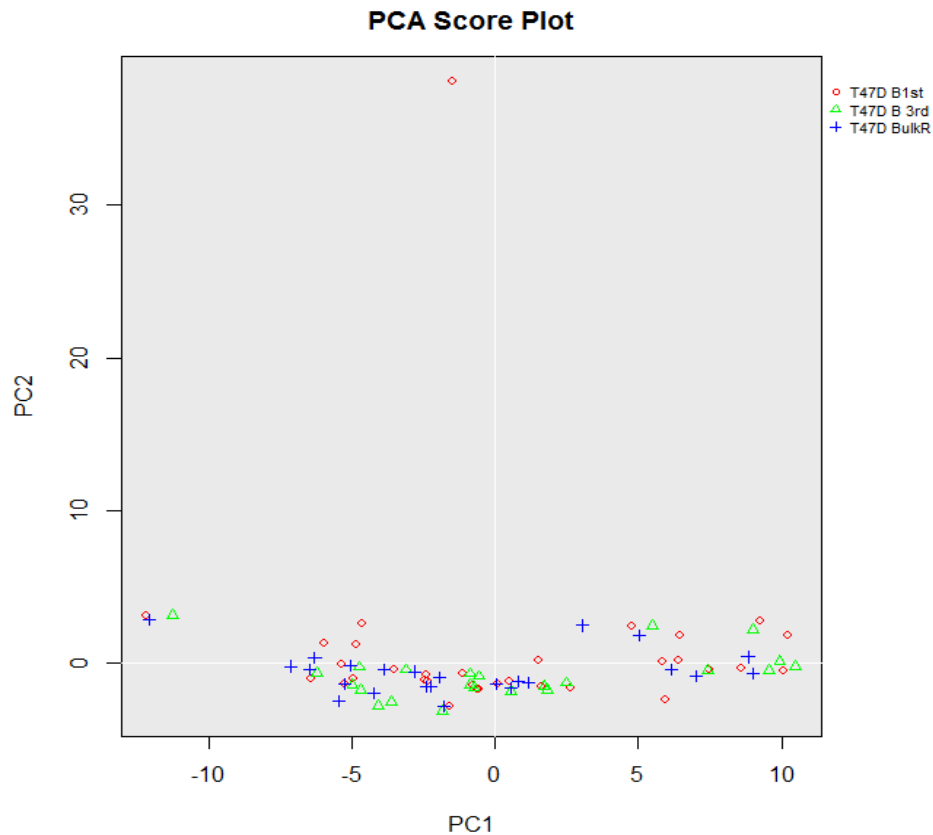


b Violin Plot of Gene Expression By the Order of PCA Gene Scores



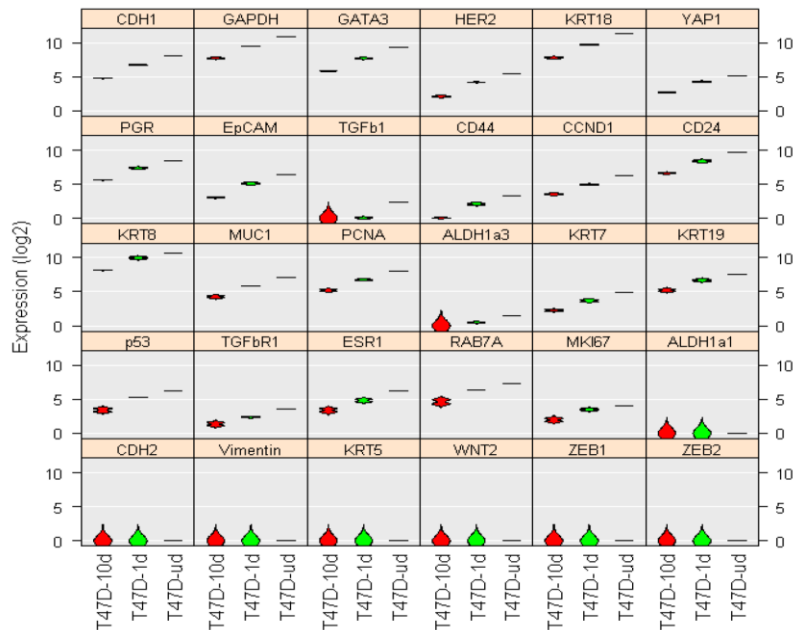
Supplementary Figure 12: Single cell expression data of Notch-ALDH1a3 high and Notch-/ALDH1a3 low cells from Notch- retrieved sphere utilizing Fluidigm C1/Biomark HD for multiplexed gene expression analysis. (a) PCA clustering of single cell expression analysis for Notch-ALDH1a3 high (N=1) and Notch-/ALDH1a3 low (N=21) cells. (b) Violin plots of the entire 96-gene panel for Notch-/ALDH1a3 high cell versus Notch-/ALDH1a3 low cells. Due to only one Notch-ALDH1a3 high cell, there was no statistical comparison possible.

Supplementary Figure 13: Single cell expression data of T47D Notch+ cells from spheres and adherent culture comparing serum and serum free media conditions utilizing Fluidigm C1/Biomark HD for multiplexed gene expression analysis. (a) PCA clustering of single cell expression analysis for Notch+ derived cells in adherent culture with and without serum in the media compared to Notch+ derived cells from spheres in serum free media. (b) Violin plots of the entire 96-gene panel for differences between these populations.

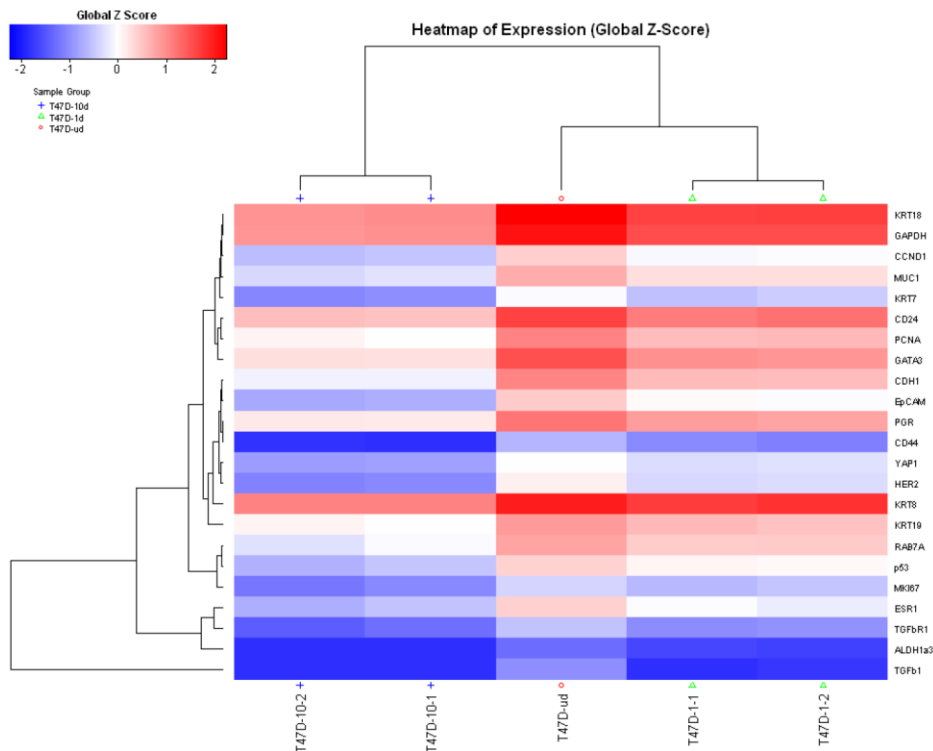


Supplementary Figure 14. Principal component analysis (PCA) plot showing log₂ gene expression data of single cells from T47D breast cancer cell line. T47D B1st (Red) and T47D BulkR (Blue) are the same single cells isolated by C1 and analyzed by BioMark HD in 2 separate RT-qPCR experiments as technical replicates. T47D B3rd (Green) and T47D B1st (Red) are biological replicates of 2 independent C1/BioMark HD experiments.

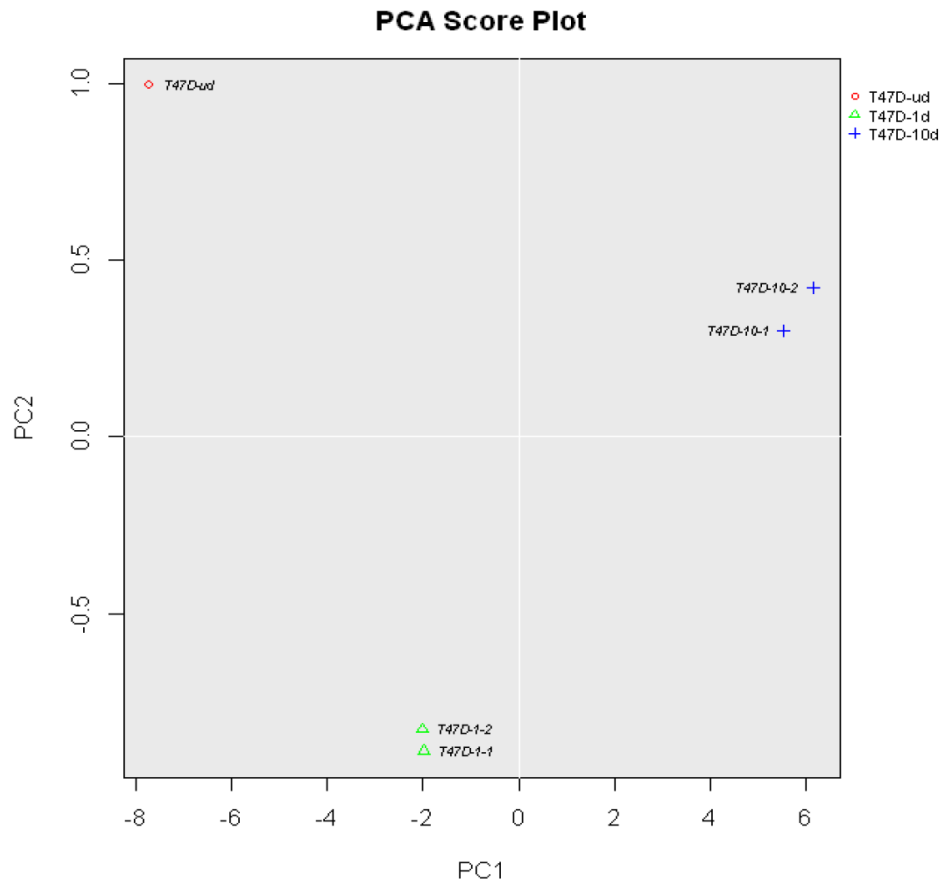
Violin Plot of Gene Expression By the Order of PCA Gene Scores



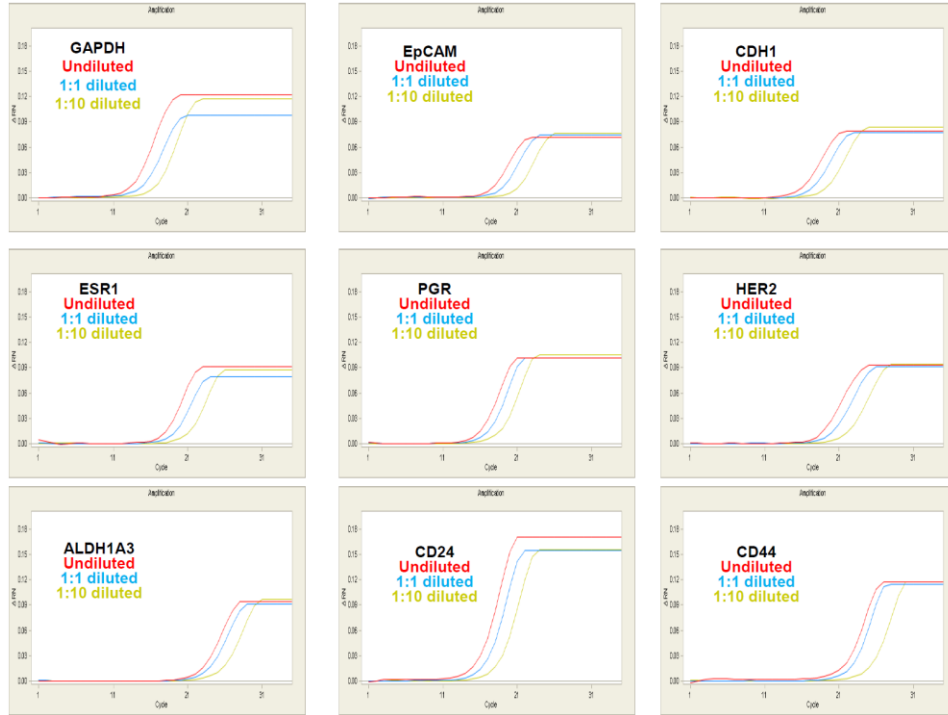
Supplementary Figure 15. Violin plot of the gene expression results of undiluted (ud), 1:1 diluted (1d), and 1:10 diluted (10d) total RNA extracted from T47D breast cancer cell line using BioMark HD system and TaqMan assays.



Supplementary Figure 16. Heatmap of the gene expression results of undiluted (ud), 1:1 diluted (1d), and 1:10 diluted (10d) total RNA extracted from T47D breast cancer cell line using BioMark HD system and TaqMan assays.



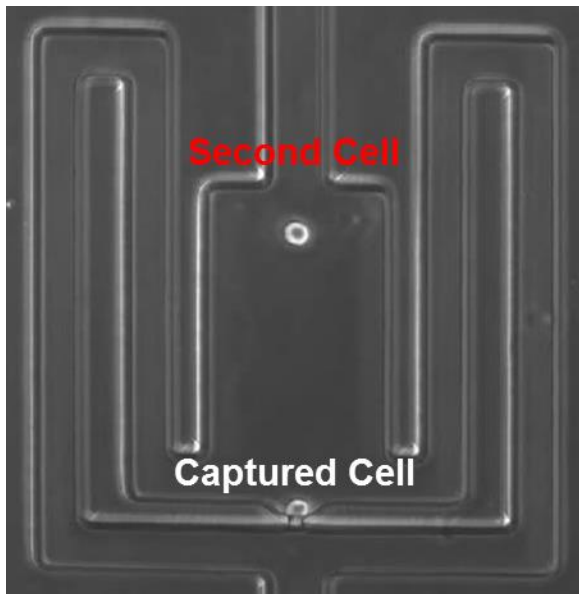
Supplementary Figure 17. PCA of the gene expression results of undiluted (ud), 1:1 diluted (1d), and 1:10 diluted (10d) total RNA extracted from T47D breast cancer cell line using BioMark HD system and TaqMan assays.



Supplementary Figure 18. Amplification plots of the critical genes of undiluted (ud), 1:1 diluted (1d), and 1:10 diluted (10d) total RNA extracted from T47D breast cancer cell line using BioMark HD system and TaqMan assays.



Supplementary Movie 1. A MDA-MB-231 cell was loaded into the chamber by gravity flow, and the first coming cell was captured at the capture site.



Supplementary Movie 2. A MDA-MB-231 cell was loaded into the chamber by gravity flow. As the capture site was occupied by a captured cell, and the next coming cell flow to downstream through serpentine path.