Supplemental information

Description of approach

Four isolates were collected between June and October 2012 in hospitals from two different cities, located 300 and 700 km away from Bogota, where the first Colombian *bla*_{NDM-1} was reported (Fig S1). Isolates were sent to the International Center for Medical Research and Training (CIDEIM) as part of a bacterial resistance surveillance program. Species identification was performed by Vitek-2® (bioMérieux, Marcy l'Étoile,France) and confirmed by MALDI-TOF MS on a Microflex LT instrument (Bruker Daltonics GmbH, Leipzig, Germany). Antimicrobial susceptibility testing was conducted using broth microdilution method (Sensititre panels; TREK Diagnostic systems, Westlake, OH, USA) and minimum inhibitory concentrations (MICs) were interpreted according to Clinical and Laboratory Standards Institute guidelines (3), except for tigecycline which was interpreted using the US Federal Drug Administration (FDA) criteria. Modified Hodge test (MHT), 3D bioassay using an imipenem disk, and double-disk synergy test using EDTA (DDST+EDTA), were performed as previously described (3, 4, 8).

DNA was isolated with the MasterPure Gram-positive DNA purification kit (Epicenter Biosciences) and subjected to SMRT sequencing on a PacBio RSII. Two SMRT cells of sequence data were collected per genome; sequencing resulted in ~9-10X coverage of error-corrected reads. Assembly was performed by using PacBio's HGAP assembler version 3.1. Plasmid content and *bla*_{NDM} location was corroborated using S1 nuclease / I-Ceul -PFGE and *bla*_{NDM-1} / 16S probe hybridizations (1, 6). Genes were annotated automatically using NCBI Prokaryotic Genome Annotation Pipeline (7) and manually using BLAST. Resistance genes where identified using ResFinder 2.1 (9), *in silico* MLST and plasmid typing was performed by MLST 1.8 (5) and PlasmidFinder 1.3 (2), respectively

Vitek-2® initially identified isolates as *Alcaligenes faecalis* 6200, Kpn 6234, Eco 6409 and *Acinetobacter baumannii-calcoaceticus complex* 6411. However, MALDI-TOF MS and whole genome BLAST confirmed isolate 6200 as *Acinetobacter baumannii* (Aba) and isolate 6411 as *Acinetobacter nosocomialis* (*Ans*).

		MIC														
Isolate	Species	CSL	PTZ	FEP	FOT	TAZ	AXO	DOR	ERT	IMI	MER	AZT	AMK	CIP	POLB	TGY
6200	Aba	>128/64	>128/4	>32	ND	>16	ND	>64	ND	64	>64	ND	16	≤0,5	1	≤0.5
6234	Kpn	64/32	>128/4	>32	>16	>16	>16	1	4	2	1	32	64	>8	0,5	≤0.5
6409	Eco	>128/64	>128/4	>32	>16	>16	>32	16	>64	16	16	32	>64	>8	8	>8
6411	Ans	>128/64	>128/4	>32	ND	>16	ND	>64	ND	>64	>64	32	16	8	4	4

Table S1. Antimicrobial susceptibility (MICs in mg/L) of NDM-producing clinical isolates

CSL: cefoperazone-sulbactam; PTZ: piperacillin/Tazobactam; FEP: cefepime; FOT:cefotaxime; TAZ: ceftazidime; AXO:ceftriaxione; DORI: doripenem; ERT:ertapenem; IMI: imipenem; MERO: meropenem; AZT:aztreonam; AMK:amikacin; CIP: ciprofloxacin; POLB:polymyxin B, TGY: tigecycline. ND: not determined

Species	% Similarity	Comment
Salmonella enterica Serovar		IncA/C Plasmid Carrying <i>bla</i> NDM-1, <i>bla</i> CMY-16, and fosA3 in a Salmonella enterica Serovar
Corvallis	83	Corvallis Strain Isolated from a Migratory Wild Bird in Germany (KR091911)
E.coli	83	This study
E.coli	80	<i>bla</i> _{KPC} -harboring IncFIA plasmid pBK32533, from <i>E. coli</i>
Enterobacter spp.	80	bla _{KPC} , bla _{TEM} and Metallo-beta-lactamase
Citrobacter freundii		
	79	bla _{VIM-4} , bla _{CTX-M}
	77	
Providencia stuartii		Novel 178-Kb Plasmid Carrying <i>bla_{NDM-1}</i> in a <i>Providencia stuartii</i> Strain Isolated in Afghanistan

Table S2. Representative IncA/C plasmids giving significant similarity to plasmid NZ_CP010373.2

Isolate	Species	City of collec- tion	Patient Origin	Age (yr)	Sex	Infection type ^a	Co- morbidity ^b	LOS (days)	Days prior to positive culture	Source	Invasive interven- tions ^c	Previous hospitali- zations ^d	Clinical Outcome	Travel history ^e
6200	Aba	Neiva	Caquetá	60	F	IAB- PNEU1- LCBI	DM, RA	48	34	Blood	SURG	Yes	Dead	NO
6234	Kpn	Neiva	Huila	53	М	ND	None	9	6	Blood	None	Yes	Alive	NO
6409	Eco	Pasto	Putumayo	74	F	PNEU1	DM	28	15	Urine	SURG, FC	Yes	Dead	NO
6411	Ans	Pasto	Putumayo	76	Μ	Sepsis	None	13	7	Urine	SURG, FC, CL, INT	Yes	Dead	NO

Table S3. Clinical features of patients infected with *bla*_{NDM-1} isolates

^a IAB: intra-abdominal infection, not specified elsewhere.; PNEU-1: Clinically-defined pneumonia; LCBI: Laboratory-confirmed bloodstream infection

^b DM: diabetes mellitus; RA : Rheumatoid arthritis

^c SURG: Surgery ; CL : Central line ; INT: Intubation; FC: Foley catheter

d Healthcare exposures in past year

e Travels in the past 12 months/ Close personal contact with people (e.g. relatives, co-workers) who have traveled in the past 12 month

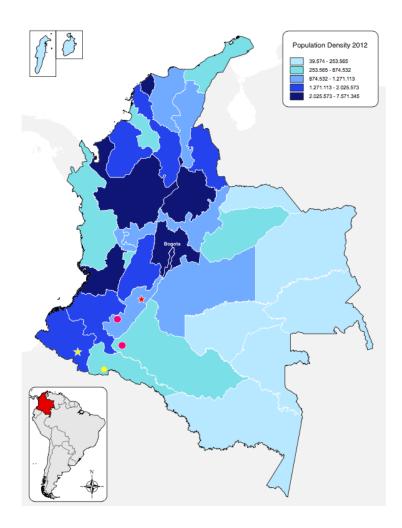


Figure S1. Map of Colombian population density. Circles indicate location of patients' origin (circle) and hospital where they were treated (star)

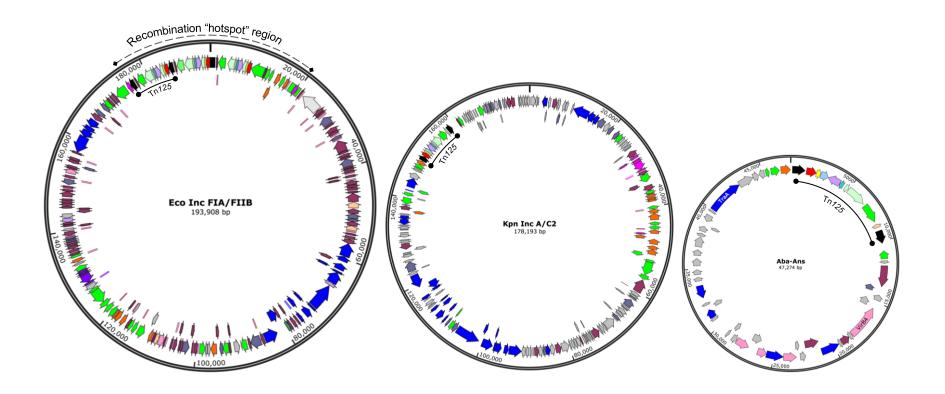


Figure S2. *bla*_{NDM-1} carrying plasmids. Selected genes are highlighted. Green, mobile genetic element-related genes; blue, genes involved in plasmid mobilization; pink, type IV secretion system; black, ISAba125; yellow, *bla*_{NDM-1}; red: *ble*_{MBL}

References

- 1. **Barton BM, Harding GP, and Zuccarelli AJ.** 1995. A General Method for Detecting and Sizing Large Plasmids. Anal. Biochem. **226:**235-240.
- 2. Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, and Hasman H. 2014. In Silico Detection and Typing of Plasmids using PlasmidFinder and Plasmid Multilocus Sequence Typing. Antimicrobial Agents and Chemotherapy 58:3895-3903.
- 3. **CLSI.** 2012. Performance standards for antimicrobial susceptibility testing; 22nd informational supplement. CLSI M100-S21. . Clinical and Laboratory Standards Institute, Wayne, PA.
- 4. **Coudron PE, Moland ES, and Thomson KS.** 2000. Occurrence and Detection of AmpC Beta-Lactamases among *Escherichia coli, Klebsiella pneumoniae*, and *Proteus mirabilis* Isolates at a Veterans Medical Center. J Clin Microbiol **38**:1791-1796.
- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, and Lund O. 2012. Multilocus Sequence Typing of Total-Genome-Sequenced Bacteria. Journal of Clinical Microbiology 50:1355-1361.
- 6. Liu S-L, Hessel A, and Sanderson KE. 1993. Genomic mapping with I-Ceu I, an intron-encoded endonuclease specific for genes for ribosomal RNA, in Salmonella spp., Escherichia coli, and other bacteria. Proceedings of the National Academy of Sciences **90**:6874-6878.
- 7. **Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Ciufo S, and Li W.** 2013. Prokaryotic Genome Annotation Pipeline. , The NCBI Handbook 2nd ed. National Center for Biotechnology Information (US), Bethesda (MD).
- 8. **Wan Nor Amilah W, Noor Izani N, Ng W, and Ashraful Haq J.** 2012. A simple screening test for the detection of metallo-beta-lactamase-producing Pseudomonas aeruginosa and Acinetobacter in a tertiary care hospital. Tropical Biomedicine **29:**588-597.
- 9. Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, and Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. Journal of Antimicrobial Chemotherapy 67:2640-2644.