Supplementary Material

Title:

Immunophenotyping of rheumatoid arthritis reveals a linkage between HLA-DRB1 genotype, CXCR4 expression on memory CD4⁺ T cells, and disease activity.

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Supplementary Figure Legends

Figure S1. Immunophenotyping by flow cytometry. Peripheral blood mononuclear cells (PBMC) CD3+CD4+ helper T cell subsets were classified based on CD45RA, CXCR3 (Th1 marker), CCR6 (Th17 marker), and CXCR5 (Tfh marker).

Figure S2. Immunophenotyping of RA patients.

Comparison of 24 immunophenotyped subset ratios among healthy donors (HD), shared epitope (SE)-negative RA patients, and SE-positive RA patients (HD; n=110, SE-RA; n=30, SE+RA; n=61). *p<0.05 **p<0.01 ***P<0.001 Kruskal-Wallis tests with post-hoc Wilcoxon

tests with Bonferroni corrections.

Figure S3. RA specific association between SE and B cell HLA-DR expression.

Comparison of HLA-DR expression on B cells between shared epitope (SE)-negative healthy donors (HD), SE-positive HD, SE-negative RA patients, and SE-positive RA patients (SE-HD; n=63, SE+HD; n=47, SE-RA; n=30, SE+RA; n=61). The analysis is based on flow cytometric HLA-DR quantitative expression per cells.

**p<0.01 Kruskal-Wallis tests with post-hoc Wilcoxon tests with Bonferroni corrections.

Figure S4. RF titer is related to dysregulated inflammatory cytokines.(A) Correlations between 11 cytokines and soluble cytokine receptors and four clinical parameters in RA patients (n=30). Representative scatter plots are also shown on the right, as in figure 1A.

(B) Comparison of serum cytokine/soluble cytokine receptor concentrations among healthy donors (HD), shared epitope (SE)-negative RA patients, and SE-positive RA patients (HD; n=34, SE-RA; n=11, SE+RA; n=19). (C) Principal component analysis of 11 cytokine and soluble cytokine receptors to summarize the difference between HD, SE-RA, and SE+RA.
PC1 explained 48% and PC2 explained 13% of the total variance (HD; n=26, SE-RA; n=10, SE+RA; n=12).

*p<0.05 **p<0.01 ***P<0.001 Spearman's tests (A) or Kruskal-Wallis tests with post-hoc Wilcoxon tests with Bonferroni corrections (B).

Figure S5. RA specific association between SE and CXCR4. Comparison of CXCR4 expression on CD4⁺ T cell subsets among SE-negative HD, SE-positive HD, SE-negative RA patients, and SE-positive RA patients (SE-HD; n=46, SE+HD; n=36, SE-RA; n=8, SE+RA; n=27). ***P<0.001 Kruskal-Wallis tests with post-hoc Wilcoxon tests with Bonferroni corrections.

Figure S6. Enhanced CXCR4 expression on CD4⁺ T cells of RA patients with high ACPA titers.

Comparison of CXCR4 expression on CD4⁺ T cell subsets among healthy donors (HD), ACPA-negative RA patients (ACPA-), RA patients with low ACPA titers (ACPA+, 4.5 U/ml \leq ACPA titer < 100 U/ml), and RA patients with high ACPA titers (ACPA++, 100 U/ml \leq ACPA titer). HD; n=82, ACPA-; n=5 (SE-; n=3, SE+; n=2), ACPA+; n=11 (SE-; n=3, SE+; n=8), ACPA++; n=16 (SE-; n=2, SE+; n=14).

*p<0.05 **p<0.01 ***P<0.001 Kruskal-Wallis tests with post-hoc Wilcoxon tests with Bonferroni corrections.

Figure S7. CXCR4 expression on B cell subsets.

Comparison of CXCR4 expression on B cell subsets among healthy donors (HD), shared epitope (SE)-negative RA patients, and SE-positive RA patients (HD; n=82, SE-RA; n=8, SE+RA; n=27). ***P<0.001

Kruskal-Wallis tests with post-hoc Wilcoxon tests with Bonferroni corrections.

Figure S8. CCR7 expression on CD4⁺ T cell subsets. Comparison of CD4⁺ T cell subsets classified according to CCR7 and CD45RA expression among healthy donors (HD), shared epitope (SE)-negative RA patients, and SE-positive RA patients (HD; n=108, SE-RA; n=25, SE+RA; n=56). CD3⁺CD4⁺ T cells were defined as naive CD4⁺ T cells (CCR7⁺CD45RA⁺ Naive), central memory CD4⁺ T cells (CCR7⁺CD45RA⁻ CM), effector memory CD4⁺ T cells (CCR7⁻CD45RA⁻ EM), and effector CD4⁺ T cells (CCR7⁻CD45RA⁺ Effector). No significant differences were observed between groups by Kruskal-Wallis tests with post-hoc Wilcoxon tests with Bonferroni corrections.

Figure S9. B cell HLA-DR levels and CXCR4 expression on memory CD4⁺ T cells in HD.

Scatter plots between B cell, monocyte (MO), and dendritic cell (DC) HLA-DR quantitative expression per cells and CXCR4-positive ratios of the MemoryTh subset in HD (n=82) compared to RA patients (Figure 3C).

Figure S10. Pathway analysis of CXCR4⁺ memory CD4⁺ T cells.

(A) Top network of differentially expressed genes in CXCR4⁺ and CXCR4⁻ memory CD4⁺ T cells by IPA pathway analysis. Red color represents upregulation and green color represents downregulation in CXCR4⁺ CD4⁺ T cells.

(B-C) Most significantly upregulated (B) and downregulated (C) pathways in CXCR4⁺ memory CD4⁺ T cells. Top 3 pathways in IPA canonical pathway analysis with -log(p-value) greater than 12.5 and an absolute z-score greater than 2 are shown. (D) TCR downstream genes differentially upregulated in CXCR4⁺ memory CD4⁺ T cells. Genes were selected by IPA knowledge database.

Figure S11. CTLA4-Ig treatment partially restores immunological abnormalities in RA.

Effects of 6 months of CTLA4-Ig treatment on PBMC immunophenotyping (A; n=20), HLA-DR expression (B; n=20), serum cytokine and soluble cytokine receptor concentrations (C; n=14), CXCR4 expression on CD4⁺ T cells (D; n=8), and clinical disease activity (E; n=20). Percent changes from baseline values are plotted.

*p<0.05 **p<0.01 ***P<0.001 Paired Wilcoxon test.

Table S1. Baseline characteristics	3
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	Shared epitope	Shared epitope		
	negative	positive		Healthy
	RA (SE-)	RA (SE+)	p-values	Donors (HD)
n	30	61		110
age, median (IQR)	56 (46-64)	64 (52-67)	0.18	39 (31-45)
sex (f/m)	26/4	47/14	0.4	88/22
disease duration, years,				
median (IQR)	3 (1-14)	6 (1-13)	0.34	
RF, IU/ml, median (IQR)	43 (21-182)	72 (31-200)	0.20	
ACPA, U/ml, median				
(IQR)	40 (1.8-94)	102 (31-348)	0.0030 **	
DAS28esr, median (IQR)	4.3 (3.4-5.3)	4.5 (3.7-5.3)	0.79	
HAQ, median (IQR)	0.63 (0.13-1.2)	0.88 (0.25-1.9)	0.29	
treatment naïve, n (%)	7 (23)	10 (17)	0.57	
treatment with				
conventional DMARDs,				
n (%)	24 (77)	50 (83)	0.57	
treatment with				
methotrexate, n (%)	12 (39)	30 (50)	0.0058 **	
treatment with biologic				
DMARDs, n (%)	7 (23)	14 (23)	1	

Categorical data were tested with Fisher's exact test. Continuous data were tested with Mann-Whitney U test. **p<0.01

RF, Rheumatoid Factor; ACPA, anti-cyclic citrullinated peptide antibody; DAS28esr, Disease Activity Score 28 joints-ESR; HAQ, Health Assessment Questionnaire Disability Index; DMARDs, disease-modifying antirheumatic drugs

rable 52. Infinutiophenotyping s	ubset definitions
subset name (abbreviations)	cell definition
CD4 ⁺ T cells	CD3+CD4+ (parent population)
naive CD4 ⁺ T cells (NaiveTh)	CD3+CD4+CD45RA+
memory CD4 ⁺ T cells (MemoryTh)	CD3+CD4+CD45RA-
T helper type 1cells (Th1)	CD3+CD4+CD45RA-CXCR5-CXCR3+CCR6-
T helper type 2 cells (Th2)	CD3+CD4+CD45RA-CXCR5-CXCR3-CCR6-
T helper type 17 cells (Th17)	CD3+CD4+CD45RA-CXCR5-CXCR3-CCR6+
T helper type1/17 cells (Th1.17)	CD3+CD4+CD45RA-CXCR5-CXCR3+CCR6+
follicular helper T cells (Tfh)	CD3+CD4+CD45RA-CXCR5+
follicular helper T cells/Th1 cells (Tfh1)	CD3+CD4+CD45RA-CXCR5+CXCR3+CCR6-
follicular helper T cells/Th2 cells (Tfh2)	CD3+CD4+CD45RA-CXCR5+CXCR3-CCR6-
follicular helper T cells/Th17 cells	
(Tfh17)	CD3+CD4+CD45RA-CXCR5+CXCR3-CCR6+
regulatory T cells (Treg)	CD3+CD4+CD45RA-CD25+CD127low
B cells	CD3-CD19+ (parent population)
naive B cells (NaiveB)	CD3-CD19+IgD+CD27-
unswitched memory B cells (unSwMB)	CD3-CD19+IgD+CD27+
switched memory B cells (SwMB)	CD3-CD19+IgD-CD27+
Double-negative B cells (DNB)	CD3-CD19+IgD-CD27-
transitional B cells (TransB)	CD3-CD19+CD24highCD38high
plasmablasts (PB)	CD3-CD19+IgD-CD27highCD38high
natural killer cells (NK)	CD3-CD19-CD14-CD56+ (parent population)
CD56 ^{high} NK cells (NK56high)	CD3-CD19-CD14-CD56highCD16-
CD56 ^{low} NK cells (NK56low)	CD3-CD19-CD14-CD56lowCD16+
	CD3-CD19-CD56-HLA-DR+CD14+ (parent
monocytes	population)
CD14 ^{bright} CD16 ⁻ monocytes (MO14b16n)	CD3-CD19-CD56-HLA-DR+CD14brightCD16-
CD14 ^{bright} CD16 ⁺ monocytes (MO14b16p)	CD3-CD19-CD56-HLA-DR+CD14brightCD16+
CD14 ^{dim} CD16 ⁺ monocytes (MO14d16p)	CD3-CD19-CD56-HLA-DR+CD14dimCD16+
	CD3-CD19-CD56-HLA-DR+CD14- (parent
dendritic cells	population)
myeloid dendritic cells (mDC)	CD3-CD19-CD56-HLA-DR+CD14-CD11c+CD123-
plasmacytoid dendritic cells (pDC)	CD3-CD19-CD56-HLA-DR+CD14-CD11c-CD123+

Table S2. Immunophenotyping subset definitions

variable	Univariate regression		Multivariate regression	
	в	P value	в	P value
age	0.19	0.08	Not selected	
sex	-0.033	0.76	Not selected	
disease duration	-0.0087	0.94	-0.18	0.26
shared epitope	0.000	Not selec		
positivity	0.028	0.79		
RF titer	0.11	0.32	Not selected	
ACPA titer	0.25	0.022*	Not selected	
NaiveTh.CXCR4	0.35	0.043*	Not selected	
MemoryTh.CXCR4	0.52	0.0018**	0.52	0.0023**
Th1.CXCR4	0.46	0.0059**	Not selected	
Th2.CXCR4	0.42	0.013*	Not selected	
Th17.CXCR4	0.41	0.016*	Not selected	
Th1.17.CXCR4	0.45	0.0078**	Not selected	
Tfh.CXCR4	0.45	0.0074**	Not selected	
Tfh1.CXCR4	0.45	0.0080**	Not selected	
Tfh2.CXCR4	0.45	0.0070**	Not selected	
Tfh17.CXCR4	0.45	0.0074**	Not selected	
Treg.CXCR4	0.43	0.012*	Not selected	

Table S3. Univariate and multivariate relationships between CXCR4 expression on CD4+ T cell subsets and DAS28esr

RA n=32. *p<0.05, **p<0.01

RF, Rheumatoid Factor; ACPA, anti-cyclic citrullinated peptide antibody;

DAS28esr, Disease Activity Score 28 joints-ESR





























D



CXCR4+ memory CD4+ T cells
 CXCR4- memory CD4+ T cells



relative expression





