Supplementary information

Nanoscale imaging of untreated mammalian cells in a medium with low radiation damage using scanning electron-assisted dielectric microscopy

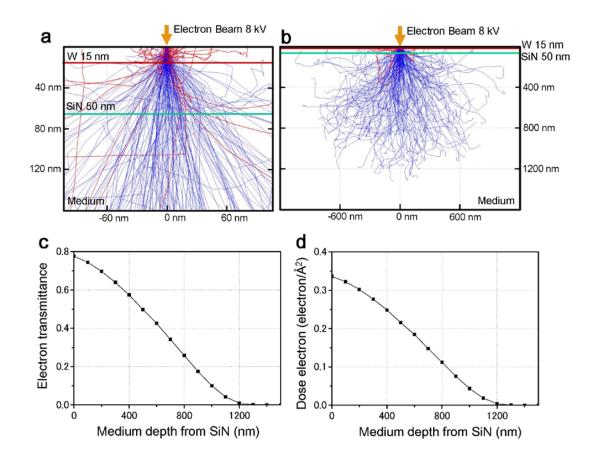
Tomoko Okada & Toshihiko Ogura*

Biomedical Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Central 2, Umezono, Tsukuba, Ibaraki 305-8568, Japan

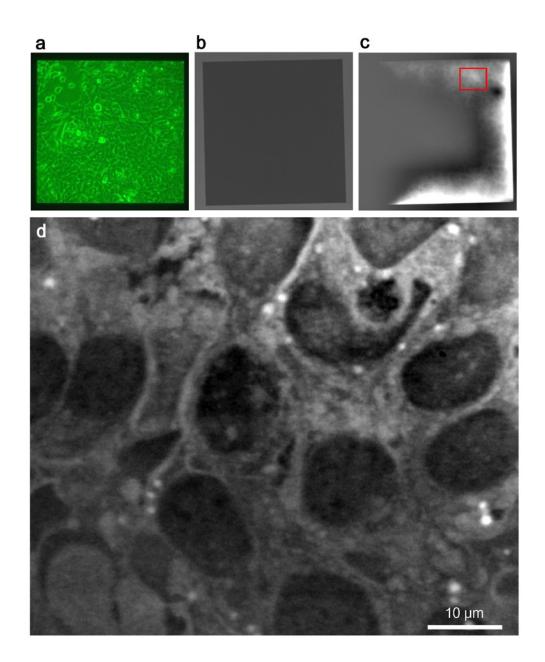
*Corresponding author: Toshihiko Ogura Biomedical Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Umezono 1-1-1, Tsukuba, Ibaraki 305-8568, Japan Tel.: +81-29-861-3408, Fax: +81-29-861-2677 E-mail: t-ogura@aist.go.jp

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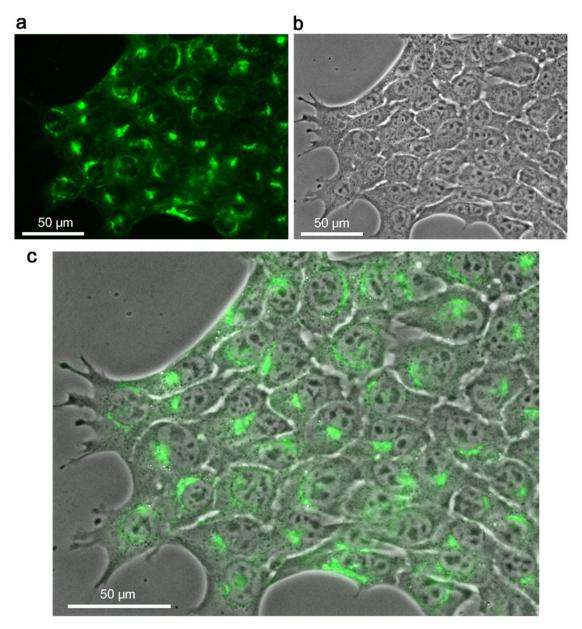
Supplementary Figs 1 to 5



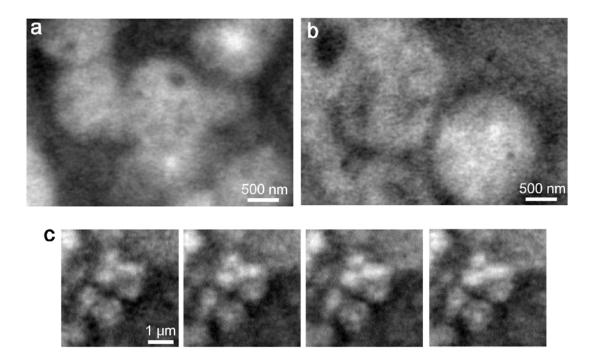
Supplementary Figure 1. Analysis of the electron trajectory in W-coated SiN film and medium in a Monte Carlo (MC) simulation. a, MC simulation analysis of W-coated SiN film, using CASINO ver. 2.42 (ref. 22); the electron trajectory area in a 15-nm W layer on 50-nm SiN film is shown. The respective densities of the W layer and SiN film were 19.3 g/cm³ and 3.12 g/cm³, and the respective thicknesses were 15 nm and 50 nm. The simulation parameters were set at 100,000 electrons, 8 kV accelerating voltage, and 3 nm EB spot diameter. **b**, Electron trajectories in W-coated SiN film with medium. Irradiated electrons were dispersed to a depth of almost 1 µm. **c**, Transmitted irradiated electrons in medium were calculated at each medium depth from the SiN film in a MC simulation. Electron transmittance just below the SiN film was 0.774; this value gradually decreased and reached 0 at a depth of 1.2 µm. **d**, The electron dose in medium was calculated at each EM condition and electron transmittance shown in (**c**), with a current of 10 pA, scanning area of 12 µm × 9.6 µm, and scanning time of 80 s. The electron dose just below the SiN film was 0.334 electron/Å²; this dose gradually decreased and reached 0 electron/Å² at a depth of 1.2 µm.



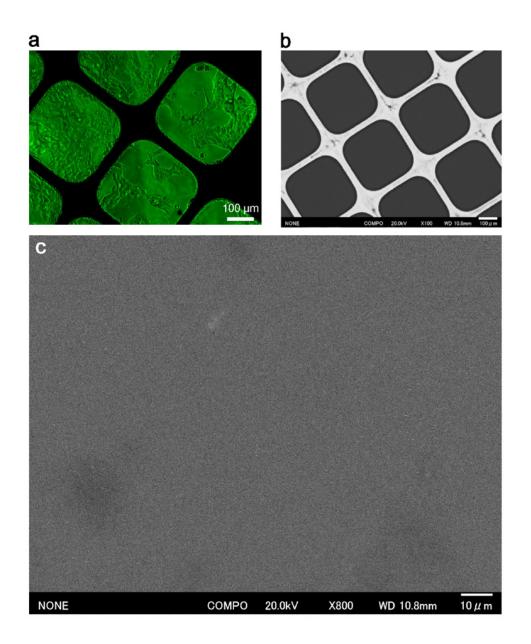
Supplementary Figure 2. Optical phase and dielectric observation images of the same cells. a, Optical microscopy image of cultured 4T1E/M3 cells in a confluent monolayer on SiN film in a dish holder. A SiN film size is 0.4×0.4 mm. b, Secondary electron detector-generated images of the same area as in (a). The image shows the SiN surface without cellular structures. c, Simultaneously observed dielectric image of the area in (b). The electric field signal was detected by the area of SiN film. d, High-magnification image of the red box in (c). Dielectric images clearly revealed cancer cells in the SiN square windows. The scale bar represents 10 μ m in (d).



Supplementary Figure 3. Optical phase and fluorescence observation images of cells stained with Golgi/ER-specific dye. a, Fluorescence image of Golgi/ER-stained cells obtained from an optical microscope with a fluorescence filter. b, Optical phase contrast image of stained cultured cells obtained with an optical microscope. c, The merged picture of a Golgi/ER fluorescence image (a) and a phase contrast image (b). The ER and Golgi apparatus are localized near the cell nucleus. The scale bars represent 50 μ m in (a) – (c).



Supplementary Figure 4. Enlarged images of vesicle regions using SE-ADM. a, Enlarged images of the vesicle regions highlighted in red boxes in Figure 3b. b, Enlarged images of the vesicle regions highlighted in red boxes in Figure 3d. c. Enlarged images of a vesicle that exhibited contrast changes throughout the four observations indicated by blue arrows in Figure 4a–d. The scale bars represent 500 nm in (a) and (b) and 1 μ m in (c).



Supplementary Figure 5. FE-SEM observation of unfixed and unstained cells in a traditional atmospheric holder (Wet SEM). **a**, Phase-contrast optical image of a culture dish in the Wet SEM capsule (ref. 10). Untreated cells in medium are clearly visible in the wet capsule membrane by optical phase microscopy. **b**, Low-magnification FE-SEM image of the same cultured cells in the wet capsule shown in (**a**) with electron backscattering. The grid supporting the membrane is visible in this image. However, unstained cells cannot be detected. **c**, High-magnification image ($800 \times$ magnification) with electron backscattering. The cell structures cannot be detected. The scale bars represent 100 µm in (**a**) and (**b**) and 10 µm in (**c**).