	MasterTaq (5 prime)	**AccuStart II PCR SuperMix (Quanta Biosciences)**	**JumpStart Taq (Sigma)**	**Terra Taq (Clontech)**	**Phusion Polymerase (NEB)**
Cycling	1. 97ºC; 30 sec 2. 94ºC; 15 sec 3. 55ºC; 30 sec 4. 72ºC; 30 sec +1 sec/cycle 39 cycles (steps 2-4) 5. 72ºC; 10 min	1. 95ºC; 2 min 2. 95ºC; 15 sec 3. 60ºC; 15 sec 4. 72ºC; 20 sec 35 cycles (steps 2-4)	1. 95ºC; 5 min 2. 95ºC; 30 sec 3. 60ºC; 30 sec 4. 68ºC; 40 sec 5. cycles (steps 2-4) 6. 68ºC; 5 min	1. 98ºC; 2 min 2. 98ºC; 10 sec 3. 60ºC; 15 sec 4. 68ºC; 1 min 26 cycles (steps 2-4)	1. 98ºC; 30 sec 2. 98ºC; 10 sec 3. 60ºC*; 30 sec 4. 72ºC; 30 sec 39 cycles (steps 2-4) 5. 72ºC; 10 min *Annealing temp of 55ºC was used for oligo cloning
PCR reaction	ddH_20 16.4 µL 10x Buffer 2.5 µL 5 mM dNTPs 1.0 µL 10 µM F primer 1.0 µL 10 µM R primer 1.0 µL DNA 3.0 µL Polymerase 0.10 µL	ddH_20 6.5 µL 2x Mix 12.5 µL 10 µM F primer 2.5 µL 10 µM R primer 2.5 µL DNA 1.0 µL	ddH_20 6.5 µL 2x Mix 12.5 µL 10 µM F primer 2.5 µL 10 µM R primer 2.5 µL DNA 1.0 µL	ddH_20 9.5 µL 2x Buffer 12.5 µL 10 µM F primer 0.75 µL 10 µM R primer 0.75 µL DNA 1.0 µL Taq 0.5 µL	ddH_20 14.25 µL 5x Buffer 5.0 µL 5 mM dNTPs 1.0 µL 10 µM F primer 1.25 µL 10 µM R primer 1.25 µL DNA or 10 ng oligo 2.0 µL Polymerase 0.25 µL

Table S2. **Cycling condition and reaction components for genotyping and cloning.**