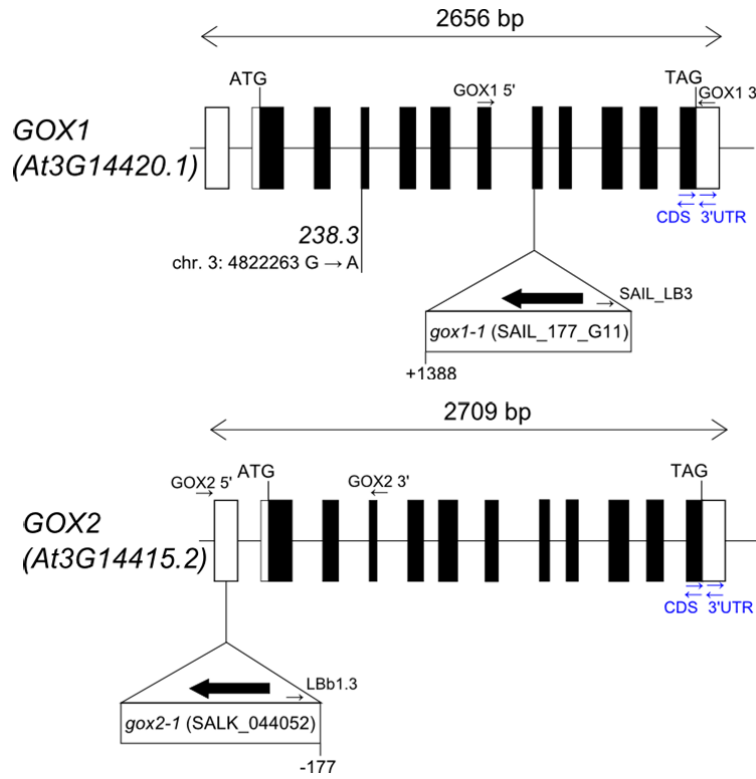


1 **Supplemental Figure S1.**

2



3

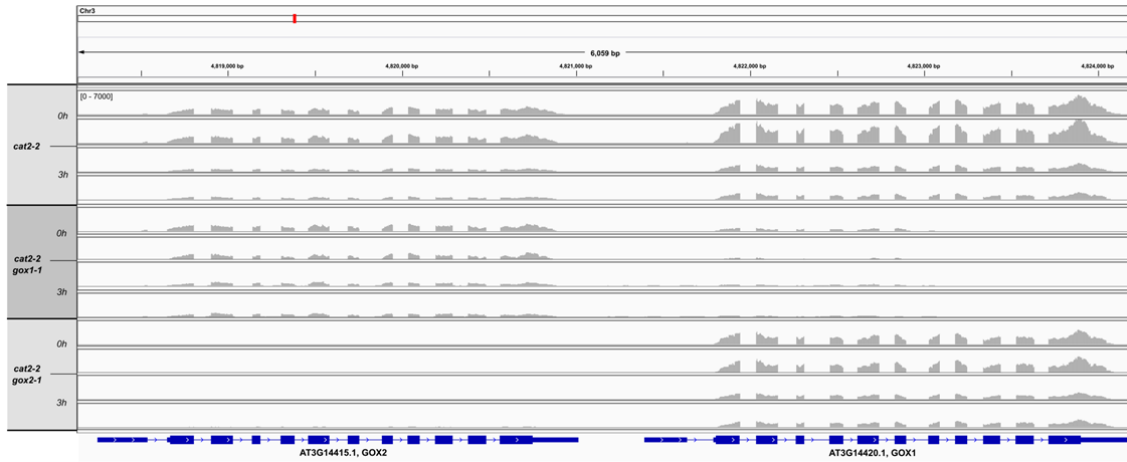
4

5 **Supplemental Figure S1.** *GOX1* and *GOX2* gene models, together with T-DNA insertion  
6 sites in the knockout SAIL\_177\_G11 (*gox1-1*) and SALK\_044052 (*gox2-1*) lines, as well as  
7 the EMS-induced single nucleotide polymorphism in line 238.3. Black and white boxes  
8 represent protein-coding and untranslated regions, respectively. Black arrows indicate primers  
9 used for genotyping and blue arrows – primers used for qRT-PCR.

10

11 **Supplemental Figure S2.**

12



13

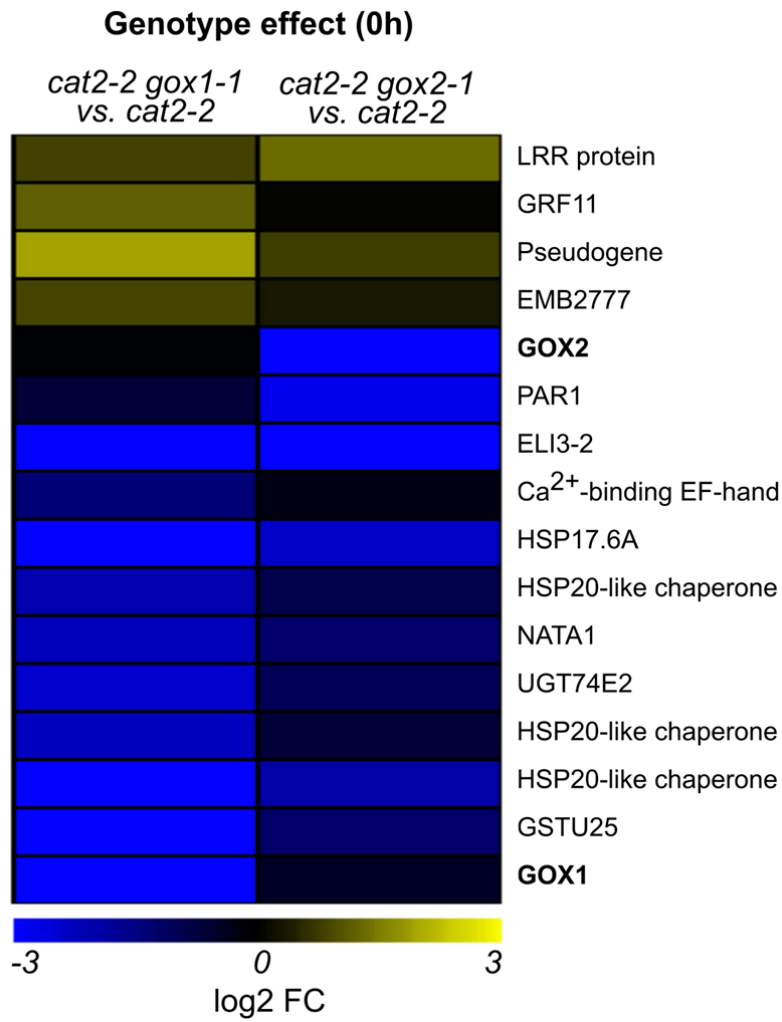
14

15 **Supplemental Figure S2.** Read coverage of *GOX1* and *GOX2* loci from RNA-seq analysis of  
16 *cat2-2 gox1-1*, *cat2-2* and *cat2-2 gox2-1* mutants. Aligning of reads obtained by RNA-seq  
17 from *cat2-2*, *cat2-2 gox1-1* and *cat2-2 gox2-2* mutants before (0h) and after exposure to  
18 photorespiratory stress (3h) to the reference sequences of *GOX1* and *GOX2*.

19

20 **Supplemental Figure S3.**

21



22

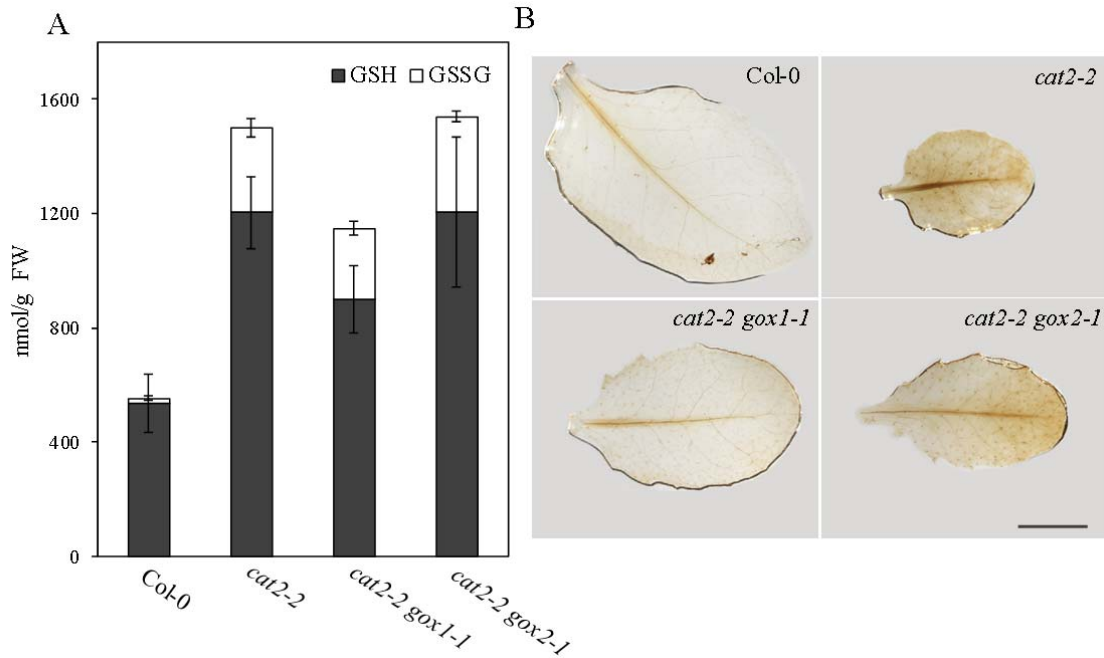
23

24 **Supplemental Figure S3.** Heat map of differentially expressed transcripts ( $|\log_2 \text{FC}| > 1$ ,  
 25  $\text{FDR} < 0.05$ ) between *cat2-2 gox1-1*, *cat2-2 gox2-1* and *cat2-2* mutants under control high  
 26  $\text{CO}_2$  conditions aimed at inhibiting the photorespiratory flux.

27

28 **Supplemental Figure S4.**

29



30

31

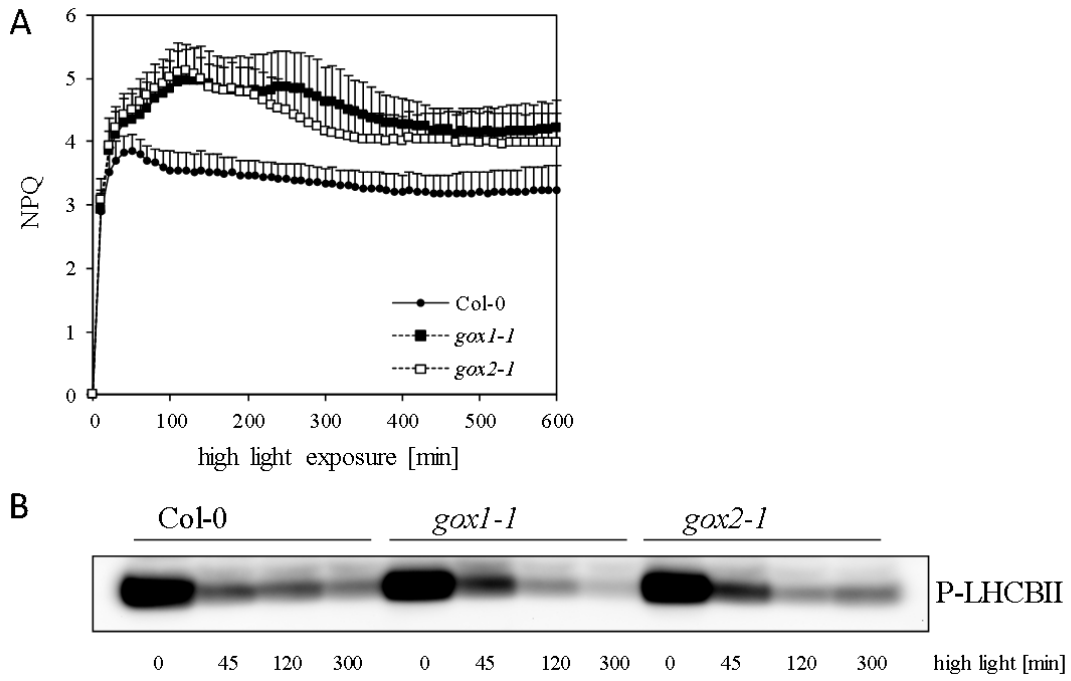
32

33 **Supplemental Figure S4.** Redox status of Col-0, *cat2-2*, *cat2-2 gox1-1* and *cat2-2 gox2-1*  
34 plants grown under ambient air and moderate light intensity ( $300 \mu\text{mol m}^{-2} \text{s}^{-1}$ ). A) Levels of  
35 reduced and oxidized glutathione (GSH and GSSG, respectively) in rosettes of three-week-old  
36 plants. Bars represent means of three biological replicates  $\pm$  SE. (B) Representative bright-  
37 field images of 3,3'-diaminobenzidine-stained mature leaves of three-week-old plants. The  
38 scale bar is 0,5 cm.

39

40 **Supplemental Figure S5.**

41



42

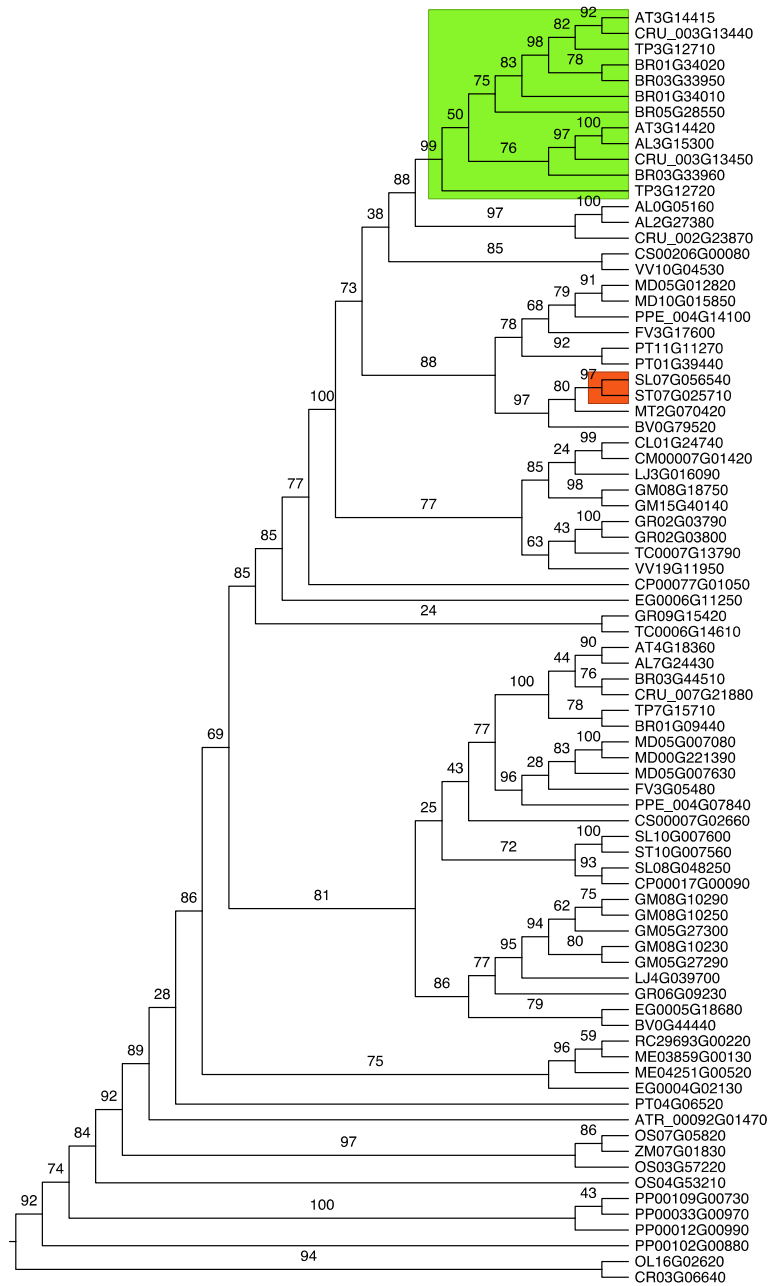
43

44 **Supplemental Figure S5.** Non-photochemical quenching (NPQ; A) and phosphorylation of  
45 LHCbII (B) upon exposure of Col-0, *gox1-1* and *gox2-1* plants grown under conditions  
46 limiting photorespiration to high light.

47

48 **Supplemental Figure S6.**

49



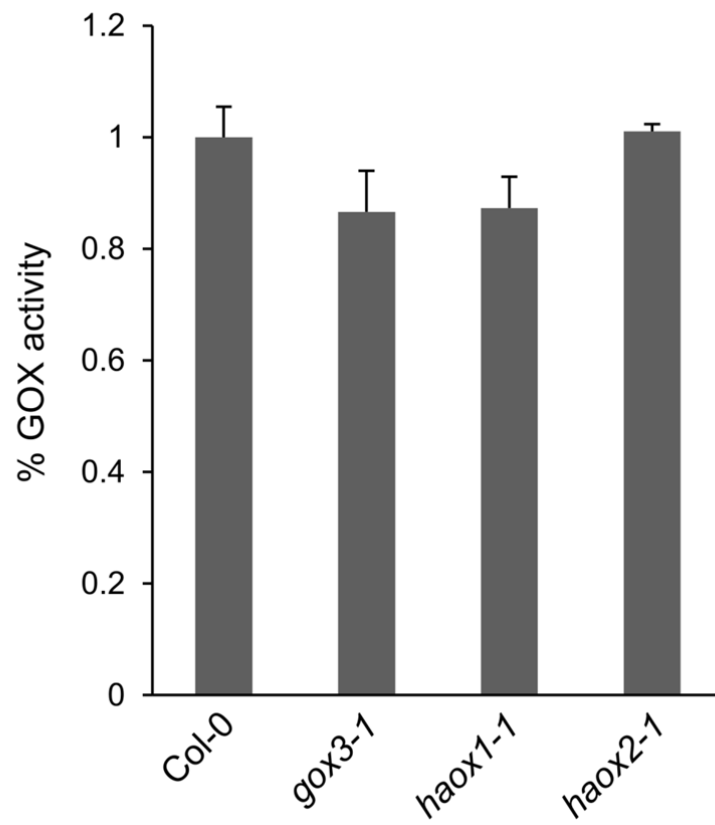
50

51 **Supplemental Figure S6.** *GOX1* and *GOX2* genes in *Brassicaceae* are clustered as  
 52 monophyletic group in the gene tree of ORTHO03D000507 from PLAZA 3.0 Dicots. *GOX1*  
 53 and *GOX2* in *Brassicaceae* are in green, and the two *GOX* genes from *Solanaceae* identified  
 54 based on synteny are shown in red. Numbers on branches denote bootstrap values.

55

56 **Supplemental Figure S7.**

57



58

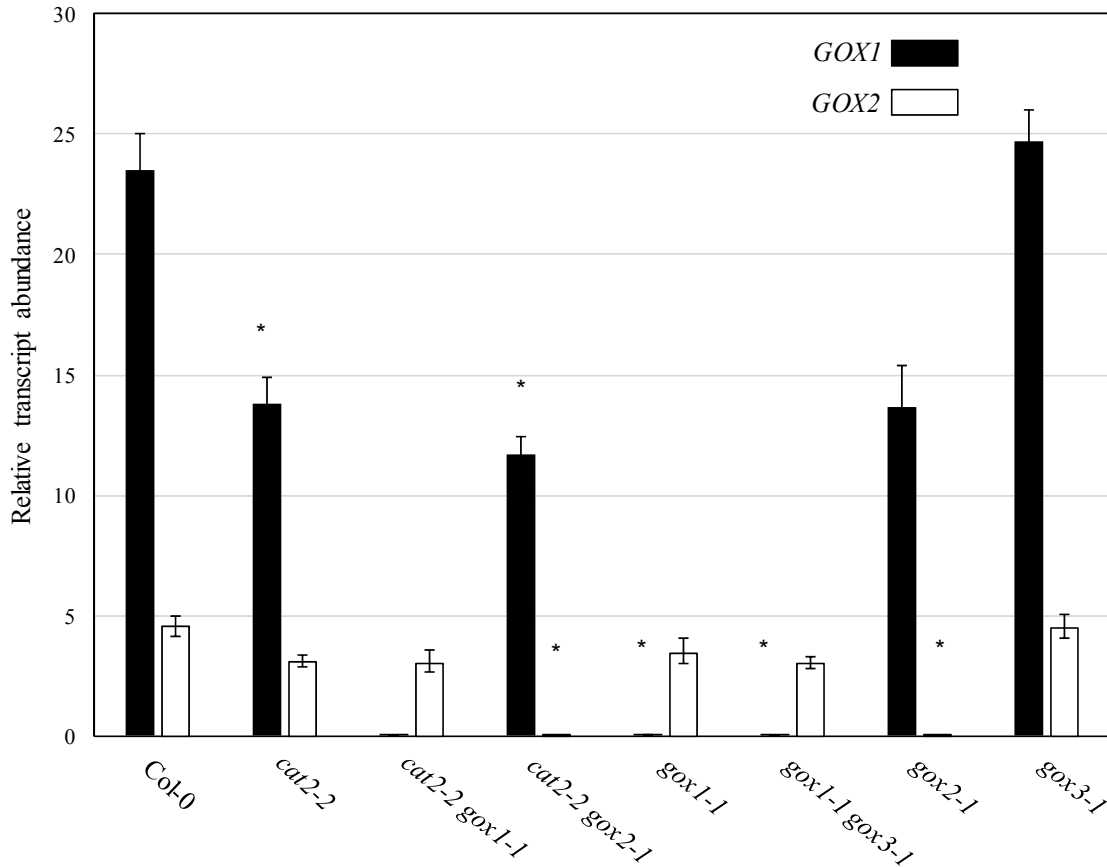
59

60 **Supplemental Figure S7.** Extractable leaf GOX activity. Bars represent averages from three  
61 biological replicates  $\pm$  SE.

62

63 **Supplemental Figure S8.**

64



65

66

67 **Supplemental Figure S8.** *GOX1* and *GOX2* transcript abundance. RNA was extracted from  
68 rosettes of three-week-old plants grown *in vitro* and used to quantify gene expression levels  
69 by qRT-PCR. Values are means  $\pm$  SD of four biological replicates. Asterisks indicate  
70 significant difference ( $p < 0.05$ ) in comparison to Col-0 according to one-way ANOVA.

71



72 **Supplemental Table S1.** Maximum likelihood estimates of parameters under branch models  
 73 on both post-duplication branches leading to the *GOX1* and *GOX2* clades.

74

<b>Model</b>	<b><math>p</math></b>	<b><math>\ell</math></b>	<b><math>\kappa</math></b>	<b><math>\omega_0</math></b>	<b><math>\omega_{GOX1}</math></b>	<b><math>\omega_{GOX2}</math></b>
A: One ratio: $\omega_0 = \omega_{GOX1} = \omega_{GOX2}$	33	-5437.64	1.71	0.070	0.070	0.070
B: Two ratios: $\omega_0 = \omega_{GOX1}, \omega_{GOX2}$	34	-5432.37	1.69	0.067	0.067	31.293
C: Two ratios: $\omega_0 = \omega_{GOX2}, \omega_{GOX1}$	34	-5436.67	1.71	0.069	0.233	0.069
D: Two ratios: $\omega_0, \omega_{GOX1} = \omega_{GOX2}$	34	-5433.50	1.71	0.067	0.291	0.291
E: Three ratios: $\omega_0, \omega_{GOX1}, \omega_{GOX2}$	35	-5432.21	1.70	0.066	0.105	3.941
F: Two ratios: $\omega_0 = \omega_{GOX1}, \omega_{GOX2} = 1$	33	-5432.55	1.70	0.067	0.067	1.000
G: Two ratios: $\omega_0 = \omega_{GOX2}, \omega_{GOX1} = 1$	33	-5437.34	1.72	0.068	1.000	0.068
H: Two ratios: $\omega_0, \omega_{GOX1} = \omega_{GOX2} = 1$	33	-5436.17	1.72	0.066	1.000	1.000
I: Three ratios: $\omega_0, \omega_{GOX1}, \omega_{GOX2} = 1$	34	-5432.30	1.70	0.067	0.116	1.000
J: Three ratios: $\omega_0, \omega_{GOX2}, \omega_{GOX1} = 1$	34	-5435.07	1.73	0.066	1.000	0.275

75

76  $p$ , number of free parameters in the model;  $\ell$ , log-likelihood;  $\kappa$ , the ratio of transition to  
 77 transversion;  $\omega_{GOX1}$  and  $\omega_{GOX2}$ , the ratio of non-synonymous substitution rate (dN) to  
 78 synonymous substitution rate (dS) on the two post-duplication branches to *GOX1* and *GOX2*  
 79 clades, respectively;  $\omega_0$ , dN/dS for rest of the branches on the tree

80

81 **Supplemental Table S2.** Likelihood ratio tests for comparing different branch models  
82 applied to both post-duplication branches leading to the *GOX1* and *GOX2* clades.

83

<b>Model comparisons</b>	<b>2<math>\Delta\ell</math></b>	<b>df</b>	<b>p value</b>
A-D	8.295754	1	0.003973788
A-B	10.542752	1	0.001166446
C-E	8.923886	1	0.00281465
A-C	1.944748	1	0.16315393
B-E	0.325882	1	0.568094036
D-H	5.342664	1	0.020809648
B-F	0.348118	1	0.555180423
E-I	0.177828	1	0.673246506
C-G	1.331848	1	0.248476723
E-J	5.729078	1	0.016686241

84

85 2 $\Delta\ell$ , likelyhood ratio statistics; df, degree of freedom

86

87 **Supplemental Table S3.** The modified branch-site models for detecting positive selection on  
88 six branches after the duplication event leading to *GOX2* in *Arabidopsis*.

89

<b>Branch</b>	<b><math>\ell_{H0}</math></b>	<b><math>\ell_{HA}</math></b>	<b><math>2\Delta\ell</math></b>	<b>df</b>	<b>p value</b>
#1	-5401.887282	-5401.781842	0.21088	1	0.32303927
#2	-5401.767414	-5401.577101	0.38062	1	0.26863420
#3	-5402.937338	-5402.85316	0.16835	1	0.34078851
#4	-5403.195792	-5403.195793	2E-06	1	0.49943581
#5	-5403.170388	-5403.170388	0	1	0.5
#6	-5402.919624	-5402.047157	1.74493	1	0.09325743

90

91  $\ell_{H0}$ , log likelihood for the branch-site model without positive selection;  $\ell_{HA}$ , log likelihood for  
92 the branch-site model with positive selection;  $2\Delta\ell$ , likelihood ratio statistics; df, degree of  
93 freedom

94

95 **Supplemental Table S4.** List of primers used in the study.

96

<b>Primer name</b>	<b>Use</b>	<b>Sequence</b>
gox1_SAIL_177_G11_newLP1	Genotyping	GGGAATCACAAGAAATTGATC
gox1_SAIL_177_G11_RP	Genotyping	GAAGGACTTGACCTCGGAAAG
gox2_SALK_044052_LP	Genotyping	ATACCATGATGGTTCCAGCAG
gox2_SALK_044052_RP	Genotyping	TTGTTATGTTTCGTCAAGCCC
gox3_GABI_523D09_LP	Genotyping	TTGGATGGGAATACTTTATTGG
gox3_GABI_523D09_RP	Genotyping	GCATCAACCTTTTGCTTGAAG
haox1_SAIL_84_A04_newLP2	Genotyping	AGAGGACAAGAGGAGCAAGATTC
haox1_SAIL_84_A04_RP	Genotyping	GCGCTTTGAAAACATCTGTTC
haox2_SALK_102409_LP	Genotyping	GGAAACTTGGTCTCCATCAGG
haox2_SALK_102409_RP	Genotyping	GCTCGACTATCCCCCTGCTAC
LBb1.3	Genotyping	ATTTTGCCGATTCGGAAC
SAIL_LB3	Genotyping	TAGCATCTGAATTCATAACCAATCTCGATACAC
GK_o8474	Genotyping	ATAATAACGCTGCGGACATCTACATTTT
SALK_newLB1	Genotyping	TGGACCGCTTGCTGCAACTCTC
CAT2_LP	Genotyping	CCCAGAGGTACCTCTTCTTCTCCCATG
CAT2_RP	Genotyping	TCAGGGAACCTTCATCCCATCGC
GOX1_F_CDS	qRT-PCR	TTCATTTGGCAGCTGAAGGA
GOX1_R_CDS	qRT-PCR	GAGTGTCCCATTTCGGTGGTA
GOX2_F_CDS	qRT-PCR	TTTGCCTAGCTGCTGAAGGA
GOX2_R_CDS	qRT-PCR	ATAACCTGGGCAAATGGCGT
GOX1_F	qRT-PCR	AGAACAGCAGCAACACAGAAC
GOX1_R	qRT-PCR	CACTAGGCTTGGTTTGTGATCTGATA
GOX2_F	qRT-PCR	ATATCTCGAAAACTTGTTTCTCCCTATAT
GOX2_R	qRT-PCR	ATCTATTGTACAAGCAATAAGAATAAACGG
EF1a_F	qRT-PCR	TCCGTCGGAGCTCAATTCTC
EF1a_R	qRT-PCR	AGGAAGCTCGAGTGCCAAGTAC
UBQ5_F	qRT-PCR	CTGCATTTCTATTTGGGAATTTTGTA
UBQ5_R	qRT-PCR	ATCCAGAACGAAAGATGTTCAACATAG

97

98

99 **Supplemental Table S5.** Mutations enriched in line 238.3.

100

Chromosome	Position	Reference base	Mutant	Coverage	Frequency	Region	AGI code	Position in gene	Position in codon	Reference amino acid	Mutant
3	4822263	G	A	65	0.9	CDS	<i>At3g14420</i>	263	2	G	E
3	7636562	G	A	40	0.89	intergenic					
3	6290758	G	A	60	0.86	intergenic					
3	3807077	G	A	54	0.84	intergenic					
3	6145283	G	A	43	0.83	five prime UTR	<i>At3g17940</i>				
4	8981113	G	A	5	0.83	intronic/noncoding	<i>At4g15780</i>				
3	6021032	G	A	50	0.82	CDS	<i>At3g17600</i>	9	3	V	V
3	436295	C	T	60	0.8	CDS	<i>At3g02260</i>	11022	3	E	E
3	3397230	G	A	37	0.8	intronic/noncoding	<i>At3g10845</i>				
3	6031593	G	A	51	0.8	CDS	<i>At3g17630</i>	2223	3	K	K

101

102

103 **Supplemental Table S6.** Data sources and sequence accessions.

104

Data source	Species	Accession	
PLAZA 3.0 Dicots	<i>Arabidopsis thaliana</i>	AT3G14415	
		AT3G14420	
	<i>Thellungiella parvula</i>	TP3G12710	
		TP3G12720	
EnsemblPlants release 30	<i>Arabidopsis lyrata</i>	fgenesh2_kg.3__1563__AT3G14420.2	
		Bra021556	
		Bra021555	
		Bra001550	
		Bra001551	
	<i>Brassica rapa</i>	Bra027338	
		Bra027339	
		<i>Solanum lycopersicum</i>	Solyc07g056540.2
		<i>Solanum tuberosum</i>	PGSC0003DMG400027654
		Phytozome 9	<i>Capsella rubella</i>
Carubv10013998m.g			
<i>Thellungiella halophila</i>	Thhalv10020979m.g		
	Thhalv10020982m.g		

105