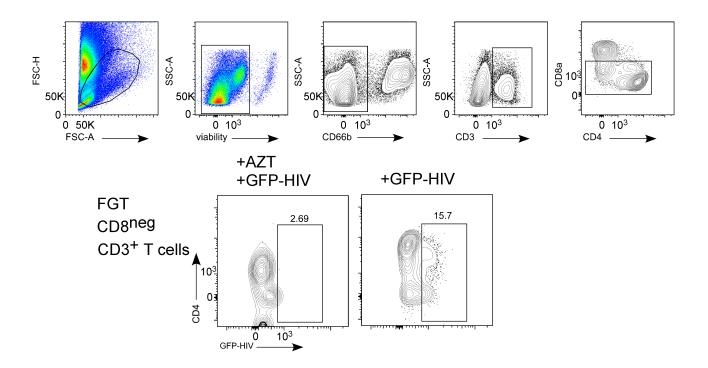
Swaims-Kohlmeier et al Supplementary Data:

Supplementary Table I

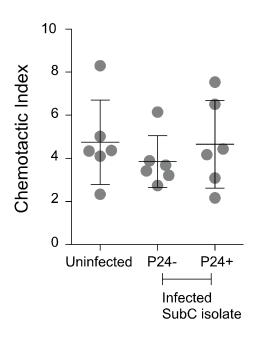
Supplementary Table I. Characteristics of CVL samples from subjects

Subject ID	Age	Race	CVL T cell number	% CD4	% CD8
1	34	AA	380	63	20
2	39	AA	678	47	44
3	24	AA	636	39	32
4	33	AA	240	49	27
5	37	AA	341	62	31
6	41	AA	591	41	50
7	22	AA	665	60	28
8	32	AA	1172	46	55
9	35	AA	295	64	23
10	31	C	827	29	45
11	38	A	8859	34	40
12	19	C	405	67	23
13	42	C	1217	65	25
14	16	AA	644	49	15
15	32	AA	220	38	44
16	23	C	2094	29	33
17	36	AA	2208	49	34
18	27	C	365	20	33
19	20	A	469	62	28
20	27	C	1043	74	18

Supplementary Figure 1: GFP-HIV gating strategy. Unstimulated CVL cells infected with R5-tropic GFP-HIV *ex-vivo* (2-10x10⁶ enriched CVL cells) were measured for GFP expression following 48 hours of cell culture with matched AZT-treated, CVL controls. CVL cells were first gated for viability and then to remove granulocytes (CD66b^{lo}). Remaining FGT cells were gated for CD3 expression, and CD3+ cells cells were then gated to remove CD8 T cells. CD3+ CD8- T cells were then analyzed for GFP expression. The gate for GFP expression was set based on the AZT control.



Supplementary Figure 2: CD4 T cells productively infected with HIV can effectively migrate to CCL19 and CCL21. PBMC stimulated with PMA/ionomycin and rested 4 days were infected with subtype C HIV isolate virus and then tested for the ability of productively infected cells to migrate to CCL19/21 chemokines by transwell assay. HIV infected T cells (labeled p24+) were detected by intracellular p24 expression in comparison to uninfected T cells or T cells in the infected culture that were negative for p24 expression. The chemotactic index represent the number of cells migrating into CCL19/21 transwells compared to cells collected from transwells containing only complete media.



Supplementary Figure 3: Total memory CD4 T cell numbers and HIV susceptibility markers CCR5 and CD38 expression on memory CD4 T cells in the FGT do not significantly increase of decrease throughout the course of a menstrual cycle. Genital lavage samples collected from participants were measured longitudinally for CCR5 (A) and CD38 (B) expression on memory CD4 T cells throughout the course of a menstrual cycle. (C) CD4 T cell numbers were calculated as the total number of lymphocyte-gated live CD3+ CD4+ cells detected during analysis by flow cytometry. Fold change was calculated by normalizing total cell numbers calculated at the first collection to all following time points. The day of cycle represents the range estimated from self-reported start of menses.

