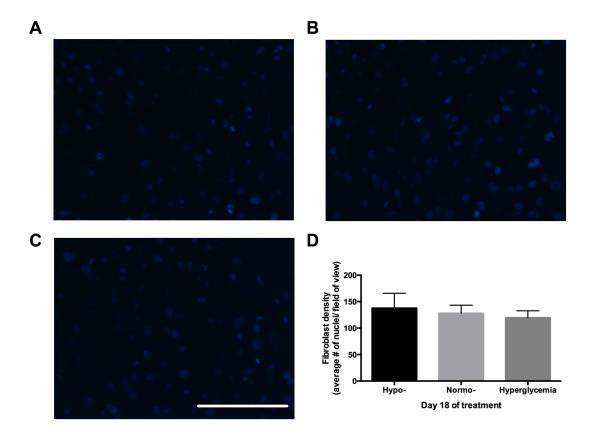
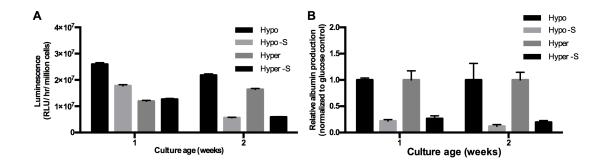
SUPPLEMENTAL INFORMATION

Long-term exposure to abnormal glucose levels alters drug metabolism pathways and insulin sensitivity in primary human hepatocytes

Matthew D. Davidson, Kimberly R. Ballinger, and Salman R. Khetani, PhD



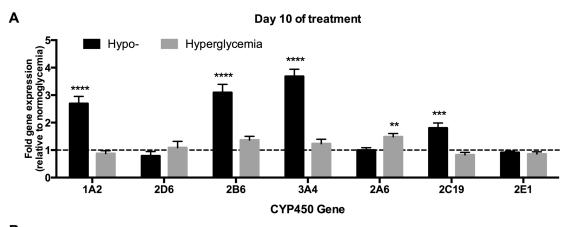
Supplemental Figure 1. Effects of glucose concentration on fibroblast numbers in MPCCs. MPCCs were fixed after 18 days of treatment with various glucose concentrations and stained with DAPI for cell counting. Images of 3T3-J2 fibroblasts between hepatocyte islands cultured in hypo- (0.4- 0.5 mM) (**A**), normo- (5 mM) (**B**) and hyperglycemic (25 mM) (**C**) culture medium on day 18 of treatment. Scale bar is 200 μm. (**D**) Average number of cells per field of view quantified using ImageJ software. Error bars represent SD.

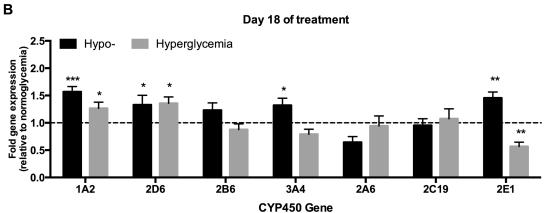


Supplemental Figure 2. Serum removal decreases hepatocyte functions in MPCCs.

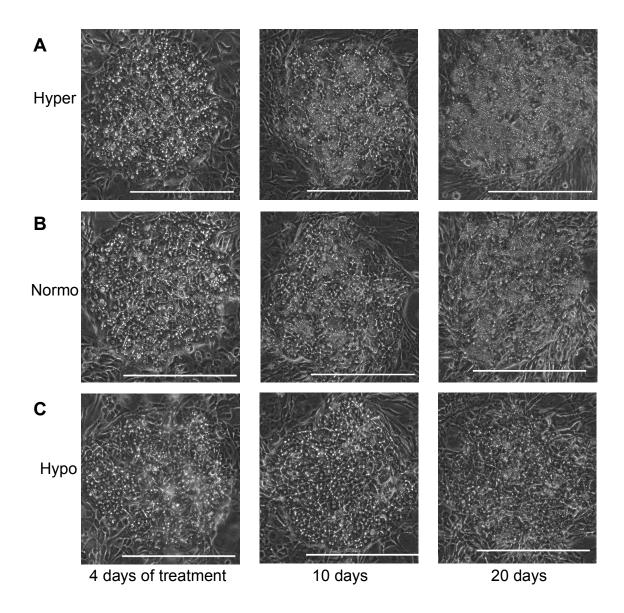
MPCCs were established for 4 days and then serum was removed from culture media.

(A) CYP3A4 activity measured with Luciferin-IPA in hypo- and hyperglycemic medium with or without 10% (vol/vol) serum (-S). (B) Relative albumin production in hypo- and hyperglycemic medium with or without serum (-S). Data is normalized to the serum-containing control for each glucose level. Error bars represent SD across 3 wells. Similar results were observed in another primary hepatocyte donor.

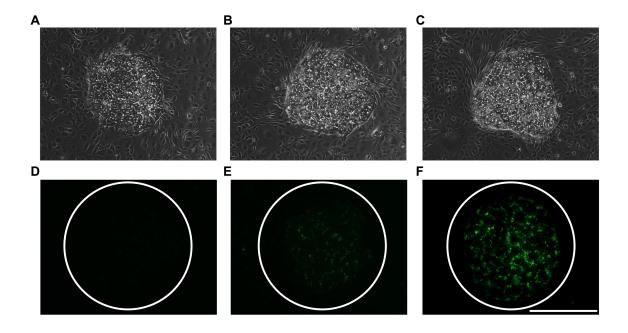




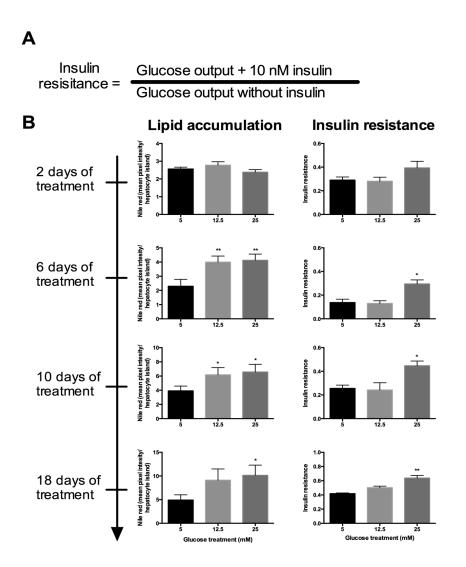
Supplemental Figure 3. Glucose-induced modulation of CYP450 gene expression in MPCCs. Levels of CYP1A2 (1A2), CYP2D6 (2D6), CYP2B6 (2B6), CYP3A4 (3A4), CYP2A6 (2A6), CYP2C19 (2C19), and CYP2E1 (2E1) mRNA transcripts in MPCCs treated with hypo- and hyperglycemic culture media for 10 days (**A**) and 18 days (**B**). Data is normalized to the normoglycemic control. Error bars represent SD. * p≤ 0.05, ** p≤ 0.01, *** p≤0.001, and **** p≤0.0001.



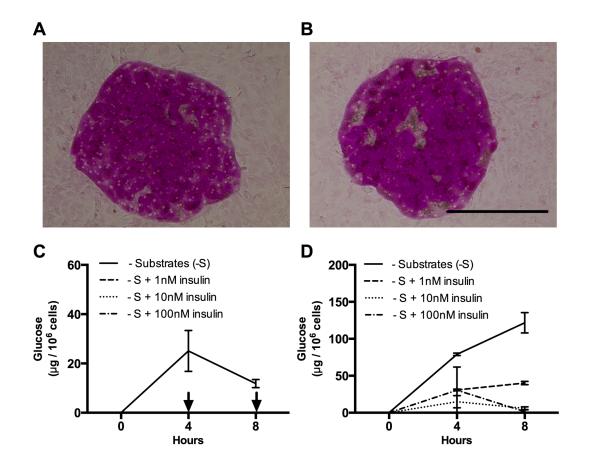
Supplemental Figure 4. Glucose-induced changes in hepatocyte morphology in MPCCs. Phase contrast image of PHHs in MPCCs treated over time with a hyper- (A), normo- (B), and hypoglycemic (C) culture medium. Scale bars represent 400 μm.



Supplemental Figure 5. Glucose-induced accumulation of neutral lipids in MPCCs after 10 days of treatment. Phase contrast images of MPCCs in hypo- ($\bf A$), normo- ($\bf B$) and hyperglycemic ($\bf C$) culture medium after 10 days of treatment. Nile red (neutral lipids) staining of MPCCs in hypo- ($\bf D$), normo- ($\bf E$) and hyperglycemic ($\bf F$) culture medium after 10 days of treatment. Circles highlight hepatocyte island location and scale bar is 400 μ m.



Supplemental Figure 6. Lipid accumulation and insulin resistance in MPCCs treated with varying glucose levels. (A) Insulin resistance is calculated by dividing the MPCC glucose output under insulin stimulation (10 nM) by the glucose output without insulin ("0" means complete inhibition of glucose output by insulin, while "1" means no effect of insulin on glucose output). (B) Time course of lipid accumulation, assessed via Nile Red fluorescence quantification (9 hepatocyte islands quantified per treatment), in MPCCs continuously treated with culture media containing 5 mM (normoglycemic), 12.5 mM or 25 mM (hyperglycemic) glucose. (C) Insulin resistance development over time in MPCCs treated as in panel (B). Error bars represent SD. * $p \le 0.05$, and ** $p \le 0.01$.



Supplemental Figure 7. Glucose-induced modulation of insulin sensitivity in

MPCCs. PAS (glycogen) staining of MPCCs treated with either normo- (A) or hyperglycemic (B) culture medium for 10 days *prior* to assessing glucose output in the supernatants as in the subsequent panels of this figure. Scale bar is 400 μm. (C) Glucose output over time in supernatants of normoglycemic MPCCs (10 days of treatment) treated *without* any gluconeogenic substrates in the presence or absence of different levels of insulin (see methods for additional details). Arrows indicate no detectable glucose in supernatants. (D) Glucose output as in panel (C) except hyperglycemic MPCCs were used. Error bars represent SD across 3 wells. Similar results were observed in another primary hepatocyte donor.

SUPPLEMENTAL TABLE 1: Sequences for the GE Healthcare Dharmacon Solaris™ brand primer/probe sets used in this study.

Gene: GAPDH

Forward Primer: GCCTCAAGATCATCAGCAATG

Reverse Primer: CTTCCACGATACCAAAGTTGTC

Probe: GCCAAGGTCATCCATGA

Gene: ARG1

Forward Primer: ACTTGCATGGACAACCTGT

Reverse Primer: ATCCTGGCACATCGGGAAT

Probe: GAACTAAAAGGAAAGATTC

Gene: ALB

Forward Primer: AATGTTGCCAAGCTGCTGA

Reverse Primer: TCATCCCGAACTTCATC

Probe: CTGTTGCCAAAGCTCGATG

Gene: AHR

Forward Primer: CGTCTAAGGTGTCTGCTGGATA

Reverse Primer: CCCTTGGAAATTCATTGC

Probe: TCATCTGGTTTTCTGGCAA

Gene: CYP3A4

Forward Primer: CCTCATCCCAATTCTTGAAG

Reverse Primer: GCTGAAGGAAATCCACTC

Probe: GAAGATACACAAAAGCACCG

Gene: CYP2A6

Forward Primer: GATGACCACGTTGAACCT

Reverse Primer: ATGGACCTTGGCCTCCAC

Probe: ATGAAGCACCCAGAGGT

Gene: MLXIPL

Forward Primer: GTTTGATGACTACGTCCGAAC

Reverse Primer: GCCGGATGAGGATGCTG

Probe: AAGTTCTGGGTGTTCAG

Gene: HMOX1

Forward Primer: ATTGCCAGTGCCACCAAGTTC

Reverse Primer: CTCCTCAAAGAGCTGGATGT

Probe: AGACTGCGTTCCTGCTCAA

Gene: G6PC

Forward Primer: CCTACAGATTTCGGTGCTTG

Reverse Primer: TTCGTGACAGACAGACATTCAG

Probe: TTCTGGGCTGTGCAGCT

Gene: PCK1

Forward Primer: TCGAGAATAACGCTGAGCTGTG

Reverse Primer: TGAGAGCCAACCAGCAGTT

Probe: CTGAAGAAGTATGACAAC

Gene: SLC2A2

Forward Primer: GCTCAACTAATCACCATGCTCTG

Reverse Primer: GCTACTAACATGGCTTTGATTCTTC

Probe: CTTGGGGACACACTTGG