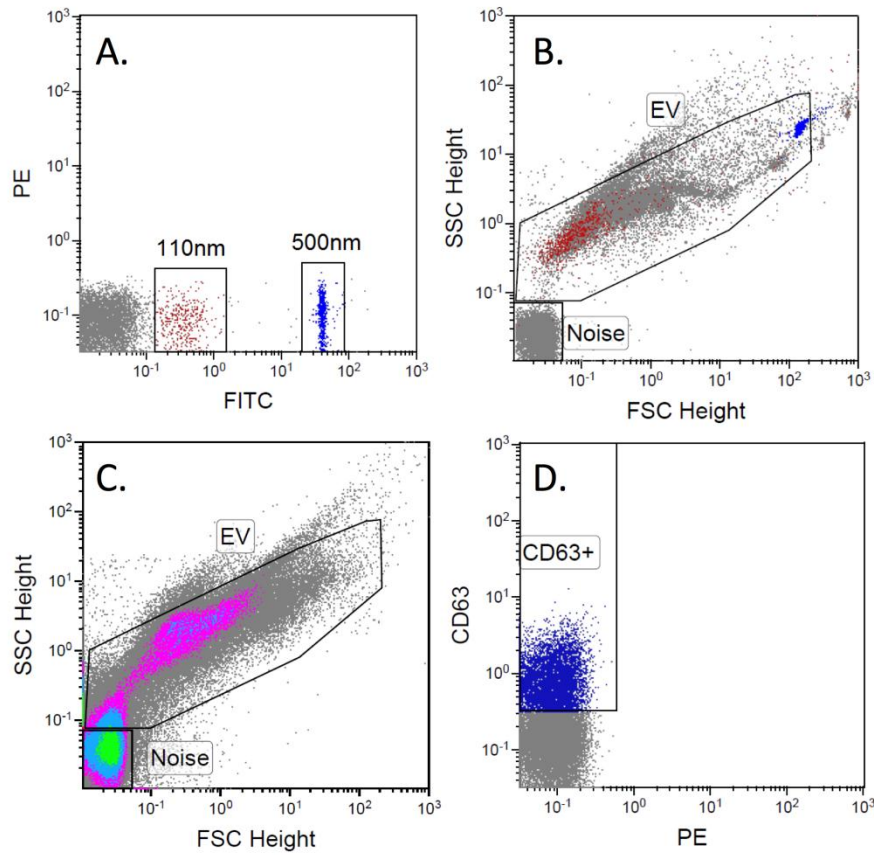
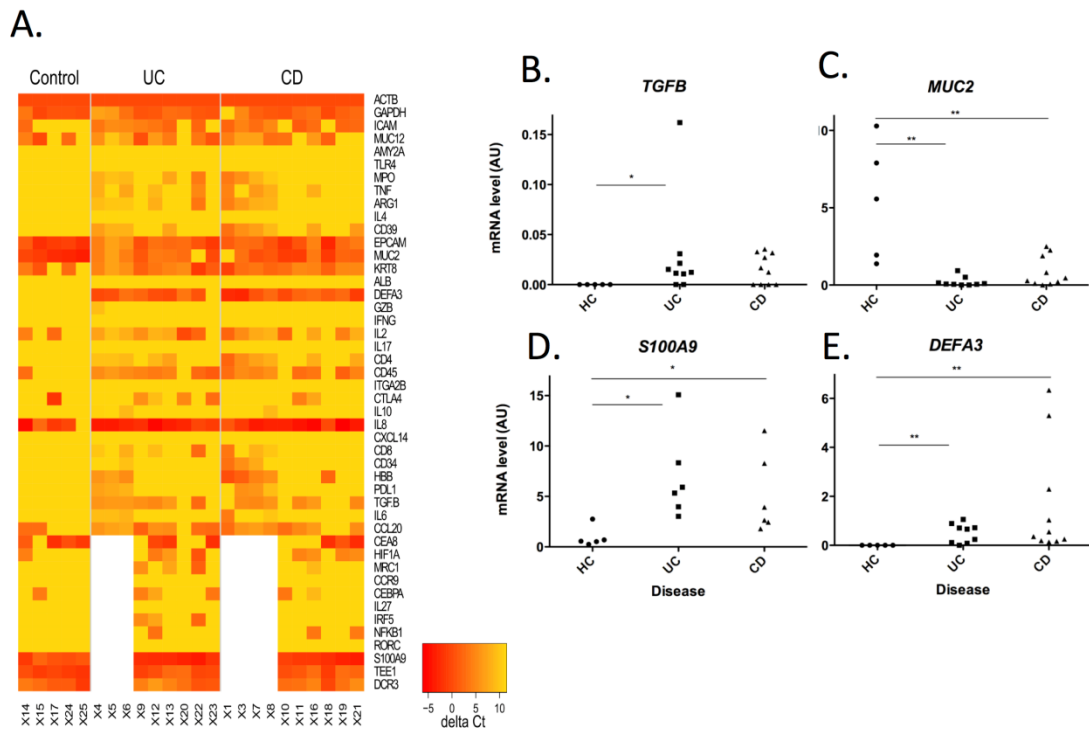


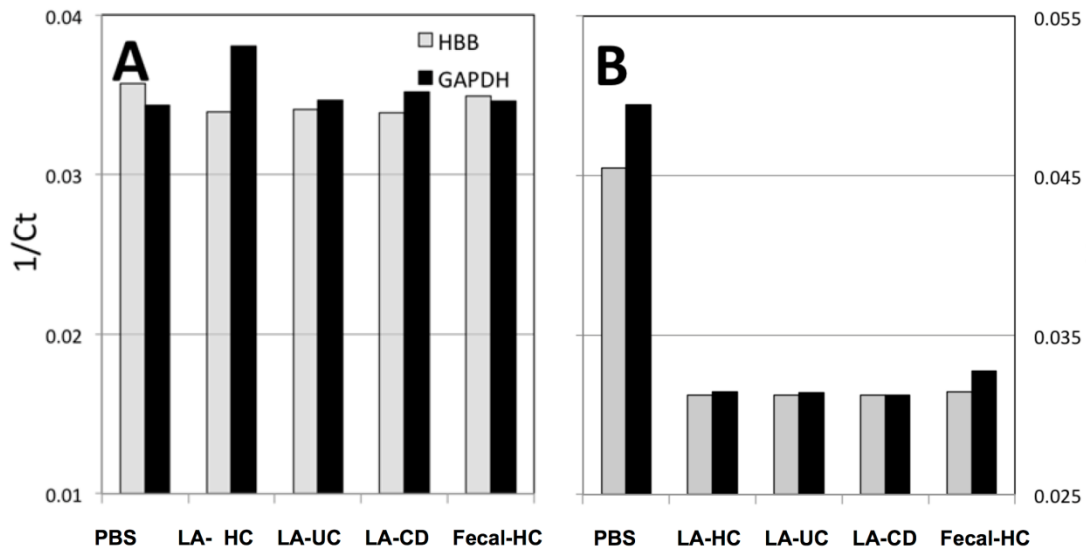
Supplementary Figure 1. Flow cytometry of the cellular fraction of luminal aspirates. Luminal aspirates were centrifuged to isolate the cellular components, and labelled with anti-CD45 and anti-14 antibodies to identify neutrophils and monocytes. **(A)** All cells / ungated. **(B)** CD45+ cells from outlined gate in **(A)**. CD14 high/positive was labeled as monocytes and CD14 low/negative was labeled as neutrophils.



Supplementary Figure 2. Particle-size-based gating strategy. The *ApogeeMix* was subjected to flow cytometric analysis. The EV gate was designed to include both population of fluorescent beads (110nm and 500nm) and the area that are most likely show particles smaller than 500nm (A & B). (C) Representative colonic EV sample from IBD patients (41%). (D) CD63 positive gate was created based on the same sample single stained with anti-CD63 (18.4%).



Supplementary Figure. 3. Levels of EV mRNA in colonic samples from IBD patients and controls. **(A)** Heatmap of mRNA, normalized by ACTB, was compared among healthy control (HC), UC, and CD; yellow indicates lower expression and red indicates higher expression. **(B-E)** Scatter plots of mRNA levels (Arbitrary Unit, AU) in samples from healthy controls (HC, N=5), UC (N=9), and CD (N=10) patients, as assessed by RT-PCR.



Supplementary Figure 4. Stability of EV mRNA in LA and stool. **(A)** PBS, LA or fecal supernatants were incubated with fetal calf serum-derived purified standard EV at 37⁰C for 30 minutes, then bovine Hemoglobin (HBB) and GAPDH mRNA levels (1/Ct) were quantified as described in the Methods. **(B)** Naked rat total RNA was incubated with PBS, LA, or fecal supernatants at 37⁰C for 30 minutes, then rat ACTB *and* GAPDH mRNA levels (1/Ct) were quantified.

Supplementary Table 1.

Primer sequences used for PCR

Gene	Forward (5'-3')	Reverse (5'-3')
Human		
<i>ACTB</i>	CCTGGCAGCCAGCACAAAT	GCCGATCCACACGGAGTACT
<i>GAPDH</i>	CCCACTCCTCCACCTTTGAC	CATACCAGGAAATGAGCTTGACAA
<i>MUC2</i>	GCGGGACATTTGTTCATGTACTC	GATGTGGGTGTAGGTGTGTGTCA
<i>MUC12</i>	GCCTTGAGAACGCCTACAACA	GAGCTCTGTGCCAGAGTCAACA
<i>ICAM</i>	TCCCCCGGTATGAGATTG	GCCTGCAGTGCCATTATG
<i>EPCAM</i>	CAGTTGTTGCTGGAATTGTTGTG	CATCTCACCCATCTCCTTTATCTCA
<i>KRT8</i>	CTGGGATGCAGAACATGAGTATTC	GCTTGTGAGGCCCCCATAG
<i>EGFR</i>	CCTTGCCGCAAAGTGTGTAA	TGAAGGAGTCACCCCTAAATGC
<i>CEACEM8</i>	AGTGCAGTGGCAGCATCTCA	AAATTAGCCGGGCGTTGTG
<i>MMP7</i>	CGGATGGTAGCAGTCTAGGGATT	GGAATGTCCCATACCCAAAGAA
<i>TFF1</i>	CCCCAGCACGGTGATTAGTC	CAGAGCAGTCAATCTGTGTTGTGA
<i>ALB</i>	TGCAAGGCTGACGATAAGGA	GTAGGCTGAGATGCTTTTAAATGTGA
<i>AMY2A</i>	ACAGAGGATTCATTGTTTTCAACAAT	ACAGTATGTGCCAGCAGGAAGA
<i>IL2</i>	GAACTAAAGGGATCTGAAACAACATTC	TGTTGAGATGATGCTTTGACAAAA
<i>IL4</i>	CACAGGCACAAGCAGCTGAT	CCTTCACAGGACAGGAATTCAG
<i>IL17</i>	CATGAACTCTGTCCCCATCCA	TCCAGCCGGAAGGAGTTG
<i>IL27</i>	CCAAGGCTGGGCACTCAGT	GATGCCAAGACTCCAGTCCATAA
<i>TNF</i>	CGAAGGCTCCAAAGAAGACAGT	CAGGGCAATGATCCCAAAGT
<i>TNFSF15</i>	TGCGAAGTAGGTAGCAACTGGTT	CCATTAGCTTGTCCCCTTCTTG
<i>DCR3</i>	CAATGTGCCAGGCTCTTCCT	TCACACTCCTCAGCTCCTGGTA
<i>GZB</i>	GCGGTGGCTTCTTGATACAA	CCAAGGTGACATTTATGGAGCTT
<i>MPO</i>	ACTGCCTGGGTTCCAATCC	TGTTTAAGGAGGGTAATTTGCTCAA
<i>DEFA3</i>	CCAGGCTCAAGGAAAAACATG	CTGGTAGATGCAGGTTCCATAGC
<i>TLR4</i>	GAAGAGTGAGTGGTGAAGTATGAA	ATGGCAGCATCATTGTTCTCATC
<i>IFNG</i>	GGAGACCATCAAGGAAGACATGA	GCTTTGCGTTGGACATTCAA
<i>IRF5</i>	CCCCAGAGCTGGTTGTTAA	CTGGAGTGTGCAGAGATGACACA
<i>IL10</i>	GCCATGAGTGAGTTTGACATCTTC	GATTTTGAGACCTCTAATTTATGTCCTA
<i>CTLA4</i>	CACTGAGGTCCGGGTGACA	GTAGGTTGCCGCACAGACTTC
<i>PDL1</i>	TCCAAGAGAGAGGAGAAGCTTTTC	GCTGTATGGTTTTCTCAGGATCT
<i>ARG1</i>	AGACACCAGAAGAAGTAACTCGAACA	TCCCAGCAAGTCCGAAAC
<i>TGFB1</i>	CTGCTGAGGCTCAAGTTAAAAGTG	TGAGGTATCGCCAGGAATTGT
<i>HBB</i>	GCCCATCACTTTGGCAAAGA	CCAGCCACCACTTCTGATAGG
<i>CD4</i>	AAATGCCACACGGCTCTCA	GGGTGCTGTGCTTCTGTGAAC
<i>CD8</i>	CCGAGAGAACGAGGGCTACTATT	GCACGAAGTGGCTGAAGTACAT
<i>CD34</i>	CAGGGAAAGGCCAGTGTGAA	ACCACGTGTTGTCTTGCTGAAT

<i>CD45</i>	AAGCTCCCTGAAGCAAAGGAA	GCAGGACCATTGACAGAATGTTC
<i>ITGA2B</i>	TGCTGCTGCTCACCATCCT	CCGGTTCCGCTTGAAGAAG
<i>IL6</i>	TCATCACTGGTCTTTTGGAGTTTG	TCTGCACAGCTCTGGCTTGT
<i>IL8</i>	TGCTAAAGAACTTAGATGTCAGTGCAT	TGGTCCACTCTCAATCACTCTCA
<i>CCL20</i>	GATACACAGACCGTATTCTTCATCCTAA	TGAAAGATGATAGCATTGATGTCACA
<i>CXCL14</i>	AAGCTGGAAATGAAGCCAAAGT	ACACGCTCTTGGTGGTGATG
<i>CCR9</i>	GGATTCCGCTTATTCCCTTGGT	TGCGAACCCAGCTGTTATAATCT
<i>CD39</i>	CCCAGCTGAGCAACCATTGT	GACCAGGGAGAATAGAACCATGA
<i>MRC1</i>	AAAGCTGACACAAGGAAGATGGA	TCAGGAGGATCACAATGATGACTAC
<i>CEBPA</i>	TTGTACTGTATGCCTTCAGCATTG	TCGGCTGATAAAGCAAAATATTTG
<i>NFKB1</i>	GGATCACAGCTGCTTTCTGTGTG	CGACCGTGATACCTTTAATGACAA
<i>HIF1A</i>	TTGCCAGCTCAAAAGAAAACAA	ACCAACAGGGTAGGCAGAACA
<i>R2RX7</i>	CTGTGAGGAAGCCCAAGCA	CAGAATCACCTGAAGGCTTCTTCT
<i>RORC</i>	CGGAAGGCAAGATCAGATCCT	CCCTGGTGTCCCTCCATGCT
<i>S100A8</i>	GGCCAAGCCTAACCGCTATAA	CTTCTGAAAGACAGCTGACAAGAGA
<i>S100A9</i>	CTGAGCTTCGAGGAGTTCATCA	CGTCACCCTCGTGCATCTT
<i>SOCS3</i>	GCTAAGAGATTGCCTTAAATGCT	CTTGGTGCCCTCCAGTGAGT
Bovine		
<i>HBB</i>	GGCATGAAGCATCTCGATGA	CAGCTTATCACAGTGCAGCTCACT
<i>GAPDH</i>	AGGTTGTCTCCTGCGACTTCA	GTGGTCGTTGAGGGCAATG
Rat		
<i>actb</i>	TCTGTGTGGATTGGTGGCTCTA	CTGCTTGCTGATCCACATCTG
<i>gapdh</i>	ACCAGGTTGTCTCCTGTGACTTC	CAGGAAATGAGCTTCACAAAGTTG

Supplementary Table 2.

Characteristics of IBD patients from who samples were obtained

Factor	N=63
Mean age (years)	48
Gender (% male)	60
Ulcerative colitis (%)	62
Crohn's disease (%)	38
Disease duration (years)	12
Mean CRP	8.3
Mean HBI / SCCAI score	3
Mean Mayo endoscopy score	1