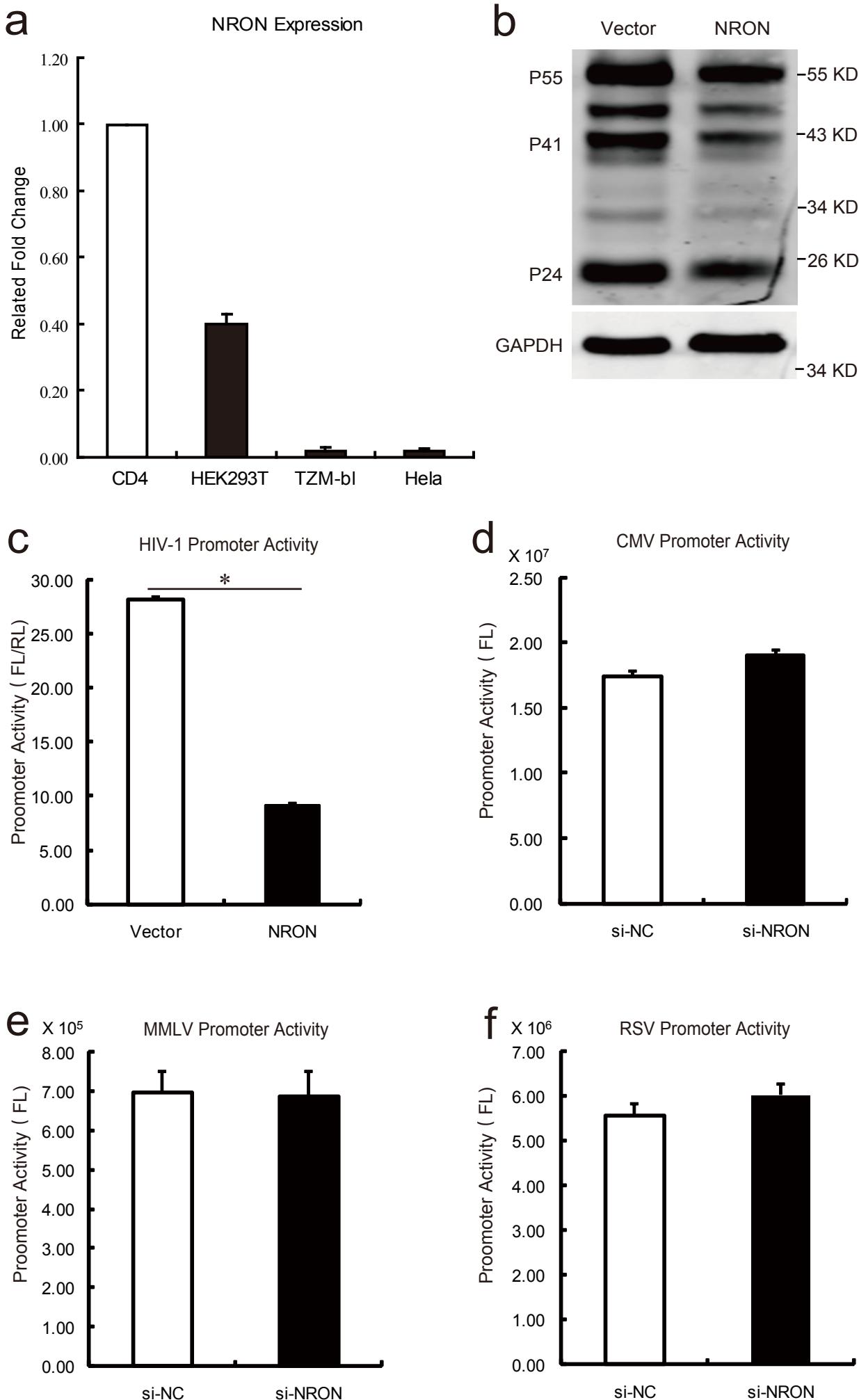


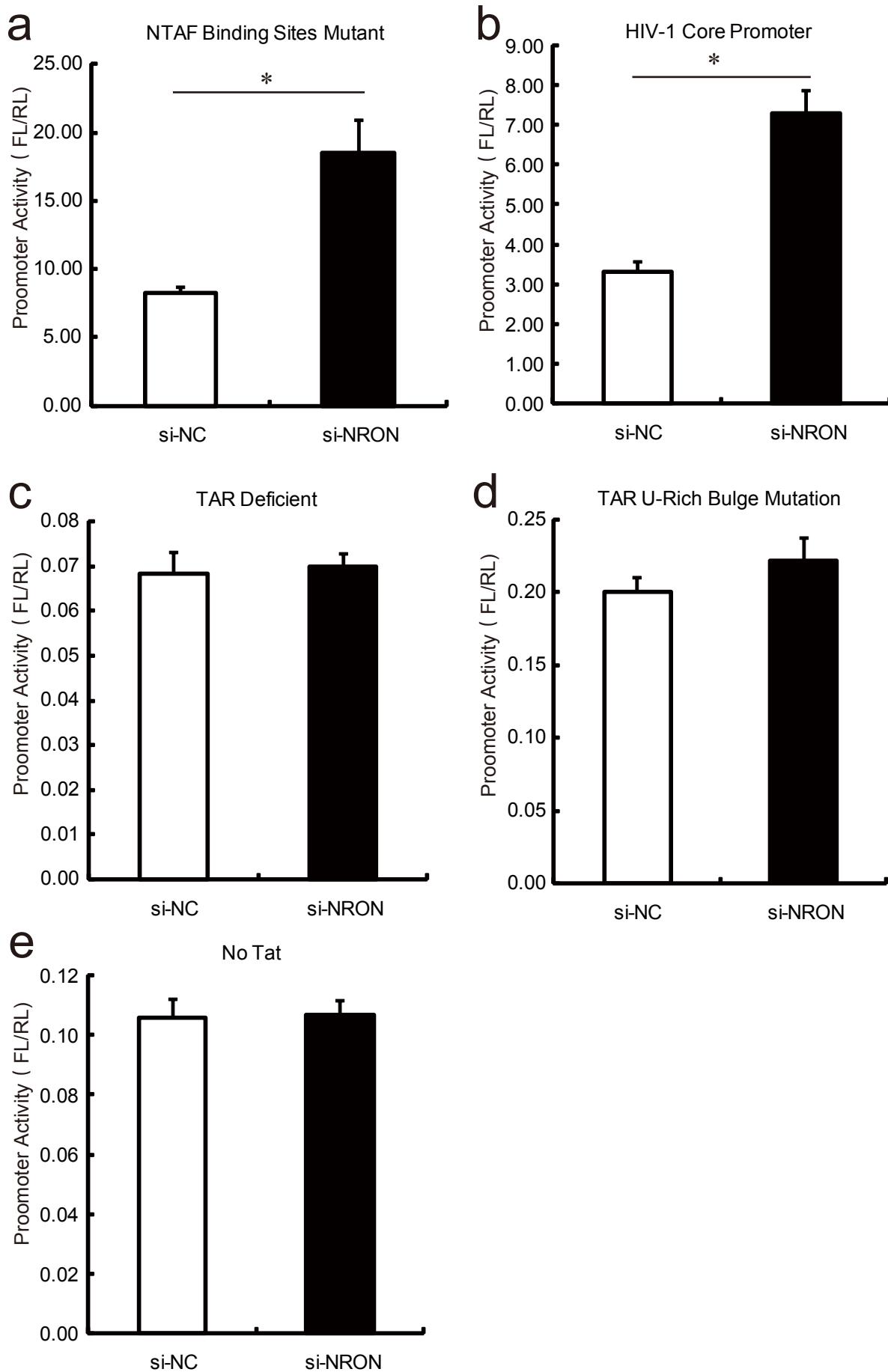
Supplementary Figure 1

Supplementary Figure 1 Knockdown of lncRNAs by siRNAs pools. The lncRNAs knockdown efficiency was detected by real-time qRT-PCR in the activated primary CD4+ T lymphocytes (n=3). The results show mean \pm S.D. (error bars).



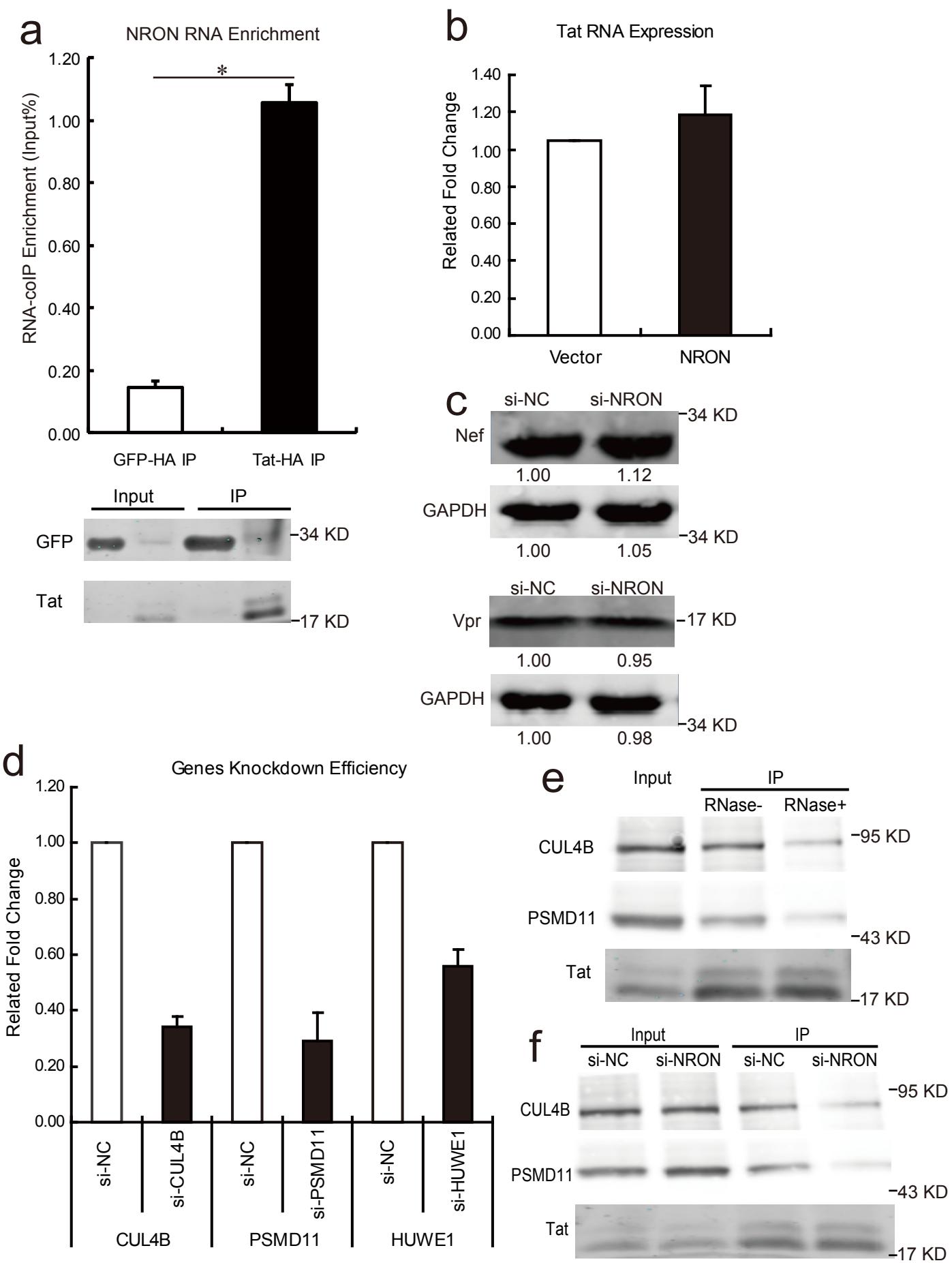
Supplementary Figure 2

Supplementary Figure 2 NRON specifically inhibits HIV-1 promoter activity. (a) NRON expression level in activated primary CD4⁺ T lymphocytes, HEK293T, TZM-bl and HeLa cells was detected by real-time qRT-PCR. The expression of NRON in activated primary CD4⁺ T lymphocytes was normalized as 1 (n=3). (b) HeLa cells were co-transfected with pNL4-3-deltaE-EGFP plasmids and NRON-expressing plasmid pcDNA3.1-NRON or empty vector control. Gag proteins expression was determined by western blotting with anti-p24 primary antibody at 72 h after transfection. (c) pcDNA3.1-Tat-HA, pRL-CMV and pcDNA3.1-NRON, or empty vector were co-transfected into TZM-bl cells, and at 48 h after transfection the integrated HIV-1 promoter activity was determined by dual-luciferase assay (n=3). (d-f) CMV, MMLV, or RSV promoter activities were examined upon NRON knockdown by luciferase reporter assay (n=3). Data in a, c-f show mean ± S.D. (error bars). Results in b represent three independent experiments. * P<0.05, Student's unpaired t test.



Supplementary Figure 3

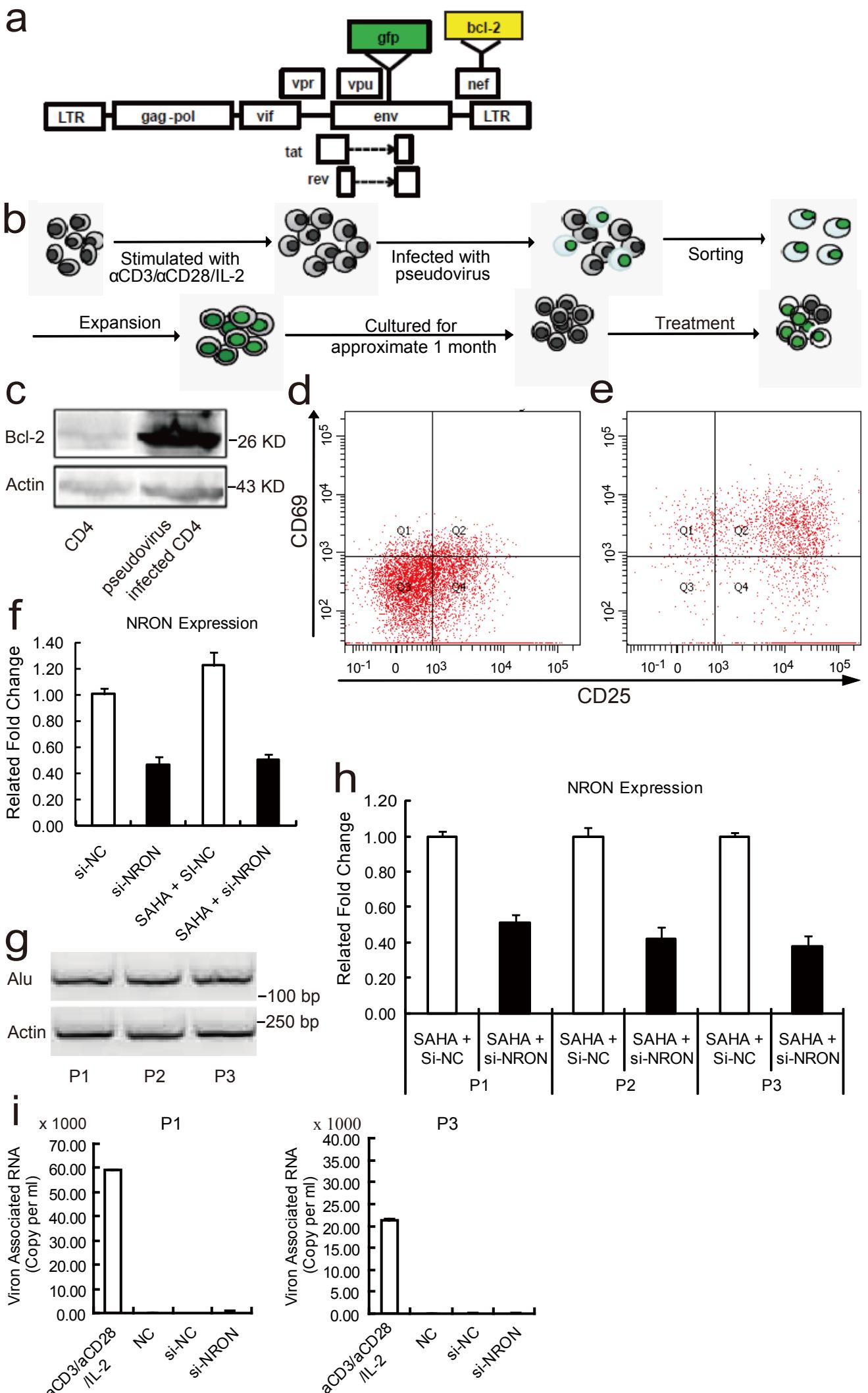
Supplementary Figure 3 NRON targets the HIV-1 Tat/TAR axis. The activities of HIV-1 NFAT binding sites mutated promoter (**a**) or core promoters (**b**) were determined by dual-luciferase reporter assay in HEK293T cells upon NRON depletion. The activities of TAR element deficient (**c**) or Tat binding U-rich bulge mutated (**d**) HIV-1 promoters were measured by dual-luciferase reporter assay. (**e**) HIV-1 promoter activity was determined upon NRON knockdown without co-transfection of the Tat-expressing plasmid pcDNA3.1-Tat-HA in HEK293T cells. The results show mean \pm S.D. (error bars) from triplicate samples. * $P<0.05$, Student's unpaired *t* test.



Supplementary Figure 4

Supplementary Figure 4 NRON recruits Tat to the ubiquitin/proteasome system. (a)

Endogenous RNA was co-immunoprecipitated by Tat in HEK293T cells, the enriched NRON RNA was determined by real-time qRT-PCR ($n=3$), and the precipitated proteins were detected by western blotting. **(b)** The pcDNA3.1-NRON or empty vector were co-transfected with pcDNA3.1-Tat-HA into TZM-bl cells, and Tat RNA level was detected by real-time qRT-PCR ($n=3$). **(c)** Nef and Vpr were detected by western blotting upon NRON knockdown in HEK293T cells. Numbers indicated the fold change related to control. **(d)** Knockdown efficiency of CUL4B, PSMD11, and HUWE1 was confirmed by real-time qRT-PCR ($n=3$). **(e)** The ectopic expressed CUL4B and PSMD11 in HEK293T cells were co-immunoprecipitated by Tat. The precipitates were washed with RNase-free or RNase A containing washing buffer and the enriched CUL4B or PSMD11 were detected by western blotting. **(f)** SiRNAs against NRON or non-specific control and pCDNA3.1-Tat-HA were co-transfected into Flag-tagged CUL4B or PSMD11-expressing HEK293T cells, and then CUL4B or PSMD11 were co-immunoprecipitated by Tat and detected by western blotting. Data in **a**, **b** and **d** show mean \pm S.D. (error bars). Results in **c**, **e** and **f** represent three independent experiments. * $P<0.05$, Student's unpaired *t* test.



Supplementary Figure 5

Supplementary Figure 5 HIV-1 latency models. (a) The *bcl-2* gene was inserted into the *nef* region of pNL4-3-deltaE-EGFP. (b) Schematic representation of generating the modified *in vitro* latency model with primary CD4⁺ T lymphocytes. (c) Western blotting detection of Bcl-2 in the activated human primary CD4⁺ T lymphocytes infected by pNL4-3-deltaE-EGFP-Bcl-2 pseudotyped viruses. After cultured for approximately 1 month (d), or then reactivated by anti-CD3 and anti-CD28 antibodies stimulation for 48 h (e), the status of the pseudotyped viruses infected primary CD4⁺ T lymphocytes were confirmed by FACS. NRON knockdown efficiency was detected in the primary CD4⁺ T lymphocytes *in vitro* latency model (f) and resting CD4⁺ T lymphocytes isolated from HIV-1 infected individuals (h). (g) Alu-PCR confirmation of the integrated HIV-1 provirus in the resting CD4⁺ T lymphocytes isolated from HIV-1 infected individuals receiving suppressive cART. (i) Resting CD4⁺ T lymphocytes isolated from HIV-1 infected individuals on suppressive cART were transfected with control or NRON-specific siRNAs. After 48 h, HIV-1 virion associated RNAs in the supernatants were isolated and detected by real-time qRT-PCR. Results in c, d, e and g represent three independent experiments. Data in f, h and i show mean ± S.D. (error bars).

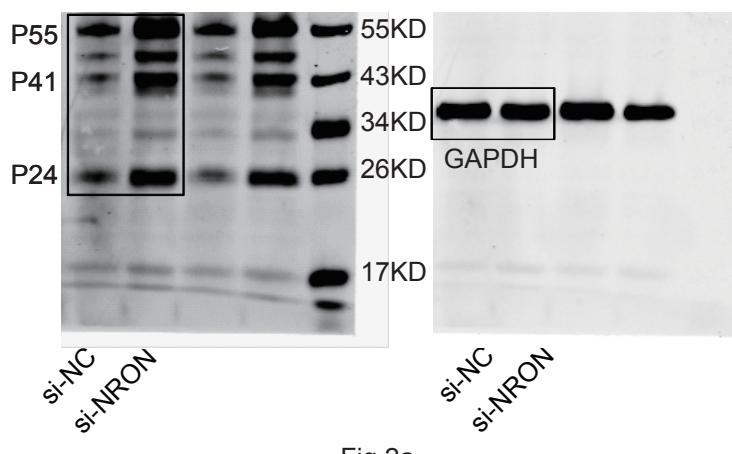
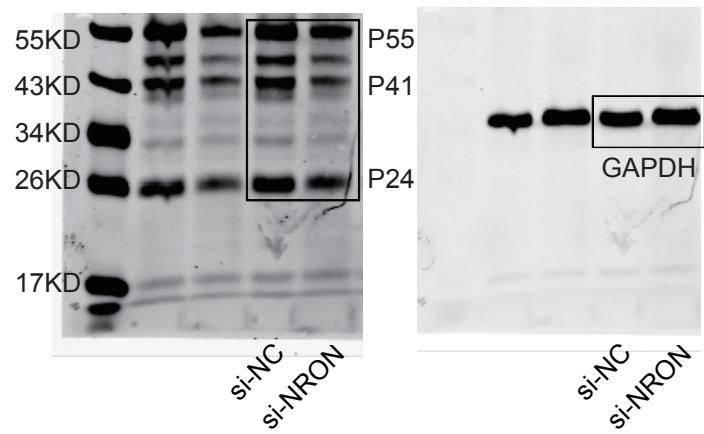


Fig 2a



Supplementary Fig 2b

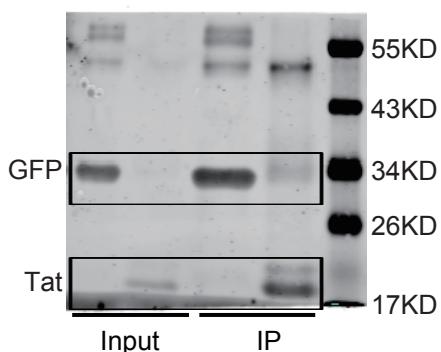
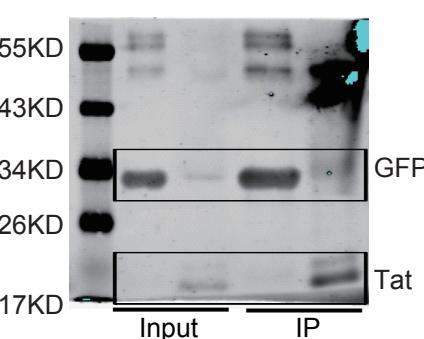


Fig 2d



Supplementary Fig 4a

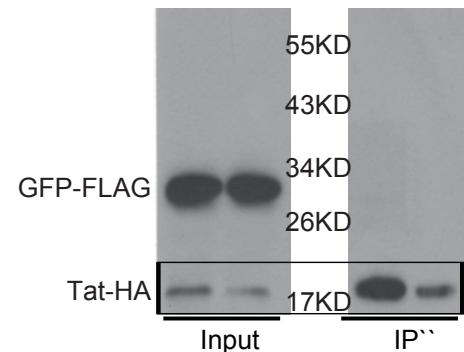


Fig 2e

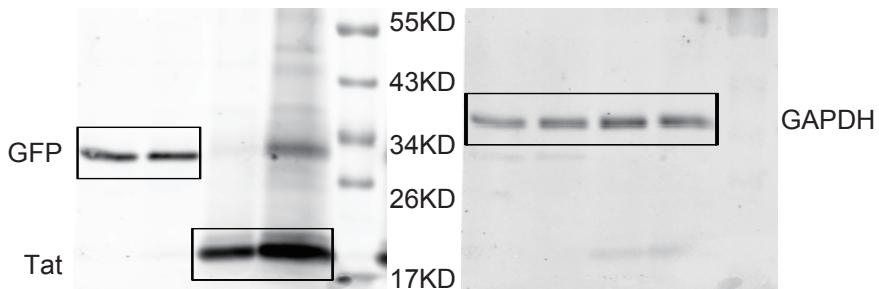


Fig 2f

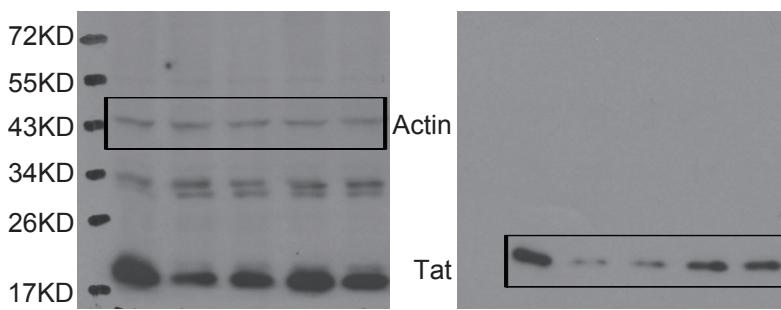
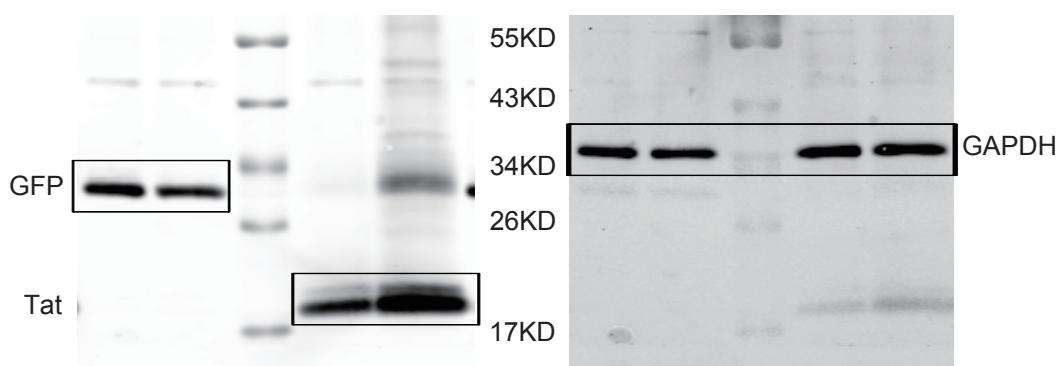
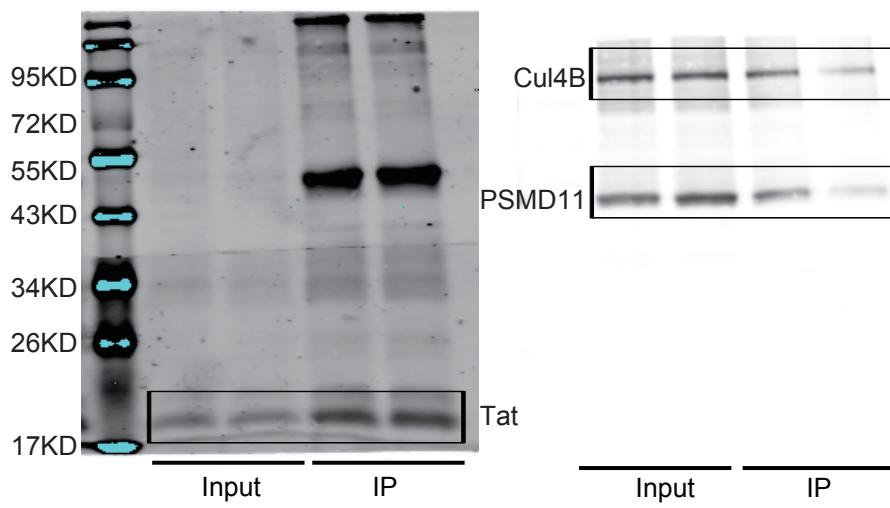
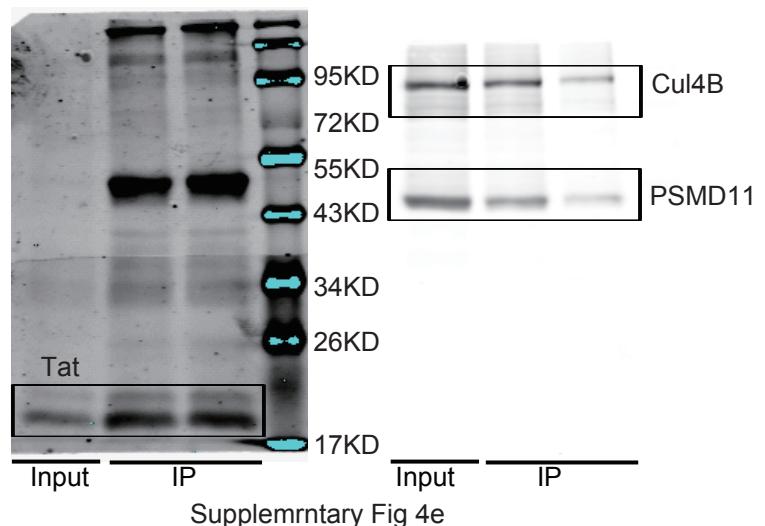
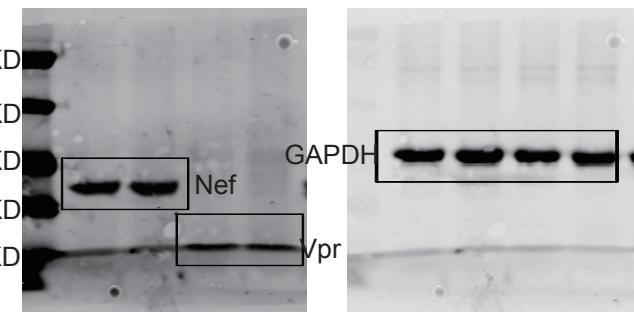


Fig 3b

Supplementary Figure 6



Supplementary Figure 6 Continued

Supplementary Figure 6 Uncropped blots scans used in the manuscript. Boxes indicate approximate portions of the films that are included in the figures. Below the blots it is indicated to which figure they correspond.

Supplementary Table 1

Table S1 Primer sets for constructs and primers for site direct mutations or deletions on HIV-1 Promoter (Red characters indicate the mutated sites)			
Constructs or mutants	Forword	Reverse	Description
pHIV-Pro-Luc	CGGGGTACCTGGAAGGGCTAATTGGTCCC	CGCGGATCCTGCTAGAGATTTCACACTGA	Full lenth of HIV-1 NL4-3 5' LTR sequence
pcDNA3.1-NR ON	AGCCCAAGCTTCACATCTCTAAACAAACCC AGC	CGGGGTACCGGAAAAAAATTCTCCTTAACATTTC	Full lenth of human NRON sequence
pHIV-Pro-mNF AT-1	GACTTGAGCTGGGACTTGAAGGGAGGTGTG GCCTGGCG	TCAAAGTCCCCAGCTCAAAGTCCCTGTAGAAAGCT	nt 351 to 380: 5'-GGACTTTGAGCTGGGG ACTTTGAAGGGAGG-3'
pHIV-Pro-mNF AT-2		CGCGGATCCTGCTAGAGATTAGAACACTGACTAAA AGGGTC	nt 607 to 634: 5'-TTTAGTCAGTGTCTAAATCTCTAGCA-3'
pHIV-Pro-Core	CGGGGTACCGGGAGGTGTGCCCTGGCGGGACT	CGCGGATCCGGGTTCCCTAGTTAGCCAGAGAGC	nt 374 to 513 of HIV-1 NL4-3 5' LTR sequence
pHIV-Pro-dTA R	GGGTCTCTCTGGTTAGTCTGGCTAACTAGGGAAC CC	TCCCTAGTTAGCCAGACTAACCAAGAGAGACCCAGTC A	Deletion of nt 470 to 492 of HIV-1 NL4-3 5' LTR sequence
pHIV-Pro-dTA R/mU	GGTTAGACCAGAACAGAGCCTGGAGCTCTCTGG C	AGAGCTCCCAGGCTCTGTTCTGGCTAACCAAGAGA GA	nt 466 to 489: 5'-TTAGACCAGAACAGAG CCTGGAG-3'
pMMLV-Pro-L uc	CGGGGTACCAATGAAAGACCCCCACCTGTAGG	CGCGGATCCAATGAAAGACCCCCGCTGACG	Full lenth of MMLV Promoter sequence
pRSV-Pro-Luc	CGGGGTACCTGCTCCCTGCTTGTGTGTTG	CGCGGATCCGGTGCACACCAATGTGGTGAA	Full lenth of RSV Promoter sequence
Tat exon 1	CGCGGATCCGCCACCATGGAGCCAGTAGATCCTA GACT	GGGATTGGGAGGTGGGTTGCTTGATAGAGAAACTTGATG	

Tat exon 2	TTCTCTATCAAAGCAACCCACCTCCAATCCGAG GGGAC	CCGCTCGAGCTAGGCGTAGTCGGCACGTCTAGG GGTAATCGAATGGATCTGTCTCTG	With HA epitope sequence tagged at the C' terminus
pcDNA3.1-CU L4B-FLAG	CGCGGATCCGCCACCATGATGTCACAGTCATCTGGAT	CCGCTCGAGTTACTTGTCATCGTCATCCTTGTAAATCT CCACCTGCAATATAAGTTGACTGGTTGGAT	Full lenth of human CUL4B cDNA sequence with FLAG epitope sequence tagged at the C' terminus
pcDNA3.1-PS MD11-FLAG	CGCGGATCCGCCACCATGGCTGCAGCGCGGTGGTGGAGTT	CCGCTCGAGTTACTTGTCATCGTCATCCTTGTAAATCT CCACCTGTCAGTTCTGGCTTGTGTTGAG	Full lenth of human PSMD11 cDNA sequence with FLAG epitope sequence tagged at the C' terminus
pFLAG-Ub	CGGAATTGCCACCATGGATTACAAGGATGACGA TGACAAGATGCAGATCTCGTAAAAACCC	TGCTCTAGATAACCACCTCTCAGACGCAGG	Full lenth of human ubiquitin cDNA sequence with FLAG epitope sequence tagged at the C' terminus

Supplementary Table 2

Table S2 Target Sequences of siRNAs agianst lncRNAs used in the study

lncRNA	Target Sequence 1	Target Sequence 2	Target Sequence 3
NRON	CTGTTCCACTACTGCTCC	ATCATTAGGCTAATAACGC	ATACTGTTCCACTACTGC
AC004840	GCACCTTGGAGATGTCATT	GGACAGAGTGCAAGGACAT	
SNHG1	GCCAGCACCTCTCTCTAA	GAGGACATCAGAACGGTCAA	
linc00630	GCTTCTACTCCTGACTTAA	GCGGTATACAACCACACTA	
TUG1	CCTCAAGCCAAATAGCTAA	GCCTCTATTCTGTATGTA	

Supplementary Table 3

Table S3 Primer sets for real-time RT-PCR used in the study

Gene or lncRNA name	Forward	Reverse
Beta-actin	GCATGGAGTCCTGTGGCA	CAGGAGGAGCAATGATCTTGA
GAPDH	CGCTCTCTGCTCCTCCTGTT	CCATGGTGTCTGAGCGATGT
NRON	ACGTTCTTAATGTACGCCTTGC	TTGCCGTGTCCTGAGTCCTT
LINC00489	CCTGCTCGATGCACCTCT	CACAGTCCCTGCCGTCTT
AC004840	AGACCCCATAACAGCAGCGACCA	CACGCTGACCCGAAATCCTCCA
RP11-86H7	TGCCTCATTATCCCCGGATGCCT	TGACACTCCCAGGACTTGACAAAGC
RP11-62I21	ATGGAACAGAGGTGGCCTACGGTC	GAACCGTCATGGCTGCTTGAGGA
SNHG6	GAGCCGTTAGTCATGCCGGTGT	GCTCAATACATGCCCGTGATCCT
SNHG1	TGCCTCCCAGGGTGAATAACAGGT	AGAGAGAAGGTGCTGCCCTTG
SCAND2	CAGAAAACGTCCAGTGCAGGTGGG	GTGCCACCTGCTCTCAAGTCCTG
RP11-67K19	CCCCAGAACATGCTGGAGGCAA	AGCAGGCCATTGTCAGTGAGAGA
LINC00900	GGGAAGTGCCAGGATGGGTATCA	TCAACCTGGTTCACTCCAGGCGAA
LINC00502	AGTAGCAGGCTTGGGACGTTTC	TGGTGCTGTCACTTCACACTGCTT
RP11-418J17	ACACTGCCCATCTGAATGCC	ACACCAGTCACCTTAGCGGCAC
LINC00426	TTCTGCAGCCTCCCTGTTAGCCA	AACTGTTGCGACATTGTTGCCGG
LINC00630	TAACCCAGTGCCTCGTATGGTGCC	GGAATCAGGCATCACATCCCACCA
AP000402	ATCCAGGCCTGCTTGGATGTCTT	AAGGGCGTGTACAATCAAGGCAGT
LINC00491	TCAGCCCCATACGAGGTTGCTT	AGACCTTGGCCTCTTTGGGGT
NEAT1	CAGAGCCCAGAACATGCCAGCAG	ACGCACAAGAAGGCAGGCAAACA
THAP7-AS	GTAGTAAGCGGCGCGAGCGTG	CATCGTGCCGGGGTGTACCGG
Gas5	TCTGCCTCACCAAGCTAGAG	TTGTGCCATGAGACTCCATCAG
TUG1	TAGCAGTTCCCCAATCCTTG	CACAAATCCCCATCATTCCC
T-bet	TGGTTGGGAAAGTGGGCTCAA	TGCTCCCAGTCCCCACACCTGA
Firefly Luciferase	CAACACCCCAACATCTCGACGC	GGACTTCCGCCCTTGGCCTT

HIV-1 total mRNA (Nef-3'LTR)	GGTGGGTTTCCAGTCACACCTCAG	GCACCATCAAAGGTCACTGG
HIV-1 Tat	CGCGGATCCGCCACCATGGAGCCAGTAGATCCTAGACT	GGGATTGGGAGGTGGGTTGCTTGATAGAGAAACTTGATG
CUL4B	CCCCTCCCCGAGGCTCTTAATTCT	CCACGTACACAGGGAAAAGAGCACA
PSMD11	GGGCTTCCAAGCAAGTCCAGACA	ACTGGACCATGCTGCCCTGGAA
HUWE1	GCCCTCGAAGGCATGAATGGCA	GCCAACAGTAGCATGTGGCGGA