A consolidated analysis of the physiologic and molecular responses induced under acid stress in the legume-symbiont model-soil bacterium *Sinorhizobium meliloti*.

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Fig. S1. Correlation analysis between the experimental and the expected (calculated) values of molecular weight (Panel B, pH 7.0 and Panel D, pH 6.1) and isolectric point (Panel A, pH 7.0 and Panel C, pH 6.1) of the protein markers identified by PMF in the gels shown in Figs. 1 and 2. The molecular weight and isoelectric-point (pI) values for each individual polypeptide were calculated from the corresponding amino-acid sequence (abscissa). Experimental values were inferred from the position in the 2-D gels by means of the *Image Master*TM software (*ordinate*). For most polypeptides a good correlation was obtained between the observed and the expected molecular weight and pI values.





Fig. S2. Image-Master detection of induced polypeptides in the cytosolic proteome of *S. meliloti* **2011 grown in chemostat at pH 6.1 compared to the expression of the corresponding polypeptides in the proteome of the same rhizobia grown at pH 7.0.** The ratio of homologous polypeptides in one pH condition compared to the other was estimated by the ratio of the spot volumes (Image Master) after each one was normalized to the total intensity (all spots) from the corresponding gel. For each pH condition four independent gels were analyzed, including those from Figs.1 and 2. The numerical codes (in black) for each red-outlined spot in the figure (differentially expressed polypeptides) correspond to the respective numbers in the first column of Table 2.



Fig. S3. Image-Master detection of induced polypeptides in the cytosolic proteome of *S. meliloti* **2011 grown in chemostat at pH 7.0 compared to the expression of the corresponding polypeptides in the proteome of the same rhizobia grown at pH 6.1.** The ratio of homologous polypeptides in one pH condition compared to the other was estimated by the ratio of the spot volumes (Image Master) after each one was normalized to the total intensity (all spots) from the corresponding gel. For each pH condition four independent gels were analyzed, including those from Figs.1 and 2. The numerical codes (in black) for each red-outlined spot in the figure (differentially expressed polypeptides) correspond to the respective numbers in the first column of Table 3.



Fig. S4. qRT-PCR estimation of 5S and 16S rRNAs in *S. meliloti* cells grown in the chemostat under steady state at pH 7.0, and at pH 6.1. The quantification of specific RNA species was carried out by quantitative reverse-transcription PCR using a KAPA SYBR FAST One-Step qRT-PCR kit (Kapa Biosystems Inc. Wilmington, MA, USA) according to the manufacturer's instructions. Analyses were performed using a same normalized amount of total RNA for all samples. M values represent the log₂ of the fold change of the indicated RNA species at pH 6.1 (red bars) and 7.4 (blue bars) both with respect to their corresponding amounts at pH 7.0 (reference condition). 5S and 16S correspond to the analysis of 5S and 16S rRNA. *degP1* (SMc02365) corresponds to the analysis of the mRNA of an acid-induced gene used as positive control (M $^{\text{pH 6.1/pH 7.0}}$ from the microarrays = 1.98). The DegP1 polypeptide was also induced in the proteome of acid-grown rhizobia (fold change *ca.* 4.8 in Table 2, so with an estimated M $^{\text{pH 6.1/pH 7.0}}$ from the proteome = 2.26). The M values are the means calculated from three technical qRT-PCR replicas. The bars represent the standard deviations.



Fig. S5. Genomic distribution of proteome and transcriptome markers differentially overexpressed at pH 6.1 *versus* **7.0.** The two outermost circular array of lines on each replicon show—in black and in gray—the respective positions of open reading frames within the two complementary DNA strands. The next array inside shows—with red and blue lines—the genomic position of genes encoding polypeptides that were observed to be induced under acidic and neutral extracellular conditions, respectively (from the proteomic data). The innermost array marks—with green and orange lines—the genomic position of genes encoding RNAs that were also observed to be induced under acidic and neutral extracellular conditions, respectively (from the transcriptome data). The pSymA region characterized by a high density of overexpressed genes at pH 6.1 (*cf.* the text) is outlined and marked by an asterisk.



Fig. S6. Vector-correlation plot between the variables examined (*i. e.*, **the amount of metabolites**) **and the PC1 and PC2 spaces of PCA.** The direction and length of the vectors are indicative of the positive contribution of each individual metabolite to the position of each sample analyzed in the plot (*i. e.*, from rhizobia grown at pH 6.1, pH 7.0, or pH 7.4).



Fig. S7. Graphical maps indicating significant increases of specific RNAs, polypeptides, or metabolites on different *S. meliloti* metabolic pathways (KEGG format); and also changes in RNAs/polypeptides related to cellular macromolecular complexes (transport, energy, translation). <u>Colored rectangles</u>: correspond to increased RNAs or polypeptides (i.e. RNAs listed in Supplementary Table 1, and polypeptides listed in the main Tables 1 and 2). <u>Colored circles</u>: correspond to increased metabolites (i.e. those listed in Supplementary Table 2, and also presented in the volcano plot from Fig. 3). Red and blue colors indicate increased amount of the corresponding species at pH 6.1 and pH 7.0, respectively. References to panels are as follows: A. Pentose phosphate pathway; B. Galactose metabolism; C. Starch and sucrose metabolism; D. Carbon metabolism; E. Arginine and proline metabolism; F. Cysteine and methionine metabolism; G. Glutathione metabolism; H. Aminoacids biosynthesis; I. Purine metabolism; J. Pyrimidine metabolism; K. Biotin metabolism; L. Ribosome components; M. RNA degradation; N. O xidative phosphorylation; O. Flagellar assembly; P. Secretion systems; Q. Two-component systems; and R. ABC transporters.































Ν





Q



R

		**			
м	ineral a	nd organi	: íon	transp	•

ABC TRANSPORTERS]										
Prokaryotic-type ABC transpor	ters										
Mineral and organic ion trans	ort	ers	CvsU	L	Ph	osphate and amino acid trans	pori	ers	PetC	L	
Sulfate	0-	CysP	CysW	CysA		Phosphate	o-	PatS	PstA	PstB	
Tungstate	0-	TupA	TupB	TupC		Phosphonate	0-	PhnD	PhoE HisM	PhnC	⊦ ►
Molybdate / Tungstate	0-	- WtpA	WtpB	WtpC NetC		Lysine / Arginine / Ornithine	0-	ArgT	HisQ	HisP	•
Nitrate / Nitrite / Cyanate	0-	NnA	NnB	NrtD	-	Histidine	0-	HisJ	HisM HisQ	HisP]→
Bicarbonate	0-	CmpA	CmpB	CmpC CmpD	*	Glutamine	0-	GlnH	GhaP	Ghq	}
Taurine	0-	TauA	TauC	TauB	⊢►	Glutamine?	0-	GhH	GlnP	GlmQ	+
Alkanesulfonate	0-	SsuA	SsuC	SsuB	⊢►	Arginine	0-	Art	Antivi ArtQ	ArtP]+
HMP / FAMP	0-	ThiY	ThiX	ThiZ	+	Glutamate / Aspartate	0-	GH	GHK	GHL	}-►
Phthalate	0-	OphF	OphG	OphH	+	Octorine / Noraline	0-	- OccT	OccM	OceP	ኈ
Molybdate	0-	ModA	ModB	ModC ModF	-				OccQ		
Iron (III)	0-	Afta	AfuB	AfuC	+	General L-Amino acid	0-	Aapi	AspM	AapP	++
Thiamin	0-	TopA	ThiP	ThiQ	-►	Glutamate	0-	GhB	GluC GluD	GluA]-►
Spermidine / Putrescine	0-	PotD	PotC	PotA	-	Cystine	0-	FhY	YecS	YecC	⊦∙
			Pots			Cystine	0-	Teyd TeyK	TcyL TcvM	TeyN	}≁
Putrescine	0-	PotF	PorH	PorG	•	Arrinine / Ornifhine	0-	Aoti	AotM	AntP	≁
Mannopine	0-	AttC	AttB AttA2	AttA1	┝►	Arginine / Lysine /	o-		RatB	Bath	1->
2. A minorthylphograponate	0-	Phys	PhnV	PhpT		Hisfidine / Ofutamine Lysine	0-	LysX	LysX	LysY	}. }≁
2-rinano ary prospiosian	Č	1100	PhnU			Branched-chain amino acid	0-	LivK	LivH	LivG	
Glycine betaine / Proline	0-	PioX	ProW	ProV					LivM	LivF	
Osmoprotectant		ComBC	OpuBB	OpuBA	-	Neutral amino acid / Histidine	0-	NatB	NatD	NatE	•
Maltose / Maltodextrin	0-	MalE	MalF	MalK		Urea	0-	UntA	UnB UnC	UnD UnE	+
Gebeteen elizemen (_	GanP			D-Methionine	0-	MetQ	Meti	MetN	⊦≁
Maltooligosaccharide	0-	- GanO	GanQ	MsmX	-	Amino acid	0-	YxeM	YxeN	YxeO]->
Raffinose/Stachyose/ Melibiose	0-	MsmE	MsmF MsmG	MsmK	► Pe	ptide and nickel transporters					_
Lactose /L-arabinose	0-	LacE	LacF	LacK	+	Oligopeptide	0-	OppA	OppB OppC	OppD OppF	►
Sochital (Manuita)	0-	Small	SmoF	- SmrV		Dipeptide / Heme /	0-	DppA	DppB DpcC	DppD DppF	┢
3000007 Walking	Č	3805	SmoG	1 3110K	-	Directicle	0	Dunk	DppB	DeeD	
a-Glucoside	0-	AglE	AglG	AgIK	*	Dippine	Č	Dppe	DppC	NWD	י ב ר
Oligogalacturonide	0-	TogB	TogM TogN	TogA	+	Nickel	0-	NikA	NikC	NikE	<u></u>
a-1.4-Digalacturonate	0-	AeuE	AguF		⊢►	Glutathione	0-	GsiB	GsiC GsiD	GsiA]-▶
, ,			AguG LnB			Microcin C	0-	YeiA	YejB	YeiF	↦
Aldouronate	0-	LplA	LpiC	12	⊢► Me	etallic cation, iron-siderophore	and	vitamin I	YejE B12 trans	porters	
Trehalose / Maltose	0-	ThuE	ThuF ThuG	ThuK	+	Iron complex	0-	FhuD	FhuB	FhuC	⊦►
Trehalose	0-	TreS	TreT	TreV	+	Vitamin B12?	0-	BtuF	BtuC	BtuD	}→
N & catula lucocamina	~	MaaF	NgcF		_	Nianganese Zinc	0-	ZnuA	ZauB	ZnuC	
11-receiping to communic	0	INGEL	NgcG	⊢∸⊥ ≀		Iron (II, III) / Manganese / Zinc	- 0-	MtsA	MtsB	MtsC	}. }►
Cellobiose	0-	CebE	CebG	MsiK	+	Iron (II) / Manganese	0-	SitA	SitC	SitB	- ⊦ ≁
Chitobiose	0-	DasA	DasB DasC	MsiK	+			_	SitD TroC		-
Chitobiase	0-	ChiE	ChiF			Zinc / Manganese / Iron (II)	0-	- TroA	TroD	TroB	
			AraP			Cobalt	0-		CbiN CbiM	(75i0)	ኈ
Arabinooligosaccharide	0-	AraN	AraQ	MsmX	+				ChiQ	1	_
Xylobiose	0-	BxlE	BxIF BxIG	2	+	Nickel	0-	CbiK	Child	Сыо]-+
Multiple sugar?	-	ChvE	GguB	GguA	+	Biotin	0-	BioY	BioN	BioM	↦
Phospholipid	0-	MlaC MlaD	MlaE	MlaF	+	Biotin	0-	BioY	EcfT	EcfA1	•
			MlaB]						ECTA2	
Monosaccharide transporters	0-	- 0165	GlcU	GRV							
010000111000000		0.00	GlcT	 1							
Glucose / Mannose	0-	GtaA	GtsC	MalK	•						
Ribose / Autoinducer 2 / D-Xylose	0-	RhsB	RbsC RbsD	RbsA Auxilia	+ rv component						
L-Arabinose	0-	AraF	AraH	AraG	+► .						
Methyl-galactoside	0-	MglB	MgIC	MglA	+						
D-Xylose D-Allose	0-	AkB	AbC	AlsA	→ →						
Fractose	0-	FrcB	FreC	FrcA	+						
Autoinducer 2	0-	LsrB	LsrC LsrD	LsrA	⊦►						
Rhamnose	0-	RhaS	RhaP	RhaT	⊢►						
Erythritol	0-	EryG	EryF	EryE							
Xylitol	0-	XIIC	XHB	XltA	+						
myo-Inositol	0-	IbpA	IatP	IatA	+						
myo-Inositol 1-phosphate	0-	InoE	inoF InoG	InoK	⊢►						
Glycerol	0-	GlpV	GlpP GlpQ	GipS GipT	•						

sn-Glycerol 3-phosphate O-UgpB UgpA UgpC →

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		Eukaryotis-type ABC transporters
ABC-2 and other transporte	в	ABCA Subfamily
Hemolysin	CyiB CyiA 🔸	ABCA1 ABCA5
Carrenlar rolamarcharida	KpsE KpsT	ABCA2 ABCA6
oupoun prijonerman	KpsM	ABCA3 ABCAS
Lipopolysarchande	ORfbA RfbB →	ABCA4 ABCA9
Teichoic acid	OTagG TagH →	ABCA7 ABCA10
Lipo-oligosaccharide	O	ABCA12
Na [*]	O NatB NatA +	ABCA13
Hemine	O-HrtB HrtA	ABCB Sub family
Oleandomycin	0	ABCB2 ABCB1 ABCB0 ABCB11 MB0A
Bacitracin	O BoeB BoeA	ABCB3 ABCB4 ABCB/ HIVB
	LaiC	ABCBO ABCBJ AINI NEE
Lipoprotein		ABCBI0 AbcA
Heme	CemD CemC CemA	StrAB
	CemB	ABCC Sub family
Cell division	FuX FuE	ABCC1 ABCC8 Mulb ABCC4
Lipopolysarcharide	O LptF LptB	ABCC2 ABCC9 HasD CFTR
	Lpto	ABCC3 ABCC11 PmD ABCC10
Fluoroquinolones	O Rv2686c Rv2687c Rv2688c ►	ABCC5 ABCC12 ReaD
YvdF peptide	Yvdl Yvdi +	ABCC6 EexD
	YtrB	ABCC13 LapB
Acetoin utilization	YttF YttC/D YttE	CydC/D
		ABCD Sub family
		ABCD1 PXA1/2
		ABCD2
		ABCD3
		ABCD4
		ABCG Sub family
		ABCG1 ABCG2 ABCG5 PDR5
		ABCG4 ABCG3 ABCG8 SNQ2
		MacB TylC MarA
		Other putative ABC transporters
		YojI
		PvdE
		SynD
		YddA

		1000	
Lipo-oligosaccharide	o	NodJ	NodI
Na [*]	o	NatB	NatA 🕨
Hemine	o	HrtB	HrtA 🔸
Oleandomycin	o	OleC5	OleC4
Bacitracin	o	BceB	BceA -
Lipoprotein	o	LolC LolE	LoID
Heme	O-CemD	CemC CemB	CemA +
Cell division		FtsX	FtsE -
Lipopolysaccharide	o	LptF LptG	LptB 🔸
Fluoroquinolones	o	Rv2686c Rv2687c	Rv2688c -
YydF peptide		YydJ	Yydi 🔸
Acetoin utilization	- YuF	YeC/D	YtrB YtrE

Glutamine	0-	GhH	GhrP	GlnQ	►
Glutamine?	0-	GlnH	GlnP	GlnQ	┝►
Arginine	0-	ArtJ	ArtM	ArtP	+
	_		GIRK		1.
Glutamate / Aspartate	0-	GH	GH	GHL	⊢►
Octopine / Nopaline	0-	OccT	OccM Occ0	OccP	⊢►
County to it	~	Ant	AapQ	A con P	
General L-Amino acid	0-	Aapu	AspM	Mapr	-
Glutamate	0-	GhuB	GluC GluD	GluA	⊦►
Cystine	0-	FhY	YecS	YecC	
Custine	0-	Teyd	TcyL	TeuN	⊨►
- ,		TcyK	TcyM	1	1
Arginine / Ornifhine	0-	Aoti	AotA	AotP	⊦►
Arginine / Lysine / Histidine / Glutamine	0-		BgtB	BgtA	⊢►
Lysine	0-	LysX	LysX	LysY	⊦►
anched-chain amino acid	0-	LivK	LivH LivM	LivG	┢
			LIVINI NetC	Nata	1
al amino acid / Histidine	0	NatB	NatD	NatE	-
Urea	0-	UrtA	UntB	UnD	+
D.Methionine	<u>0</u> -	Marc	Meti	Diffet N	∣ ⊢∎-
Amino ariid	~ ~	VyaM	VyaN	Vieut	,. ∟ ⊳
nd nickel transporters		1 40144	1 401	1400	, -
na n			OpeB	OppD	1.
Oligopeptide	0-	ОррА	OppC	OppF	-
Dipeptide / Heme / δ-Aminolevulinic acid	0-	DppA	DppB DppC	DppD DppF	-
Direntide	0-	DownE	DppB	Drad	L-
2 qo pan		- oppe	DppC	DppD	1
Nickel	0-	NikA	NikC	NikE	┢╸
Glutathione	0-	GsiB	GaiC	GsiA	⊦►
		_	VeiB		,
Microein C	0-	YejA	YejE	YejF	┝►
ation, iron-siderophore	and	vitamin I	312 trans	porters	1.
Iron complex	0-	FhuD	FhuB	FhuC	⊢ ►
Vitamin B12?	0-	1 BtuF	BtuC	BtuL)	⊢≁ ∟⊷
ivanganese 7ma	~	Znuč	ZauR	ZnuC	1 * L e
Zinc III) (Manzanese / Zinc	0-	Mite	MiteP	MbC	 -
,, . manganese / 2200	č	TANK .	SitC	11100) * 1 •
iron (II) / Manganese	0-	SitA	SitD	SitB	⊢►
ic / Manganese / Iron (II)	0-	TroA	TroC TroD	TroB	⊦►
			ChiN		
Cobalt	0-		ChiM	CbiO	⊢►
			ChiNP		
Nickel	0-	Chik	ChiM	Сыо	⊢►
pinin	0-	PinV	ChiQ	DiaM	L .
niohn		BIOY	DION	Esorial EcfA1	1.
Biotin	0-	BioY	EcfT	EcfA2	►