

**Supplementary Figure 1 Map of AGM (*Chlorocebus sabaeus*) immunoglobulin heavy chain locus on chromosome (CHR) 24.** Representation of the identified functional and potentially functional, open reading frames, and pseudogenes of AGM  $V_H$ ,  $D_H$  and  $J_H$  germline segments along with the Ig constant region genes of Ig M, D, G, A and E located on CHR24. Four additional functional AGM  $V_H$  gene segments that were identified from the unassembled contigs are not showed here (see Supplementary Data 1).

a

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                                     50                                     100
                                     |                                     |
HuIgG1  ASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSQVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQTYICNVNHKPSNTKVKDKKVE-
RhIgG1  .....R...ES.....S.....V.....R..I
AGMIgG1 .....R..ES.....S.....T.....R..I
HuIgG2 .....C.R..ES.....NF.....T..D.....T..R
RhIgG2 .....SC.R..QS.....Q.....V..V.E.....T.GL
AGMIgG2 -----
HuIgG3 .....C.R.....T.....R..L
HuIgG4 .....C.R..ES.....K..T..D.....R..S
RhIgG3 .....SC.R..QS.....V..V.E.....R..F
RhIgG4 .....S..R..ES.....V..V.E.....R..F

                                     200
                                     |
HuIgG1  -----FKSCDKTHTCPCPAPELLGGPSVFLFPPPKPKDTLMISRTPEVTCVVVDVSHED
RhIgG1  KTCG-----GGG.PP.....Q..
AGMIgG1 KTCG-----GGG.PP.....N.....Q..
HuIgG2 -----C---VE-----PVA.....
RhIgG2 PCR-----S.....Q..E
AGMIgG2 -----Q..E
HuIgG3 KTPLGDTTHTCPRCPEPKSCDTPPPCPRCPEPKSCDTPPPCPRCPE...TPPP..R.....
HuIgG4 -----YG---PP..S...F.....Q..
RhIgG3 TPPC-----GDTT-PP.....Q..
RhIgG4 TP-----P.....Q..

                                     250                                     300
                                     |                                     |
HuIgG1  PEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQVSLTCLVK
RhIgG1  .D.....N.A..H.Q...T.....T.....Q..D.....E.....
AGMIgG1 ..Q.....Q.....T.....R.....T.....Q..D.....A..E.....
HuIgG2 ..Q.....F..F.....V.....G.....T.....E.M.....
RhIgG2 .D.....Q.....F.....T.....T.....RQ..V..T.....P.E.....
AGMIgG2 ..Q.....Q.....N.....T.....T.....RQ..V..T.....P.E.....I.
HuIgG3 ..Q..K.....F.....T.....T.....E.M.....
HuIgG4 ..Q.....F.....G..SS.....QE.M.....
RhIgG3 ..Q.....A..H.Q...R.F.....T.....T.....G.....I..PQE.....T
RhIgG4 ..Q.....Q.....R.F.....T.....T.....G.....I..PQE.....T

                                     350
                                     |
HuIgG1  GFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGK
RhIgG1  .....V.....S.....T.....Y.....
AGMIgG1 .....V.....T.....Y.....T.....V.....
HuIgG2 .....M.....
RhIgG2 .....V..A.....T.....Y.....T.....
AGMIgG2 .....V.....R.....T.....Y.....T.....V.....
HuIgG3 .....S.....N..M.....I.....RF.....
HuIgG4 .....R.....E.....L.....
RhIgG3 .....T.....Y.....T.....
RhIgG4 .....T.....YL.....N.....P..I..T.....
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b

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                    50                                     100
HuIgA1      SPTSPKVFPLSLCSTQPDGNVVIACLVQGFPPQEPPLSVTWSESGQGVARNFPSPQDASGDLYTTSSQLTLPATQCLAGKSVTCHVKHYTNPSQDVTVPC
HuIgA2      .....D.PQ.....V.....N.....PD.....
RMiGA_AI    ...K.....EG.S~.V.....K.N..K.A..VI..R...G.....A.P.SE.....E...R...A...
RMiGA_AII   ..R.....EG.S~.V.....N...K.A..VI..R...G.....A.P.SE.....E...R...G...
RMiGA_BI    ...R.....N...S~.V.....N...K.A..VI..R...G.....A.P.SE.....E...A...
RMiGA_BII   ..R.....S~.....N...K.A..VI..R...G.....A.P.SE.....E...A...
RMiGA_CI    ...R.....N...S~.V.....N...K.AD.VI..R...G.....A.P.SE.....E...A...
RMiGA_CII   ..R.....EG.S~.V.....N...K.A..VI..R...G.....A.P.SE.....E...AR..A...
RMiGA_DI    ..R.....EG.S~.V.....N...K.A..VI..R...G.....A.P.SE.....E...R...G...
RMiGA_DII   ..R.....EG.S~.V.....N...K.A..VI..R...G.....A.P.SE.....E...AR..A...
RMiGA_RJZ7  .....KG.S...V.....N..NK.A..VI.....G.....P.TE.....E.....A.R...
RMiGA_RUA8  .....N...S~.V.....N...K.AD.VI..R...G.....A.P.SE.....E...A...
AGMiGA     .....EG.S...V.....N...KN.E~~VI...A..G.....A.P.SE...R.E.....A...

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                    150                                     200
Hinge region
HuIgA1      PVPSTPPPTSPSTPPPTSPSPSCCHPRLSLHRPALEDLLLGSEANLTCTLTGLRDASGVFTTWTTPSSGKSAVQGPPEPRLCGCYSVSSVLPGCAEPWNHGKT
HuIgA2      .....P.P.....A.....Q.....E...
RMiGA_AI    R~VPP.KC.-----LK.DK...R.....P.A.....N...QS.H.P.....NRV...
RMiGA_AII   R~VPP.NCR-----L.DK...R.....P.A.A.....N...QS.H.P.....NRV...
RMiGA_BI    VP.I..C.-----E...R.....K.P.A.....N...QS.K.P.....NRE...
RMiGA_BII   V~QPK.CL-----DK...R.....P.A.....N...QS.H.P.....NRV...
RMiGA_CI    VP.I..C.-----E...R.....P.A.....N...QS.K.P.....NRE...
RMiGA_CII   V~SQ.K.CL-----DE...R.....P.A.....N...QS.K.P.....NRV...
RMiGA_DI    R~VPP.NCR-----L.DK...R.....P.A.A.....N...QS.H.P.....NRV...
RMiGA_DII   V~SQ.K.CL-----DE...R.....P.A.....N...QS.K.P.....NRV...
RMiGA_RJZ7  RD.QPK.CL-----DK.Q...R.....P.A.A.....N...QS.H.P.....NRE...
RMiGA_RUA8  VP.I..C.-----E...R.....K.P.A.....N...QS.K.P.....NRE...
AGMiGA     V~.PKTCF-----DE...R.....P.A.....N...QS.H.P.....N.V...

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                    250                                     300
HuIgA1      FTCTAAYPESKTPLTATLSKSGNTFRPEVHLLPPPSEELANLNLVTLTCLARGFSPKDVLRWLQGSQELPREKYLTVASRQEPSQGTTFFAVTSILRVA
HuIgA2      .....H.L.....NIT.....
RMiGA_AI    ...EH.LE.Q...I.....E.....K.TEQ..D...E.K.....
RMiGA_AII   ..N..NH.L...I.....E.....K.TEQV..D...E.K.....
RMiGA_BI    ...EH.LE...I.....E.....K.TEQ..D...E.K.....
RMiGA_BII   ..EH.QL...I.....E.....K.TEQ..D...E.K.....
RMiGA_CI    ...NH.LE...I.....E.....K.TEQ..D...E.K.....
RMiGA_CII   ..K.EH.LE...I.....E.....K.TEQ..D...E.K.....
RMiGA_DI    ..N..NH.L...I.....E.....K.TEQV..D...E.K.....
RMiGA_DII   ..K.EH.LE...I.....E.....K.TEQ..D...E.K.....
RMiGA_RJZ7  ..NH.L...I.....E.....K.TEQ..D...E.K.....
RMiGA_RUA8  ...NH.LE...I.....E.....K.TEQ..D...E.K.....
AGMiGA     ...NH.LE.Q...IT.....E.....K.TEQ..D...E.K...P.....

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                    350
HuIgA1      AEDWKKGDTFSCMVGHEALPLAFTQKTIDRLAGKPTHVNVSVVMAEVDGTCY
HuIgA2      .....
RMiGA_AI    .....
RMiGA_AII   .....
RMiGA_BI    .....
RMiGA_BII   .....
RMiGA_CI    .....
RMiGA_CII   .....
RMiGA_DI    .....
RMiGA_DII   .....
RMiGA_RJZ7  .....
RMiGA_RUA8  .....
AGMiGA     .....

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**c**

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                                     50                                     100
                                     |                                     |
HuIgD  APTKAPDVFPIISGCRHPKDNSPVVLAACLITGYHPTSVTVTWYMGTSQPQRTFPEIQRRDSYYMTSSQLS--TPLQQWRQGEYKCVVQHTASKSKKE-I
RhIgD  -----A.....A.QL.....N.K.....HL...I.N.IM...TE.EG.-.T....APTP..S.QH.....T.K..P.DTS..KT
AGMIgD  .S.....-T.A.EL.....N.K.....HL...I.N.IM...TDTQG.-.T....V-TP..S.RH.....T.K..P.NTN..KT

                                     150                                     200
                                     |                                     |
HuIgD  FRWPESPKAQASSVPTAQPOAEGSLAKATTAPATTRNTGRGGEKKKEKEEQEERETKTPECPHSHTQPLGVYLLTPAVQDLWLRDKATFTCFVVGSDL
RhIgD  .....YP...V.....G.S...RP...N.....--RE.EDE.EK..Q.G.....P..L...FQ..V.....
AGMIgD  .H.....YP..S.....G.S...RS...N.....--RE.ED..EK..Q.G.....R.....I...P..L...F.....

                                     250                                     300
                                     |                                     |
HuIgD  KDAHLTWEVAGKVPVTGGVEEGLLERHSNGSQSQRSLTLPRSLWNAGTSVCTLNHPSLPPQRLMALREPAQAQPVKLSLNLLASSDPPEAASWLLCEVS
RhIgD  Q...S.....K..M...P..Q.....A.....S.K.....R.....
AGMIgD  Q...S.....K.DM...P..Q.....A.....S.K.....R.....

                                     250
                                     |
HuIgD  GFSPFNILLMWLEDQREVENTSGFAPARPPPQPRSTTFWAWSVLRVPAPPSQPATYTCVVSHEDSRTLLNASRSLEVSIVTDHGPMK
RhIgD  D.....N.....W..TTH.T...G..M.....G.T.....
AGMIgD  D.....S.W..TTH.T...G..M.....G.T.....V.YLTD RGP.KX-----
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**d**

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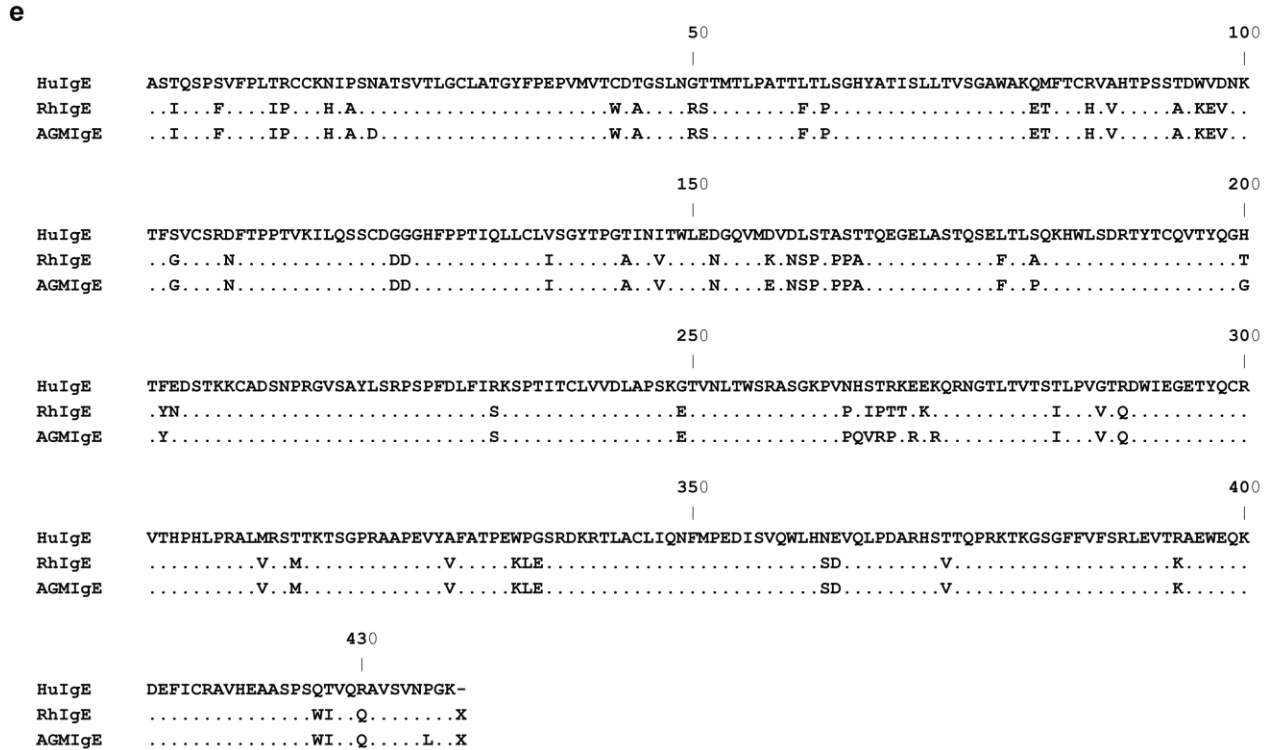
                                     50                                     100
                                     |                                     |
HuIgM  GSASAPTLFPLVSCENSPTSSTSSVAVGCLAQDFLPSITLSWKYKNSDIS--TRGFPSVLRGGKYAATSQVLLPSKDVMOGTDEHVVCVKVQHPNGNKEK
RhIgM  .....A.L..NE.....F..F...N..KGVW.....A.....Q
AGMIgM  .....ALL..NK.....F..F...KGVW.....D..V.....A.T.....Q

                                     150                                     200
                                     |                                     |
HuIgM  NVPLPVIAELPPKVSFVPPRDGFFGNPRKSKLICQATGFSPRQIQVSWLREGKQVSGVTTDQVQAEAKESGPTTYKVTSTLTIKESDWLGQSMFTCRV
RhIgM  .....L..R..N.....V...E.....E.....I...R.E.....F.....VS.R...S..V....
AGMIgM  .....V..Q..N...I...V...E.....E.....I...E.....F.....VS.R...S..V....

                                     250                                     300
                                     |                                     |
HuIgM  DHRGLTFQQNASSMCPDQDTAIRVFAIPPSFASIFLTKSTKLTCLVTDLTTYSVTISWTRQNGEAVKTHTNISESHPNATFSAVGEASICEDDWNNGE
RhIgM  .....K.V..V.G.NP.....A.....T.....L.....G.....
AGMIgM  .....K.V..V.G.NP.I.....A.....L.....G.....

                                     350                                     400
                                     |                                     |
HuIgM  RFTCTVTHDPLSPLKQTIISRPKGVALHRPDVYLLPPAREQLNLRRESATITCLVTGFSPADVFVQWMQRGQPLSPEKYVTSAPMPEPQAPGRYFAHSILT
RhIgM  ..R.....M.....I.....I.....
AGMIgM  W.R.....PM.....I.....

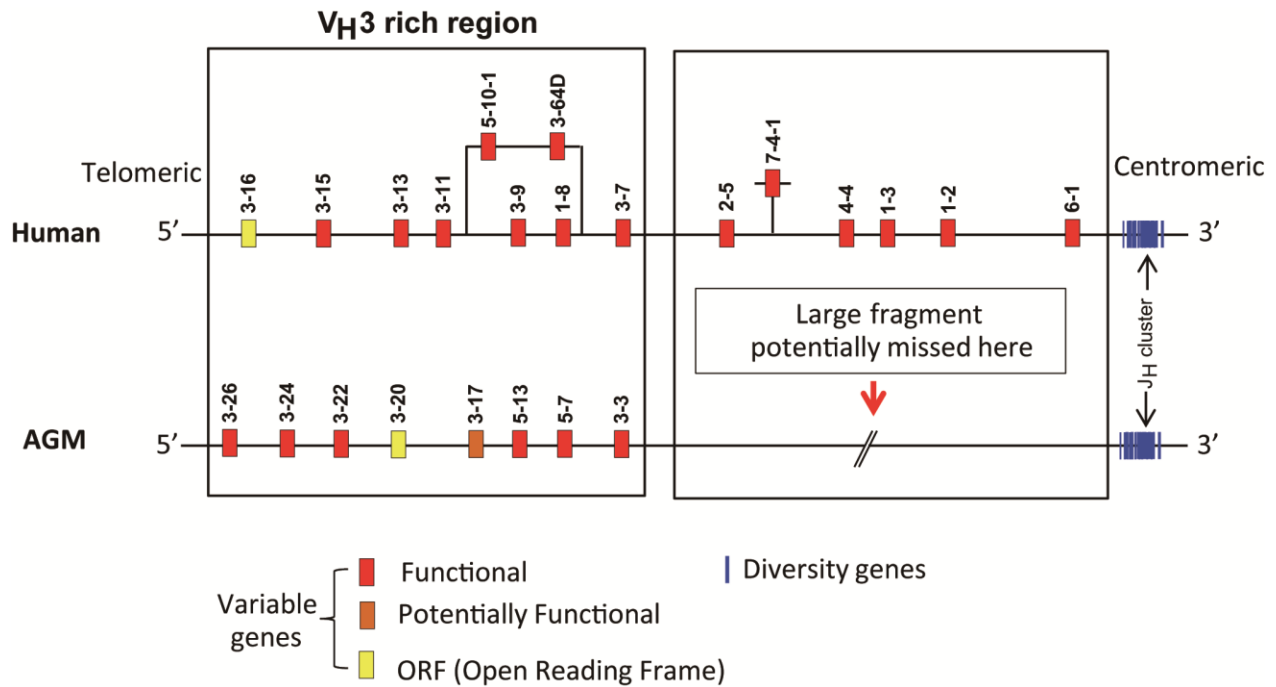
                                     430
                                     |
HuIgM  VSEEEWNTGETYTC-VAHEALPNRVTERTVDKST
RhIgM  ....D.....V.....
AGMIgM  ....D.....V.....
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**Supplementary Figure 2 Sequence alignment of AGM, RM, and human Ig constant regions.** Panel **a** shows sequence alignment of IgGs from the three species. The underlined text shows the hinge region of IgGs of three species. Sequences highlighted in yellow represent the identical residues in the hinge region between AGM IgG1 and RM IgG1. Sequences in red show the shared unique mutations in the C<sub>H</sub>3 domain of AGM IgG2 and RM IgG2. Panel **b** shows sequence alignment of IgAs of three species. The hinge region of human IgA1 is highlighted in red. Ten RM intra-species heterogenous IgA alleles were used in the alignment. The sequence alignments of IgD (**c**), IgM (**d**) and IgE (**e**) of three species are also represented.

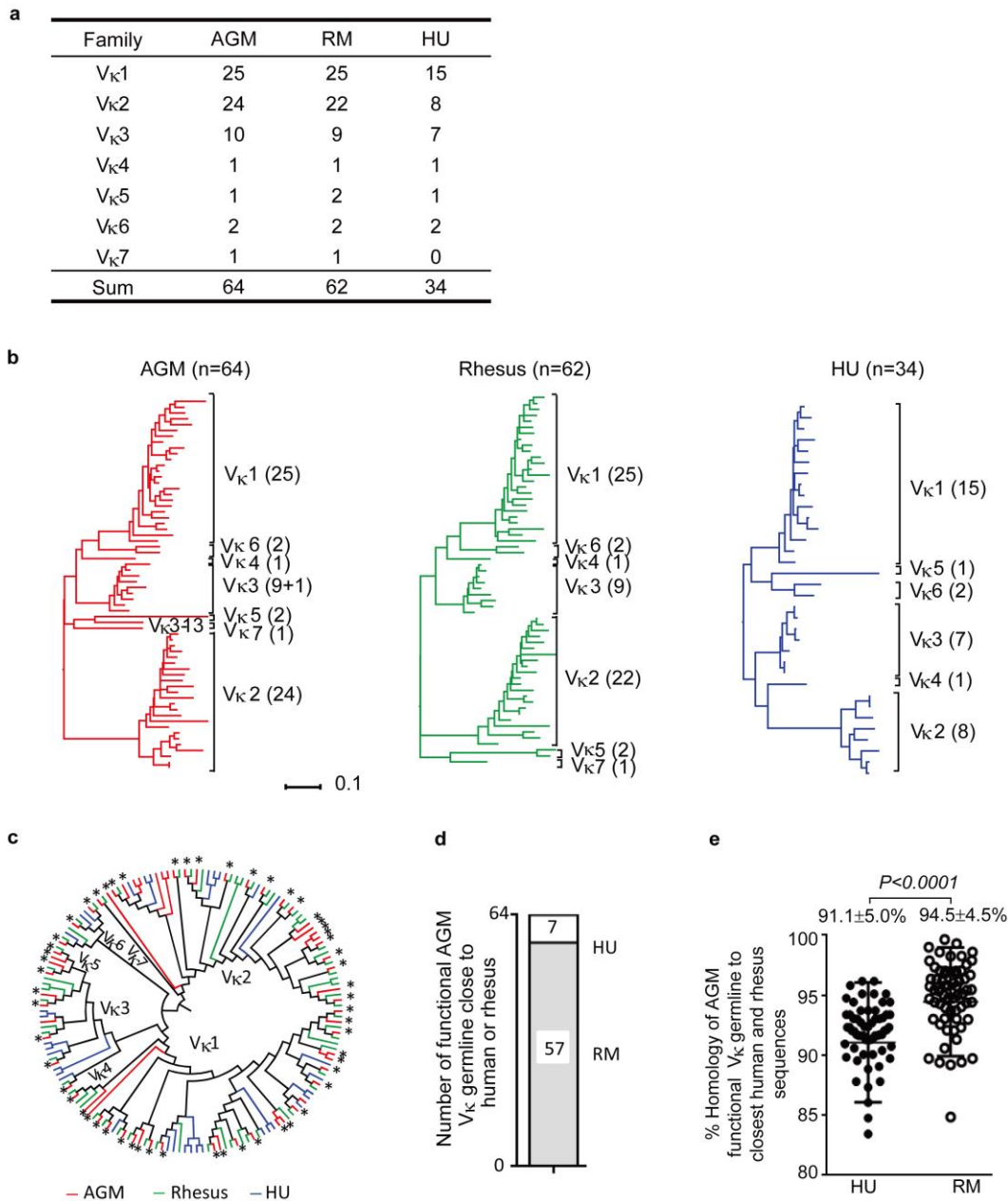
		50		100
			μE1	μE5
AGM_IgH_Eμ	CATCATAAATCAAGTTTATTTTTTTAATTAATTCAGTCAACCTGGGACGGGATGATGAGTTAGAGTCAAGATGGCTGGCATGGGGTCTTTGGCAACCA			
Rhesus_IgH_Eμ	.....G.....C.....G			
Human_IgH_Eμ	.....C.....A..A.....A.....CC.....			
		150		200
	μE2	μA	μE3	μB
AGM_IgH_Eμ	<u>GCAGGTGGCAGGAAGCA</u> <u>GGTCA</u> <u>CCGGCAAGAGTCTA</u> <u>TTTTAGGA</u> GCAAAAAACAACAACCTGGTACATTTATCATTCTCGTTGIGAAAGAGGIGGTTTCGC			
Rhesus_IgH_Eμ	.....G.....T.....-.....C...G.T...A...G.....			
Human_IgH_Eμ	.....G.....T...A.....T..			
			μE4	Octamer motif
AGM_IgH_Eμ	CTGGGOCAGATCTGAAGTGTCTCTACAGCAAAACA <u>CACC</u> <u>GGCA</u> <u>CGATTTGGGT</u> TTCTAAATAAGGCGAGGCTGACCGAAACIGGAAAGGCTC-TT			
Rhesus_IgH_Eμ	.....C.....A.....A.....T..			
Human_IgH_Eμ	.CA.....A.....A.....---			
		350		
			CACA Box	TATA Box
AGM_IgH_Eμ	TTTTTTAACTATCCGATTTTCATTT <u>CCAAT</u> CTAGCTTATCAACCTAGTCTGTTGTCAAACAGG <u>ATA</u> TCAACTTCTAAACTGCATTTCATTTT			
Rhesus_IgH_Eμ	.....A.....C.....C.....			
Human_IgH_Eμ	.....T.....C.....			

**Supplementary Figure 3 Comparison of IgH intronic enhancer core region (cEμ) of AGM, RM and human.** The cEμ elements, Octamer motif, CACA box, and TATA box were highlighted and underlined. RM cEμ intronic enhancer region was identified from the assembly on Oct. 2010 (BGI CR\_1.0/rheMac3).



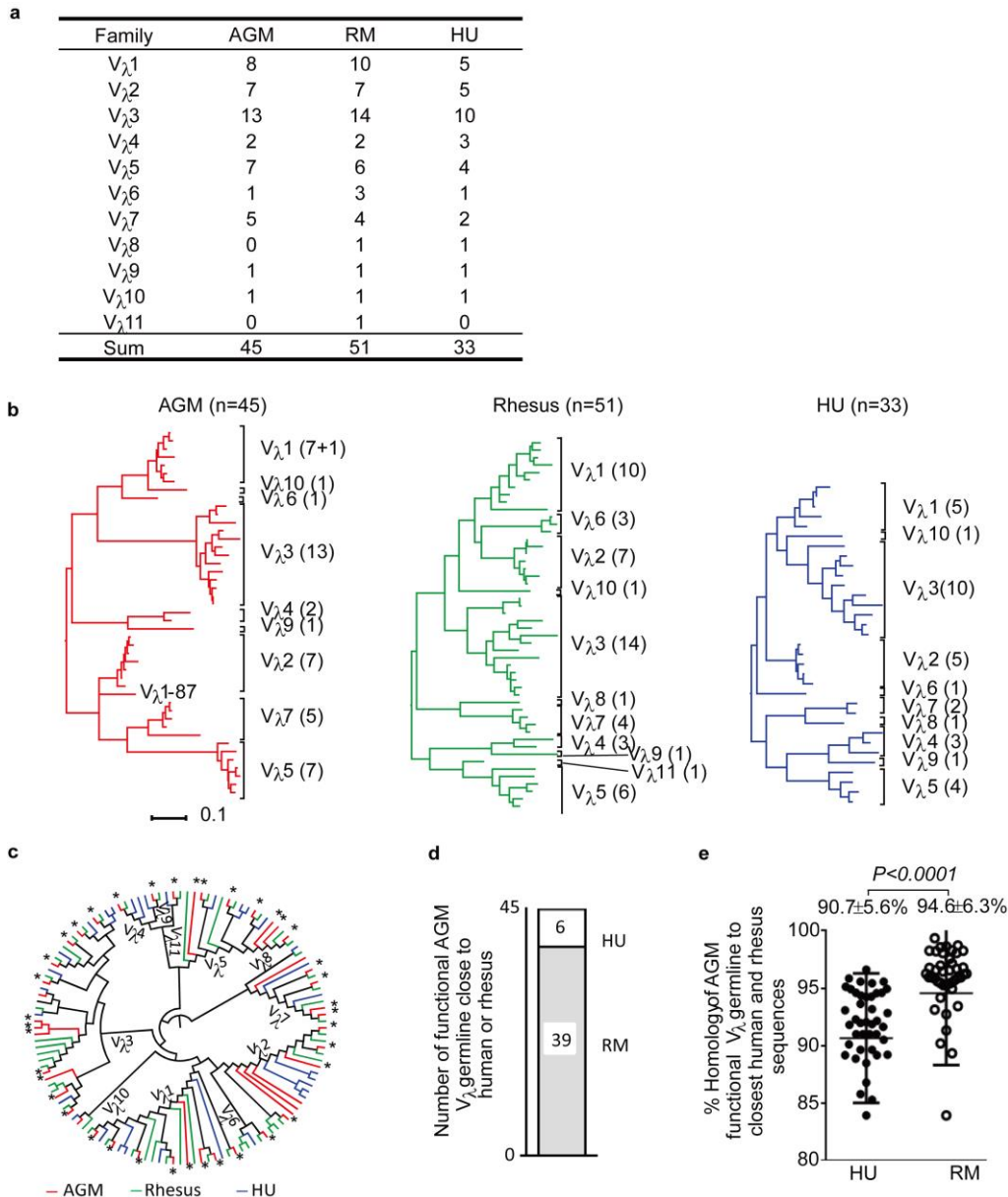
**Supplementary Figure 4 Comparison of the centromeric region of V<sub>H</sub> locus in AGM and human genomes.** Mapping of AGM V<sub>H</sub> gene segments in reference of human genome indicated that a large DNA fragment is potentially missing in the centromeric side of AGM V<sub>H</sub> locus, in which V<sub>H</sub>6, V<sub>H</sub>7 and V<sub>H</sub>2 might be located by comparing human V<sub>H</sub> locus.



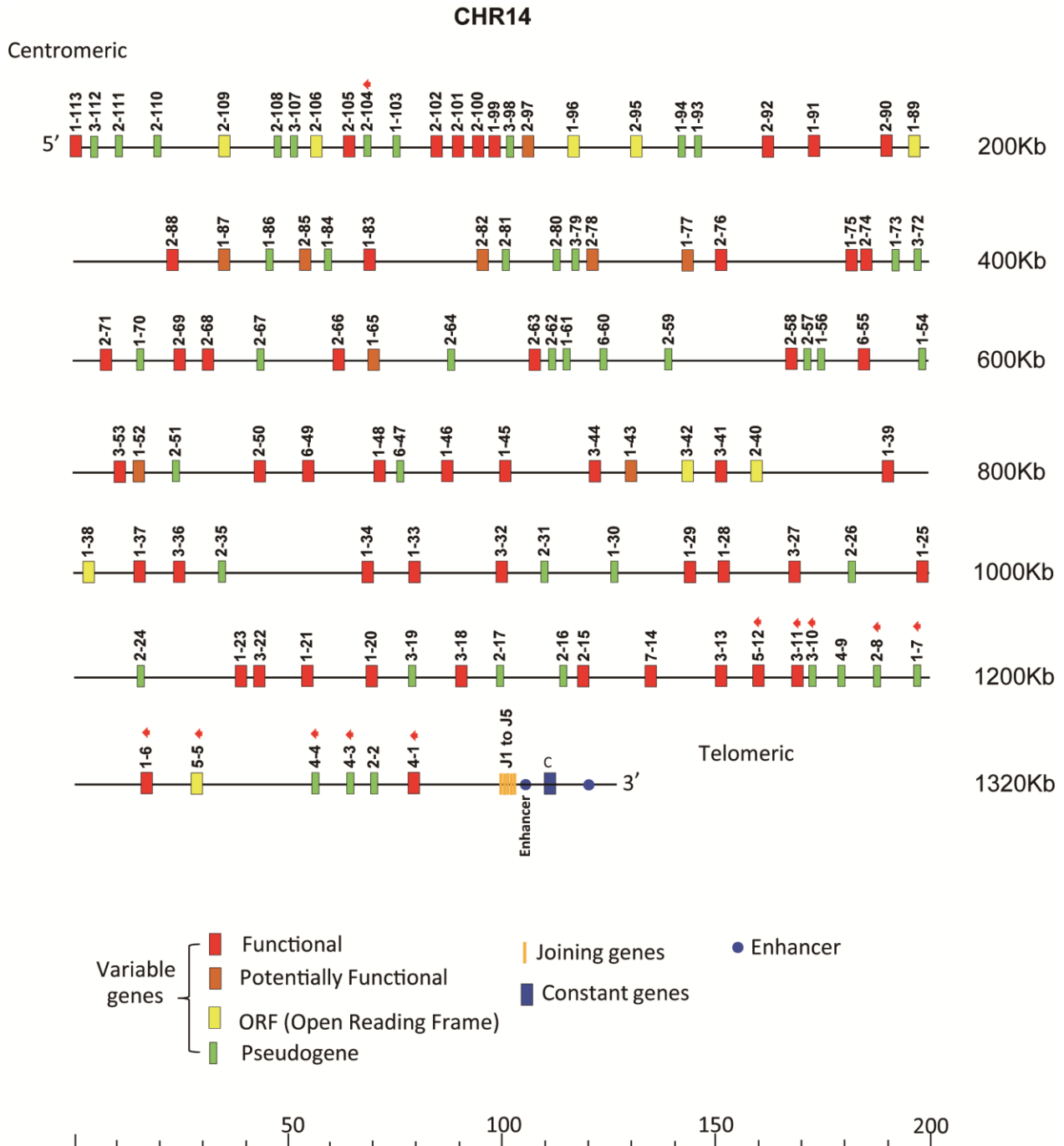


**Supplementary Figure 5 AGM, RM and human immunoglobulin kappa chain variable gene (V<sub>κ</sub>) homology.** AGM V<sub>κ</sub> genes are genetically more similar to those of RM than human. **Panel a** shows numbers of functional V<sub>κ</sub> genes from each species. **Panel b** shows phylogenetic comparison of V<sub>κ</sub> families of AGM, RM and human. AGM V<sub>κ</sub> 3-13 is located out of V<sub>κ</sub> 3 family branch and has a human homologue gene as IGKV3-NL5\*01 F. **Panel c** shows joint radial phylogenetic tree of AGM (red lines), RM (green lines), and human (blue lines) V<sub>κ</sub> genes; the AGM V<sub>κ</sub> genes that are genetically more similar to RM than human V<sub>κ</sub> homologs are indicated with star (\*). **Panel d** indicates the number of AGM V<sub>κ</sub> genes that are genetically more similar to RM (grey box) or human (white box) than AGM. **Panel e** indicates percent homology of AGM V<sub>κ</sub> genes to those of RM and human. Percent homology between V<sub>κ</sub> germline genes between AGMs, humans and RMs were compared ( $p < 0.0001$ ) using an Exact Wilcoxon rank sum test. The data in Panel e shows means $\pm$ sd.

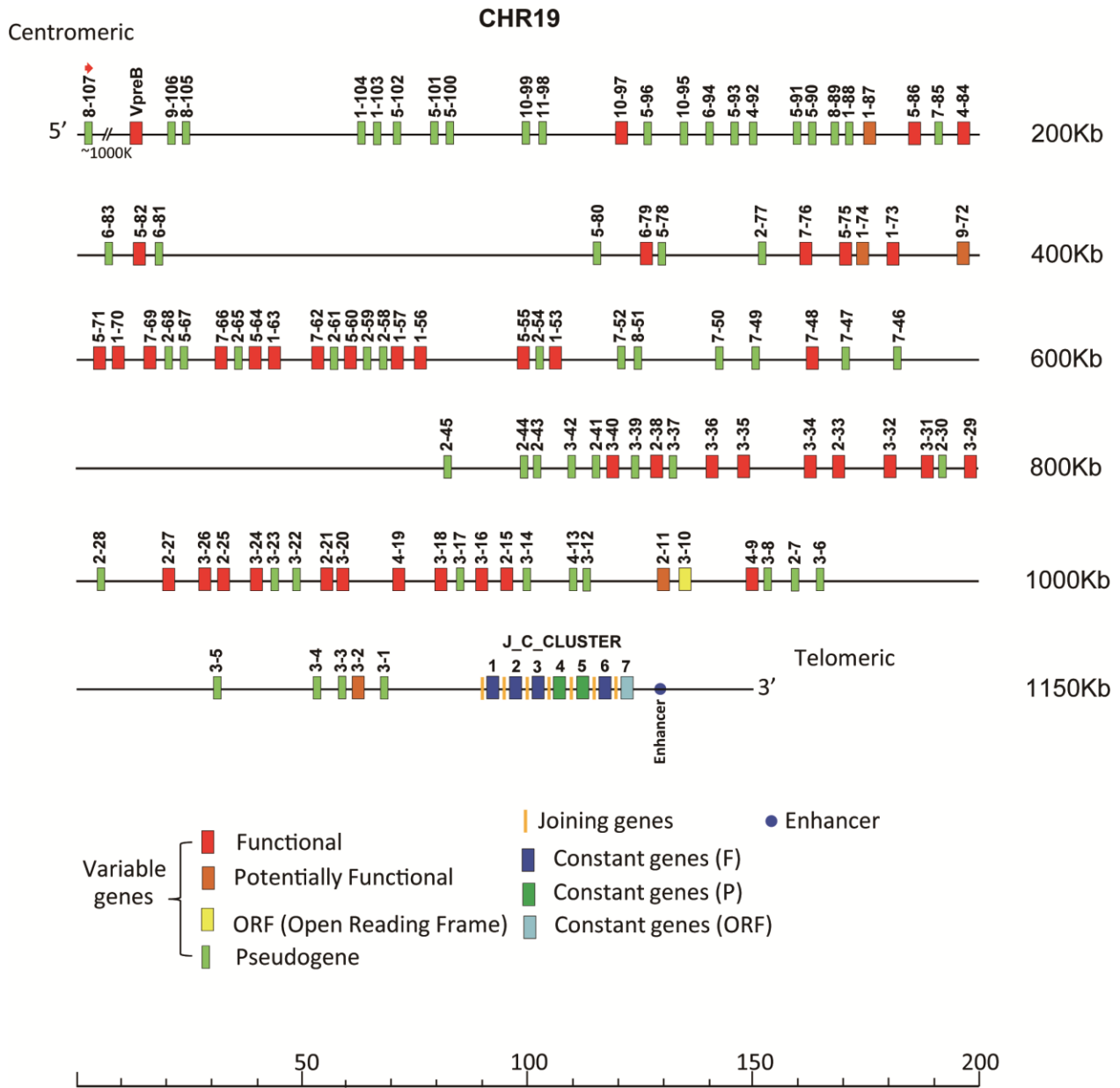




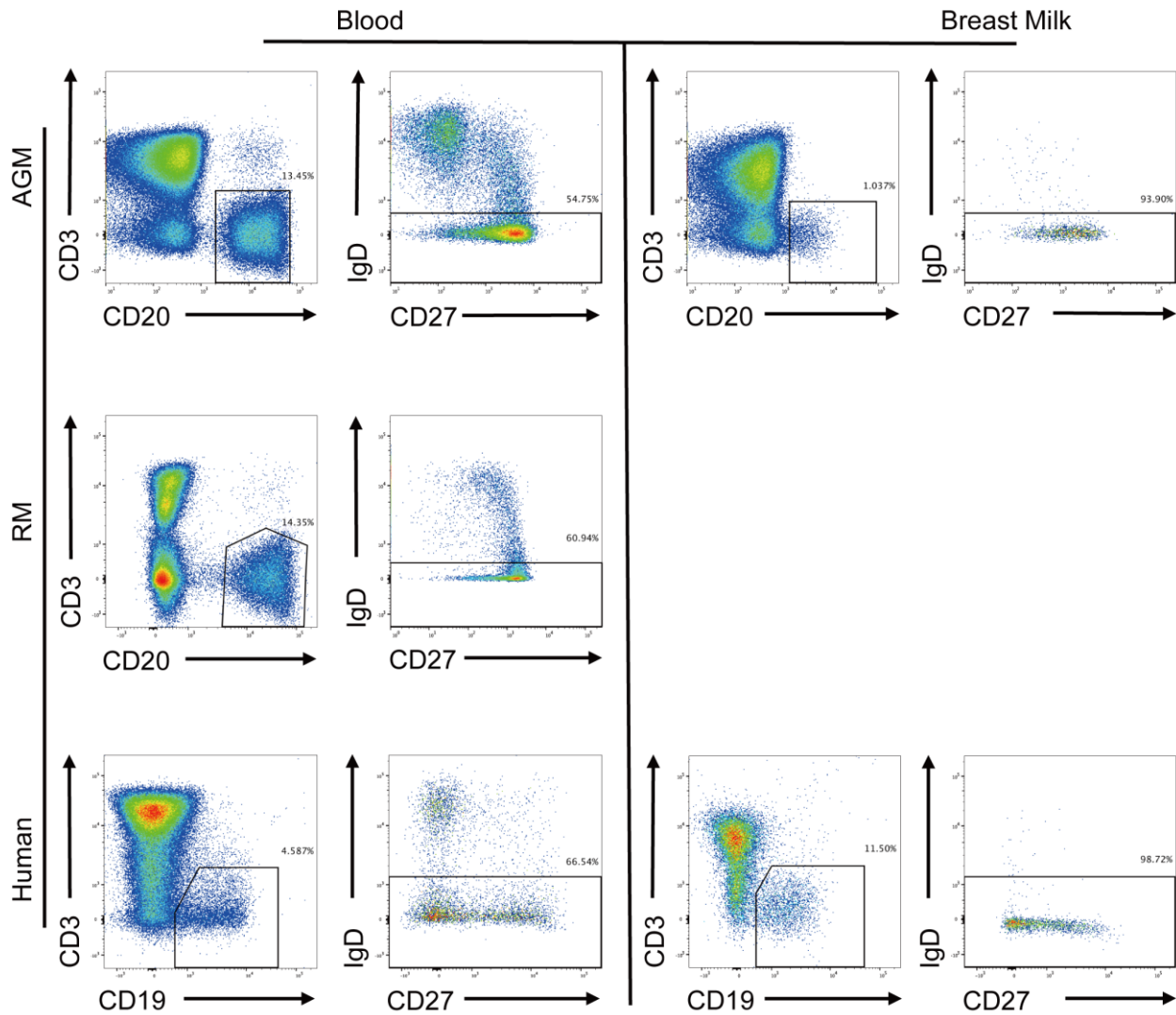
**Supplementary Figure 6 AGM, RM and human immunoglobulin lambda chain variable gene (V<sub>λ</sub>) homology.** AGM V<sub>λ</sub> genes are genetically more similar to those of RM than human. **Panel a** shows number of functional V<sub>λ</sub> genes in each species. **Panel b** shows phylogenetic comparison of AGM, RM and human V<sub>λ</sub> families. AGMLV1-87 is located out of V<sub>λ</sub>1 family branch, and has the human homologue gene as IGLV1-62\*01 P. **Panel c** shows joint radial phylogenetic tree of AGM (red lines), RM (green lines), and human (blue lines) V<sub>L</sub> genes; the AGM V<sub>λ</sub> genes that are genetically more similar to RM than human V<sub>λ</sub> homologs are indicated with star (\*). **Panel d** shows number of AGM V<sub>λ</sub> genes genetically more similar to RM (grey box) or human (white box) than AGM. **Panel e** shows percent homology of AGM V<sub>λ</sub> genes to those of RM and human. Percent homology between V<sub>λ</sub> germline genes between AGMs, humans and RMs were compared ( $p < 0.0001$ ) using an Exact Wilcoxon rank sum test. The data in Panel e shows means ± sd.



**Supplementary Figure 7 Map of AGM (*Chlorocebus sabaues*) immunoglobulin  $\kappa$  locus on chromosome (CHR) 14.** Shown are the identified functional, potentially functional, open reading frames, and pseudogenes of AGM  $V_{\kappa}$  and  $J_{\kappa}$  germline segments along with the Ig kappa chain constant region gene located on CHR14.

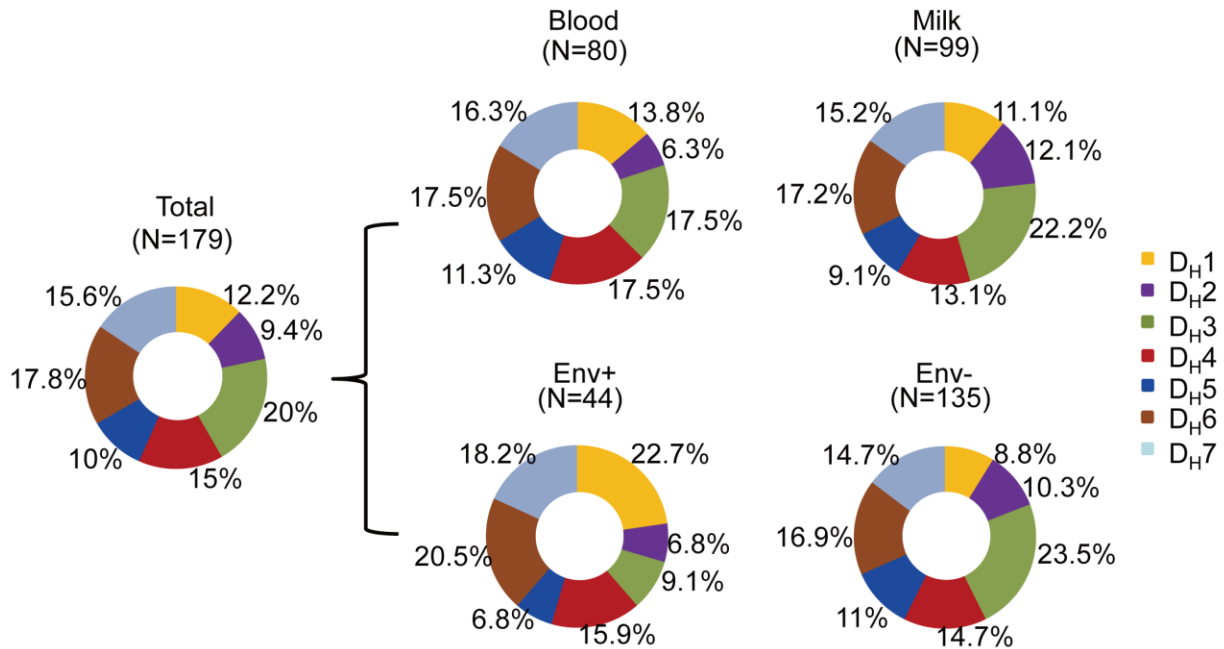


**Supplementary Figure 8 Map of AGM (*Chlorocebus sabaeus*) immunoglobulin  $\lambda$  locus on chromosome (CHR) 19.** Shown are the identified functional, potentially functional, open reading frames, and pseudogenes of AGM  $V_\lambda$  and  $J_\lambda$  germline segments along with the Ig lambda chain constant region gene located on CHR19.

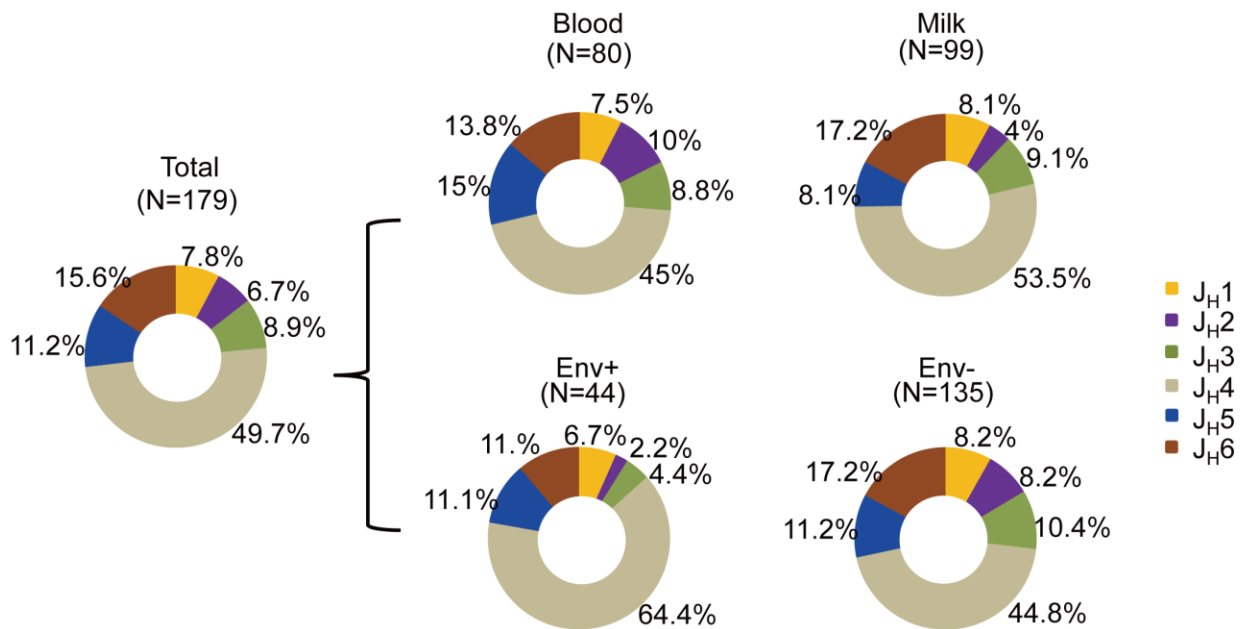


**Supplementary Figure 9 Gating strategy for total memory B cell populations in blood and milk of SIV/HIV-infected, AGM, RM, and human.** Total memory B cells of SIV-infected AGM and RM were gated as CD3<sup>-</sup>/CD20<sup>+</sup>/IgD<sup>-</sup> (top two rows). Total memory B cells from blood and milk of HIV-infected human were gated as CD3<sup>-</sup>/CD19<sup>+</sup>/IgD<sup>-</sup> (bottom row).

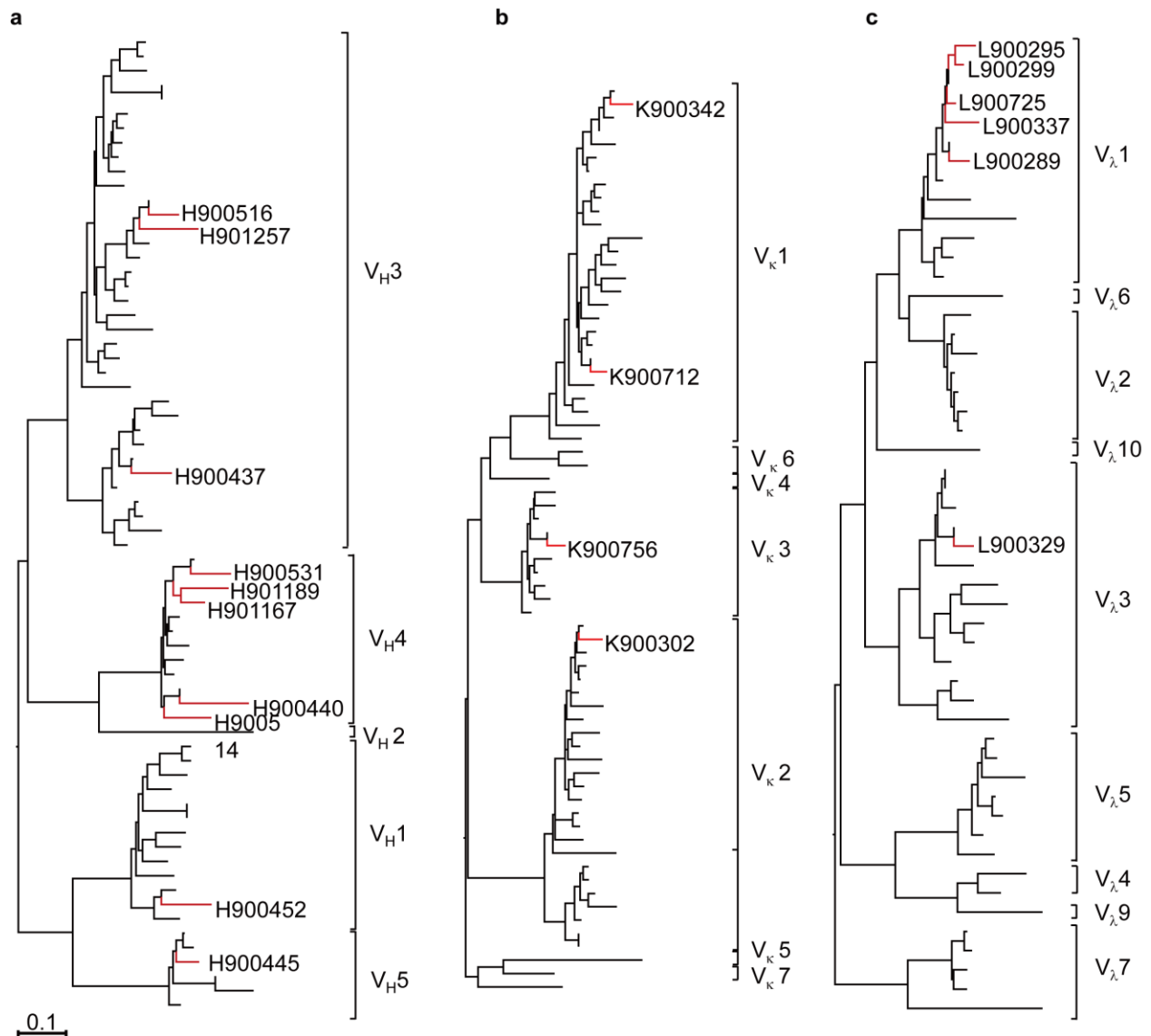
a



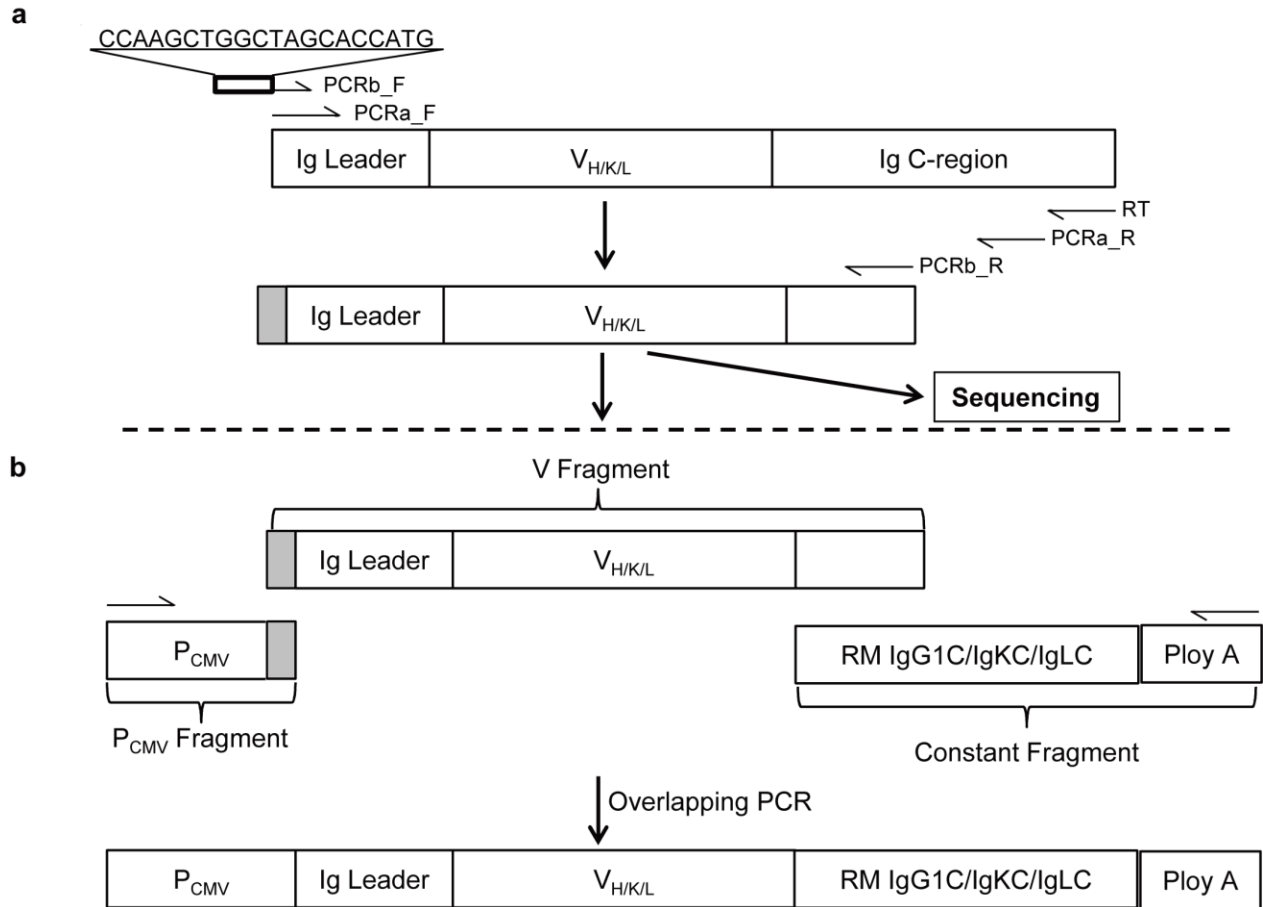
b



**Supplementary Figure 10 Genetic distribution of D<sub>H</sub> and J<sub>H</sub> of AGM Env-specific and non Env-specific mAbs.** The distribution of D<sub>H</sub> gene families (Panel a) and J<sub>H</sub> gene families (Panel b) isolated from AGM blood and milk samples.



**Supplementary Figure 11 The phylogenetic relationship of 10 purified SIV Env-reactive mAbs to AGM heavy and light chain variable germline genes.** The phylogenetic relationship of V<sub>H</sub> (Panel a), V<sub>κ</sub> (Panel b) and V<sub>λ</sub> (Panel c) genes of 10 purified SIV Env-reactive mAbs to AGM V<sub>H</sub> germline genes.



**Supplementary Figure 12 Schematic diagram for the generation of linear full-length Ig heavy- and light-chain gene constructs. (a)** Schematic representation of cloning rhesus immunoglobulin heavy and light chain variable genes by semi-nest PCR. **(b)** The Ig H/L PCR products were sequenced and the functional genes were then assembled into full-length linear expression cassettes with CMV promoter and BGH Poly A by overlapping PCR.



**Supplementary Table 1 Demographic, virologic, and immunologic characterization of HIV-1-infected individuals used for antibody isolation from blood and breast milk**

#	Patient ID	Number of Antibodies Isolated			Patient Characteristics						
		Blood	Milk	Total	Age (yrs)	Sex	Country of Origin	Specimen Draw Date	Viral Load (RNA copies/mL)	CD4 count (cells/mm <sup>1</sup> )	ARV
1	711-12-071-8	93	0	93	21	M	USA	24-Jun-10	<40	528	Yes
2	700-12-051-1	68	0	68	59	M	USA	9-Oct-09	16,800	957	No
3	714-90-060-8	22	0	22	41	F	Malawi	3-Dec-07	<400	275	Yes
4	715-90-060-5	0	3	3	28	F	Malawi	2-Apr-08	32,164	519	No
5	714-09-360-1	0	31	31	18	F	Malawi	10-Mar-08	60,335	452	No
6	715-90-030-1	6	5	11	32	F	Malawi	26-Mar-08	75,934	406	No
7	715-90-120-9	14	2	16	25	F	Malawi	18-Jun-08	174,976	153	No
Antibodies Isolated		Blood		203							
		Milk		41							
		Total		244							

<sup>1</sup> VL and CD4 count performed at the time of PBMC/milk cell isolation

**Supplementary Table 2 Amino acid sequences of the upper, core and lower hinge of IGG molecules from AGM (*Chlorocebus Sabaeus*), RM, and human.**

	Upper Hinge	Core	Lower Hinge
HU_IgG1	EPKSCDKTHT	CPPC	PAPELLGGP
RM_IgG1	EIKTCGGGSKPPT	CPPC	PAPELLGGP
AGM_IgG1	EIKTCGGGGKPPT	CPPC	PAPELLGGP
HU_IgG2	ERK	CCVECPCP	PAPPVAGP
RM_IgG2	GLP	CRSTCPPC	PAELLGGP
AGM_IgG2	not available	not available	ELLGGP (partial)
		CPRCP	
HU_IgG3	ELKTPLGDTTHT	EPKSCDTPPPCPRCP	APELLGGP
		EPKSCDTPPPCPRCP	
		EPKSCDTPPPCPRCP	
RM_IgG3	EFTPPCST	CPPC	PAPELLGGP
HU_IgG4	ESKYGPP	CPSC	PAPEFLGGP
RM_IgG4	EFTPP	CPPC	PAPELLGGP

**Supplementary Table 3 Primer list for amplifying both AGM and rhesus macaque Ig pairs**

Primers	Sequences	Tm
RT primers		
G_RT	TCTGTGCCACCTTGGTGTTG	51.8
M-RT	TGAATTTCCAGGAGAAAGTGATG	51.7
A_RT	TGTTTCCRGATTTTGAGATGGTG	53.5
D_RT	AAGGTCTTCTCCTTGCTTGTATC	53.5
E_RT	TTGGAATCTGCACACTTCTTG	50.5
CK_RT	TCTCTGGGATAGAAGTTATTCAG	51.7
CL_RT	CTTGTTGTTGCTCTGTTTGGAG	53
PCR1 primers		
IgH primers		
VH1+7_EXT1	CACCATGGACTGGACCTGGAGGMTCCCTC	67.9
VH1+7_EXT2	CACCATGGACCTGACCCGGAGGATCCTTTTC	72.4
VH2_EXT	CACCATGGACACGCTTTGCTCCACRCTC	68
VH3_EXT1	CATGGAGTTgGGGCTGAGCTGGGTYTTCC	71.3
VH3_EXT2	CATGGAGTTgGGGCTGAGYTGGGTTTTCC	69.7
VH3_EXT3	CATGGAGTTTGGGCTGAGCTGGR	61.1
VH3_EXT4	CATGGAGTTTGGGCTGAGCTKGGTTTTYC	66.8
VH4_EXT1	ACCATGAAGCACCTGTGGTTCTBCCTCCTCC	69.7
VH4_EXT2	CACCATGAAGCACCTGKGGTTCTTY	60.1
VH5_EXT	CACCATGGGGTCAACTGCCMTCCTC	65.3
VH6_EXT	CCATGTCTGTCTCCTTCCTCATCGTCC	63.8
A_EXT	GAAGAAGCCCTGGACCAGGCAGGC	66.6
G-EXT	AAGGTGTGCACGCCGCTGGTCAG	66.9
M_EXT	GTCGGGAAGGAAGTCCTGTGCGAGG	66.5
D_EXT	TCCCCAGGTGCCAGGTGACAGTCAC	66.8
E_EXT	ACGGTCAGCAAGCTGATGGTGGCA	66.3

IgK primers		
VK1_EXT1	CACCATGGACATGAGGGYCCC	58.4
VK1_EXT2	CACCATGGACATGAGGGTCCCCAGTC	66.9
VK1_EXT3	CACCATGGACATGAGGGTCCCCGGTtAtC	70.2
VK1_EXT4	CACCATGGACATGAGGGTTCYCCGGTCAG	69.5
VK1_EXT5	CACCATGGACATGAGGGTCCCCGGTCAGCTYC	75.9
VK2_EXT1	CACCATGAGGCTCCCWGCTCAG	60.6
VK2_EXT2	CACCATGAGGCTCCCTGCTCAGCTYCTGGGGC	76.8
VK3_EXT1	CACCATGGAAGCCCCAGCTCRGCTTCTC	69.7
VK3_EXT2	CACCATGGAAGCCCCAGCACAGCTTCTC	70.4
VK4_EXT	CACCATGGTGTACAGACCCAAGWCTTC	64.4
VK5_EXT	CACCATGGGATCCCAGGTTACCTCCTCAG	71.4
VK6_EXT1	CACCATGGTGTCCCCATTGCAACTCCTG	69.9
VK6_EXT2	CACCATGtTGTCtCCATCACAACTCATtG	63.2
VK7_EXT	CACCATGGGGTCTGGGGCTCCTTTCTG	72.2
CK_EXT	ACCTGATCCTCAGATGGCGGGAAGATG	66.7
IgL primers		
VL1_EXT1	CACCATGGCCTGGTCTCCTCTCSTCCTCAC	69.7
VL1_EXT2	CACCATGGCCTGGTCTCCTCTCCTtCTC	67.4
VL2_EXT	CACCATGGCCTGGGCTCTGSTCCTC	67.2
VL3_EXT1	CACCATGGCCGGGACCCYTCTCCTCCTC	73.1
VL3_EXT2	CACCATGGCCTGGACCCCTGTTCTGCTC	71.8
VL3_EXT3	CACCATGGCCtGGACCCcTCcCCTRCTC	73.1
VL4_EXT	CACCATGGCCTGGACCCACTCCTCCTC	72.6
VL5_EXT	CACCATGGCCTGGACTCYTCTC	57.3
VL6_EXT	CACCATGGCCTGGGCTCCACTCCTCCTC	72.7
VL7_EXT	CACCATGGCCTGGACTCTGCTCCTCCTCC	72
VL8_EXT	CACCATGGCCTGGATGATGCTTCTCCTCG	71.6
VL11_EXT	CACCATGGCCCTGACTCCTCTCCTCCTC	69.1
CL_EXT	TGCCATCTGCCTTCCAGGCCACTT	66.3
Primers	Sequences	Tm

PCR2 primers

IgH primers		
VH1+7_INT1	CCAAGCTGGCTAGCACCATGGACTGGACCTGGAGGMTCTC	80.1
VH1+7_INT2	CCAAGCTGGCTAGCACCATGGACCTSAcccGGAGSATCCTTTTC	78.4
VH2_INT	CCAAGCTGGCTAGCACCATGGACACGCTTTGCTCCAC	78.9
VH3_INT1	CCAAGCTGGCTAGCACCATGGAGTTGGGGCTGAGYTG	78.2
VH3_INT2	CCAAGCTGGCTAGCACCATGGAGTTTGGGGCTGAGCTKG	78.5
VH4_INT	CCAAGCTGGCTAGCACCATGAAGCACCTGKGGTTC	75.3
VH5_INT	CCAAGCTGGCTAGCACCATGGGGTCAACTGCCMTCCTC	79
VH6_INT	CCAAGCTGGCTAGCACCATGTCTGTCTCCTTCTCATCGTC	77.8
A_ACD_Int	CAGGGCCGCTGTGCTCTCGGAGGTGCTCCTGCCCTCGAGG CTCAGCGGGAAGAC	76.3
A_BC_Int	CAGGGCCGCTGTGCTCTCGGAGGTGCTCCTGCTGCAGAGG YTCAGCGGGAAGAC	75.5
G_Int	CAGGGCCGCTGTGCTCTCGGAGGTGCTCCTGGAG	80.9
M_Int	CAGGGCCGCTGTGCTCTCGGAGGTGCTCCTACAGGAGACG AGGGGAAAAGGG	61.2
D_Int	CAGGGCCGCTGTGCTCTCGGAGGTGCTCCTCGCTGATATG ATGGGGAACACATCC	62.1
IgK primers		
VK1_INT1	CCAAGCTGGCTAGCACCATGGACATGAGGGYCCC	76.6
VK1_INT2	CCAAGCTGGCTAGCACCATGGACATGAGGGTCYCCG	78
VK2_INT1	CCAAGCTGGCTAGCACCATGAGGCTCCCWGCTC	75.8
VK3_INT1	CCAAGCTGGCTAGCACCATGGAAGCCCCAGCWC	76.7
VK4_INT	CCAAGCTGGCTAGCACCATGGTGTACAGACCCAAG	76.4
VK5_INT	CCAAGCTGGCTAGCACCATGGGATCCCAGGTTACCTCC	80.3
VK6_INT1	CCAAGCTGGCTAGCACCATGGTGTCCCCATTGCAACTC	79
VK6_INT2	CCAAGCTGGCTAGCACCATGTTGTCCATCACAACTC	74.9
VK7_INT	CCAAGCTGGCTAGCACCATGGGGTCCTGGGCTCCTTTCC	81.5
CK_INT	TGGCGGGAAGATGAAGACAGATGGTG	66.2
IgL primers		
VL1_INT	CCAAGCTGGCTAGCACCATGGCCTGGTCTCCTCTC	76.7
VL2_INT	CCAAGCTGGCTAGCACCATGGCCTGGGCTCTGSTCC	79.5
VL3_INT1	CCAAGCTGGCTAGCACCATGGCCGGGACCCYTC	78.8

VL3_INT2	CCAAGCTGGCTAGCACCATGGCCGGACCCCTS	77.4
VL4_INT	CCAAGCTGGCTAGCACCATGGCCTGGACCCACTCC	80.6
VL5_INT	CCAAGCTGGCTAGCACCATGGCCTGGACTCYTCTC	75.4
VL6_INT	CCAAGCTGGCTAGCACCATGGCCTGGGCTCCTC	79.4
VL7_INT	CCAAGCTGGCTAGCACCATGGCCTGGACTCTGCTCCTC	79.5
VL8_INT	CCAAGCTGGCTAGCACCATGGCCTGGATGATGCTTCTC	78.6
VL11_INT	CCAAGCTGGCTAGCACCATGGCCCTGACTCCTCTCCTC	78.9
CL_INT	GTCAGTATCAGACACACTAGTGTGG	54.4
Sequencing primers		
VHKLSEQ_F	CCAAGCTGGCTAGCACCATG	54.7
VHSEQ_R	CTGTGCTCTCGGAGGTGCTCC	55.7
VKSEQ_R	GGAAGATGAAGACAGATGGTG	48.4
VLSEQ_R	TGATCAGACACACTAGTGTGG	45.1
Primers for the construction of linear cassettes		
Primers for P <sub>CMV</sub> fragment <sup>1</sup>		
HV13220CMV-P-F262	AGTAATCAATTACGGGGTCATTAGTTCATAG	57.9
CMV-P_R	CATGGTGCTAGCCAGCTTGGGTC	60.6
Primers for Rh IgG1C fragment <sup>2</sup>		
RhIGC_HF	GGAGCACCTCCGAGAGCACAGC	62
BGH-R1235	TCCCAGCATGCCTGCTATTGTC	61.9
Primers for Rh IgKC fragment <sup>3</sup>		
RhIGC_KF	CACCATCTGTCTTCATCTTCCCGCCA	65.7
BGH-R1235	TCCCAGCATGCCTGCTATTGTC	61.9
Primers for Rh IgLC fragment <sup>4</sup>		
RhIGC_LF	CCACACTAGTGTGTCTGATCAGTG	51.3
BGH-R1235	TCCCAGCATGCCTGCTATTGTC	61.9
Primers for overlapping PCR		
HV13220CMV-P-F262	AGTAATCAATTACGGGGTCATTAGTTCATAG	57.9
BGH-R1235	TCCCAGCATGCCTGCTATTGTC	61.9

<sup>1</sup> pcDNA3.1 plasmid as template

<sup>2</sup> Plasmid with rhesus whole IgG1 gene as template

<sup>3</sup> Plasmid with rhesus whole IgK gene as template

<sup>4</sup> Plasmid with rhesus whole IgL gene as template

**Supplementary Table 4 Statistical analysis Raw p values and false discovery rate p values.**

Figure	Category	Test	Comparison	Variable	Compartment	Germline	Raw_P	fdr_p
Figure 2E	Homology	Wilcoxon	AGM:HU vs AGM:RM			V-Heavy	0.0000	0.0000
Figure 2E	Homology	Wilcoxon	AGM:HU vs AGM:RM			V-Kappa	0.0000	0.0000
Figure 2E	Homology	Wilcoxon	AGM:HU vs AGM:RM			V-Lambda	0.0000	0.0000
Figure 3B	Genomic	Fisher's Exact	HIV+/-: AGM vs HU		All		0.0000	0.0000
Figure 3B	Genomic	Fisher's Exact	HIV+/-: AGM vs RM		All		0.3095	0.5364
Figure 3B	Genomic	Fisher's Exact	HIV+/-: AGM vs HU		BM		0.0037	0.0113
Figure 3B	Genomic	Fisher's Exact	HIV+/-: AGM vs HU		PBMC		0.0000	0.0000
Figure 3B	Genomic	Fisher's Exact	HIV+/-: AGM vs RM		PBMC		1.0000	1.0000
Figure 3B	Genomic	Fisher's Exact	gp120+/-: AGM vs HU		All		1.0000	1.0000
Figure 3B	Genomic	Fisher's Exact	gp120+/-: AGM vs RM		All		0.0014	0.0052
Figure 3B	Genomic	Fisher's Exact	gp120+/-: AGM vs HU		BM		0.5403	0.7236
Figure 3B	Genomic	Fisher's Exact	gp120+/-: AGM vs HU		PBMC		0.4375	0.6319
Figure 3B	Genomic	Fisher's Exact	gp120+/-: AGM vs RM		PBMC		0.0004	0.0018
Figure 4	Genomic	Kolmogorov-Smirnov	ENV+ vs ENV-	HCDR3_Length			0.8996	1.0000
Figure 4	Genomic	Wilcoxon	ENV+ vs ENV-	H_Mutated			0.6547	0.7738
Figure 4	Genomic	Kolmogorov-Smirnov	gp120 vs Env cross-reactive	HCDR3_Length			0.3934	0.6016
Figure 4	Genomic	Exact Wilcoxon	gp120 vs Env cross-reactive	H_Mutated			0.5566	0.7236
Figure 4	Genomic	Kolmogorov-Smirnov	gp120 vs gp41	HCDR3_Length			0.9733	1.0000
Figure 4	Genomic	Exact Wilcoxon	gp120 vs gp41	H_Mutated			0.0401	0.0745
Figure 4	Genomic	Kolmogorov-Smirnov	gp41 vs Env cross-reactive	HCDR3_Length			0.6483	0.7738
Figure 4	Genomic	Exact Wilcoxon	gp41 vs Env cross-reactive	H_Mutated			0.3369	0.5475



**Supplementary Table 5 % blocking by AGM gp120-specific ADCC mediating antibodies with undefined SIVsabgp120 fine-epitope specificity**

	% blocking			
	DH546	DH550	DH549	DH552
DH546-biotin	73.9	0	0	17
DH550-biotin	0	35.4	48.6	6.9
DH549-biotin	0	19.7	57.3	5.1
DH552-biotin	22.7	0	3.6	53.23

**Supplementary Table 6 AGM and RM total B cell (CD3<sup>-</sup> CD20<sup>+</sup>) and memory B cell (CD20<sup>+</sup> IgD<sup>-</sup> CD27<sup>+</sup>) count (cells/ $\mu$ l) at week 45 post infection**

	animal ID	total B cell count (cells/ $\mu$ l) <sup>1</sup>	Mann-Whitney test	total memory B cell count (cells/ $\mu$ l) <sup>2</sup>	Mann-Whitney test
AGM	013-89-10	220	p = 0.07	118	p = 0.26
	013-90-10	21		8	
	013-91-10	36		11	
	013-92-10	116		59	
	013-93-10	12		4	
	013-94-10	149		83	
RM	206-96	289		109	
	257-97	95		40	
	402-98	662		376	
	403-98	251		66	

<sup>1</sup> denotes CD3<sup>-</sup> CD20<sup>+</sup> cells

<sup>2</sup> denotes CD20<sup>+</sup> IgD<sup>-</sup> CD27<sup>+</sup>