

A consolidated analysis of the physiologic and molecular responses induced under acid stress in the legume-symbiont model-soil bacterium *Sinorhizobium meliloti*.

Draghi, W. O.¹; Del Papa, M. F.¹; Hellweg, C.²; Watt, S. A.²; Watt, T. F.²; Barsch, A.².
Lozano, M. J.¹; Lagares, A. (Jr.)⁴; Salas, M. E.¹; López, J. L.¹; Albicoro, F. J.¹; Nilson, J. F.¹; Torres Tejerizo, G. A.¹; Luna, M. F.³; Pistorio, M.¹; Boiardi, J. L.³; Pühler, A.²; Weidner; S.²; Niehaus, K.²; and Lagares A.^{1*}

¹ IBBM - Instituto de Biotecnología y Biología Molecular, CONICET - Departamento de Ciencias Biológicas, Facultad de Ciencias Exactas, Universidad Nacional de La Plata, calles 47 y 115, 1900-La Plata, Argentina

² CeBiTec - Centrum für Biotechnologie, Universität Bielefeld, Bielefeld, Germany

³ CINDEFI – Centro de Investigación y Desarrollo en Fermentaciones Industriales, CONICET - Departamento de Química, Facultad de Ciencias Exactas, Universidad Nacional de La Plata, calles 47 y 115, 1900-La Plata, Argentina

⁴ Laboratorio de Bioquímica, Microbiología e Interacciones Biológicas en el Suelo, Departamento de Ciencia y Tecnología, Universidad Nacional de Quilmes, Roque Sáenz Peña 352, Bernal B1876BXD, Buenos Aires, Argentina

Key words: acid stress, rhizobia, legume

***Corresponding author**

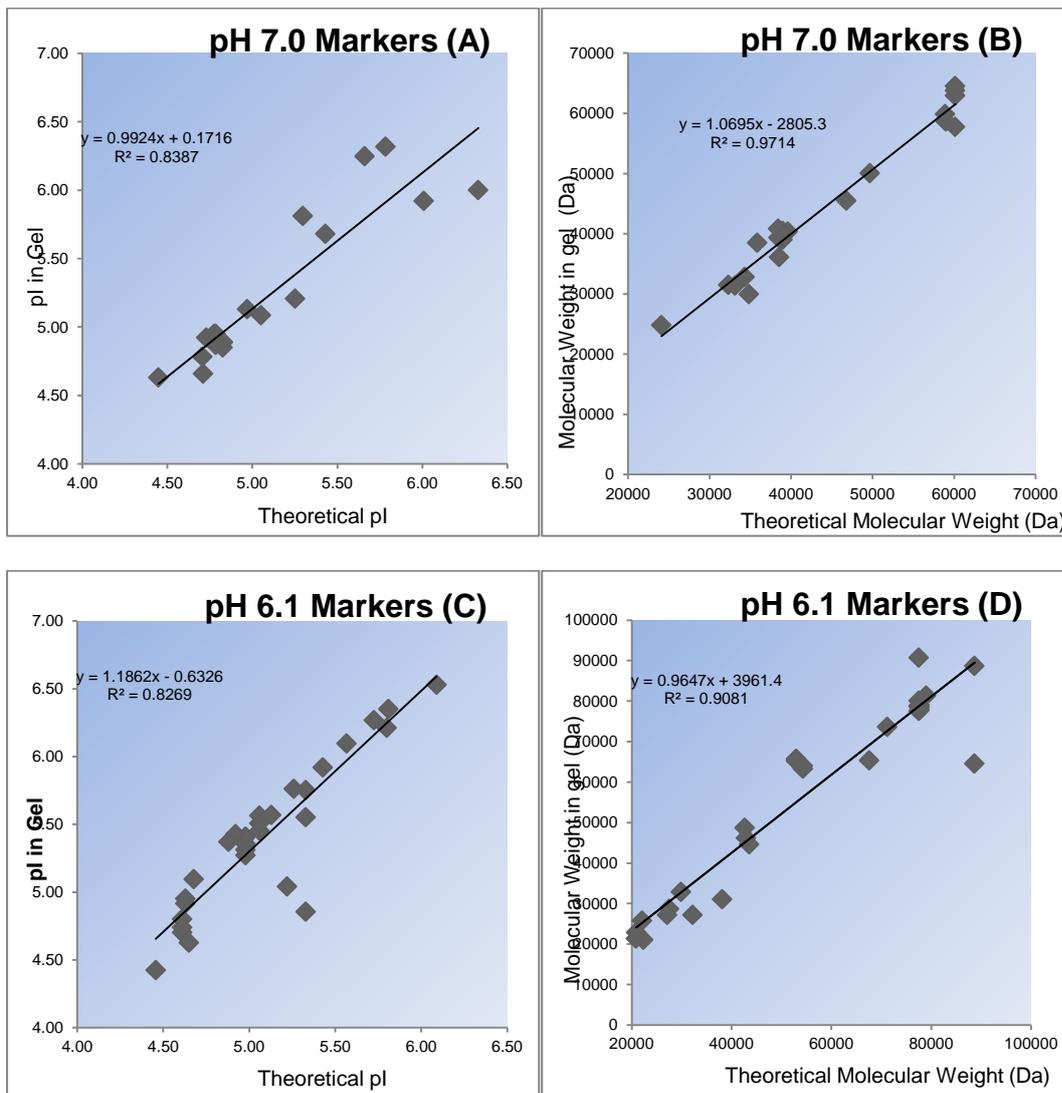
Phone: +54-221-425-0497 ext. 31

Fax: +54-221-422-3409 ext. 56

E-mail:lagares@biol.unlp.edu.ar

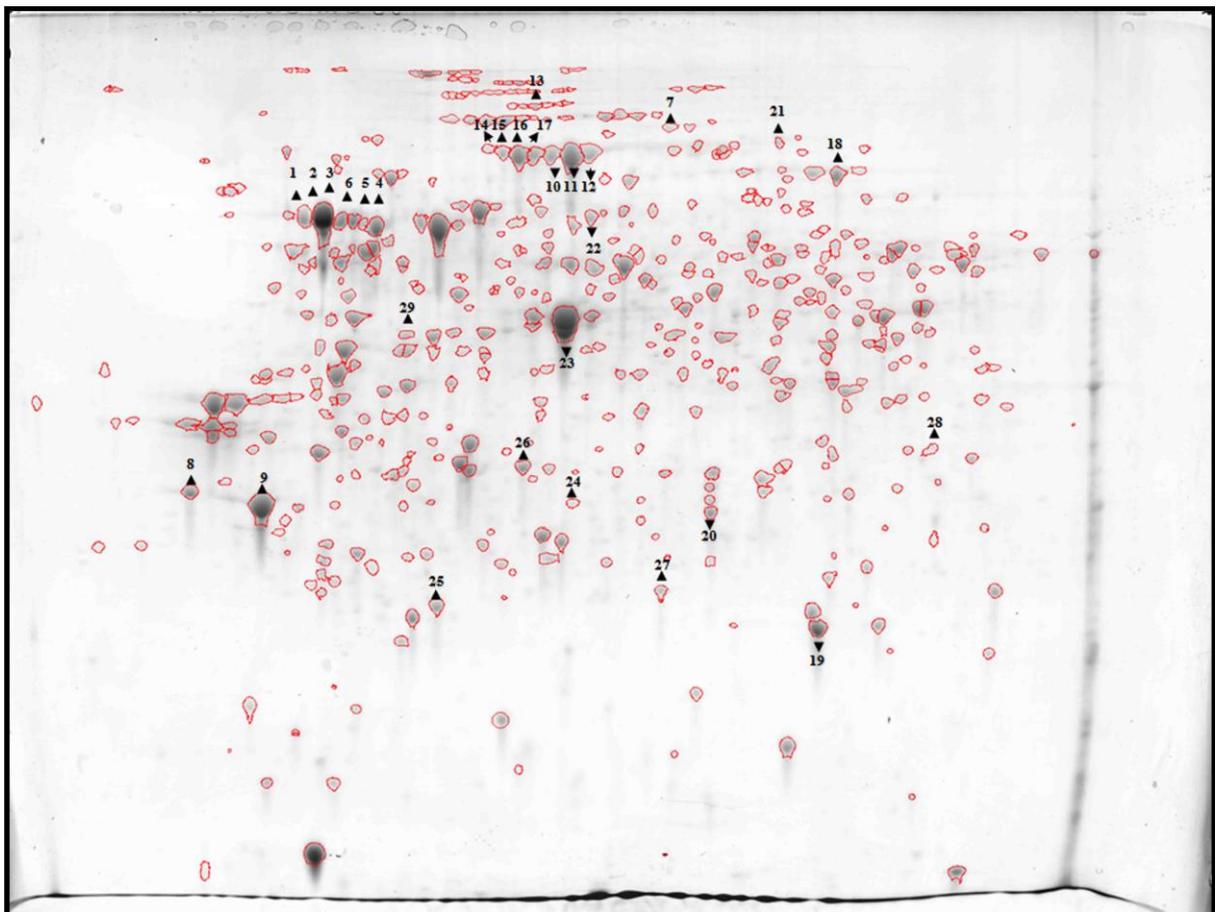
SUPPLEMENTARY DATA

Fig. S1. Correlation analysis between the experimental and the expected (calculated) values of molecular weight (Panel B, pH 7.0 and Panel D, pH 6.1) and isoelectric point (Panel A, pH 7.0 and Panel C, pH 6.1) of the protein markers identified by PMF in the gels shown in Figs. 1 and 2. The molecular weight and isoelectric-point (pI) values for each individual polypeptide were calculated from the corresponding amino-acid sequence (abscissa). Experimental values were inferred from the position in the 2-D gels by means of the *Image Master*TM software (*ordinate*). For most polypeptides a good correlation was obtained between the observed and the expected molecular weight and pI values.



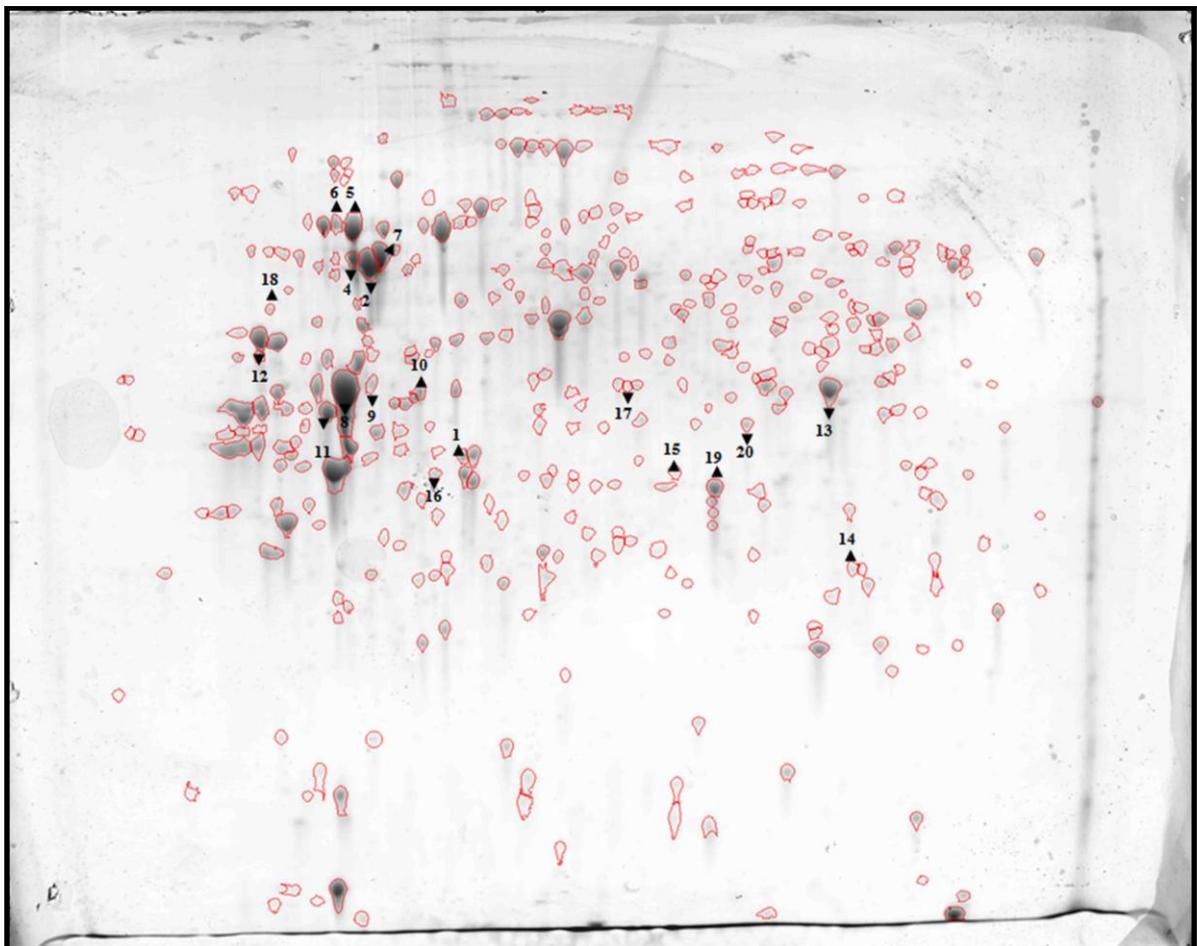
SUPPLEMENTARY DATA

Fig. S2. Image-Master detection of induced polypeptides in the cytosolic proteome of *S. meliloti* 2011 grown in chemostat at pH 6.1 compared to the expression of the corresponding polypeptides in the proteome of the same rhizobia grown at pH 7.0. The ratio of homologous polypeptides in one pH condition compared to the other was estimated by the ratio of the spot volumes (Image Master) after each one was normalized to the total intensity (all spots) from the corresponding gel. For each pH condition four independent gels were analyzed, including those from Figs.1 and 2. The numerical codes (in black) for each red-outlined spot in the figure (differentially expressed polypeptides) correspond to the respective numbers in the first column of Table 2.



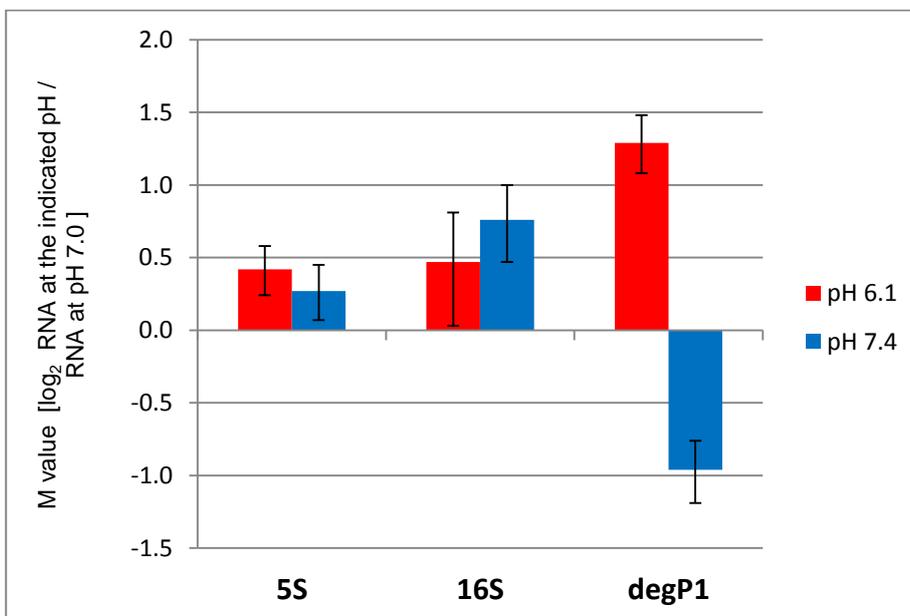
SUPPLEMENTARY DATA

Fig. S3. Image-Master detection of induced polypeptides in the cytosolic proteome of *S. meliloti* 2011 grown in chemostat at pH 7.0 compared to the expression of the corresponding polypeptides in the proteome of the same rhizobia grown at pH 6.1. The ratio of homologous polypeptides in one pH condition compared to the other was estimated by the ratio of the spot volumes (Image Master) after each one was normalized to the total intensity (all spots) from the corresponding gel. For each pH condition four independent gels were analyzed, including those from Figs.1 and 2. The numerical codes (in black) for each red-outlined spot in the figure (differentially expressed polypeptides) correspond to the respective numbers in the first column of Table 3.



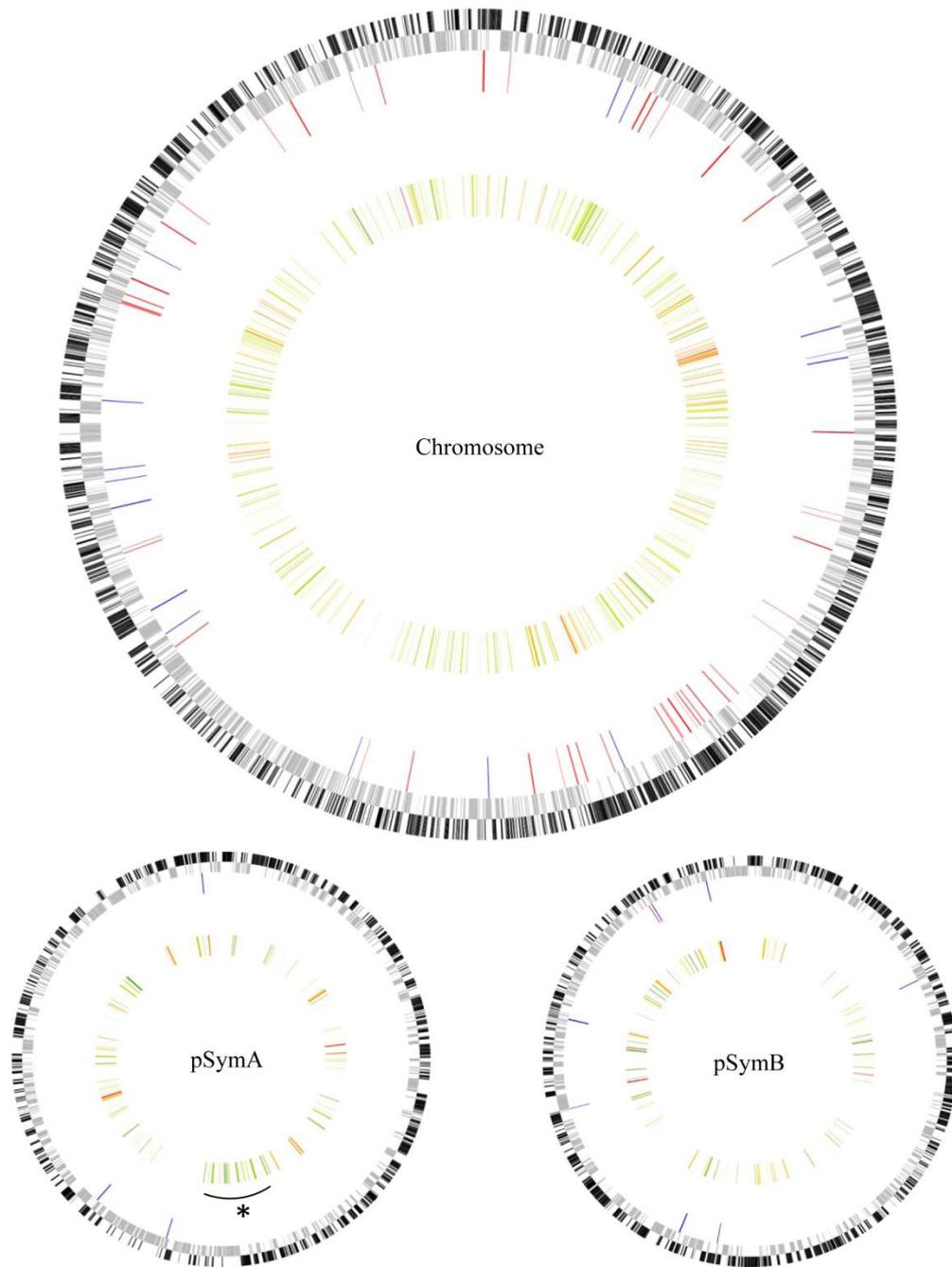
SUPPLEMENTARY DATA

Fig. S4. qRT-PCR estimation of 5S and 16S rRNAs in *S. meliloti* cells grown in the chemostat under steady state at pH 7.0, and at pH 6.1. The quantification of specific RNA species was carried out by quantitative reverse-transcription PCR using a KAPA SYBR FAST One-Step qRT-PCR kit (Kapa Biosystems Inc. Wilmington, MA, USA) according to the manufacturer's instructions. Analyses were performed using a same normalized amount of total RNA for all samples. M values represent the \log_2 of the fold change of the indicated RNA species at pH 6.1 (red bars) and 7.4 (blue bars) both with respect to their corresponding amounts at pH 7.0 (reference condition). 5S and 16S correspond to the analysis of 5S and 16S rRNA. *degP1* (SMc02365) corresponds to the analysis of the mRNA of an acid-induced gene used as positive control ($M^{\text{pH } 6.1 / \text{pH } 7.0}$ from the microarrays = 1.98). The DegP1 polypeptide was also induced in the proteome of acid-grown rhizobia (fold change *ca.* 4.8 in Table 2, so with an estimated $M^{\text{pH } 6.1 / \text{pH } 7.0}$ from the proteome = 2.26). The M values are the means calculated from three technical qRT-PCR replicas. The bars represent the standard deviations.



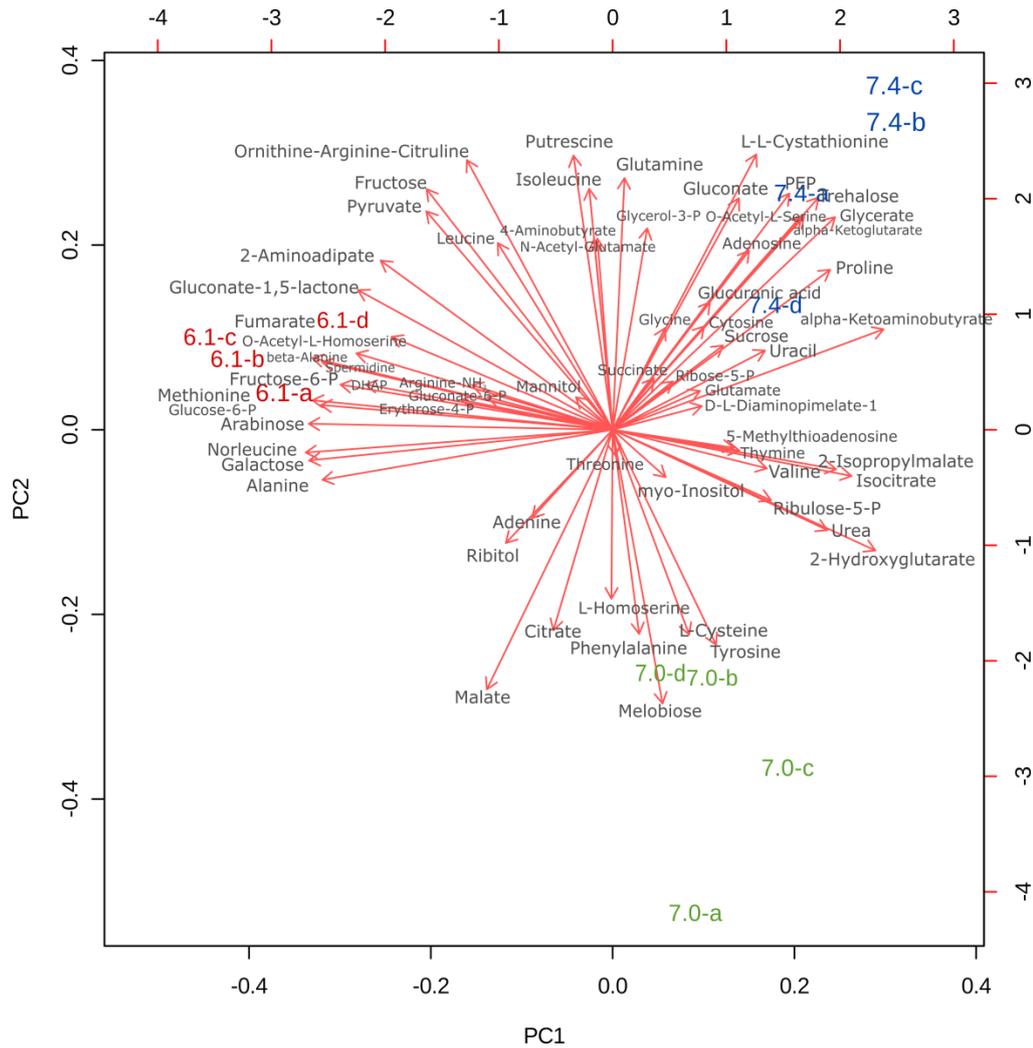
SUPPLEMENTARY DATA

Fig. S5. Genomic distribution of proteome and transcriptome markers differentially overexpressed at pH 6.1 versus 7.0. The two outermost circular array of lines on each replicon show—in black and in gray—the respective positions of open reading frames within the two complementary DNA strands. The next array inside shows—with red and blue lines—the genomic position of genes encoding polypeptides that were observed to be induced under acidic and neutral extracellular conditions, respectively (from the proteomic data). The innermost array marks—with green and orange lines—the genomic position of genes encoding RNAs that were also observed to be induced under acidic and neutral extracellular conditions, respectively (from the transcriptome data). The pSymA region characterized by a high density of overexpressed genes at pH 6.1 (*cf.* the text) is outlined and marked by an asterisk.

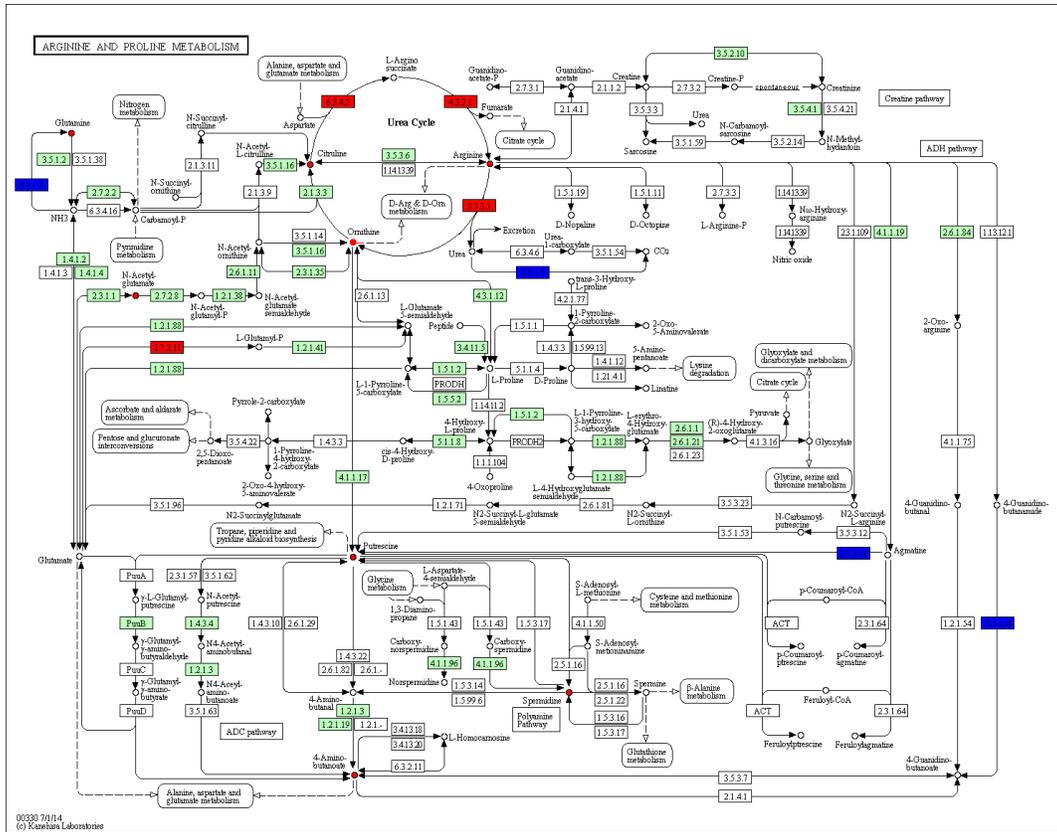


SUPPLEMENTARY DATA

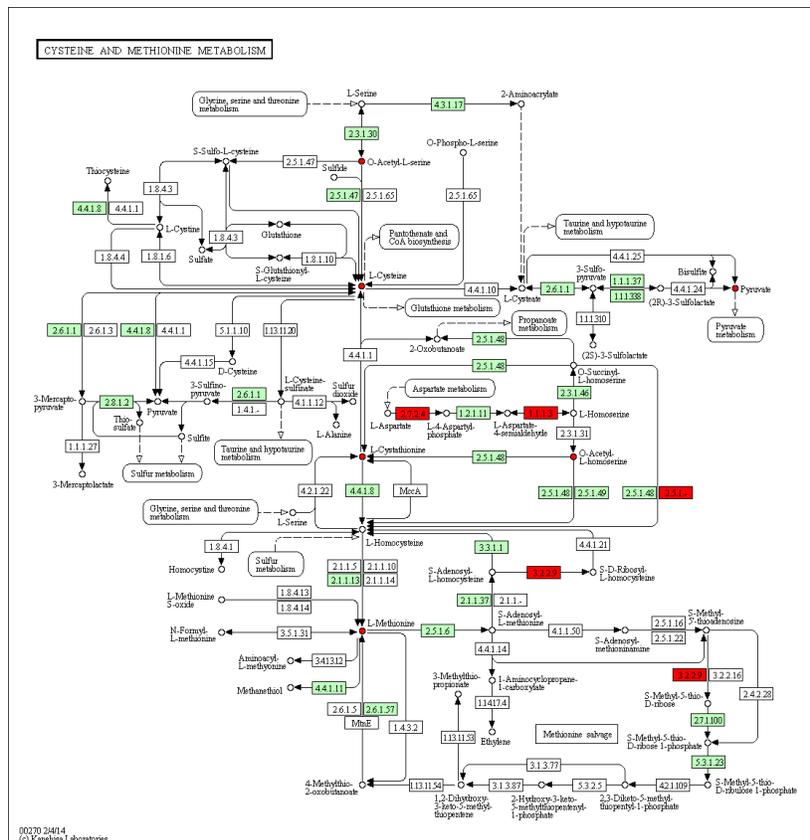
Fig. S6. Vector-correlation plot between the variables examined (*i. e.*, the amount of metabolites) and the PC1 and PC2 spaces of PCA. The direction and length of the vectors are indicative of the positive contribution of each individual metabolite to the position of each sample analyzed in the plot (*i. e.*, from rhizobia grown at pH 6.1, pH 7.0, or pH 7.4).



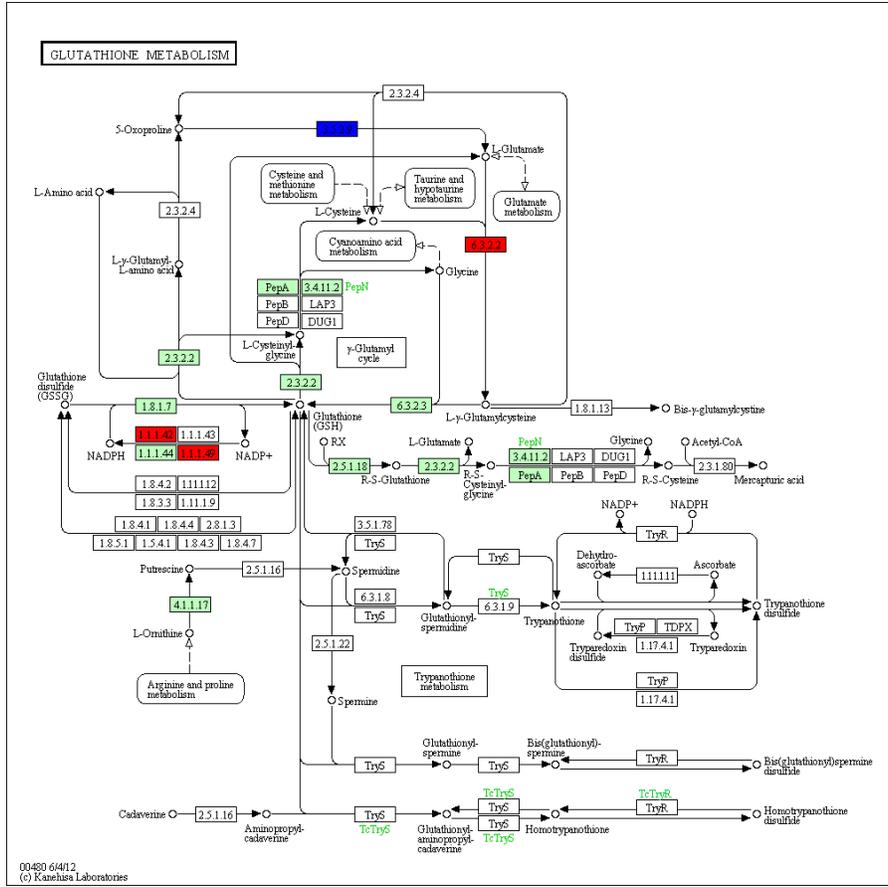
E



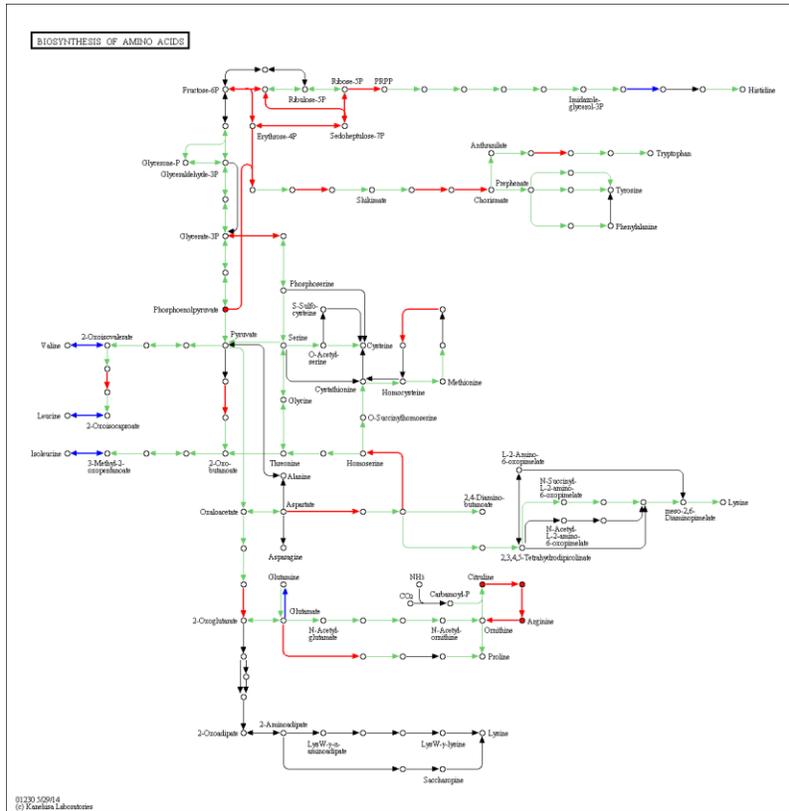
F



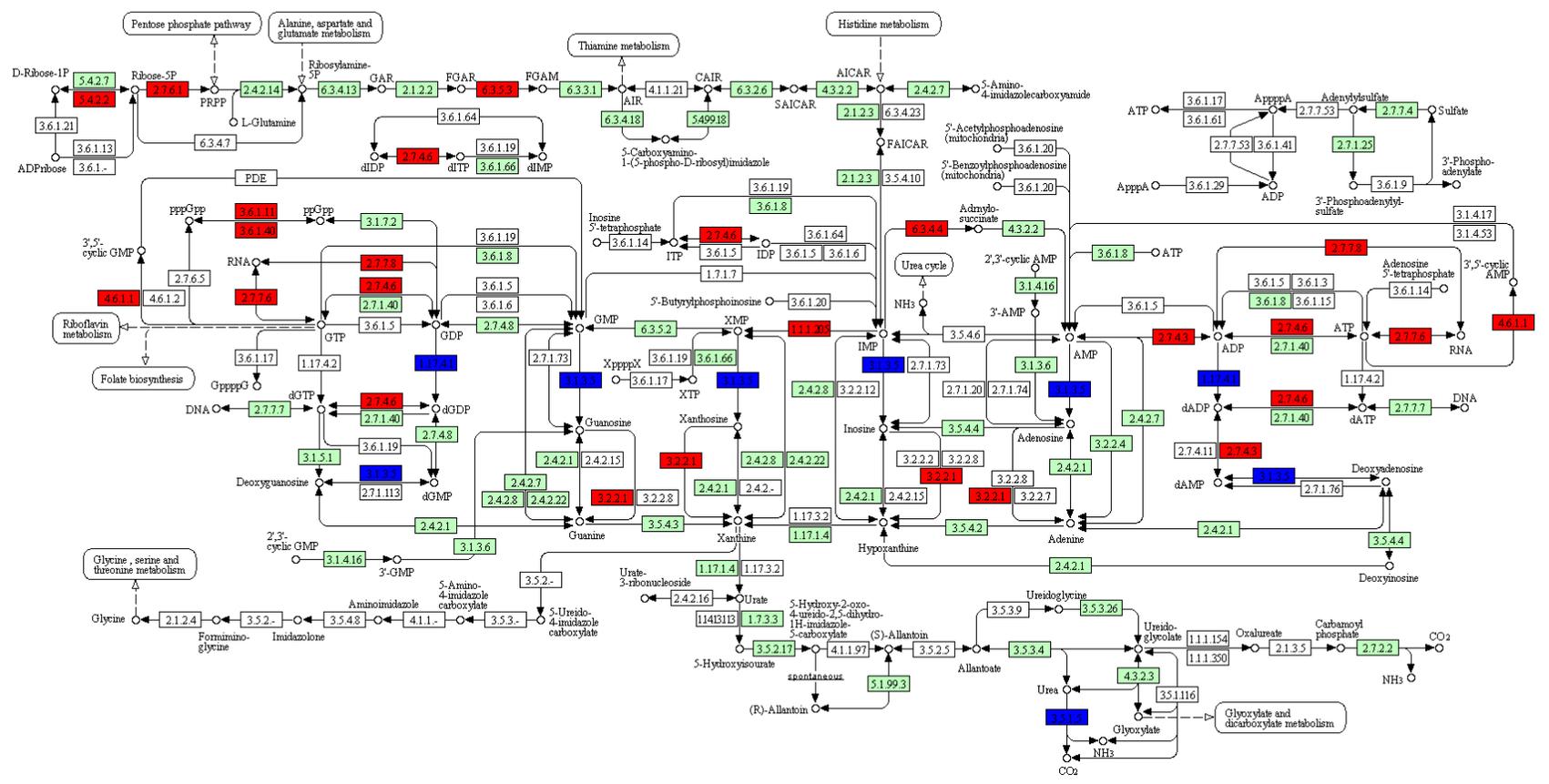
G



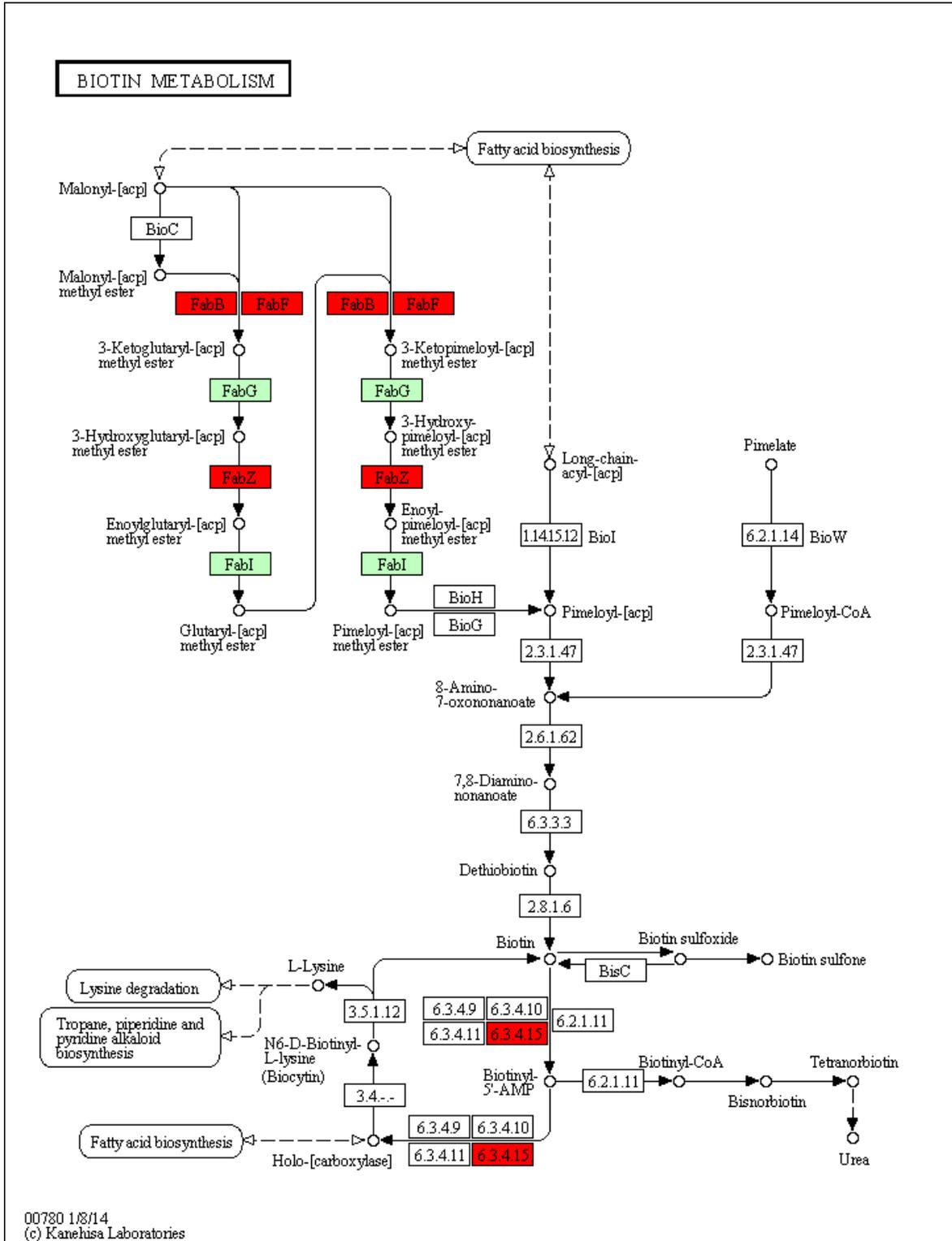
H



PURINE METABOLISM

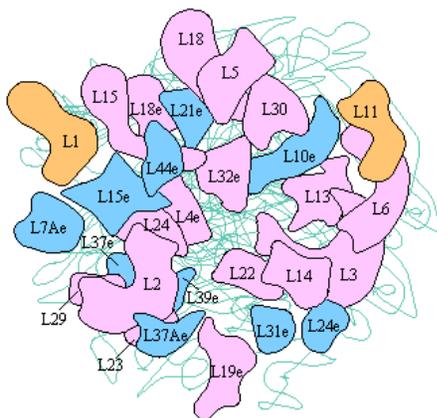


K

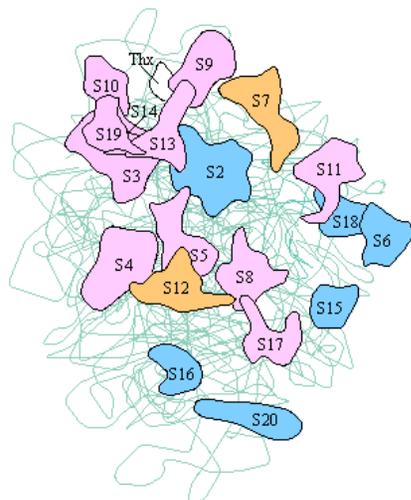


L

RIBOSOME



Large subunit (*Haloarcula marismortui*)

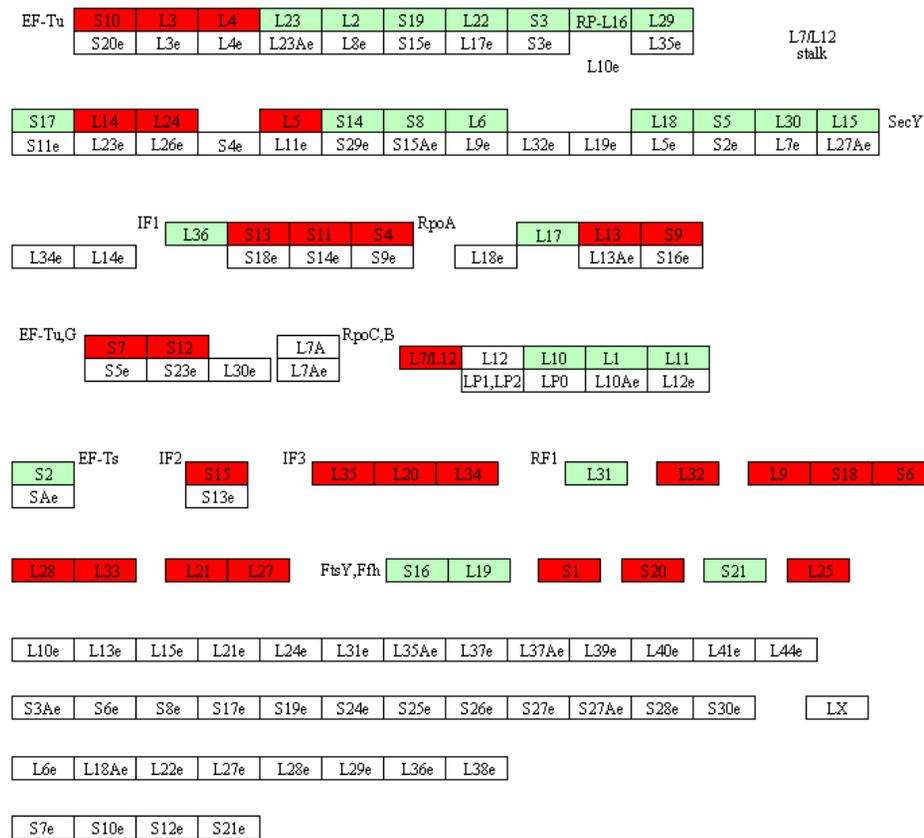


Small subunit (*Thermus aquaticus*)

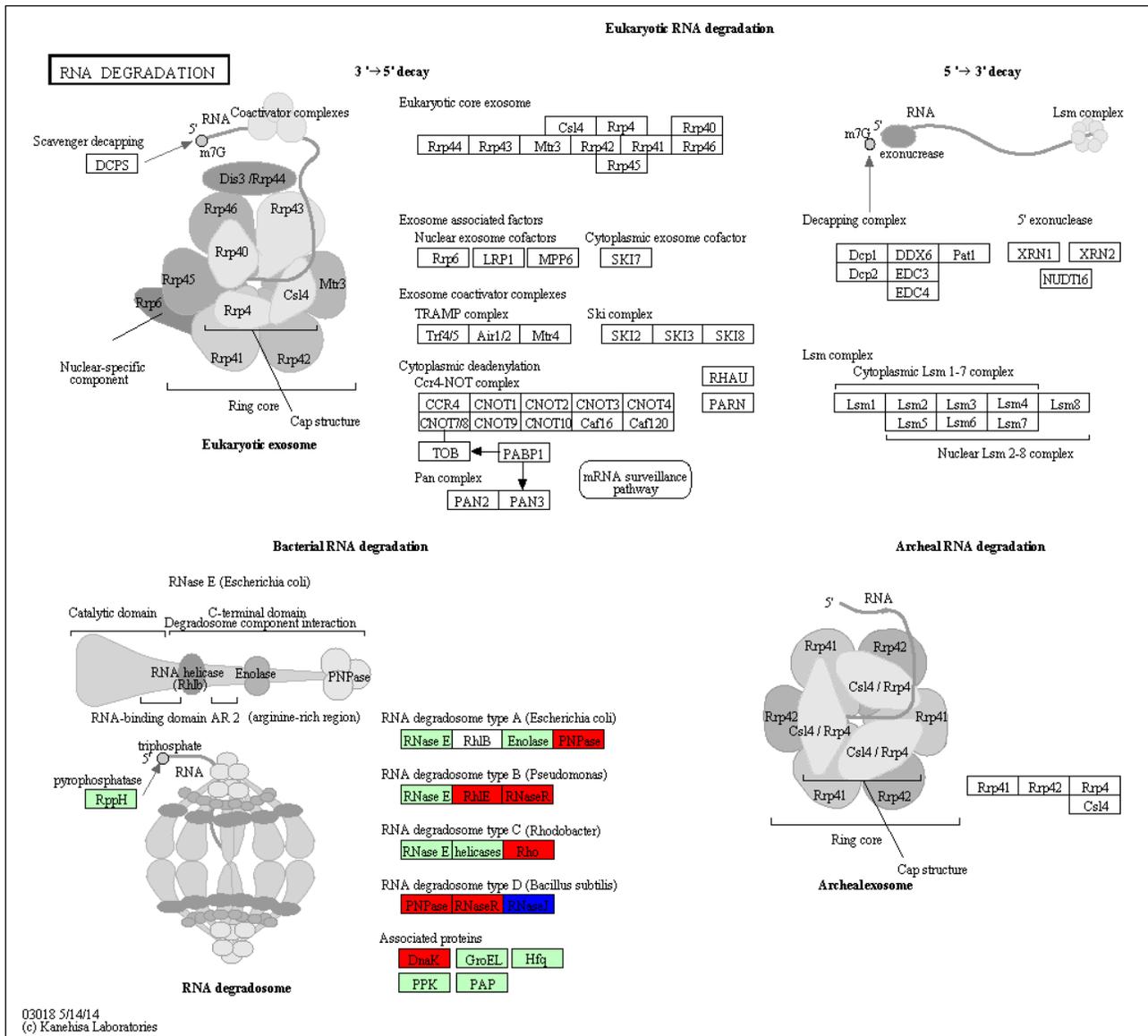
Ribosomal RNAs

| | | | | |
|--------------------|-----|----|------|-----|
| Bacteria / Archaea | 23S | 5S | | 16S |
| Eukaryotes | 25S | 5S | 5.8S | 18S |

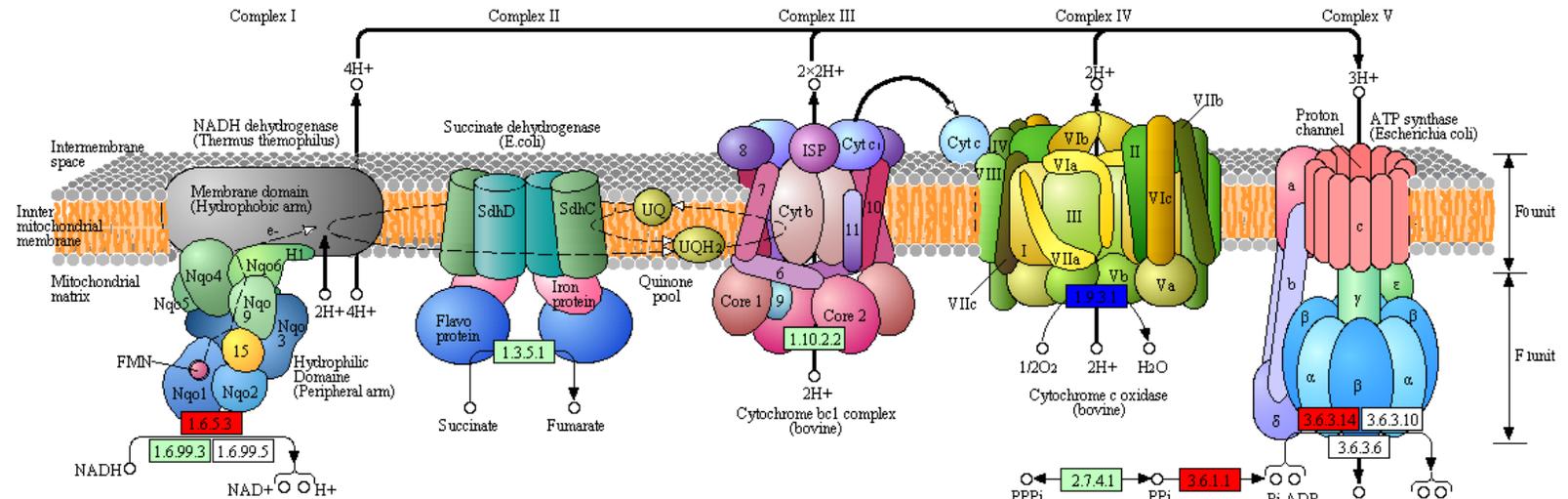
Ribosomal proteins



M



OXIDATIVE PHOSPHORYLATION



NADH dehydrogenase

| | | | | | | | | | | | | | | | | | |
|-----|--------|--------|--------|--------|--------|--------|--------|--------|--------|---------|---------|---------|---------|---------|------|------|------|
| E | ND1 | ND2 | ND3 | ND4 | ND4L | ND5 | ND6 | | | | | | | | | | |
| E | Ndufs1 | Ndufs2 | Ndufs3 | Ndufs4 | Ndufs5 | Ndufs6 | Ndufs7 | Ndufs8 | Ndufv1 | Ndufv2 | Ndufv3 | | | | | | |
| B/A | NuoA | NuoB | NuoC | NuoD | NuoE | NuoF | NuoG | NuoH | NuoI | NuoJ | NuoK | NuoL | NuoM | NuoN | | | |
| B/A | NdhC | NdhK | NdhJ | NdhH | NdhA | NdhI | NdhG | NdhE | NdhF | NdhD | NdhB | NdhL | NdhM | NdhN | HoxE | HoxF | HoxU |
| E | Ndufa1 | Ndufa2 | Ndufa3 | Ndufa4 | Ndufa5 | Ndufa6 | Ndufa7 | Ndufa8 | Ndufa9 | Ndufa10 | Ndufab1 | Ndufa11 | Ndufa12 | Ndufa13 | | | |
| E | Ndufb1 | Ndufb2 | Ndufb3 | Ndufb4 | Ndufb5 | Ndufb6 | Ndufb7 | Ndufb8 | Ndufb9 | Ndufb10 | Ndufb11 | Ndufc1 | Ndufc2 | | | | |

Succinate dehydrogenase / Fumarate reductase

| | | | | |
|-----|------|------|------|------|
| E | SDHC | SDHD | SDHA | SDHB |
| B/A | SdhC | SdhD | SdhA | SdhB |
| | FrdA | FrdB | FrdC | FrdD |

Cytochrome c reductase

| | | | | | | | |
|-------|------|-------|-------|------|------|------|-------|
| E/B/A | ISP | Cyt b | Cyt 1 | | | | |
| E | COR1 | QCR2 | QCR6 | QCR7 | QCR8 | QCR9 | QCR10 |

Cytochrome c oxidase

| | | | | | | | | | | | | | | | | | | |
|-----|-------|------|------|------|------|-------|-------|-------|-------|-------|-------|-------|-------|------|-------|-------|-------|-------|
| E | COX10 | COX3 | COX1 | COX2 | COX4 | COX5A | COX5B | COX6A | COX6B | COX6C | COX7A | COX7B | COX7C | COX8 | E/B/A | COX11 | COX15 | COX17 |
| B/A | CyoE | CyoD | CyoC | CyoB | CyoA | CoxD | CoxC | CoxA | CoxB | QoxD | QoxC | QoxB | QoxA | | | | | |

Cytochrome c oxidase, cbb3-type

| | | | | |
|---|---|----|----|-----|
| B | I | II | IV | III |
|---|---|----|----|-----|

Cytochrome bd complex

| | | |
|-----|------|------|
| B/A | CydA | CydB |
|-----|------|------|

F-type ATPase (Bacteria)

| | | | | |
|---------|---------|----------|----------|---------|
| 2.7.4.1 | 3.6.1.1 | 3.6.3.14 | 3.6.3.10 | 3.6.3.6 |
| alpha | beta | gamma | delta | epsilon |
| a | b | c | | |

F-type ATPase (Eukaryotes)

| | | | | | |
|-------|------|-------|-------|---------|---|
| alpha | beta | gamma | delta | epsilon | |
| OSCP | a | b | c | d | e |
| f | g | f6/a | j | k | 8 |

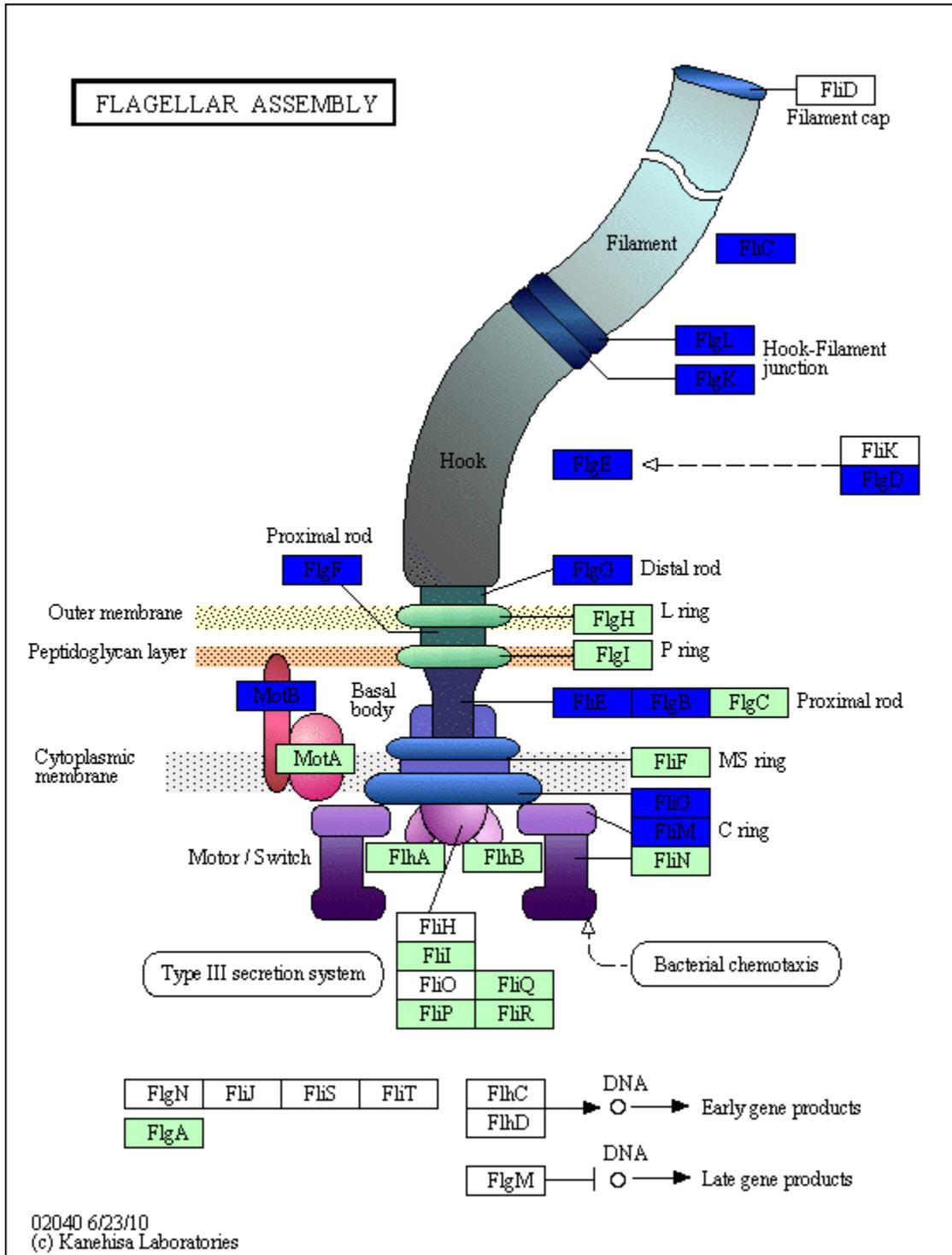
V/A-type ATPase (Bacteria, Archaeas)

| | | | | | | |
|---|---|---|---|---|---|-----|
| A | B | C | D | E | F | G/H |
| I | K | | | | | |

V-type ATPase (Eukaryotes)

| | | | | | | | |
|---|---|---|---|----|---|---|---|
| A | B | C | D | E | F | G | H |
| a | c | d | e | S1 | | | |

○



R

