Title *Caenorhabditis elegans* employs innate and learned aversion in response to bacterial toxic metabolites tambjamine and violacein

Francesco Ballestriero¹, Jadranka Nappi¹, Giuseppina Zampi³, Paolo Bazzicalupo^{2,3}, Elia Di Schiavi^{2,3#} and Suhelen Egan^{1#}

¹School of Biological, Earth and Environmental Science and Centre for Marine Bio-Innovation, University of New South Wales, Australia, ²Institute of Genetics and Biophysics, National Research Council, Naples, Italy, ³Institute of Biosciences and BioResources, National Research Council, Naples, Italy

Running title: Nematode avoidance of bacterial toxic metabolites

[#]Corresponding authors email: <u>s.egan@unsw.edu.au</u> and <u>elia.dischiavi@ibbr.cnr.it</u>



Figure S1. Schematic representation of the protocol for the food choice assay. (A) training plates, and (B) test plates. (C) Data from the food choice assay plotted as choice index or Learning index. A choice index of -1.0 represents complete preference for the control bacterium, an index of 1.0 represents an equal distribution. A positive learning index indicates a learned avoidance of toxic bacteria with the value 2 as maximum learning index. Adapted from (Zhang et al., 2005).



Figure S2. *C. elegans* assessed for aversive olfactory learning behavior in the food choice assay tested using 20G8vioA- and 20G8vioA- supplemented with pure violacein. Naïve (white bars) and trained (black bars). Nematode preference was tested against three and 300 μ M of pure violacein. Each data point represents means \pm the standard error of three replicates. * denotes 0.001<p<0.05.

Strain/Vector	Relevant characteristic or genotype	Source or reference
Bacterial strains		
E. coli EPI300- T1 ^R	F-mcrA Δ (mrrhsdRMSmcrBC) φ 80dlacZ Δ M15 Δ lacX 74 recA1 endA1 araD139 Δ (ara, leu) 7697galU galK λ - rpsL nupG trfA tonA dhfr	Epicentre
E. coli AA11	Fosmid AA11 cloned in <i>EPI300-T1^R</i> ; Cm ^r	(Burke, et al. 2007)
E. coli AA11tamG ⁻	Fosmid AA11 mutated in <i>tamG</i> gene and cloned in <i>EPI300-T1</i> ^{<i>R</i>} ; Cm ^{<i>r</i>} , Tet ^{<i>r</i>}	(Ballestriero, et al. 2014)
<i>E. coli</i> 20G8	Fosmid 20G8 cloned in <i>EPI300-T1</i> ^{<i>R</i>} ; Cm ^r	(Penesyan, et al.
E. coli 20G8vioA-	Fosmid 20G8 mutated in <i>vioA</i> gene and cloned in <i>EPI300-T1^R</i> ; Cm ^r , Kan ^r	2012) (Ballestriero, et al. 2014)
E. coli 20G8vioC-	Fosmid 20G8 mutated in <i>vioC</i> gene and cloned in <i>EPI300-T1^R</i> ; Cm ^r , Kan ^r	(Ballestriero, et al. 2014)
<i>E. coli</i> OP50	Uracil auxotroph	(Brenner 1974)
<u>Nematode</u> <u>strains</u>		
N2 Bristol	<i>C. elegans</i> wild type isolate	CGC b
PR811	<i>osm-6(p811)</i> V. <i>osm-6</i> mutants are defective in the ability to avoid high <u>osm</u> olarity. <i>osm-6</i> is required for proper sensory cilium structure. In <i>osm-6</i> mutants most of the ciliated neurons have severely shortened cilia that are not exposed to the environment. <i>osm-6</i> mutant shows reduced avoidance responses to repellents and fail to stain amphids with FITC.	CGC b
GR1321	<i>tph-1(mg280) cam-1(vs166)</i> II. Mutation in <i>tph-1</i> , which encodes the enzyme tryptophan hydroxylase that catalyses the rate-limiting first step in serotonin biosynthesis. Some phenotypic defects originally attributed to <i>mg280</i> in this strain are likely due to <i>vs166</i> , a deletion in the <i>cam-1</i> gene.	CGC b
MT15434	<i>tph-1(mg280)</i> II. Backcrossed strain carrying <i>mg280</i> allele without the <i>cam-1</i> mutation.	(Flavell, et al. 2013) °
<u>Vectors</u>		
pCC1FOSª	Fosmid backbone for genomic library; Cm ^r	Epicentre

Table S1. Bacterial and nematode strains and vectors used in this study.

Bargmann, Rockfeller University New York

References

Ballestriero, F., Thomas, T., Burke, C., Egan, S. & Kjelleberg, S. Identification of compounds with bioactivity against the nematode *Caenorhabditis elegans* by a screen based on the functional genomics of the marine bacterium *Pseudoalteromonas tunicata* D2. *Appl. Environ. Microbiol.* **76**, 5710-5717 (2010).

Ballestriero, F. *et al.* Antinematode activity of violacein and the role of the insulin/IGF-1 pathway in controlling violacein sensitivity in *Caenorhabditis elegans. PLoS One* **9**, e109201 (2014).

Brenner, S. The genetics of *Caenorhabditis elegans*. *Genetics* **77**, 71-94 (1974).

Burke, C., Thomas, T., Egan, S. & Kjelleberg, S. The use of functional genomics for the identification of a gene cluster encoding for the biosynthesis of an antifungal tambjamine in the marine bacterium *Pseudoalteromonas tunicata*. *Environ. Microbiol.* **9**, 814-818 (2007).

Flavell, S. W., *et al* (2013). Serotonin and the neuropeptide PDF initiate and extend opposing behavioral states in *C. elegans*. *Cell* **154**: 1023-1035.

Penesyan, A. *et al.* Assessing the effectiveness of functional genetic screens for the identification of bioactive metabolites. *Mar Drugs.* **11**, 40-49 (2012).

Zhang, Y., Lu, H. & Bargmann, C. Pathogenic bacteria induce aversive olfactory learning in *Caenorhabditis elegans*. *Nature* **438**, 179-184 (2005).