

**Supplement to:**

**Quantitative analysis of dynamic protein interactions during transcription reveals a role for casein kinase II in PAF complex phosphorylation and regulation of H2B monoubiquitylation**

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## SUPPLEMENTAL TABLE LEGENDS

Table S1: Yeast strains used in this study.

Table S2: Primers and plasmids used in this study.

Table S3: SAINT analysis of Spt16-TAP interactions (n=4). Known interactions were annotated as such based on the iRef index ([irefindex.org](http://irefindex.org)). Prey names are the standard gene names as defined in the *Saccharomyces* genome database ([www.yeastgenome.org](http://www.yeastgenome.org)).

Table S4: Phosphorylated peptides identified by PTM analysis of Cdc73-TAP. Both the preceding and following amino acids for each tryptic peptide are shown. The phosphorylation sites are indicated by 'PO4'. Note that some sites were detected with multiple unique peptides.

Table S5: Peptide ratios for Ctr9-FLAG SILAC experiments from WT and CKII temperature sensitive strains. The peptides sequence identified, the sequence match to a putative CKII phosphorylation site, and the SILAC ratio of WT (heavy) and *cka1Δ cka2-8* (light) peptides are given.

## SUPPLEMENTAL TABLES

Table S1:

Strain Name	Genotype	Reference
<b>BY4717</b>	<i>MATa his3D leu2D met15D ura3D</i>	Ghaemmaghami, S et al. (2003)
<b>Spt16-TAP</b>	<i>MATa his3D leu2D met15D ura3D Spt16-TAP:HIS3MX6</i>	Ghaemmaghami, S et al. (2003)
<b>Ckb2-TAP</b>	<i>MATa his3D leu2D met15D ura3D Ckb2-TAP:HIS3MX6</i>	Ghaemmaghami, S et al. (2003)
<b>Cdc73-TAP</b>	<i>MATa his3D leu2D met15D ura3D Cdc73-TAP:HIS3MX6</i>	Ghaemmaghami, S et al. (2003)
<b>DY150</b>	<i>MATa ade2 can1 his3 leu2 trp1 ura3</i>	Biswas, D et al. (2006)
<b>DY8107</b>	<i>MATa spt16-11 ade2 can1 his3 leu2 lys2 met15 trp1 ura3</i>	Biswas, D et al. (2007)
<b>DDY4010</b>	<i>MATa his3 Δ200 leu2-3,112 ura3-52</i>	Peng, Y et al. (2011)
<b>DDY4011</b>	<i>MATa his3 Δ200 leu2-3,112 ura3-52 cka1::CgURA3</i>	Peng, Y et al. (2011)
<b>DDY4013</b>	<i>MATa his3 Δ200 leu2-3,112 ura3-52 cka1::CgURA3 cka2-8::NatR</i>	Peng, Y et al. (2011)
<b>YJJ662</b>	<i>MATa ura3-52 leu2Δ1 his3Δ200</i>	Shi X, et al. (1997)
<b>YJJ577</b>	<i>MATa ura3-52 leu2Δ1 his3Δ200 paf1Δ::HIS3</i>	Porter, S et al. (2002)
<b>yJK002*</b>	<i>MATa his3 Δ200 leu2-3,112 ura3-52 CTR9-3xFLAG::kanMX</i>	This study
<b>yJK004†</b>	<i>MATa his3 Δ200 leu2-3,112 ura3-52 cka1Δ::CgURA3 cka2-8::NatR CTR9-3xFLAG::kanMX</i>	This study
<b>yJK006‡</b>	<i>MATa ura3-52 leu2Δ1 his3Δ200 paf1::natNT2</i>	This study
<b>yJK008§</b>	<i>MATa ura3-52 leu2Δ1 his3Δ200 ctr9::natNT2</i>	This study

\* isogenic to DDY4010 (Peng Y, et al. 2011 (1))

† isogenic to DDY4013 (Peng Y, et al. 2011)

‡ isogenic to YJJ662 (Shi X, et al. 1997 (2))

Table S2:

Table S2. Plasmids and primers used in this study.		
Plasmids	Features	Source
pRS316	CEN URA3	Sikorski R.S. and Hieter P. (1989)

pRS313-3HA-SSN6	<i>CEN HIS3 p3xHA-SSN6</i>	Michael Keogh
pJK001	<i>CEN HIS3 pPAF1(WT)-3XHA-SSN6</i>	This study
pJK002	<i>CEN HIS3 pPAF1(S147A, T385A, T422A, S426A)-3XHA-SSN6</i>	This study
pJK003	<i>CEN HIS3 pPAF1(S147E, T385E, T422E, S426E)-3XHA-SSN6</i>	This study
pJK007	<i>CEN HIS3 pCTR9(WT)-3XHA-SSN6</i>	This study
pJK008	<i>CEN HIS3 pCTR9(S974A, S977A, S1015A, S1017A, S1046A, S1054A, S1056A)-3XHA-SSN6</i>	This study
pJK009	<i>CEN HIS3 pCTR9(S974E, S977E, S1015E, S1017E, S1046E, S1054E, S1056E)-3XHA-SSN6</i>	This study
pFA6a – natNT2	cassette plasmid (deletion)	Janke C., <i>et al.</i> (2014)
p3FLAG-kanMX	cassette plasmid (3x-FLAG tagging)	Gelbart M.E., <i>et al.</i> (2001)
Primers	Sequence (5'-3')	Application
oGGW05 4	GAAATGTATTCACTACAATAGAACAGTGCTCATAATAGTATA AAGGGTCACAATGCGTACGCTGCAGGTCGAC	Paf1 deletion cassette
oGGW05 5	GTAAAAAAGAACTACAGGTTAAAATCAATCTCCCTTCACTTCT CAATATTCTAATCGATGAATTGAGCTCG	Paf1 deletion cassette
oGGW05 6	CAAAGTTCTAATTGTCTGGTCCATTGTGTTGAGAGCAAGAA AAAAAAACATGCGTACGCTGCAGGTCGAC	Ctr9 deletion cassette
oGGW05 7	GTATAATATTAAGTTCTTAAAAGTCTGATTCTAACCCCTCG CCTCTTCTTAATCGATGAATTGAGCTCG	Ctr9 deletion cassette
oJLK087	CGACGAAAACAATGATAATGATGATAACGACGGATTGTCAG GGAACAAAAGCTGGAG	Ctr9-3xFLAG cassette
oJLK088	TTTCTTAAAAGTCTGATTCTAACCCCTCGCCTCTTACTAT AGGGCGAATTGGGT	Ctr9-3xFLAG cassette
oJLK025	AAAGGATCCGCCTGCCAACAGATAAAA	Paf1 cloning
oJLK026	AAACCCGGTTCTCTTGAAAGTTCTTTCTCC	Paf1 cloning
oJLK055	AAAGGATCCCTCCACTGCAAACGTTTC	Ctr9 cloning
oJLK056	AAAGATATCGAACAACTCGTCGTTATCATCA	Ctr9 cloning
oJLK018	GCGCTTGGATGATGGAGATGCGGATGATGAAAACC	Paf1 S147A mutagenesis
oJLK019	AGAAAGAACGAAAGGCGATTCAAAAGCAGAAGGTTCTGAG	Paf1 T385A mutagenesis
oJLK020	GATGAAAACCGTGCAGCTGATGCGCCCCGAAACTTC	Paf1 T422A mutagenesis
oJLK021	CTGATGCGCCCAGAACTGCAGATGCTGTTCTACT	Paf1 S426A

		mutagenesis
oJLK034	AGGCCTGGATGATGGAGATGAGGATGATGAAAACCTGA	Paf1 S147E mutagenesis
oJLK035	GAAAGAACCGAAGGCGATTCAAAAGAGGAAGGTTCTGAGC AAGAAGGA	Paf1 T385E mutagenesis
oJLK036	CAAGATGAAAACCGTGCAGCTGATGAGGCCGAAACTGAGGA TGCTGTTCATACTGAACAAAAAA	Paf1 T422E, S426E mutagenesis
oJLK061	CCATAAGTGAACATAATGTTAAAGATGATGCAGATCTGGCTG ATAAAGACAATGAATATGAC	Ctr9 S974A, S977A mutagenesis
oJLK062	GAAAGCTGCTAAAAAAACTCTGCCGACGCCGACGAGGATGA CGAT	Ctr9 S1015A, S1017A mutagenesis
oJLK063	ATCTCAACTATCGAACATGAATTATTGAAGATGCTGATGAGGA AGAAGCTAAA	Ctr9 S1046A mutagenesis
oJLK064	GATGAGGAAGAACGCTCAAATGGCCGGTGCAGAACAAAATAA AAACGATG	Ctr9 S1054A, S1056A mutagenesis
oJLK072	CCATGGCCATAAGTGAACATAATGTTAAAGATGATGAGGATC TGGAGGATAAAGACAATGAATATGACGAAGAAAAGCCAAG	Ctr9 S974E, S977E mutagenesis
oJLK073	CGGTGAATCTAAGAGAACGGAAAGCTGCTAAAAAAACTCTGA GGACGAGGACGAGGATGACGATGATGTG	Ctr9 S1015E, S1017E mutagenesis
oJLK074	ATCTCAACTATCGAACATGAATTATTGAAGATGAGGATGAGGA AGAAGCTCAAATGGAG	Ctr9 S1046E mutagenesis
oJLK076	AAGATAGTGATGAGGAAGAACGCTCAAATGGAGGGTGAGGAA CAAAATAAAACGATGACAACGACG	Ctr9 S1054E, S1056E mutagenesis

Table S3:

BAIT	PREY	FC_A (average mean of replicates)	FC_B (geometric mean of replicates)	SAINT probability Score	Known interaction (iREF)
SPT16	POB3	3475.31	3257.34	1	Yes
SPT16	SPT16	1295.52	998.31	1	Yes
SPT16	HIS1	409.69	358.56	1	No
SPT16	CKA2	175.66	110.49	1	Yes
SPT16	LEO1	96.61	90.75	1	Yes
SPT16	CKB2	150.5	82.3	1	Yes
SPT16	CKB1	125.24	80.7	1	Yes
SPT16	CTR9	68.58	61.82	1	Yes
SPT16	HHF1	36.39	31.42	1	Yes
SPT16	RPS6A	35.2	29.02	1	No

SPT16	PSH1	28.94	25.01	1	Yes
SPT16	PAF1	24.5	19.73	1	Yes
SPT16	IKI3	29	16.12	1	No
SPT16	RPS8A	14.99	13.37	1	No
SPT16	RTF1	13.56	11.5	0.98	Yes
SPT16	CKA1	164.19	69.92	0.93	Yes
SPT16	MSH6	56.6	20.17	0.75	No
SPT16	YFL054C	36.15	16.29	0.75	No
SPT16	HTB2	28.54	11.4	0.75	Yes
SPT16	LAP4	38.59	11.37	0.75	No
SPT16	MRC1	19.34	8.72	0.75	No
SPT16	RRM3	13.39	8.43	0.75	No
SPT16	RPS17A	14.24	8.11	0.75	No
SPT16	HTA1	32.53	7.7	0.75	Yes
SPT16	RAV1	13.24	7.31	0.75	No
SPT16	YTA6	15.62	7.27	0.75	No
SPT16	PTK2	12.24	7.12	0.75	No
SPT16	CTI6	11.8	7.03	0.75	No
SPT16	SFL1	15.68	6.93	0.75	No
SPT16	YOR1	13.32	6.89	0.75	No
SPT16	RPS18A	10.9	6.55	0.75	No
SPT16	RPL1A	8.05	6.35	0.75	No
SPT16	YKL105C	9.22	6.22	0.75	No
SPT16	CDC73	17.92	6.1	0.75	Yes
SPT16	SNT1	9.09	5.88	0.75	No
SPT16	SKI2	8.08	5.28	0.75	No
SPT16	BOI2	8.45	5.28	0.75	No
SPT16	ZIP1	10.87	4.93	0.75	No
SPT16	PKH2	5.45	4.17	0.75	No

Table S4:

Protein	Site	Peptide	XCorr	X!Tandem
Cdc73	S157	K.STTSASLEND\$ <sup>P04</sup> EVSDPVVVETMK.H	4.35	6.15
Ctrl9	S974, S977	R.EAMAISEHNVKDDS <sup>P04</sup> DLS <sup>P04</sup> DK.D	5.55	3.15
		K.DDS <sup>P04</sup> DLS <sup>P04</sup> DKDNEYDEEKPR.Q	4.57	5.49
	S977	K.DDS DLS <sup>P04</sup> DKDNEYDEEKPR.Q	4.54	4.89
		K.DDS DLS <sup>P04</sup> DKDNEYDEEKPR.Q	4.86	2.39
	S1015	K.AAKTLS <sup>P04</sup> DSDED-DDDVVK.K	5.6	6.12
	S1017	K.AAKTLS DSD <sup>P04</sup> DEDDDDVVVK.K	5.14	5.77
	S1015, S1017	K.AAKTLS <sup>P04</sup> DS <sup>P04</sup> DEDDDDVVVK.K	5.63	3.06
		K.KTLS <sup>P04</sup> DS <sup>P04</sup> DEDDDDVVVK.K	6.03	5.7
	S1046, S1054, 1056	K.SQLSNEFIEDS <sup>P04</sup> DEEEAQMS <sup>P04</sup> GS <sup>P04</sup> EQNK.N	8.11	3.89

	S1046, S1054	K.SQLSNEFIEDS <sup>PO4</sup> DEEEAQMS <sup>PO4</sup> GSEQNK.N	5.67	3.51
		K.SQLSNEFIEDsDEEEAQmsGSEQNK(N)	7.36	5.96
	S1054	(K)SQLSNEFIEDSDEEEAQMS <sup>PO4</sup> GSEQNK.N	6.94	5.64
Leo1	S25	K.EQISNNVGVTTNS <sup>PO4</sup> TSNEETSR.S	5.96	7.21
	T26	K.EQISNNVGVTTNST <sup>PO4</sup> SNEETSR.S	5.65	6.96
	S105	R.SRHRES <sup>PO4</sup> LGLDDDEAEEQAMYTR.K	6.46	2.1
	S132	R.KFYGEDANNFS <sup>PO4</sup> DQDETTHTFK.E	7.6	4.32
		K.FYGEDANNFS <sup>PO4</sup> DQDETTHTFKEENVELVR.H	7.04	5.26
		R.KFYGEDANNFS <sup>PO4</sup> DQDETTHTFK.E	7.6	4.32
	S358	K.EKQES <sup>PO4</sup> PDAAFETGFR.K	3.77	1.96
		K.QES <sup>PO4</sup> PDAAFETGFR.K	4.39	2.49
	S372	R.KQNS <sup>PO4</sup> PTTYGASR.R	3.87	1.68
Paf1	S32	K.LLVYPES <sup>PO4</sup> PETNADSSQLINSLYIK.T	5.24	5.1
	S147	K.RRLDDGDS <sup>PO4</sup> DDENLDVNHIISR.V	5.83	4.17
	S388 or T385	K.TEGS <sup>PO4</sup> EQEGENEKDEEIK.Q	5.29	2.52
		K.T <sup>PO4</sup> EGSEQEGENEKDEEIKQEK.E	6.04	1.66
	S388, T385	K.ESEGDSKT <sup>PO4</sup> EGS <sup>PO4</sup> EQEGENEKDEEIK.Q	4.61	3.85
	T422	R.AADT <sup>PO4</sup> PETSDAVHTEQKPEEEEK.E	5.49	4.42
	T422, S426	R.AADT <sup>PO4</sup> PETS <sup>PO4</sup> DAVHTEQKPEEEEK.E	5.62	2.4
Rtf1	S90	K.DES <sup>PO4</sup> DREHLESLPEMER.E	4.48	
	S181	R.HYS <sup>PO4</sup> DNEDEDDEEDYR.E	4.77	5.24
		R.HYS <sup>PO4</sup> DNEDEDDEEDYREEDYK.D	6.12	3.85
	S203	K.DDEGS <sup>PO4</sup> EYGDDEEYNPFDR.R	5.5	5.05

Table S5:

Protein	Potential CKII site(s)?	Peptide sequence	<i>cka1Δ</i> <i>cka2-8 / WT ratio</i>	Modification status
Ctr9	Yes	EAMAISEHNVKDDSDLSDKDN EYDEEKPR	8.69	S974 and 977 unmodified
Ctr9	Yes	EAMAISEHNVKDDS <sup>PO4</sup> DLS <sup>PO4</sup> DKDNEYDEEKPR	0.39	S974 and 977 PO4
Ctr9	Yes	KTLSDSDEDDDDVVKKPSHNK	41.31	S1015 and 1017 unmodified
Ctr9	Yes	TLSDSDEDDDDVVKKPSHNKG K	48.78	S1015 and 1017 unmodified
Ctr9	Yes	TLSDSDEDDDDVVKKPSHNK	39.64	S1015 and 1017 unmodified
Ctr9	Yes	KTLSDS <sup>PO4</sup> DEDDDDVVKKPSHNK	3.20	S1017 PO4
Ctr9	Yes	TLSDS <sup>PO4</sup> DEDDDDVVKKPSHNKGK	3.48	S1017 PO4
Ctr9	Yes	TLSDS <sup>PO4</sup> DEDDDDVVKKPSHNK	3.20	S1017 PO4

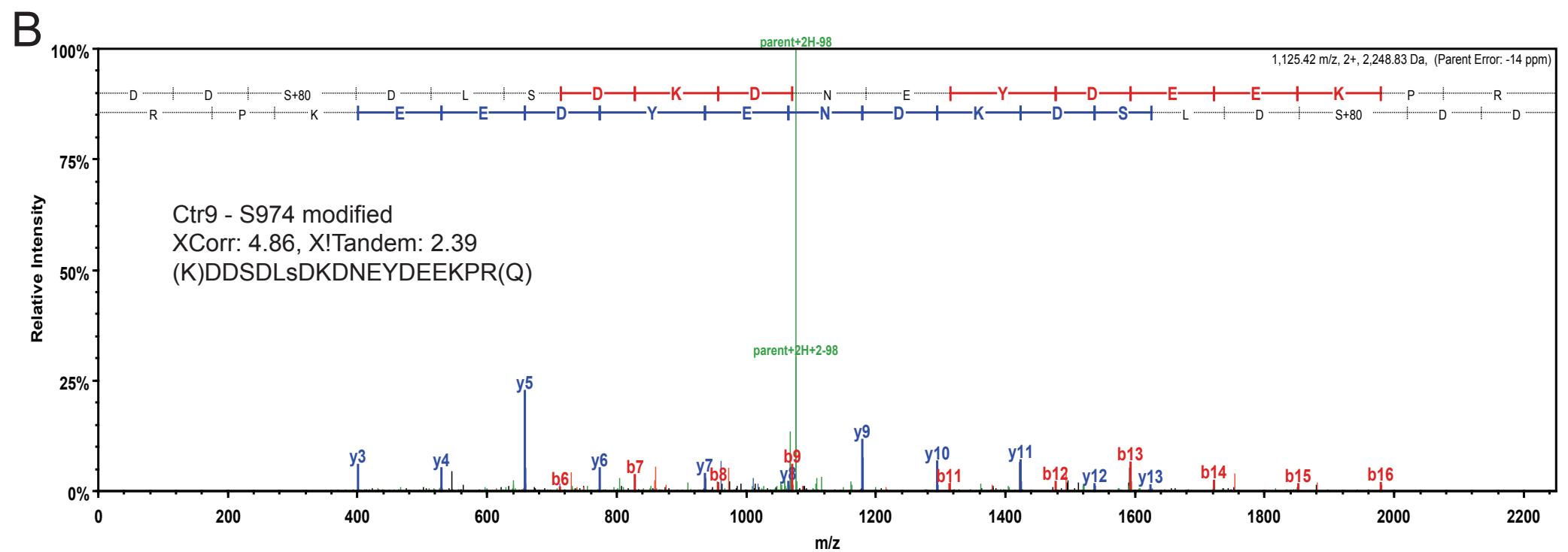
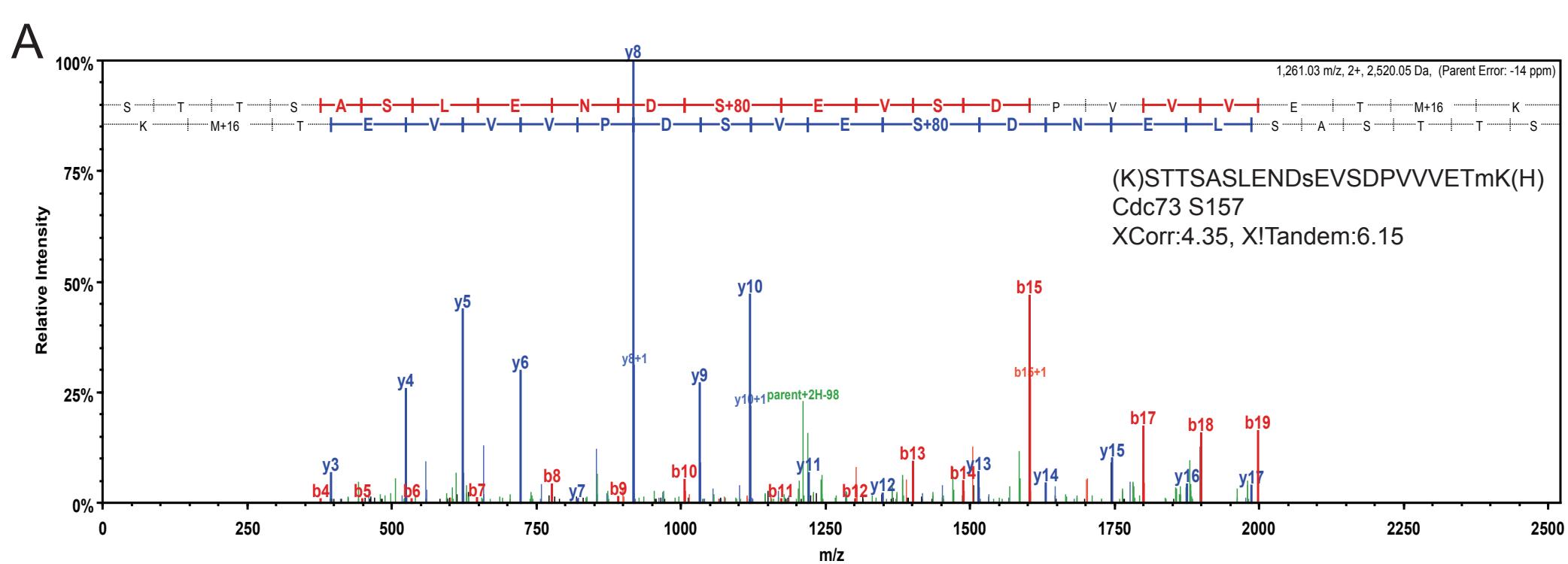
Ctr9	Yes	TLS <sup>PO4</sup> DS <sup>PO4</sup> DEDDDDVVK	0.13	S1015 and 1017 PO4
Paf1	No	LLVYPESPETNADSSQLINSLYIK	1.14	S32 unmodified
Paf1	No	LLVYPES <sup>PO4</sup> PETNADSSQLINSLYIK	0.64	S32 PO4
Paf1	No	RTEYVSNT <sup>PO4</sup> IAAHDNTSLKR	3.15	T127 PO4
Paf1	No	TEYVSNTIAAHDNTSLKR	0.79	T127 unmodified
Paf1	Yes	RLDDGDSDDENLDVNHIISRVEGTFNK	44.19	S147 unmodified
Paf1	Yes	RRLDDGDSDDENLDVNHIISR	29.57	S147 unmodified
Paf1	Yes	RLDDGDSDDENLDVNHIISR	29.38	S147 unmodified
Paf1	Yes	LDDGDSDDENLDVNHIISR	28.10	S147 unmodified
Paf1	Yes	RLDDGDS <sup>PO4</sup> DDENLDVNHIISRVEGTFNK	0.25	S147 PO4
Paf1	Yes	RRLDDGDS <sup>PO4</sup> DDENLDVNHIISR	0.31	S147 PO4
Paf1	Yes	LDDGDS <sup>PO4</sup> DDENLDVNHIISR	0.24	S147 PO4
Leo1	No	HRESLGLDDDEAEEQAMYTR	0.82	S105 unmodified
Leo1	No	HRES <sup>PO4</sup> LGLDDDEAEEQAMYTRK	1.42	S105 PO4
Leo1	No	SRHRES <sup>PO4</sup> LGLDDDEAEEQAMYTR	1.35	S105 PO4
Leo1	Yes	KFYGEDANNFSDQDETTHFK EENVELVR	2.47	S132 unmodified
Leo1	Yes	FYGEDANNFSDQDETTHFK ENVELVR	2.40	S132 unmodified
Leo1	Yes	KFYGEDANNFS <sup>PO4</sup> DQDETTHFK KEENVELVR	0.34	S132 PO4
Leo1	Yes	FYGEDANNFS <sup>PO4</sup> DQDETTHFK KEENVELVR	0.37	S132 PO4
Rtf1	Yes	RIEVGSVEDDDEDDYNPYSV GNADYGSEEEEANPFPLEGK	6.92	S71 unmodified
Rtf1	Yes	RIEVGSVEDDDEDDYNPYSV GNADYGS <sup>PO4</sup> EEEEANPFPLEGK	0.33	S71 PO4

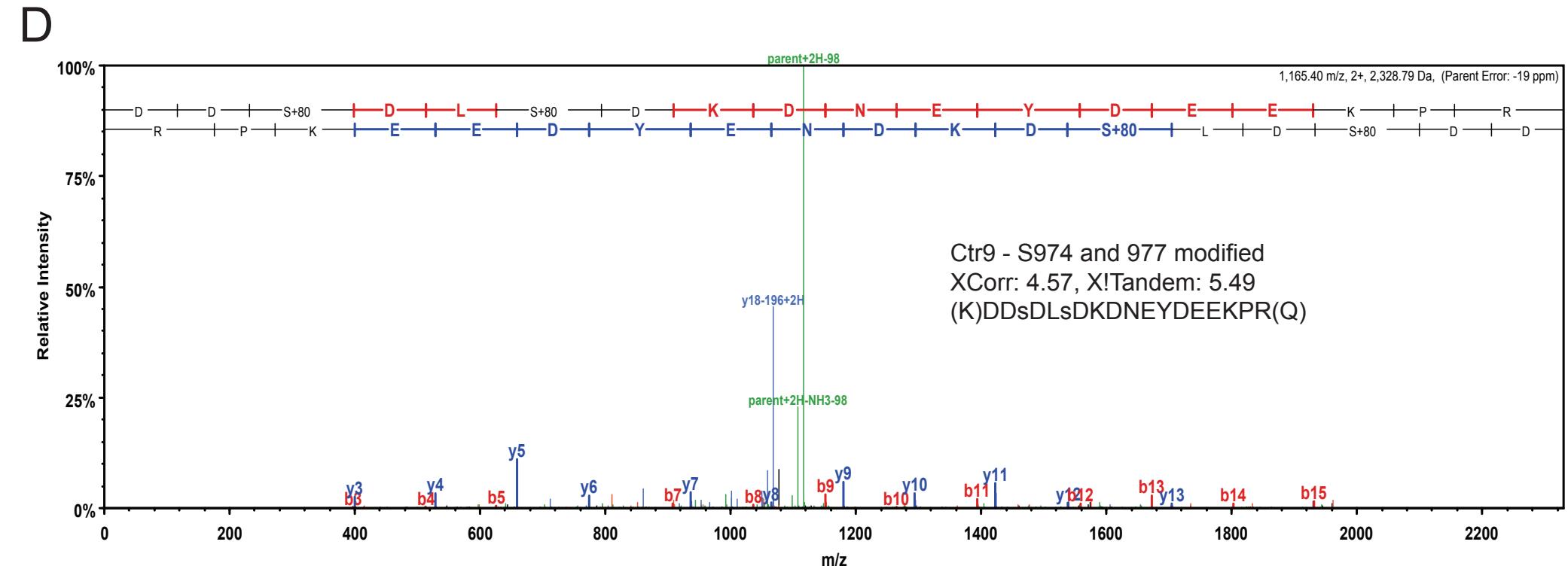
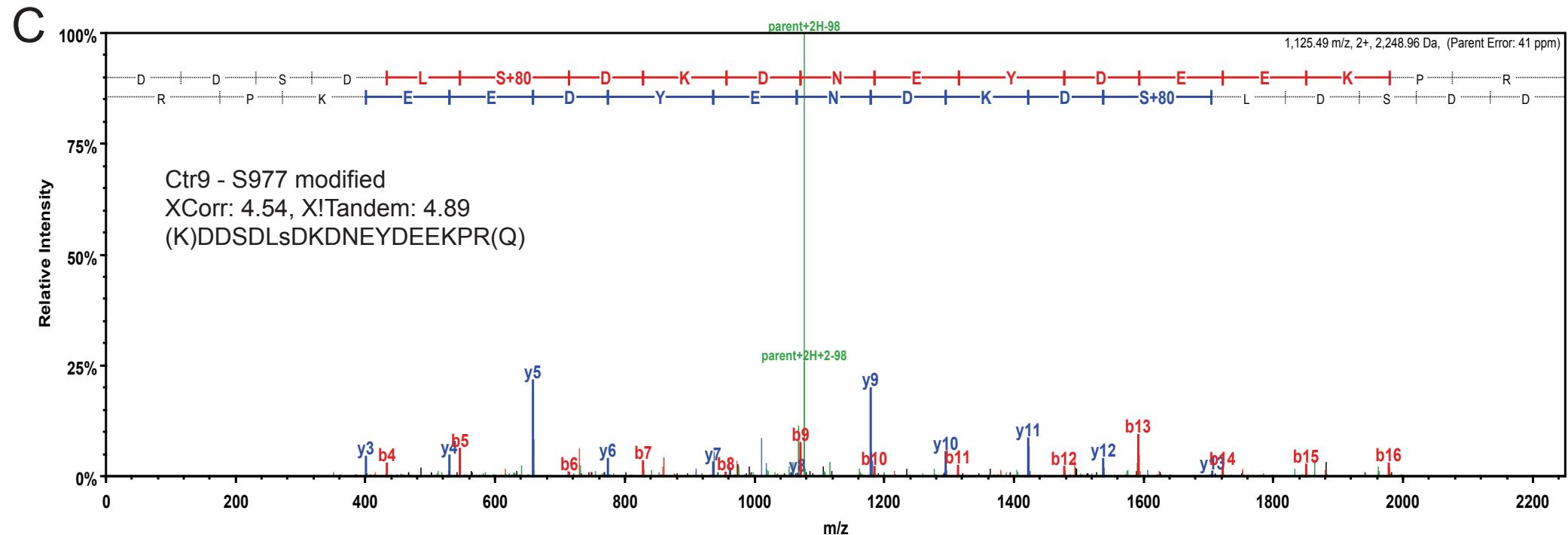
## SUPPLEMENTAL FIGURES

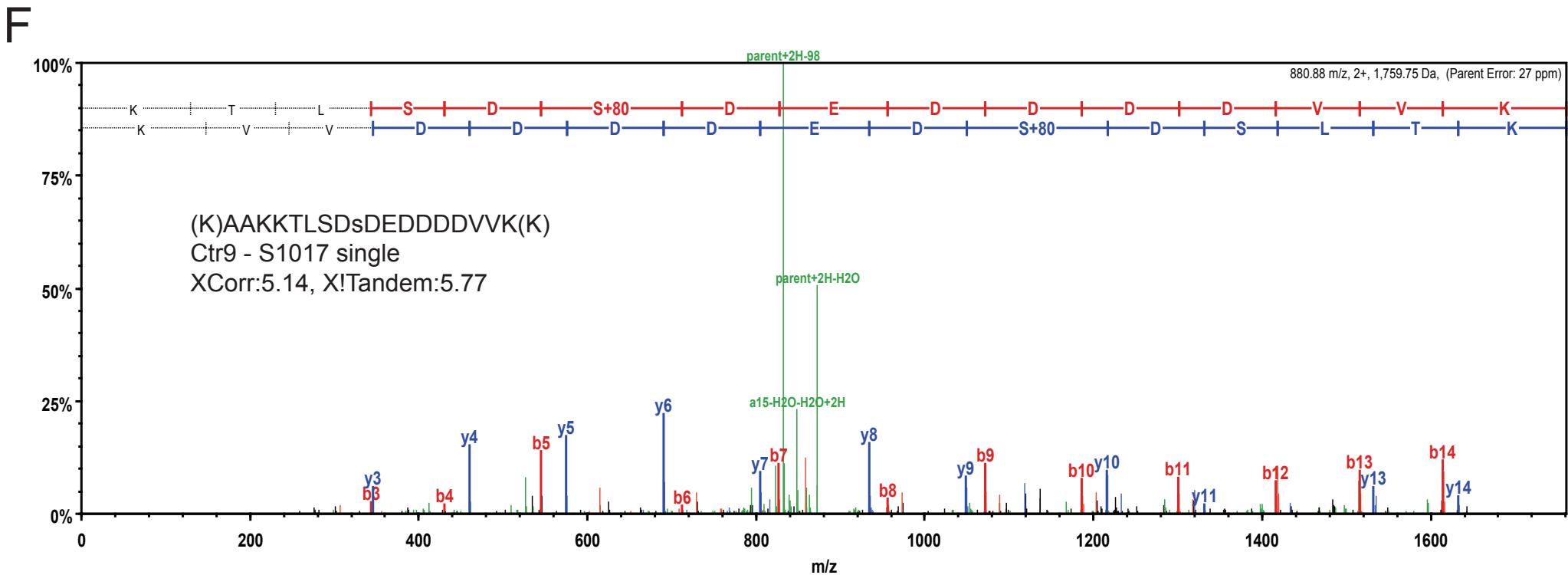
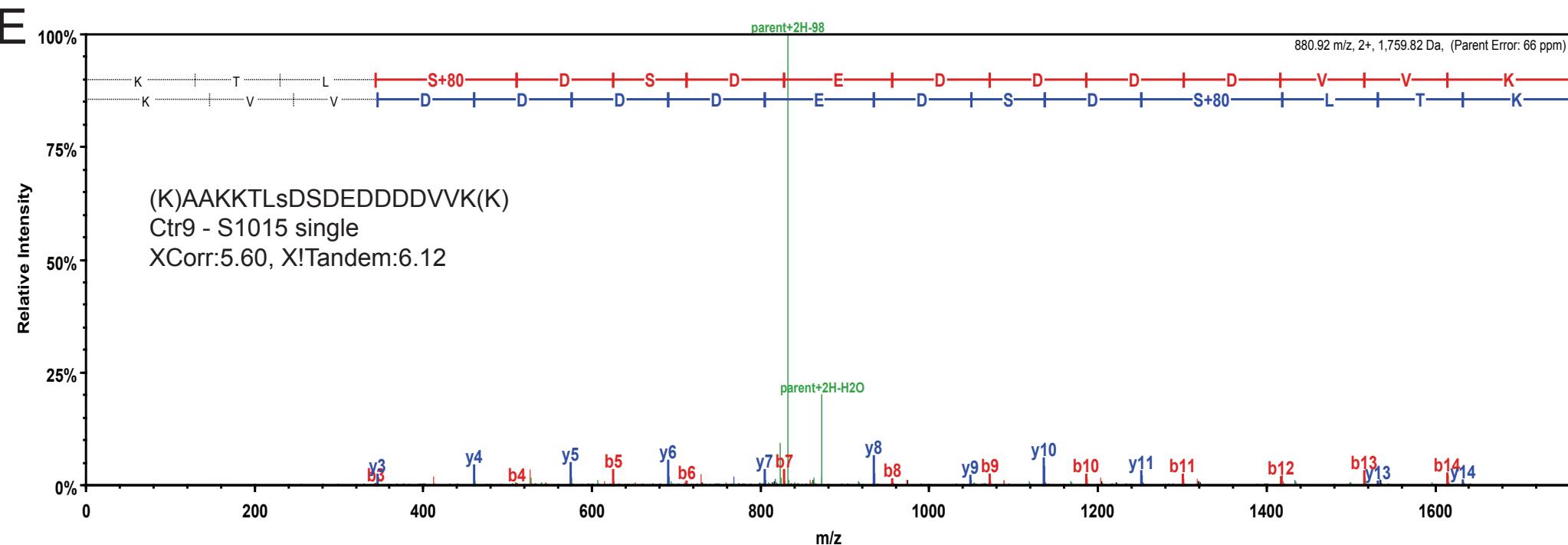
Figure S1: Representative spectra for the phosphorylation sites in the five subunits of PAF-C: Cdc73 (A), Ctr9 (B-K), Leo1 (L-Q), Paf1 (R-W), and Rtf1 (X-AA). In all spectra, b-ions are labeled in red and y-ions are labeled in blue. Neutral loss and other parent ion fragments are shown in green. The XCorr and X!Tandem scores and ppm values are given for reference.

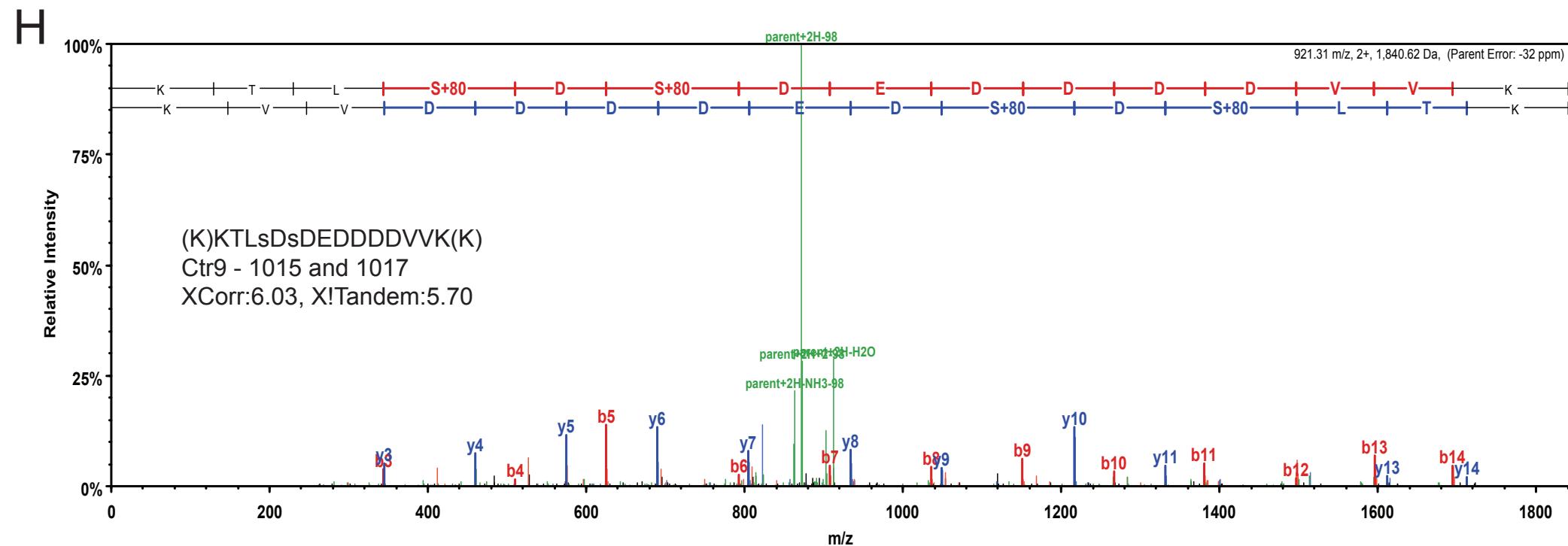
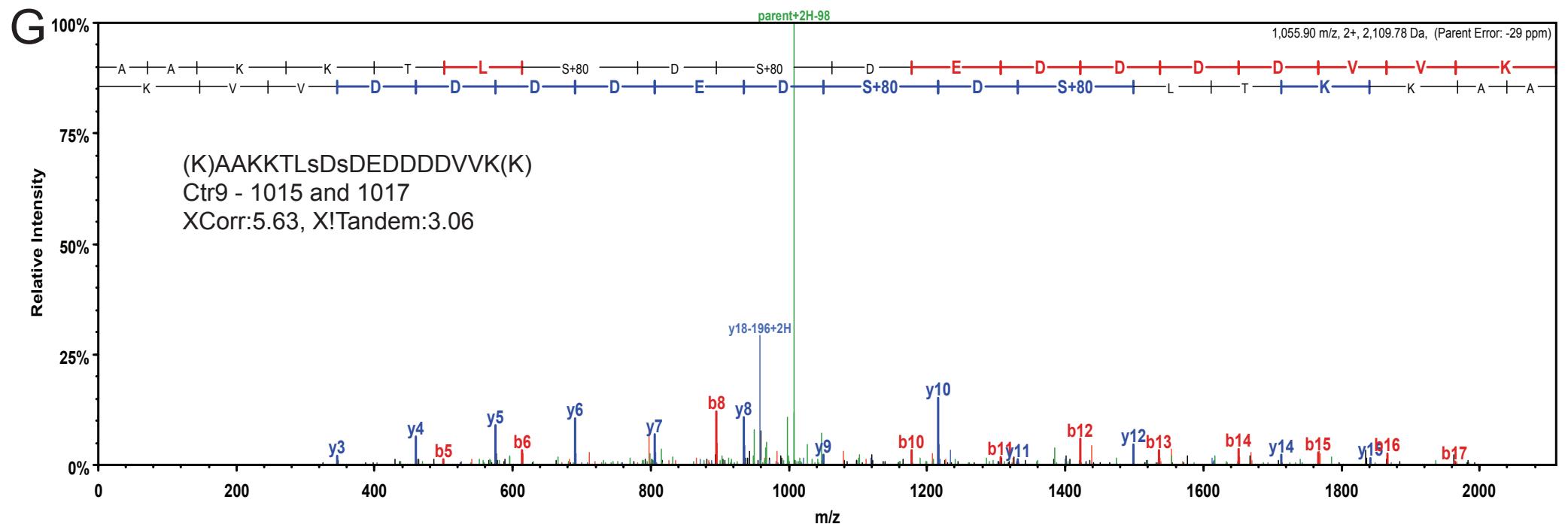
## REFERENCES

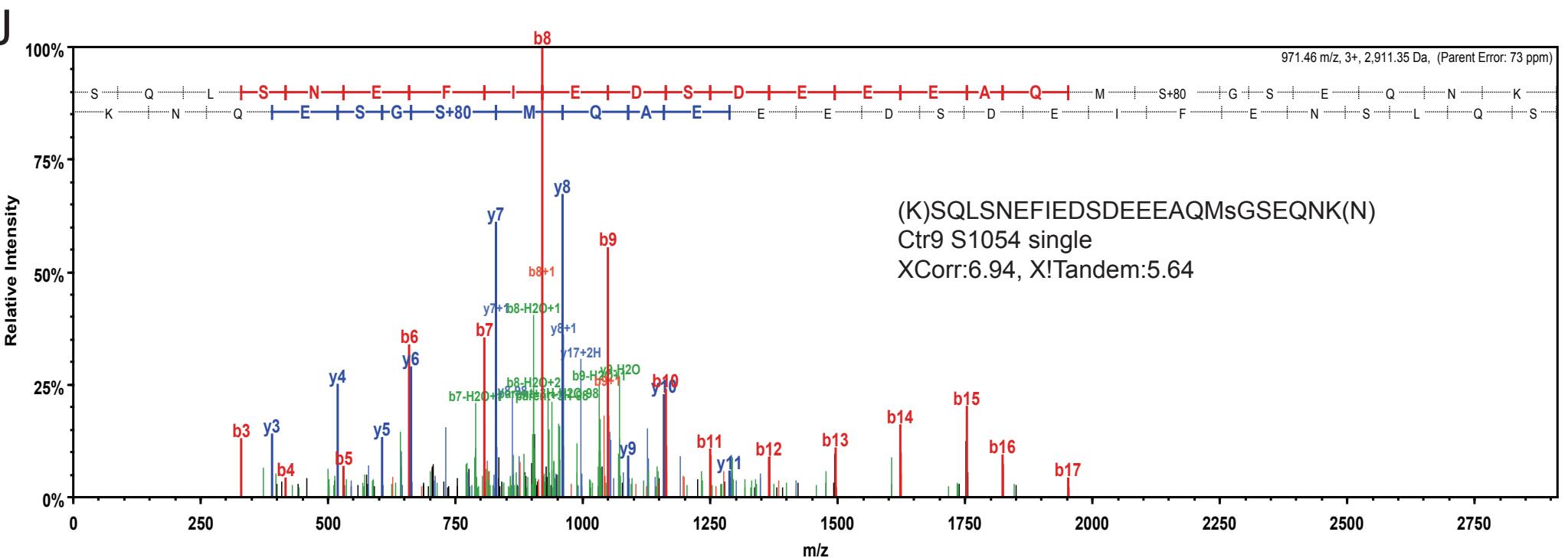
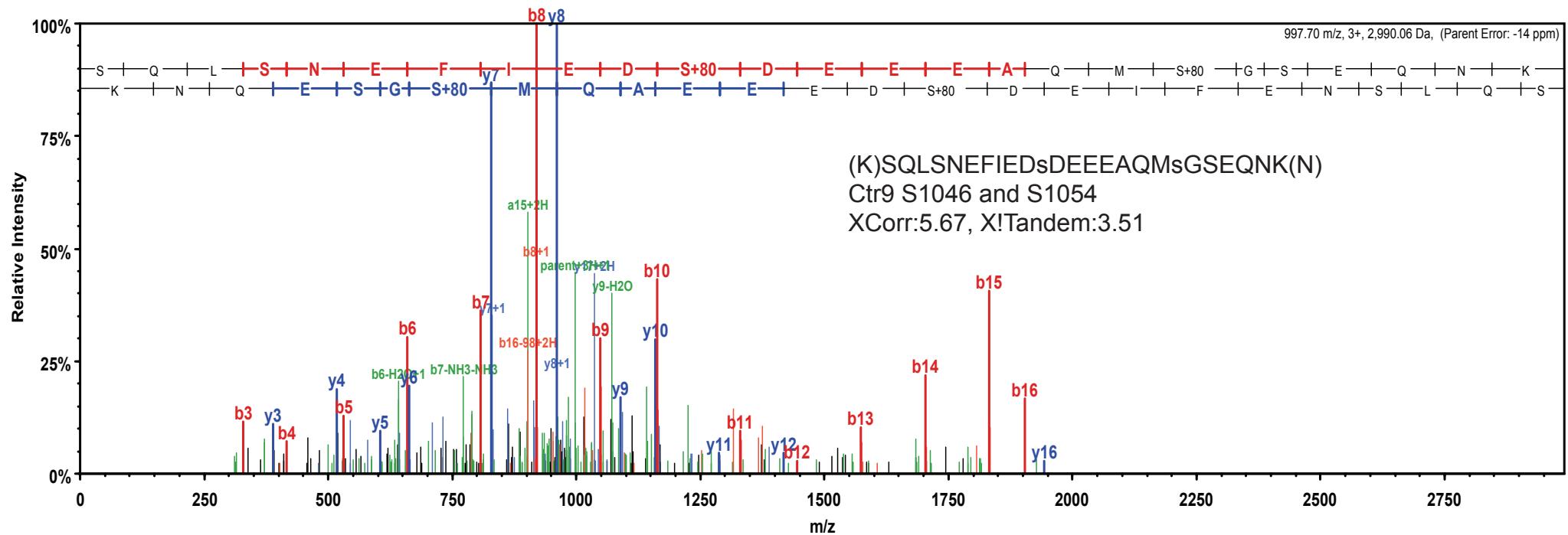
1. Peng, Y. B., Fan, B., Han, X. L., Xu, X. W., Rothschild, M. F., Yerle, M., and Liu, B. (2011) Molecular characterization of the porcine JHDM1A gene associated with average daily gain: evaluation its role in skeletal muscle development and growth. *Molecular biology reports* **38**, 4697-4704
2. Shi, X., Chang, M., Wolf, A. J., Chang, C. H., Frazer-Abel, A. A., Wade, P. A., Burton, Z. F., and Jaehning, J. A. (1997) Cdc73p and Paf1p are found in a novel RNA polymerase II-containing complex distinct from the Srbp-containing holoenzyme. *Molecular and cellular biology* **17**, 1160-1169

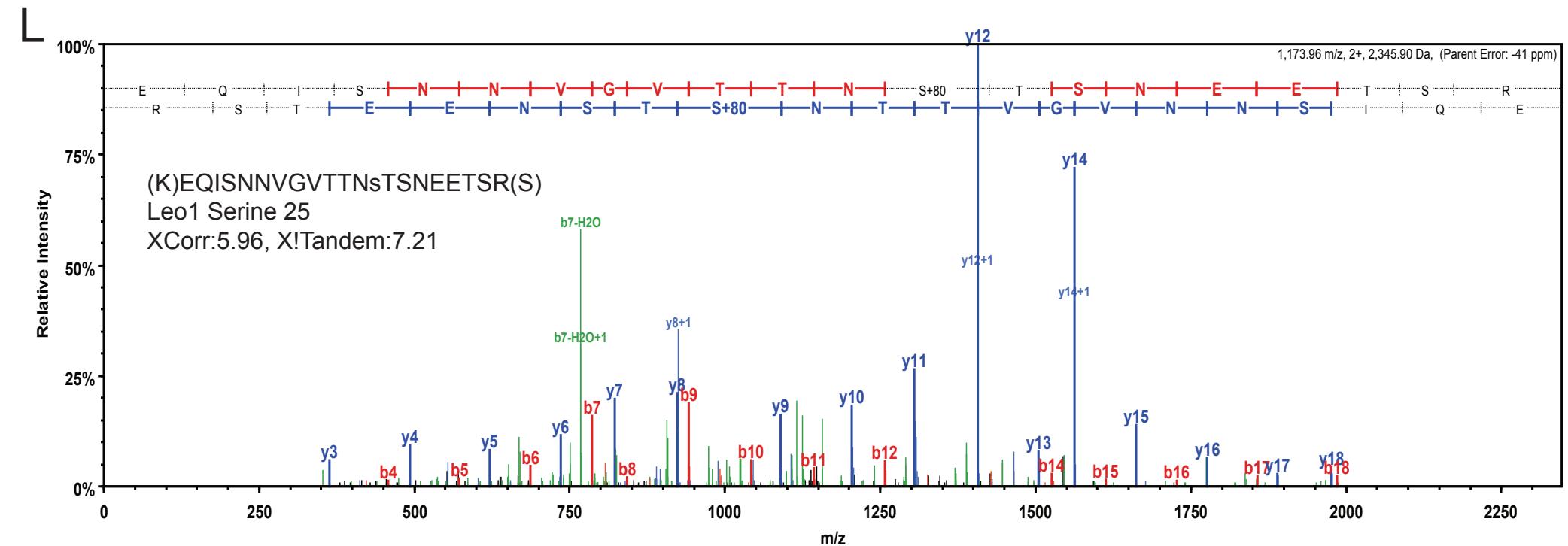
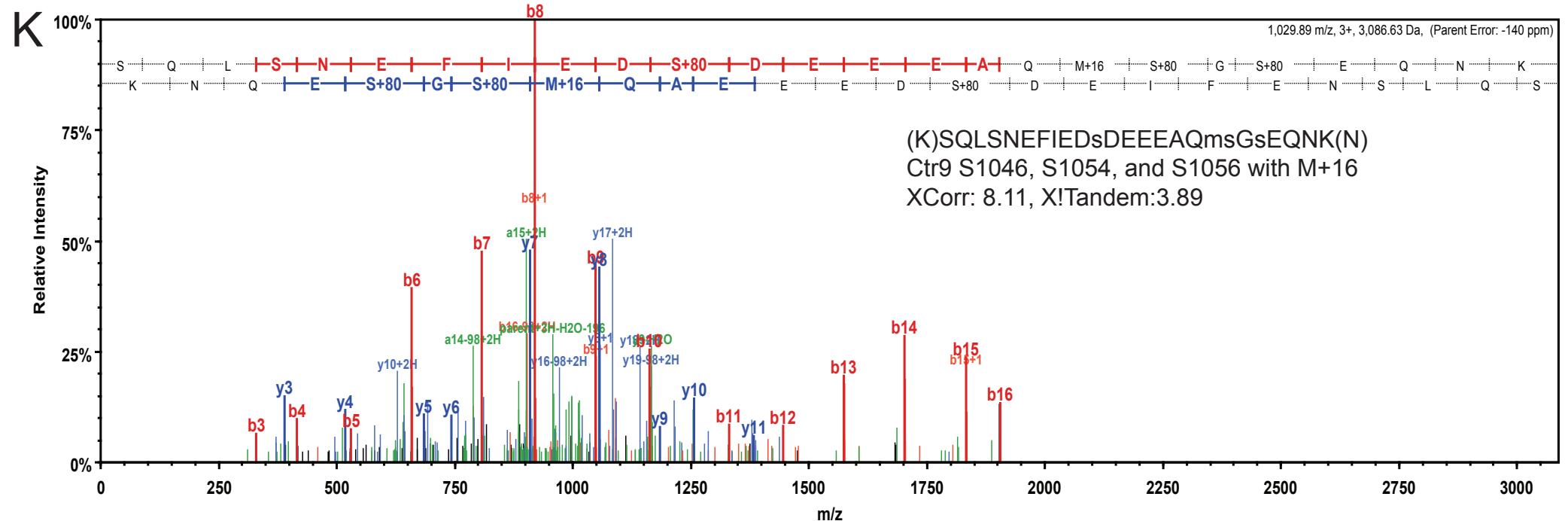


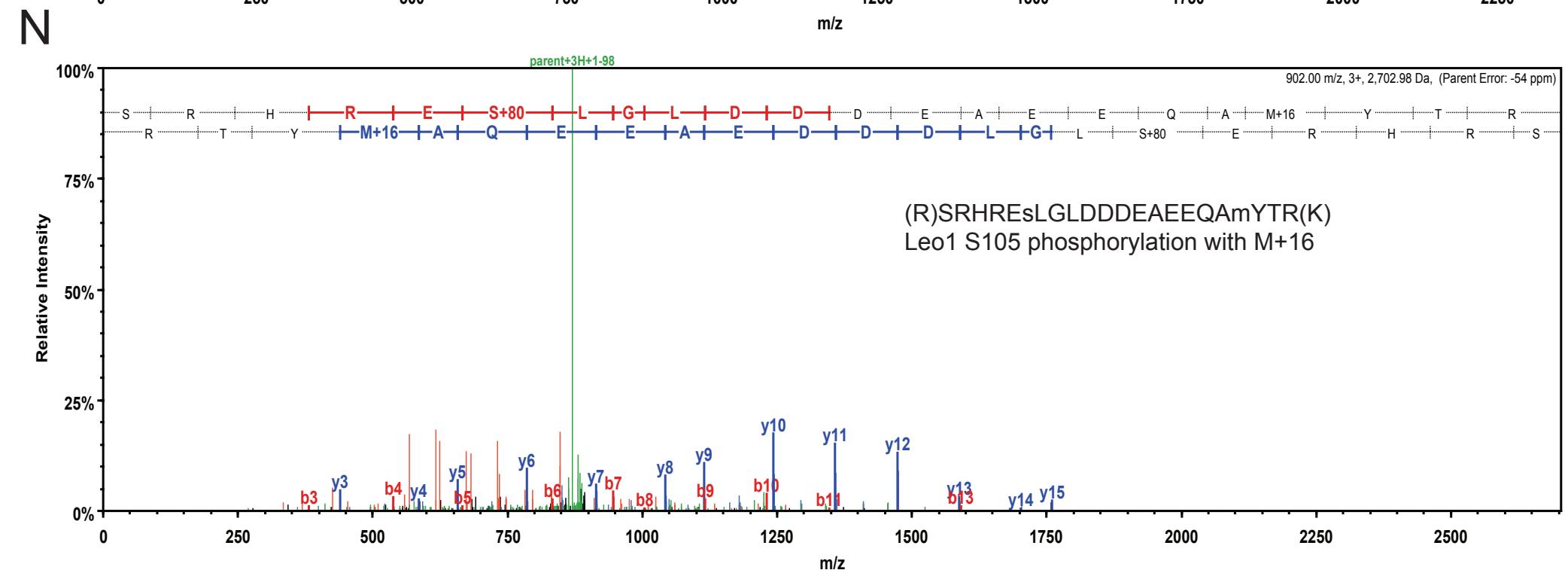
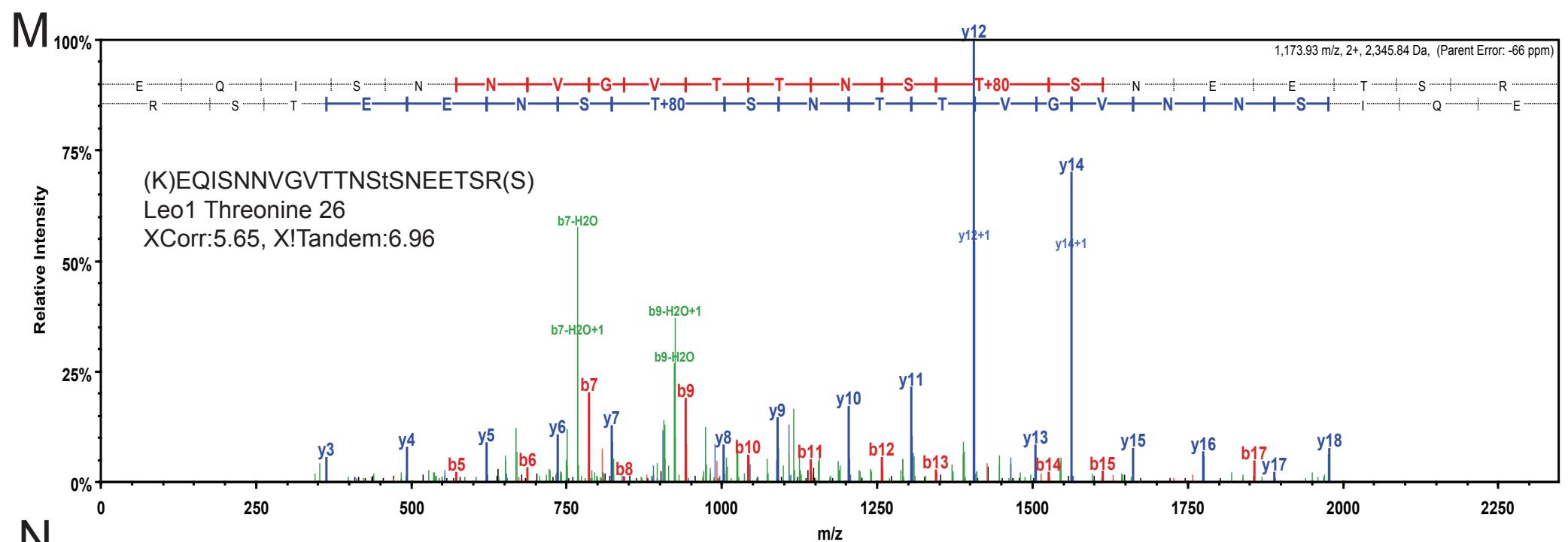


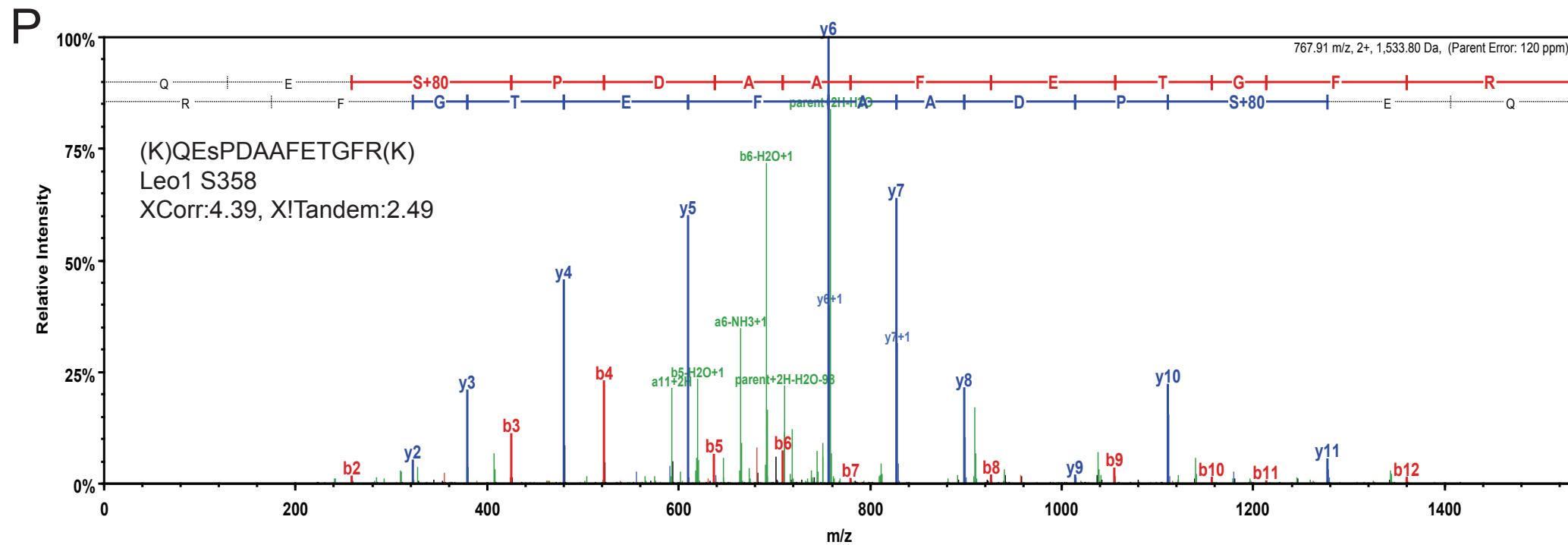
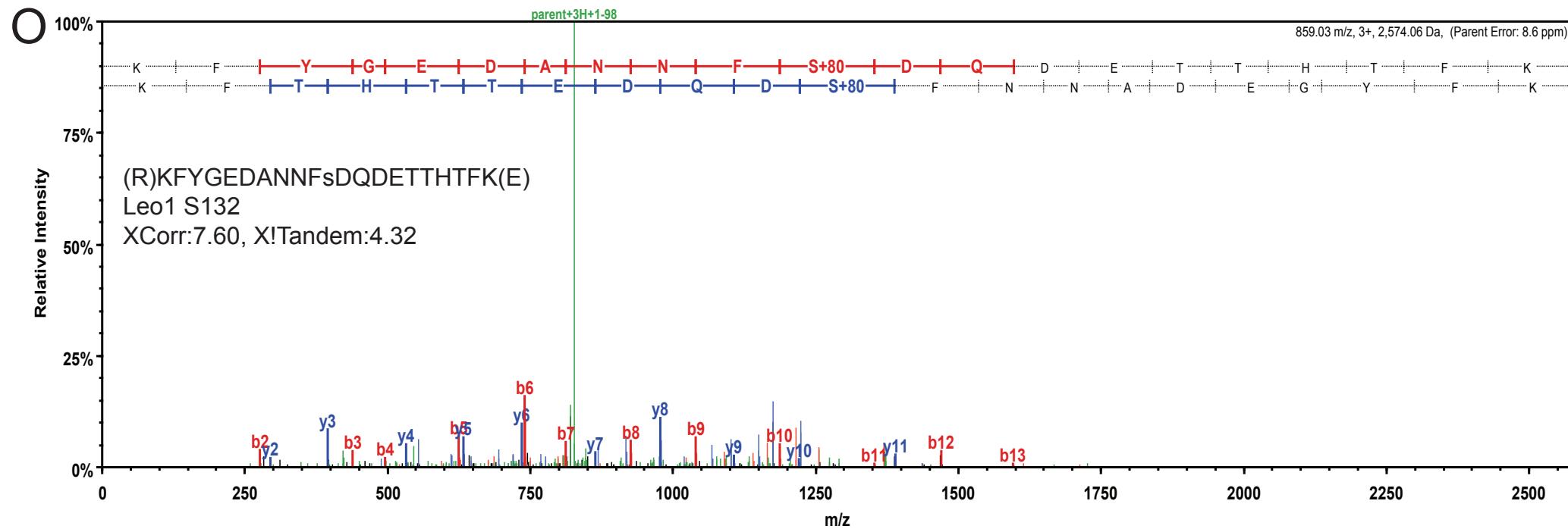


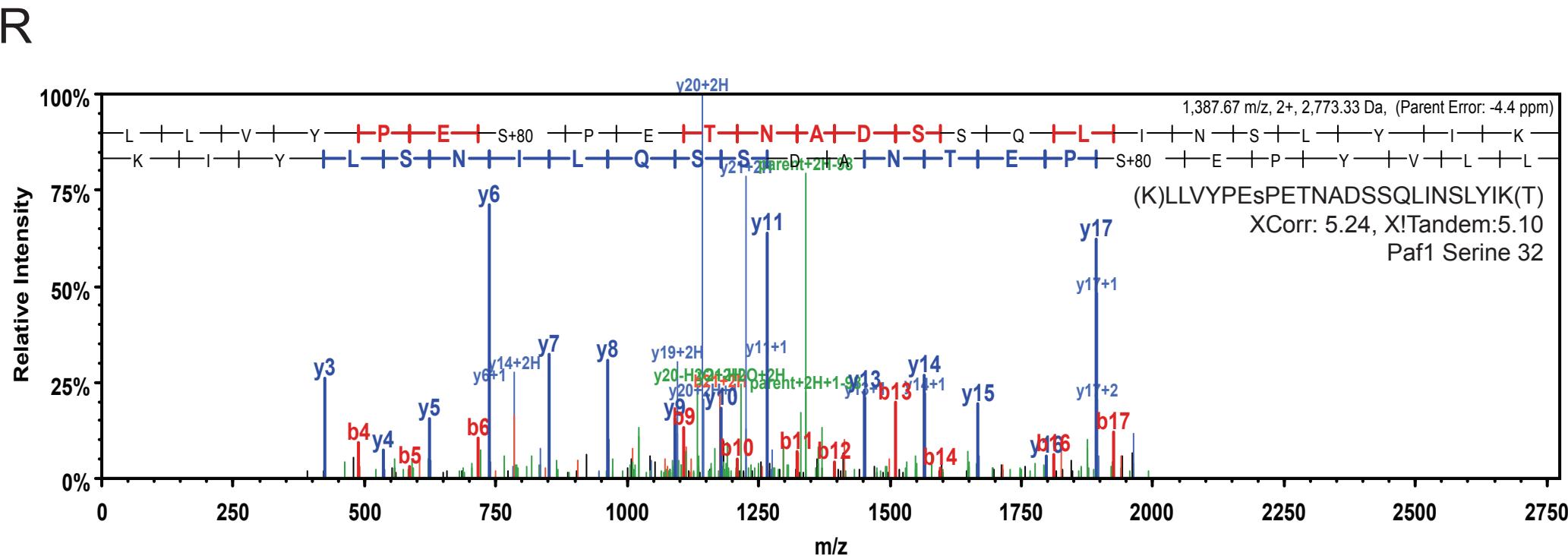
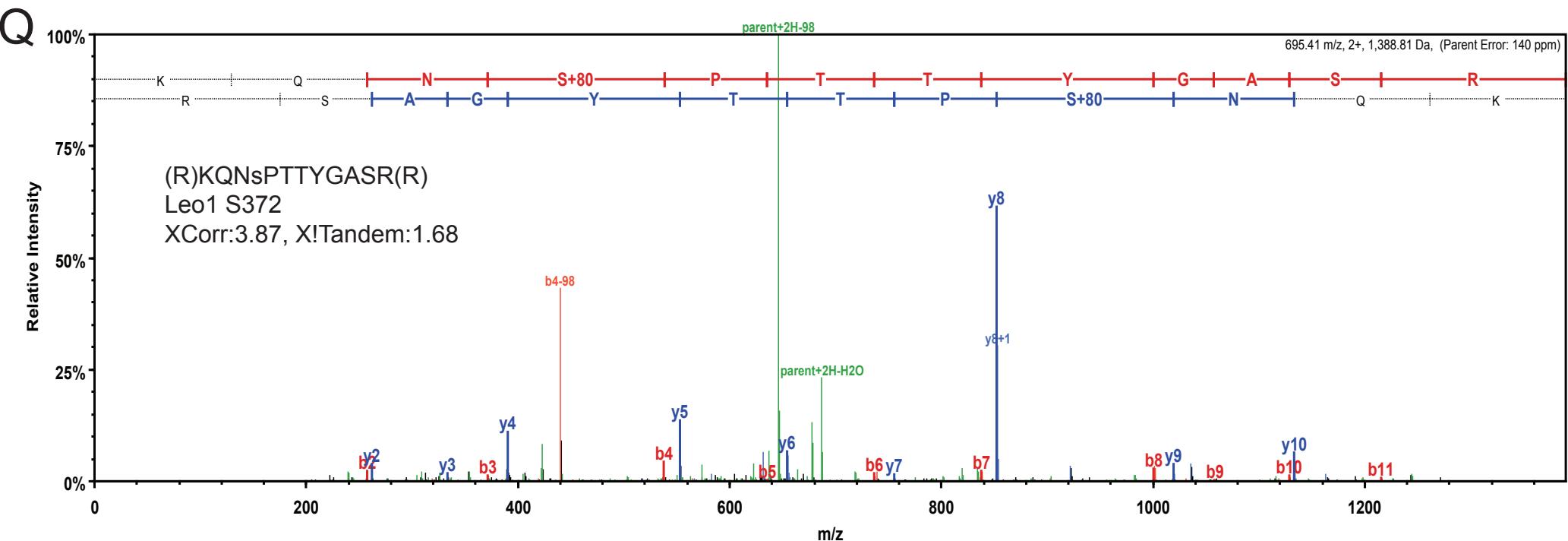


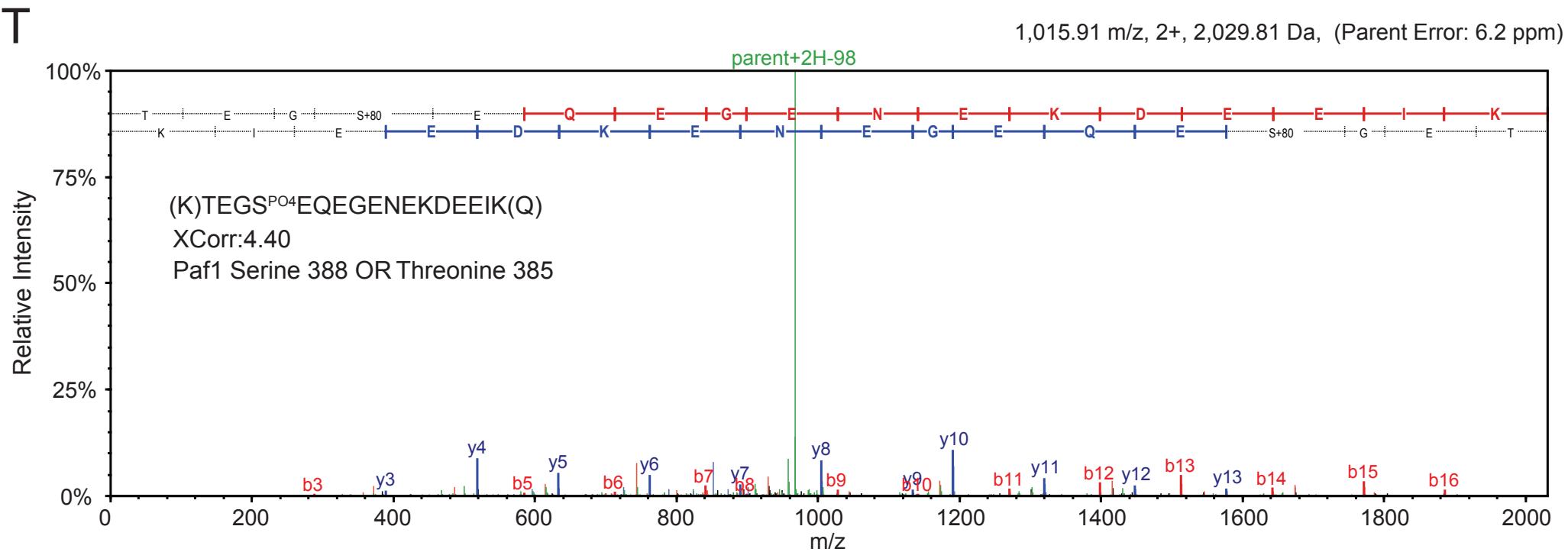
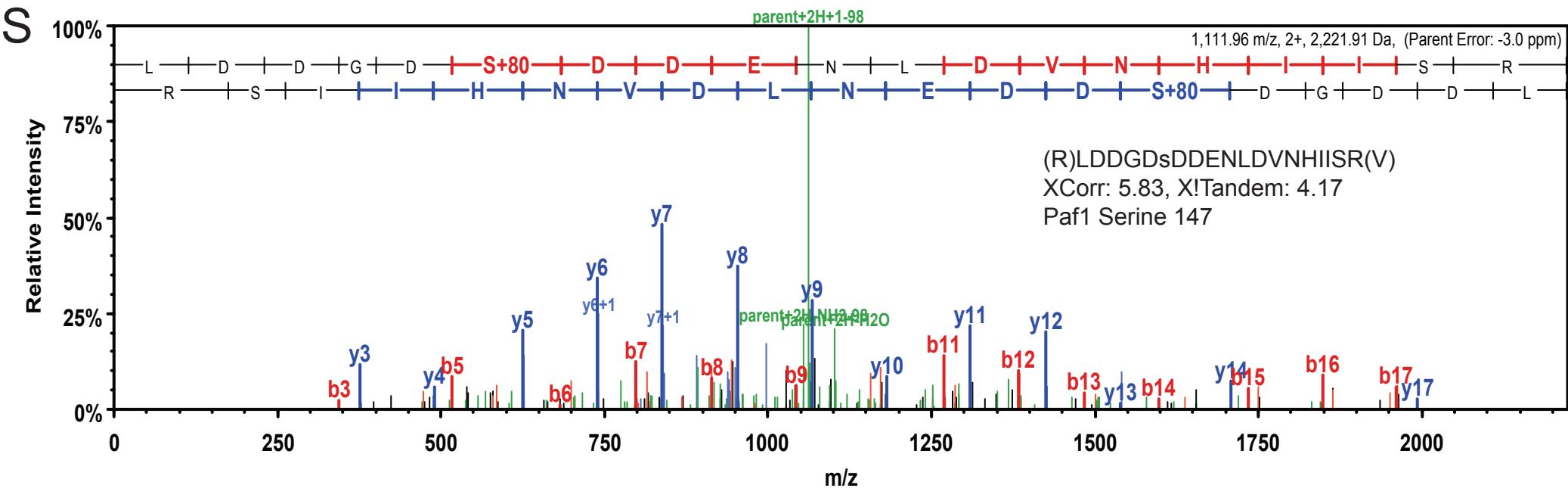




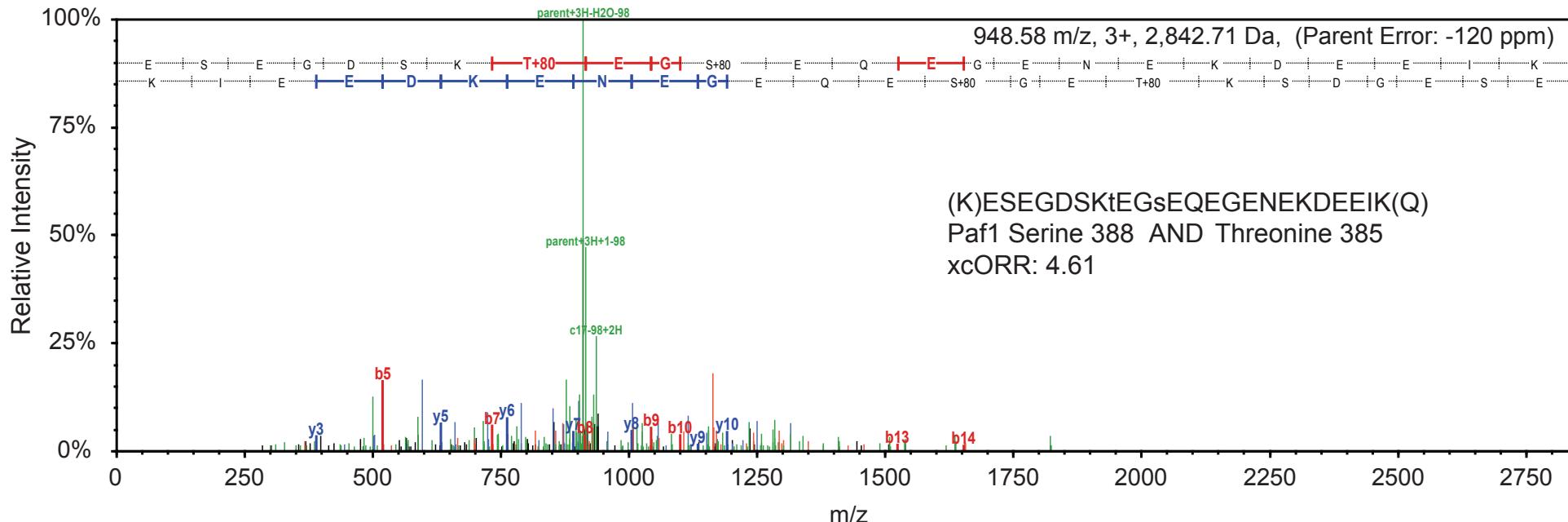




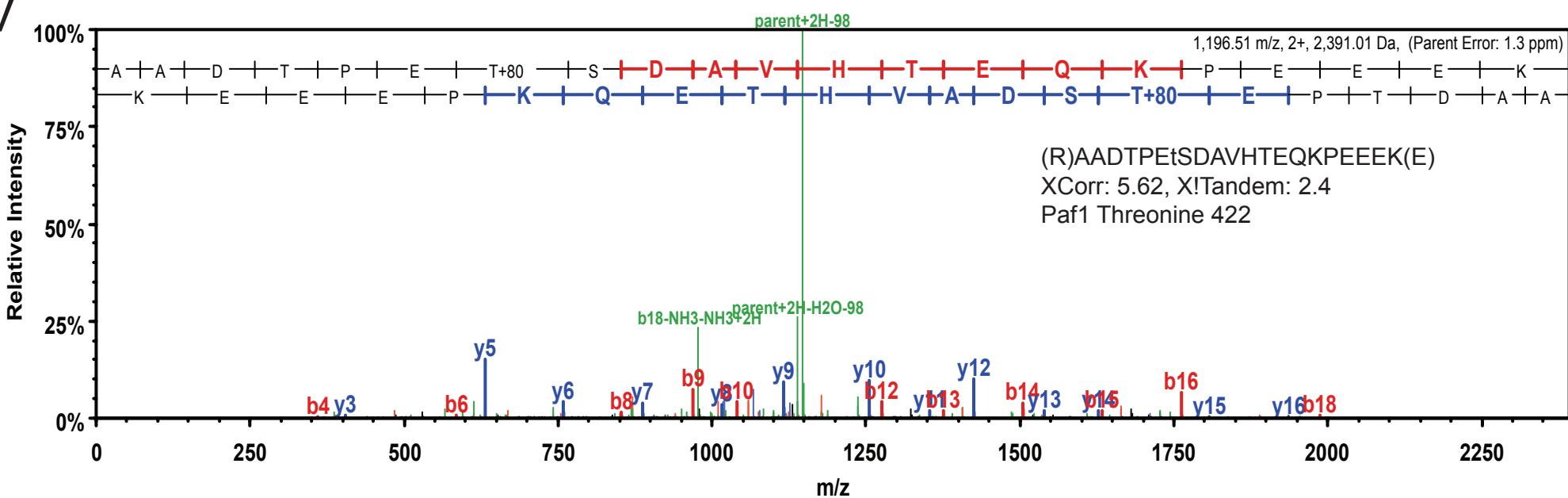


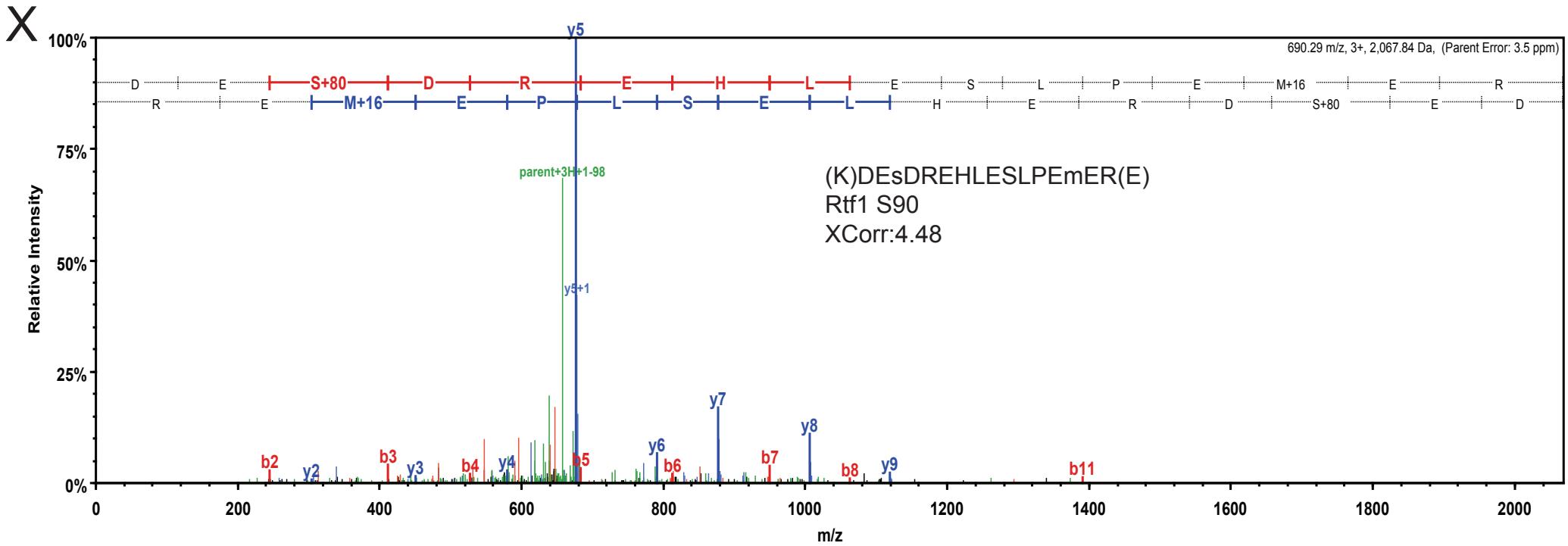
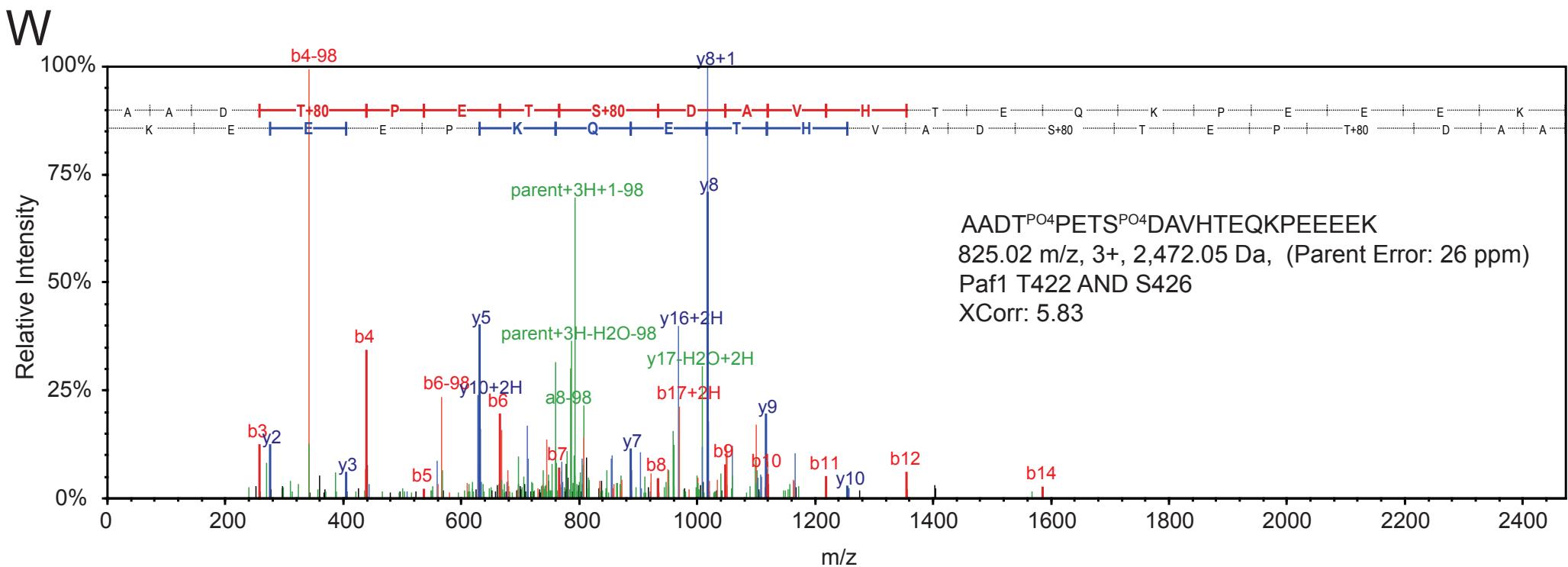


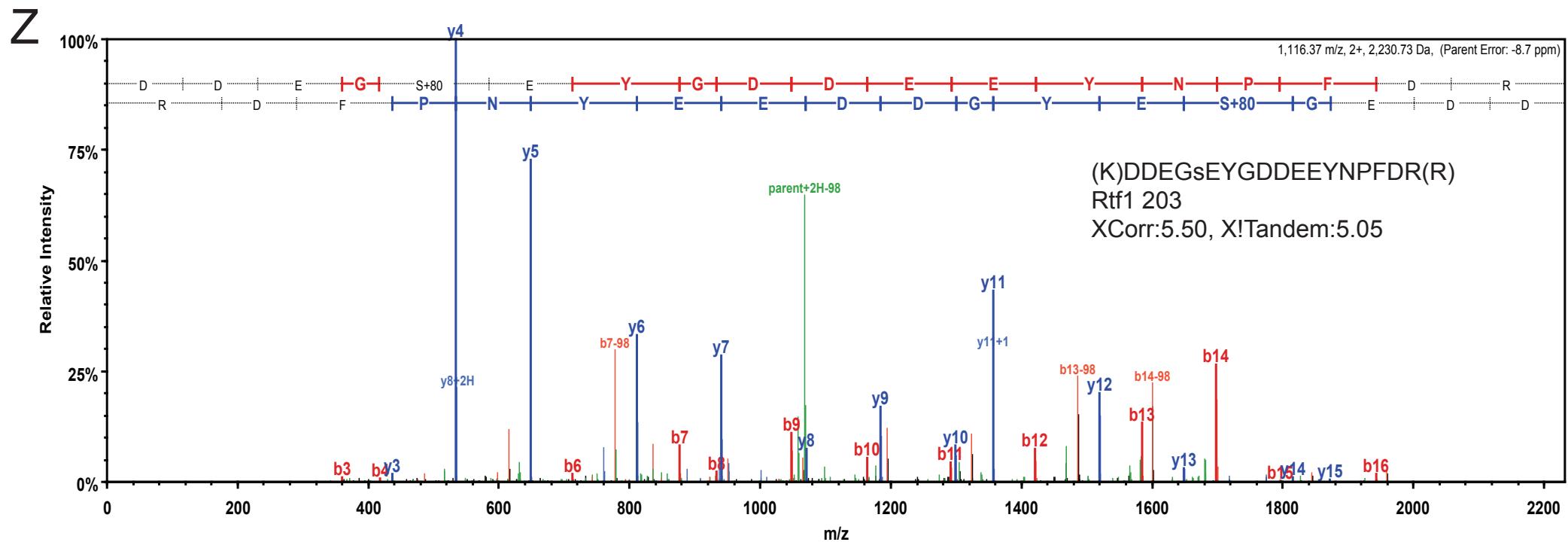
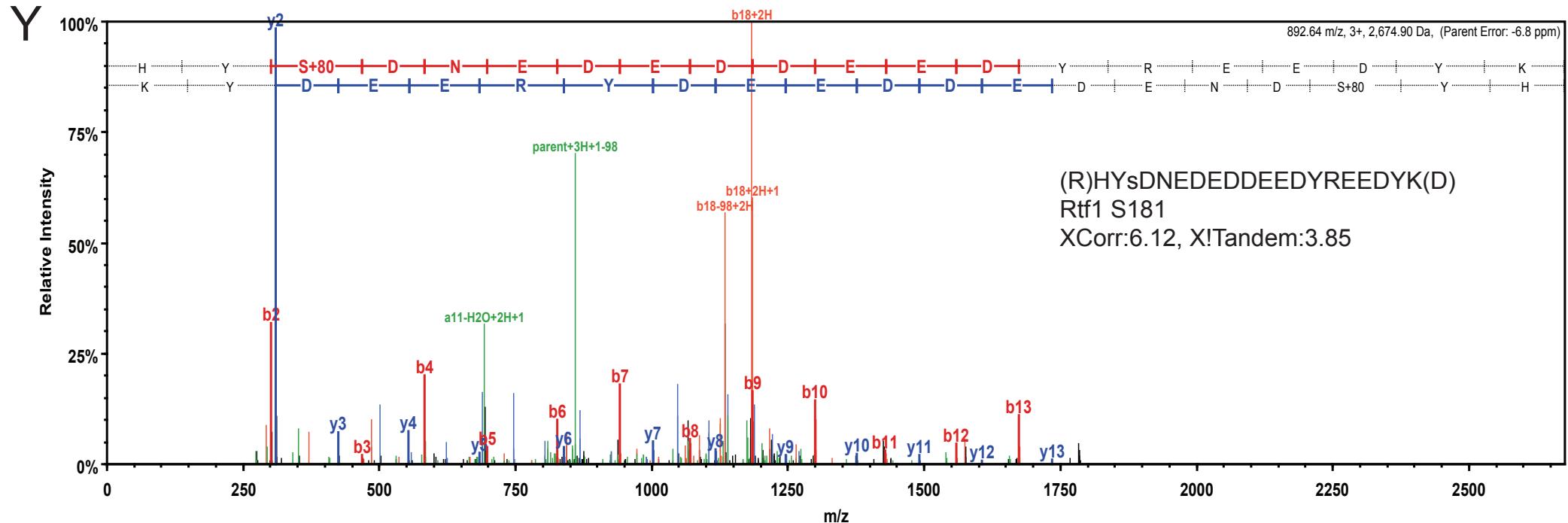
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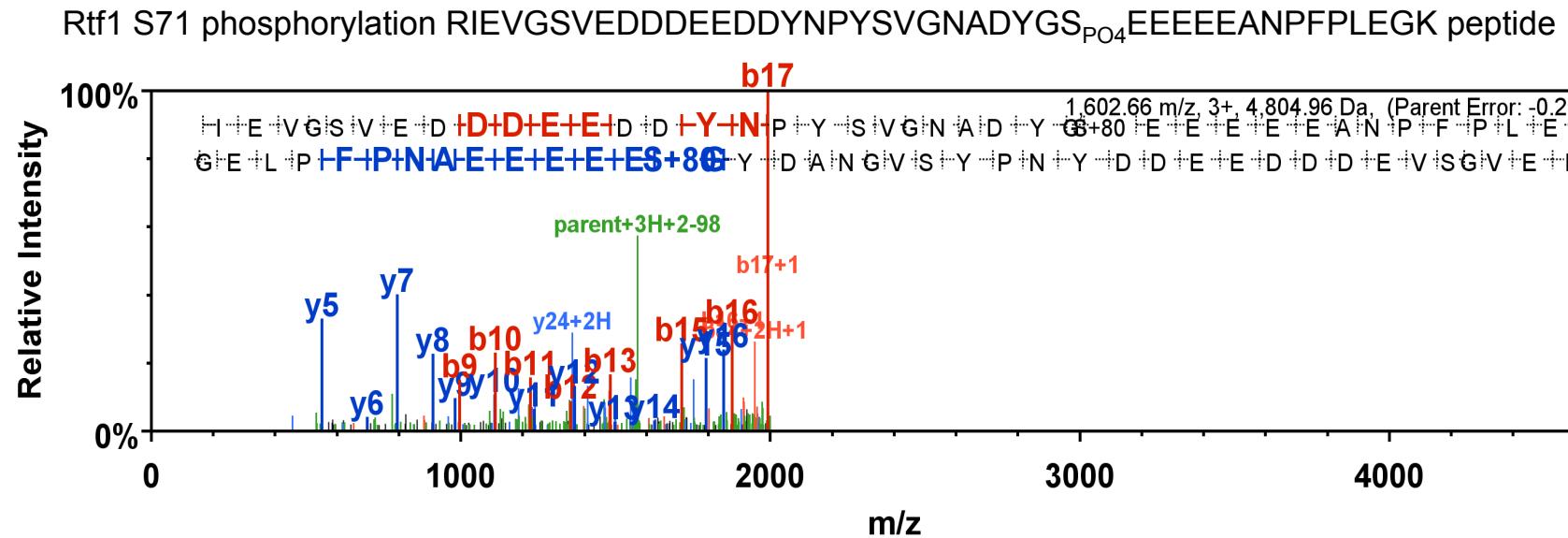
V







AA



*End of Figure S1*