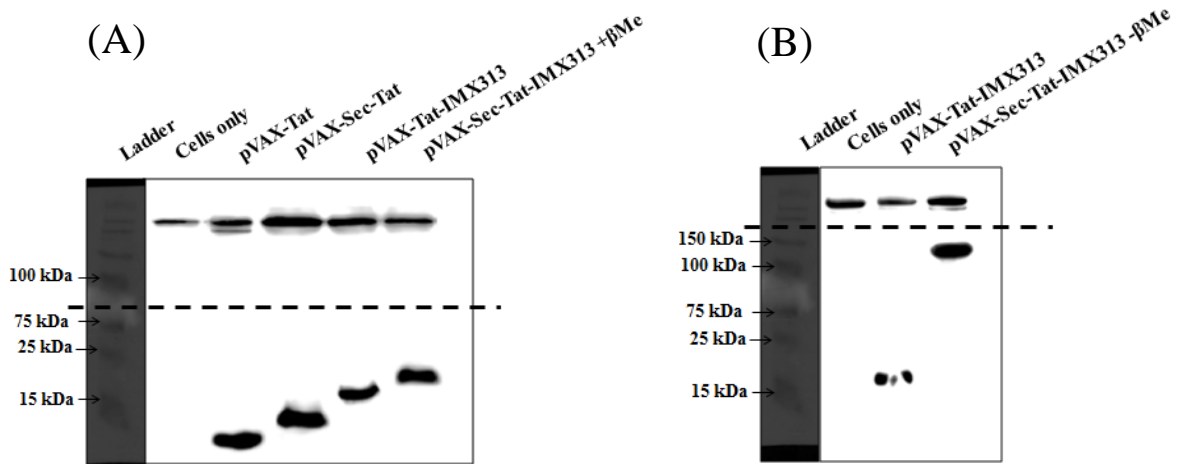


1 **A HIV-Tat/C4-binding protein chimera encoded by a DNA vaccine is highly**  
2 **immunogenic and contains acute EcoHIV infection in mice**

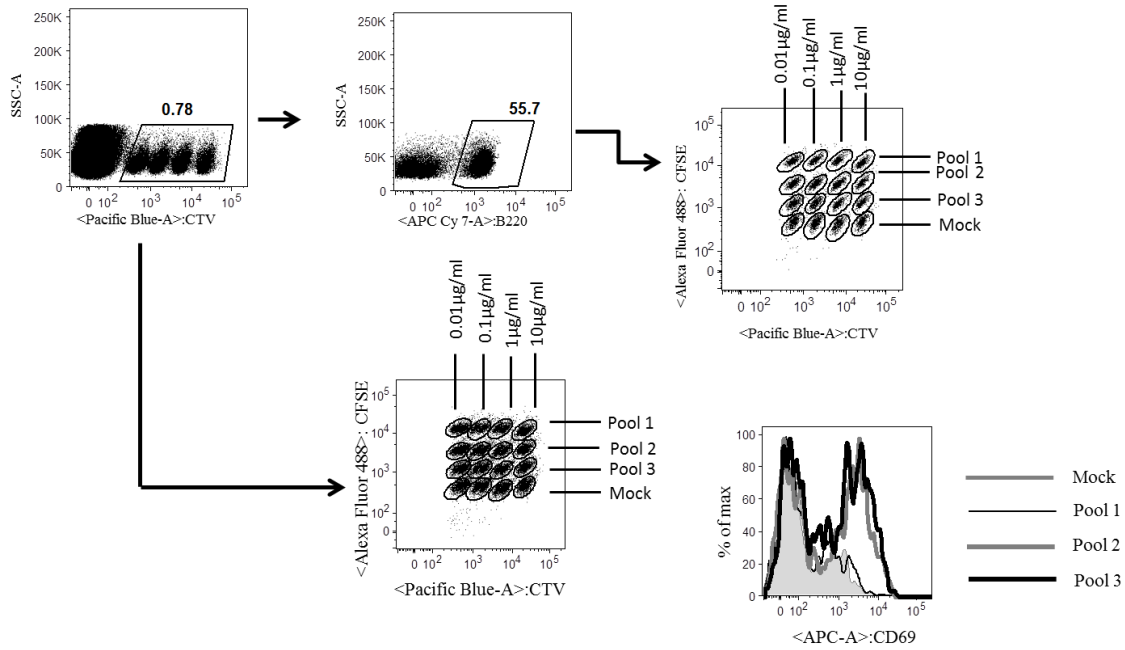
3 Khamis Tomusange<sup>1</sup>, Danushka Wijesundara<sup>1</sup>, Jason Gummow<sup>1</sup>, Tamsin Garrod<sup>2</sup>, YanruiLi<sup>1</sup>,  
4 Lachlan Gray<sup>3, 4</sup>, Melissa Churchill<sup>3</sup>, Branka Grubor-Bauk<sup>1</sup> and Eric J. Gowans<sup>1#</sup>



5  
6 **Supplementary Figure S1. Tat expression.** (A) reducing Western blot analysis of Tat in cell  
7 lysates from HEK293T cells transfected with plasmid DNA encoding the different forms of  
8 Tat (tracks 2-5) and (B) non-reducing Western blot analysis of Tat in supernatant fluids of  
9 HEK293T cells transfected with pVAX-Tat-IMX313 (track 2) or pVAX-sTat-IMX313 (track  
10 3) DNA. The dotted lines indicate the positions where the blots were cropped for figure IB  
11 and IC in the main manuscript.

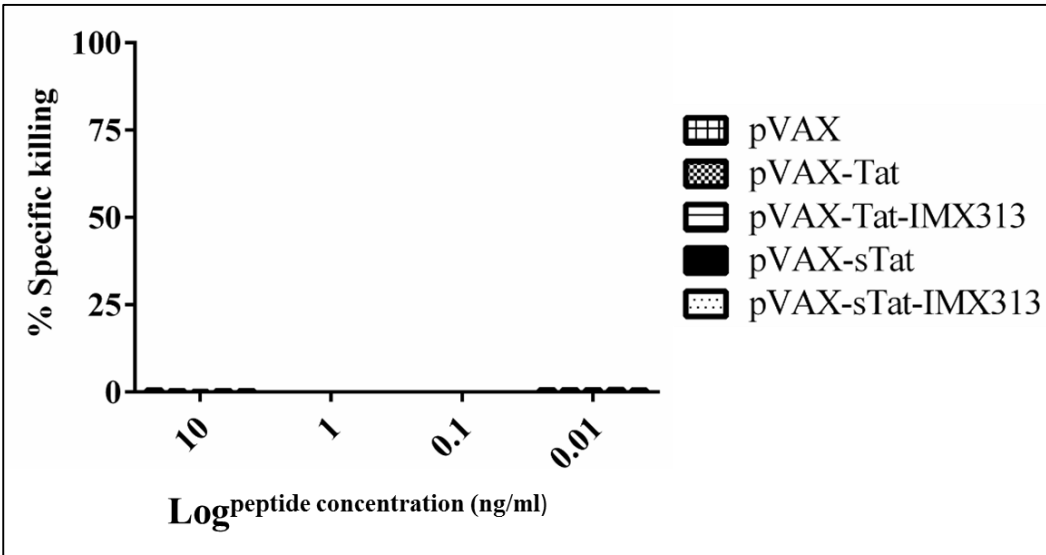
12

13 (A)



14

15 (B)



16

17 **Supplementary Figure S2. Gating strategy for the FTA analysis conducted in Fig 2B**  
 18 **and 2C.** RBC-depleted splenocytes from naïve mice were labelled with titrated  
 19 concentrations of CTV or CFSE dyes, and pulsed for 4h at 37 °C in 5% CO<sub>2</sub> with 3 Tat  
 20 peptide pools (pool 1-3) at concentrations ranging from 10µg/ml to 0.01µg/ml or media only

21 (mock control) before adoptive transfer into vaccinated mice. Representative plots depict the  
22 gating strategy for FTA analysis on doublet discriminated lymphocytes from a Tat DNA  
23 vaccinated animal (A). Total FTA targets were delineated based on CTV labelling and the  
24 total percentage of target cells in each of the peptide pulsed-clusters relative to the mock  
25 clusters was analysed to determine the magnitude of *in vivo* killing of FTA cells (B). To  
26 determine the magnitude of Th cell responses, B220<sup>+</sup> cells within the FTA were gated and  
27 CD69 up-regulation was determined on peptide pulsed B cells relative to mock B cells within  
28 the FTA.

## 29 **Supplementary materials**

### 30 **Primers used to amplify Tat inserts and quantify EcoHIV RNA levels**

#### 31 **a) For pVAX-Tat**

32 1- Tat-forward: GTGCTAGCGCCAGCATGGAACCCGTGGACCCCAGAC

33 2- Tat- reverse: ACGAATTCGTCCTCGGGGTCTGTCTCTGTC

#### 34 **b) For pVAX-sTat**

35 3- secr tat CMV fwd, forward: GAGAGAAAGCTT ATGGAACCCGTGGACCCC

36 4- secr tat CMV rev, reverse: GAGAGAGAATTCTCAGTCCTCGGGGTCTGT

#### 37 **c) For pVAX-sTat-IMX313**

38 5- secr tat CMV fwd: GAGAGAAAGCTTATGGAACCCGTGGACCCC

39 6-Wt\_Tat\_nostop reverse:GAGAGAGAATTTCGTCCTCGGGGTCTGTCTCTGTC

#### 40 **d) For pVAX-Tat-IMX313**

41 **7- Wt\_Tat\_no stop forward:**TCT CTC GCT AGC GCC ACC ATG GAA CCC GTG GAC

42 CCC AGA C

43 8- New Tat\_IMX313 con\_Rev-2: GAGAGACTCGAGTCACTCTTTGCTCAGGCCCTGC

#### 44 **e) For EcoHIV qPCR**

45 9-RPL13a FWD\_forward: GAGGTCGGGTGGAAGTACCA

46 10-RPL13a REV\_reverse: TGCATCTTGGCCTTTTCCTT

47 11-MLV FWD\_forward: TAGGGCCAAACCCCGTTCTG

48 12-MLV REV\_reverse: GCCGGTGGAAAGTTGGGTAGG