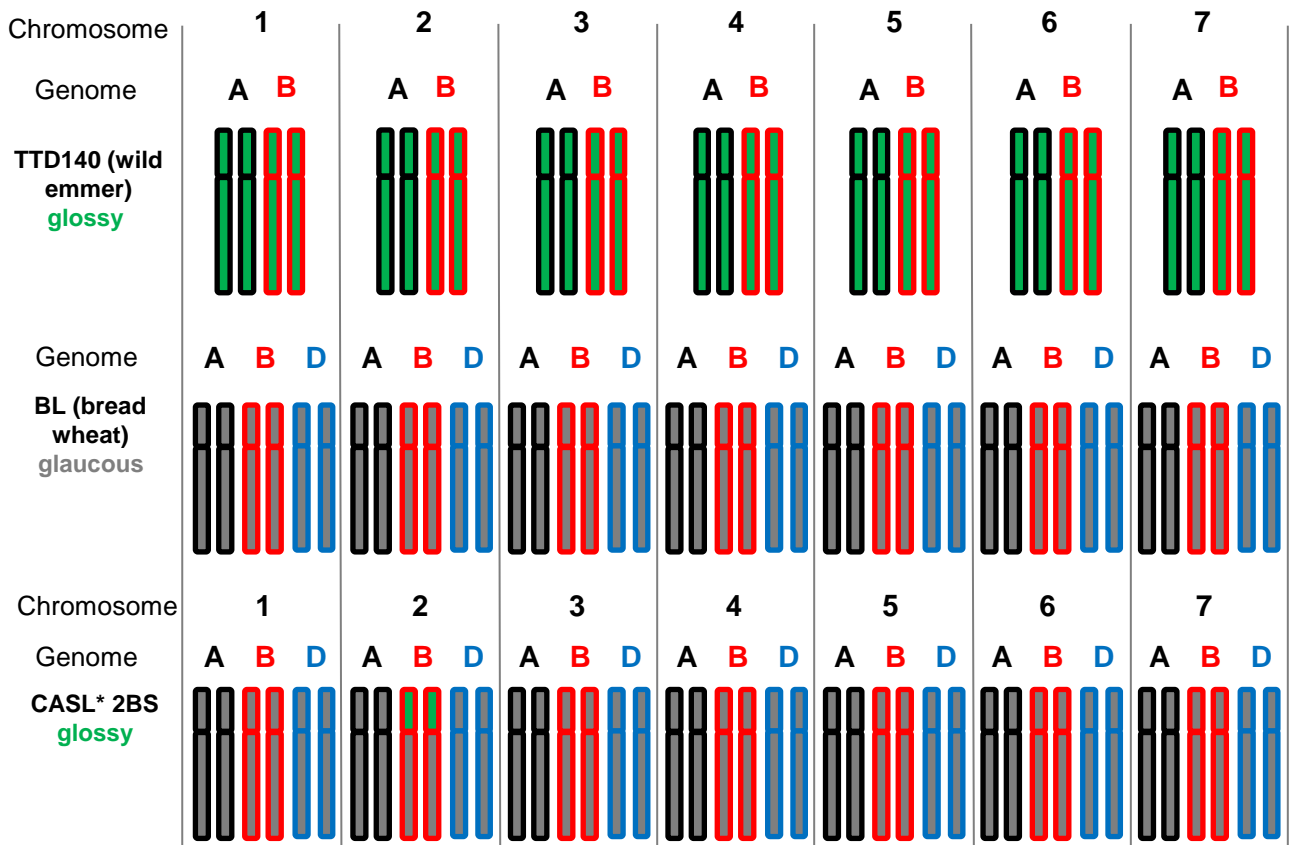
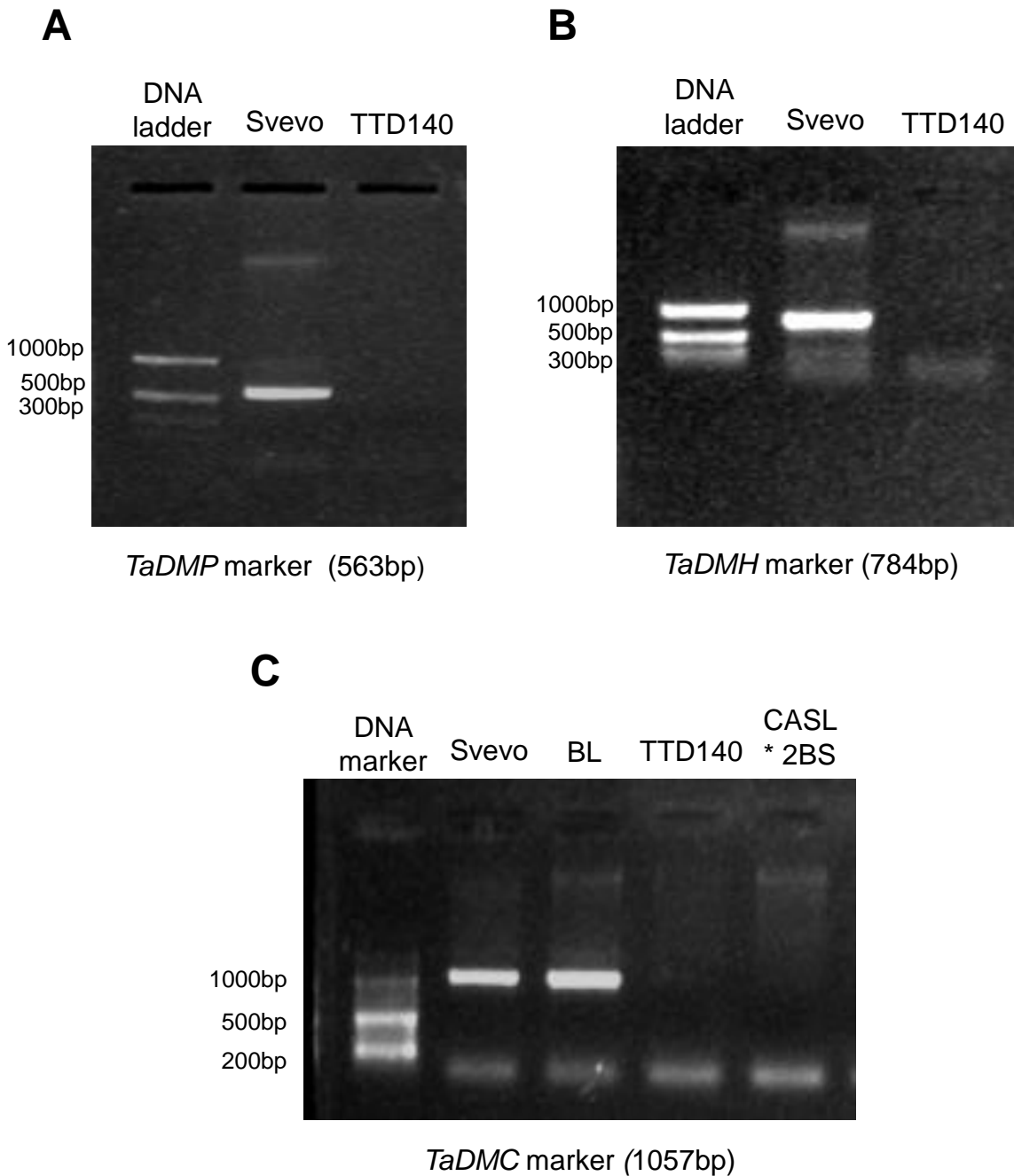


Supplemental Fig. 1

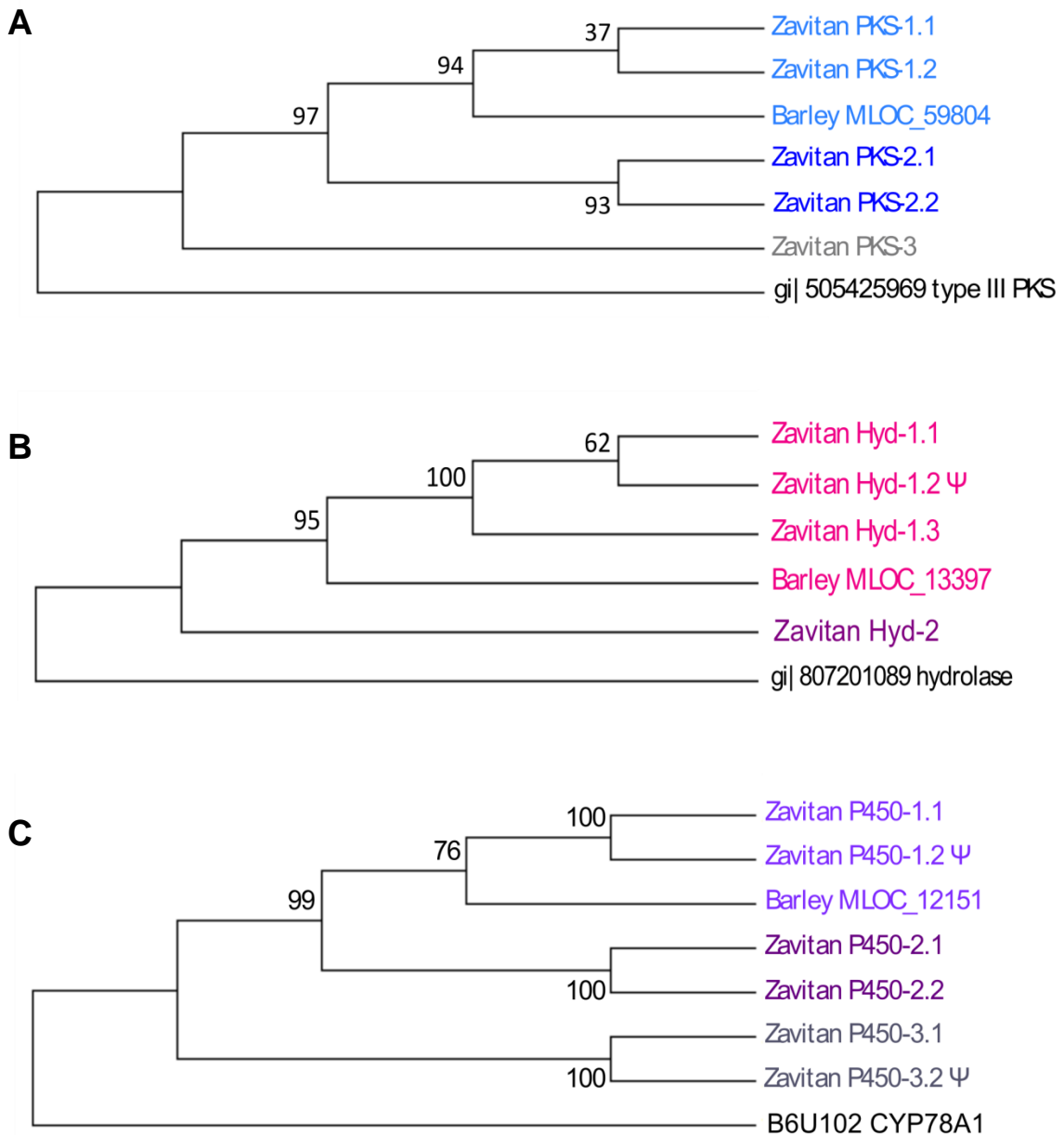
Supplemental Figure 1. General scheme of biosynthetic pathways generating different cuticular wax components. The alkane-forming (in purple) and alcohol-forming (in blue) pathways of cuticular wax biosynthesis. Many Gramineae species have additional wax compounds (in red). β -diketones and hydroxy- β -diketones, synthesized through an independent pathway, are shown in a red rectangle. Methylketones, 2-alkanols and 2-alkanol esters are side products of the β -diketone biosynthesis pathway. The β -diketones may be derived from FAS products or FAE products. Alkylresorsinols are polyketides also found in many Gramineae species. FAS- fatty acid synthase system, FAE- fatty acid elongase system. n, m indicate varying chain lengths.



Supplemental Figure 2. Illustration of wheat parental and chromosome arm substitution lines used in this study. Wheat substitution lines used in this study were produced by Millet et al. (2013). A series of wild emmer [*Triticum turgidum* ssp. *dicoccoides* (line TTD140)] chromosome arm substitution lines (CASLs) were produced in the background of the modern common wheat [*Triticum aestivum* ssp. *aestivum* cv. Bethlehem (BL)]. In CASL* 2BS, the whole 2BS arm from TTD140 was inserted to the BL genome background. The wax phenotype is indicated for each line. Grey and green chromosome colors indicates BL and TTD140-derived chromosomes, correspondingly.

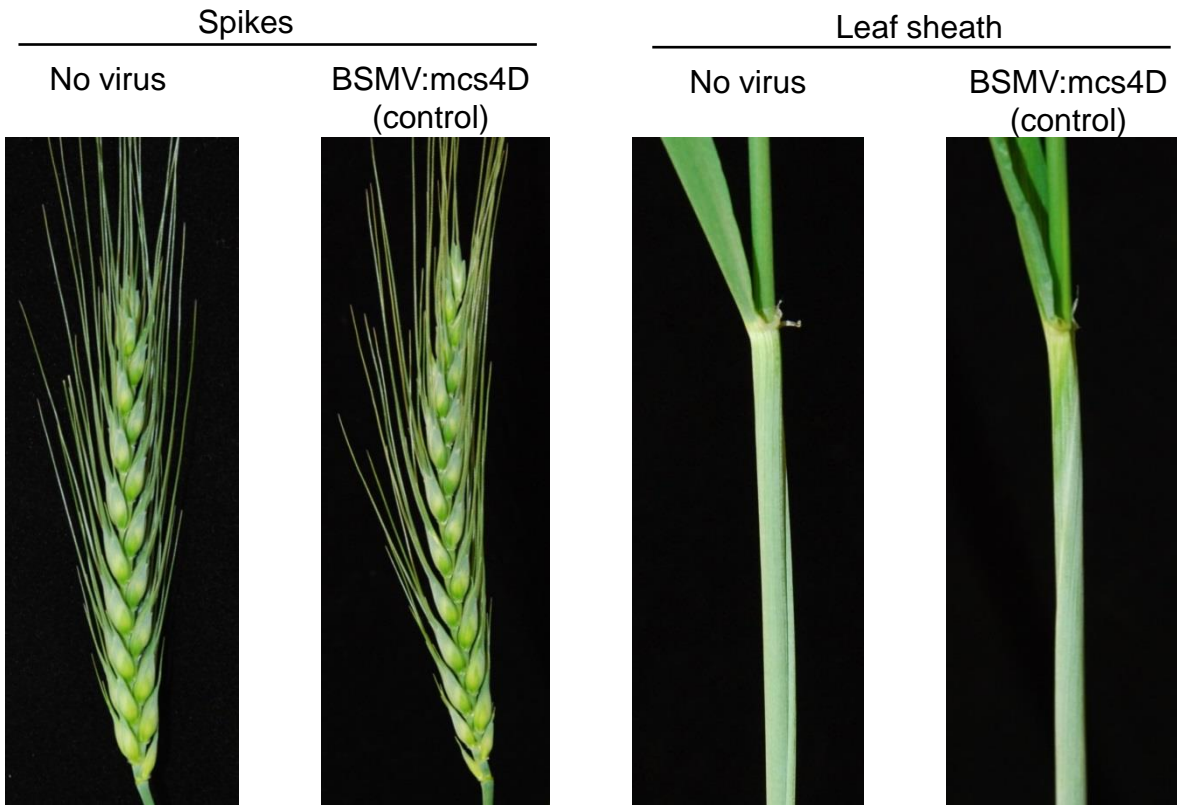


Supplemental Figure 3. Amplification of markers developed for the three candidate genes in glaucous and glossy wheat. The β -diketone biosynthesis candidate genes are present in the glaucous parental line 'Svevo' however absent in glossy parental line 'TTD140'. Specific markers were developed for *TaDMP* (**A**), *TaDMH* (**B**) and *TaDMC* (**C**). The marker developed for the *TaDMC* gene was also polymorphic in the hexaploid population while the other two were not, apparently due to homologues in the D genome.

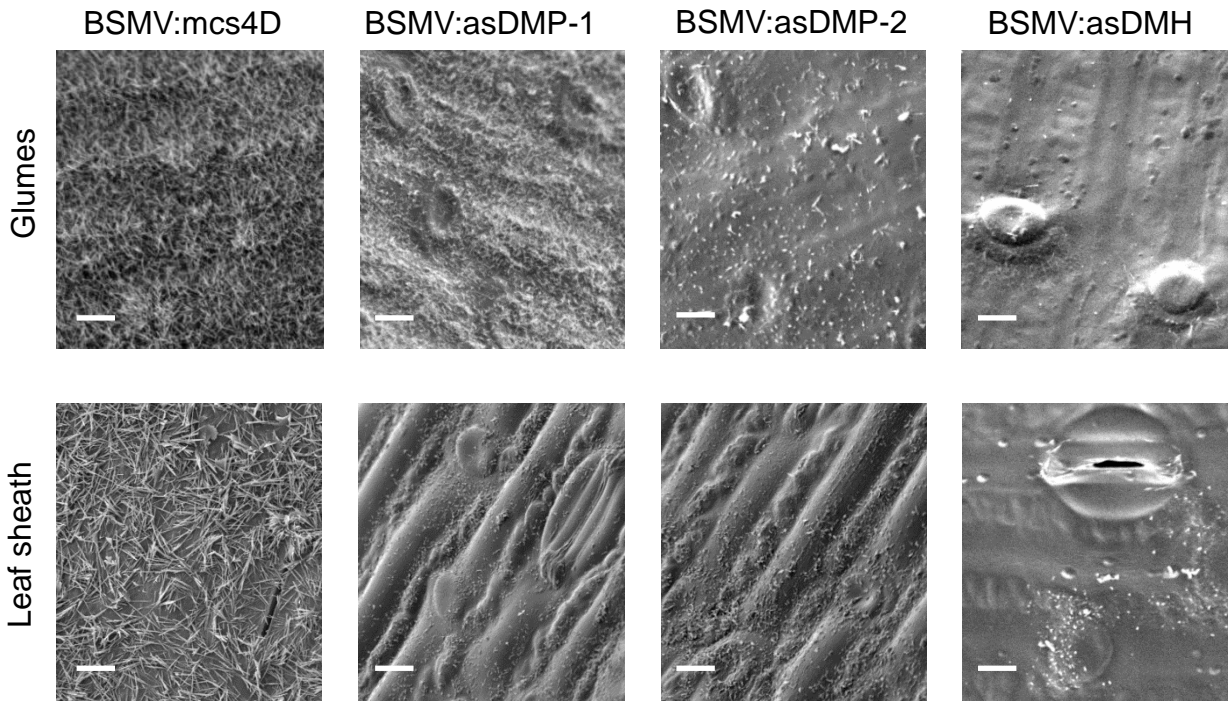


Supplemental Figure 4. Phylogenetic trees of Zavitan proteins. Phylogenetic trees describe the similarity between Zavitan and barley β -diketone proteins. **(A)** PKS, **(B)** Hydrolase (HYD) and **(C)** P450 phylogenetic trees. Sequences for the Zavitan proteins can be found in Supplemental File 1. Outgroup protein accessions are: For PKS tree: gj|505425969 (type III PKS from *Streptomyces fulvissimus*), for HYD: gj|807201089 (adenosylhomocysteinase from *Solanum lycopersicum*), for P450: B6U102 (CYP78A1 from *Zea mays*). Ψ - Premature termination codon (PTC). Closely related proteins are denoted with the same color.

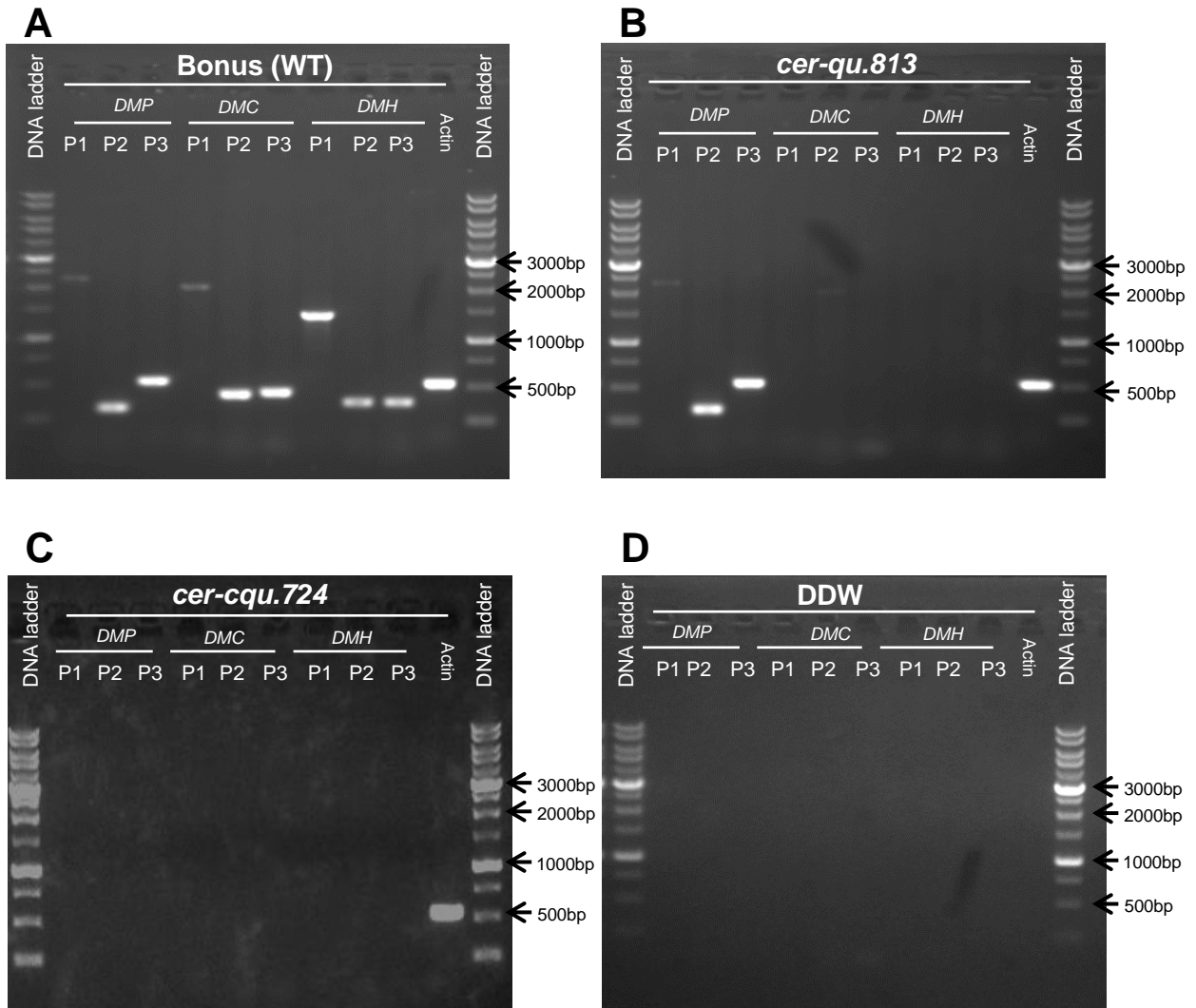
A



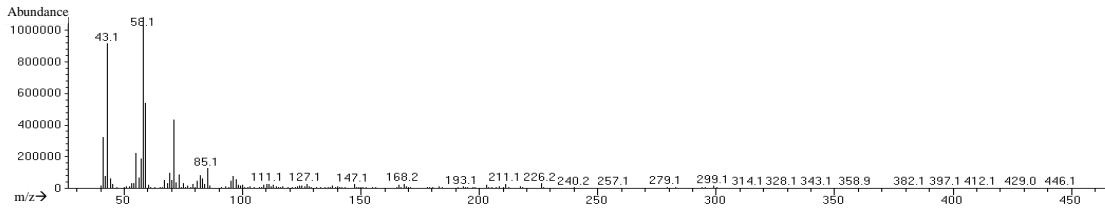
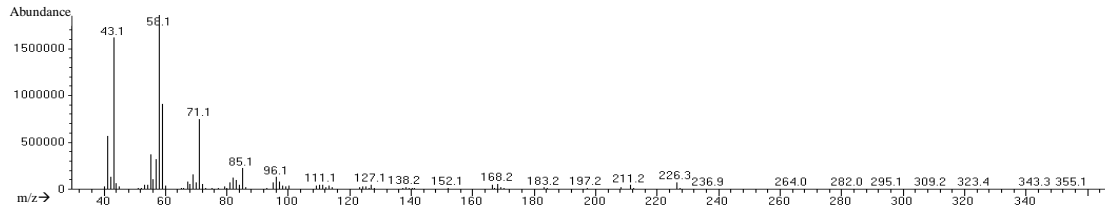
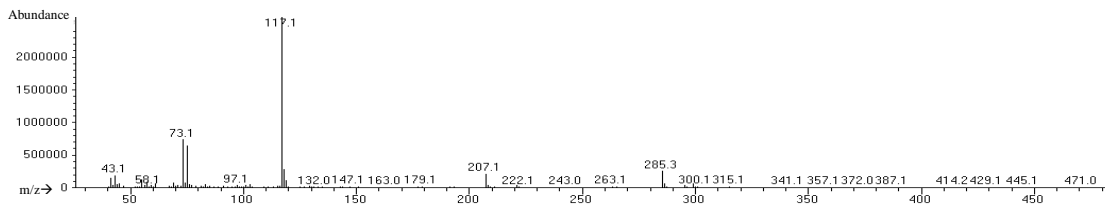
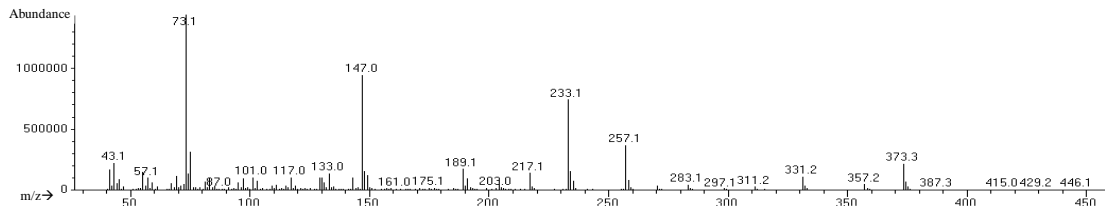
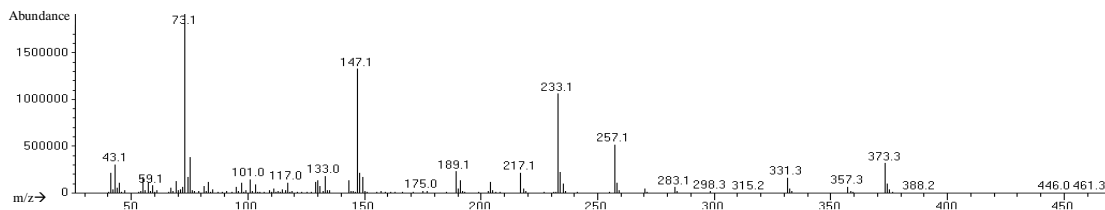
B



Supplemental Figure 5. Virus-induced gene silencing (VIGS) of the β -diketone genes reduces wax accumulation and deposition of epicuticular wax crystals. (A) Control infection with BSMV:mcs4D does not reduce glaucousness. Non-infected plants (no virus) and BSMV:mcs4D-infected (control) plants both have glaucous phenotype in glumes and leaf sheath (B) Epicuticular wax on surfaces of flag leaf sheath and glumes tissues was examined using scanning electron microscopy (SEM). Plants infected with constructs targeting the β -diketone biosynthesis gene transcripts (BSMV:*asDMP-1*, BSMV:*asDMP-2* and BSMV:*asDMH*) displayed reduced wax amount as well as reduced deposition of tubular wax crystals, typical for β -diketone-rich wax. Magnification: X500. Scale bar indicates 2 μ m.



Supplemental Figure 6. The barley *cer-qu.813* double and *cer.cqu-724* triple mutants have deletions that include the metabolic gene cluster genes. Amplifications of the three candidate genes, *DMP*, *DMH* and *DMC*, were performed using three different primer pairs (P1- P3) per gene and genomic DNA template. Actin oligonucleotides were used as positive control, DDW as negative control. Agarose gels are shown for reactions using the following templates: **(A)** bonus (WT), **(B)** *cer-qu.813*, **(C)** *cer-cqu.724*, **(D)** DDW control. An unspecific band appears in the reaction mixture of *DMC* P2 primers and *cer-qu.813* gDNA, however has different product size from the corresponding reaction with WT template.

A Peak at 7.5 min: pentadecan-2-one (C₁₅ 2-ketone)**B Peak at 7.5 min: pentadecan-2-one (C₁₅ 2-ketone) standard****C Peak at 8.4 min: pentadecan-2-ol (C₁₅ 2-alkanol)****D Peak at 10.8 min: 3-OH tetradecanoic acid (C₁₄ 3-hydroxy-acid)****E Peak at 10.8 min: 3-OH tetradecanoic acid (C₁₄ 3-hydroxy-acid) standard**

Supplemental Figure 7. Compound identification of *E-coli* expressing DMH metabolites. Gas chromatography mass spectrometry (GC-MS) spectra of increased compounds in lipids extracted from *E-coli* expressing DMH from barley (**A**, **C-D**) and commercial standards (**B**, **E**). (**A**) C₁₅ 2-ketone, RT 7.5 min (**B**) C₁₅ 2-ketone standard (Sigma, cat. W372404), RT 7.5 min. (**C**) C₁₅ 2-alkanol, RT 8.4 min. (**D**) C₁₄ 3-hydroxy-acid, RT 10.8 min. (**E**) C₁₄ 3-hydroxy-acid standard (Sigma, cat. H4148), RT 10.8 min.

Supplemental Text 1: Fine genetic map of 2BS *W1/lw1* loci in different segregating populations

Kofa+Lr19 (glaucous)x AUS2499 (glossy):

Out of 352 F₂ progeny screened we observed 271 individuals with a glaucous phenotype and 81 individuals with a glossy phenotype. This correlates well with the expected 3:1 segregation ratio for a single locus ($\chi^2=0.74$, $df=1$, $P=0.3889$). Using genetic markers *W1_1*, *W1_2*, *W1_3* and *W1_4* (corresponding to markers *JIC005*, *JIC007*, *JIC009* and *JIC010*, respectively) we have mapped this single locus to the short arm of chromosome 2B between markers *W1_3* and *W1_4*; two recombinants were identified between markers *W1_3* and *W1_4*, giving a genetic distance of 0.57 cM. We then proceeded to screen another 4,423 F₂ progeny using markers *W1_3* and *W1_4*, giving a total of 4,775 F₂ plants screened (9,550 gametes) out of which 15 plants were shown to be recombinant between markers *W1_3* and *W1_4*. This represents a genetic distance of 0.16 cM. By selfing the 15 recombinants we obtained homozygous recombinants and verified both the mapping data as well as the phenotype. These results suggest that the dominant locus for β -diketone production designated *W1* is confined in a 0.16 cM interval flanked by markers *W1_3* and *W1_4*.

Svevo (glaucous) x TTD140 (glossy)

In this population *lw1* derived from TTD140 segregates. Out of 299 F₂ progeny screened we observed 77 individuals with a glaucous phenotype and 222 individuals with a glossy phenotype, fitting a single locus 1:3 Mendelian segregation ratio ($\chi^2=0.09$, $df=1$, $P=0.7674$). Using genetic markers *Xwmc661*, *W1_3* and *W1_4* we have mapped this single locus to the short arm of chromosome 2B completely linked to *W1_3*; seven recombinants were identified between markers *Xgwm614* and *W1_4*, giving a genetic distance of 1.17 cM. By selfing the 7 recombinants we obtained homozygous recombinants and verified both the mapping data as well as the phenotype.

Supplemental Text 2: Analysis of the Zavitan *W1/lw1* interval

Using the wild emmer Zavitan genome sequence (unpublished) we analysed the genes within the interval between genetic markers *JIC036* and *JIC032* which include the *W1* locus and the three β -diketones candidate genes (see Figure 2C). We predicted 22 genes in this region, 15 out of the 22 were annotated similarly to the β -diketones candidate genes; six *CYP450* family (*P450*) genes, five *Type III polyketide synthase* (*PKS*) family members and four members of the *hydrolase/lipase* (*Hydrolase*; *HYD*) family.

Among the five *PKS* genes there are three distinct genes, two of which have been duplicated in tandem. One of the tandem duplicated genes has not been fully captured in the Zavitan genome sequence and only the first exon is present, but the gene is likely intact. The similarity between the tandemly duplicated genes is ~99.5%, indicating a relatively recent duplication event. Comparing the coding sequence (CDS) of these five genes against that of barley *MLOC_59804* shows that one of the duplicated gene pairs (*PKS-1.1* and *PKS-1.2*) is more closely related than the other three genes, indicating that these are the likely orthologues. However, *MLOC_59804* is also the best hit for *PKS-2.1* and *PKS-2.2*, suggesting that either their barley orthologue is missing from the database or that these are older duplicated versions of *PKS-1.1* and *PKS-1.2* (Supplemental Table 3 and Supplemental Figure 4A).

Among the four *HYD* genes there are two distinct genes, one of which has been duplicated twice (*HYD-1*). The similarity between these three genes is >99.6% indicating relatively recent duplication events. Comparing the CDS of these four genes against that of the barley *MLOC_13397* showed that the triplicated gene pair (*HYD-1*) is more closely related than the other gene (*HYD-2*), indicating that the triplicated genes are the likely orthologues (Supplemental Table 3 and Supplemental Figure 4B). Interestingly, *HYD-1.2* has a single base pair deletion within its CDS, causing a frameshift and premature termination codon (PTC), suggesting that *HYD-1.1* and *HYD-1.3* are the functional orthologues of *MLOC_13397*.

Among the six *P450* genes, three distinct genes are represented, all of which have been duplicated in tandem. The similarity between the tandemly duplicated genes

is >99.3% indicating relatively recent duplication events. Comparison of the CDS of these six genes against that of *MLOC_12151* showed that one of the duplicated gene pairs (*P450-1.1* and *P450-1.2*) is more closely related than the other four genes, indicating that genes in this pair are the likely the orthologues (Supplemental Table 3 and Supplemental Figure 4C). Similar to the *HYD* example, *P450-1.2* has a single base pair deletion within its CDS, causing a frameshift leading to a PTC. This suggests that *P450-1.1* is the only functional orthologue of *MLOC_12151*. The orthologous loci of the other *P450* genes appear to be *MLOC_13649* and *MLOC_71974*. Notably, *P450-3.2* also has a PTC.

The pattern of gene duplications within the *Zavitan* interval has occurred after the divergence from barley based on the high sequence similarity of the duplicated genes (>99.3 % in all cases). Moreover, the position of the genes within the interval provides clues as to how those duplications occurred. The arrangement of the genes within the interval precludes a large scale duplication event and is most consistent with a series of independent small scale local duplication events.

One duplication consists of *PKS-1.1* and *PKS-1.2*, the two genes most closely related to *MLOC_59804*. Another block includes the duplication of *P450-1.1*, *HYD-1.1* and *Hly-1.1* into a second block (*P450-1.2*, *HYD-1.3* and *Hly-1.2*) which was followed by the separate duplication of *HYD-1.1* into *HYD-1.2*. A third duplication block within this interval encompasses *PKS-2*, *P450-2* and *P450-3* (see Figure 2C).

In addition to β -diketones candidate genes, two more relevant genes were found to be located in the *W1/lw1* *Zavitan* interval: wax ester synthase (*WES*) and *CYP96B30* (*P450-3.1*). The genes annotation as wax related and their genomic localization suggests that they might also be involved in β -diketones or related metabolites biosynthesis.

Supplemental Text 3: Comparison of the *W1//lw1* interval between the wild emmer accessions TTD140 and Zavitan

Genetic markers *JIC007/IWB7407* (distal) and *JIC010/CD927782* (proximal) (see Figure 2C) which are flanking both the *W1* and *lw1* interval were used to anchor genomic sequence of Zavitan (glaucous) and TTD140 (glossy). The distal TTD140 and Zavitan sequences are 97.6% similar over ~8 kb of intergenic, exon and intron sequence. Proximal to the *W1//lw1* interval the similarity is only 93% over ~18 kb of intergenic, exon and intron sequence, but this rises to 96% if corrected for gaps in the alignment. (Figure 2C, Supplemental Table 3). However, the interval between these highly similar regions is completely different between TTD140 and Zavitan, both for the intergenic space and also for the gene content, with the exception of two NB-Arc-like genes (NB-Arc-like_1 and _2; Figure 2C). These two genes are located at the distal border of the conserved interval in Zavitan and TTD140. However, in the latter the NB-Arc like genes have undergone a series of tandem duplication events; the available sequence of TTD140 suggests at least 13 copies, but there are likely more. The duplications in TTD140 are not restricted to the open reading frames themselves, but also include intergenic sequence and transposable elements.

None of the other genes identified in Zavitan can be found in the TTD140 physical map covering the *W1//lw1* interval. This could be attributed to the incomplete state of the TTD140 physical map. But the reverse is also true, i.e. none of the genes identified in TTD140 can be found in the Zavitan whole genome assembly. Interestingly, the TTD140 physical map contains two P450 and two PKS genes, but these have low similarity (~85 % in the nucleotide level) to the *P450* and *PKS* genes identified in the Zavitan interval (see Supplemental Table 3). In summary, at least 808,361 base pairs of TTD140 sequence (excluding the region of tandem duplicated NB-Arc-like genes) are completely different in the Zavitan whole genome assembly, suggesting a divergence in haplotype between these two wild emmer accessions across the *lw1//W1* interval.

The haplotype divergence is supported by the observation that no recombinant plants between markers *JIC036* and *JIC032*, the two markers closest to the interval, were observed in F₂ progeny of a cross between three different mapping populations in

which TT140 is a parent or the 2BS is derived from TTD140 (CASL*2BS). The studied populations were TTD140 and *T. durum* cultivar Langdon (glaucous; *iw1/iw1*; *W1/W1*), TTD140 and *T. durum* cultivar Svevo and CASL*2BS and *T. aestivum* cultivar BL. Based on the disparity in sequence content between TTD140 and Zavitan we hypothesise that Langdon and TTD140 also have equivalent sequence disparity, i.e. different haplotypes between markers *JIC036* and *JIC032*. Recombination numbers from Langdon X TTD140 population are given in purple in Figure 2C.

Supplemental Table 1: Linked deleted markers in the TTD140 accession detected by the 90K chip analysis

Number	SNP name	Reside inside β -diketone gene	SNP ID	Marker sequence	Index on iSelect 90K SNP bead chip
1	wsnp_Ra_rep_c106727_90434958	DMP	IWA8128	AGGTCCATGGTGTCTGCTACCAGTGTGGGCAGTTGGTGTAAACTTTCCCATGAGTGC CATTGCTCCCCAAGTTTCAGGGTGCAGACATTCTCAGTGCTC[A/G]GCACGAGGGTCTGA GAGACGGACCCATCTCAAACAGAGGATTCTCGATGGGGTGCATGGGGTTGGCGCCG ACGATTACCGCGCTGCCCATCACAAA	81290
2	RAC875_rep_c109471_154	DMP	IWB61884	CCCACCTTATCGTCAGCACCAACTCTGACACTGGCGCCCCAAGCGCTGAC[A/G]TACGC TTGGTTTCACTCCTTGGCCTTCGCGCCGATGCTCTGCATACCATG	61884
3	RAC875_rep_c115433_378	DMP	IWB62322	CAAACCTGCCACACTGGTAGCAGACACCATGGAACCTTGTCTTCTGGAAG[T/C]GTTTG GTCCACTTGAGATGGACTTCCAAT	62322
4	wsnp_Ex_c851_1654297	DMC	IWA4808	GCAACCGKAKGACTGGAAGCGCGCGCAAATTCACCCCGGCTTTTCAGCCAAAG AGAAGATCAAGTCCATGTCAGCAATAACGTTGGAGTGCACAC[A/G]GCAGATGATGGAA CAGTGGCGCACTCAATGCAGGAGACACATGCAGCAAGCCGAGATCGACATGAGG TACGACTCCGATGACATAGCAATGCGTGTC	78868
5	wsnp_Ra_c1660_3275687	DMC	IWA7656	TTGATGATGGTTCGCTATCACAATCTGCACCTCAACCATGGCGAAGTTTTGCCCGGCACA AACCTCGGCCGAATGAGAAGGCCAGCAGCGCTGTAAT[A/G]CTTGGCGGCTCTC GATAAGCCATTCTGGAACCTCATCGGATTAACCTGTCGGCGTGGGTCCCCAAATCTC CTTGCCCGGTGCAACAAAATAATGGT	80948
6	Ra_c2544_1566	DMC	IWB51601	CTCTTGGCTGAAAGCCGGGTGGATGAATTTGCGCCGCGCTCCAGTCGT[A/C]TCCGT TTGCAATATTACCCCGTTCCCTAGAAATTGCTTCCAAACTGGGAT	51601
7	RAC875_c9013_185	DMC	IWB60992	AAGACTGCCTCAGACACTATCTAACGCACGTGAAGGTGCCGAAAGGAAC[A/G]ATGAT AACAAACCAATTAGTTTTGTGTCACCGGACAAGGAGATTTGGGG	60992
8	BS00084667_51		IWB11378	GTCACCCTCTGCCCGCAGCTGAGGCCACTAAACCAACATTCATTGGTGT[C]GAAGG ATTTGAGGGCGACTATGGTGAATTGAGTTTGGTAACGAGGAAGC	11378
9	Ra_c491_902		IWB52168	ATACCATTAGTTTTGTTGCACCGGGACAAGGAGATTTGGGG	52168
10	wsnp_Ex_c1996_3754394		IWA2482	CGTGCATGATCATTGGCTACATGTTTCGCTGCTAGGATACAGTTGGAACAACCCTGA CATGGATCTTCTACAACTCGCCAGAACCTAACATTGTTTT[C]GAATATCCGAAAAG AACTCTACCCATTGCATCAGCAAGCAGCGGTTGGTGTGGATGCCATGTTGATCTTT GAGCCAAAAGAGACCAGATCTCTAGTA	77079
11	Ra_c11464_294		IWB50940	GCATCAACGCTATGGAACCTCGATGTGGAGGTGATGGACGGGCAACAAT[T/C]CAGCC CAAGCAAGCTTGTATACAGCAGATGAAAAATGGGCTCATAGTTAA	50940
12	BS00023068_51		IWB7407	TACTTGCTTTGCACTGGCACTTCTAGTAGTAGTTCCCCGAACGTTGACTGT[C]AAATATG CGAGGATTTCCAACCTTGTGGTTCATCACCAGGACAGTAGTT	7407

Supplemental Table 2: Markers and oligonucleotides used for linkage analysis

Kofa+Lr19 x AUS2499 population

	Marker	Kofa+Lr19 allele	AUS2499 allele	Common oligonucleotide	corresponds to
1	W1_1	TAACGCCGCTCCTCTCGT	TAACGCCGCTCCTCTCCTT	TCAGTTGGGTATTTCTCTCCAG	JIC005
2	W1_2	ACTTCAAATGTGATGTGCTTGCTT	ACTTCAAATGTGATGTGCTGCTC	CGACAGTCGGATTCTTCTTACC	JIC007
3	W1_3	CAGAAGACTGCACACACATTACTTG	CAGAAGACTGCACACACATTACTTT	CCAGTTAAGTGAACACAGTGATCT	JIC009
4	W1_4	CGTGAGCTGCTAAACAAGGTG	CGTGAGCTGCTAAACAAGGTA	AGAGAAGTGCCTTGAGACATATTTAC	JIC010

Svevo x TTD140:

	Marker	Forward oligonucleotide (5'-3')	Reverse oligonucleotide (5'-3')	Tm (°C)	Type	Amplicon length (bp)
1	<i>Xwmc764</i>	CCTGGAACCTGAAGCTCTGA	TTCCGAAGGACTCCGTAACA	61	SSR	
2	<i>RAC875_c38665_247 (WB57438)</i>	TGCTGAGTGCACAAAGCATTGC	TGGGTATAAGTTGAGCATGCAGTT	59	SNP (HRM)	
3	<i>Xwmc661</i>	CCACCATGGTGTAAATAGTGTG	AGCTCGTAACGTAATGCAACTG	60.8	SSR	
4	<i>Xqwm614</i>	GATCACATGCATGCGTCATG	TTTTACCGTTCGGCCTT	60	SSR	
5	<i>BS00027693_51 (WB7661)</i>	GCTGTGTAACCTGAAGACCATC	TCATCATGCACCATCAGTCA	59	SNP (HRM)	
6	<i>CD893659 (JIC009)</i>	CAAATTAAGCAACAATAACACGA	GCTAGGGTATTACTGTACACC	58	PCR	723bp
7	<i>TaDMC gene^A</i>	CTGACCCATCAAGGCAGTT	CTTCAGTTGCTCCCTCCTG	68	PCR	1057bp
8	<i>TaDMH gene^A</i>	CCGGTGCCGACGCTCCTACGAT	TGGGAGCTTGCTTCAAAGGTA	65	PCR	784bp
9	<i>TaDMP gene^A</i>	GGTTACTGCACCGAGACCAT	TGAAACCAGCGTACGCTGAG	67.5	PCR	563bp
10	<i>CD927782 (JIC0010, WB43910)</i>	CTGTGTAACCTGAAGACCATCACC	GCACCATCAGTCACATGCTTA	60	SNP assay	
11	<i>CD927782 (JIC0010, WB43910) probes</i>	allele one- CAACGATGCCGCTCT	allele two-CAACGATGCTCGTCTTG	60	SNP assay	

^A Dominant in Svevo, null in TTD140

Supplemental table 3: Homology between Zavitan, TTD140 and the corresponding barley genes**TTD140 vs Zavitan flanking sequence similarity**

	Intergenic/exon/intron	
	Identities	gaps
Distal side	7827/8019 (97.6%)	72/8019 (0%)
Proximal side	17008/18287 (93%)	475/18287 (3%)

TTD140 vs Zavitan gene similarity

	PKS 1.1		PKS 1.2		PKS 2.1		PKS 2.2		PKS 3	
	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps
TTD140_CHS-1	no alignment		no alignment		no alignment		no alignment		no alignment	
TTD140_CHS-2	969/1192 (81%)	26/1192 (2%)	966/1187 (81%)	26/1187 (2%)	966/1192 (81%)	26/1192 (2%)	116/135 (86%)	0/135 (0%)	768/1167 (66%)	59/1167 (5%)

	P450 1.1		P450 1.2		P450 2.1		P450 2.2		P450 3.1		P450 3.2	
	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps
TTD140_P450-1	1326/1556 (85%)	9/1556 (0.58%)	1322/1556 (85%)	10/1556 (0.64%)	957/1117 (86%)	30/1117 (3%)	955/1117 (86%)	30/1117 (3%)	no alignment		no alignment	
TTD140_P450-2	no alignment		no alignment		no alignment		no alignment		no alignment		no alignment	

Barley vs Zavitan gene similarity

	PKS 1.1		PKS 1.2		PKS 2.1		PKS 2.2		PKS 3	
	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps
MLOC_59804/AK375690 1182 bp	1107/1181(94%)	0/1181(0%)	1107/1182(94%)	0/1181(0%)	1084/1181(92%)	0/1181(0%)	131/147(89%)	0/147(0%)	785/1154(68%)	33/1154(2%)

	P450 1.1		P450 1.2		P450 2.1		P450 2.2		P450 3.1		P450 3.2	
	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps
MLOC_12151/AK373499 1548 bp	1444/1545(93%)	0/1545(0%)	1440/1545(93%)	1/1545(0.006%)	947/1116(85%)	21/1116(1%)	945/1116(85%)	21/1116(1%)	no alignment		no alignment	

	HYD 1.1		HYD 1.2		HYD 1.3		HYD 2	
	Identities	gaps	Identities	gaps	Identities	gaps	Identities	gaps
MLOC_13397 1230 bp	1110/1235(90%)	7/1235(0.056%)	1108/1235(90%)	8/1235(0.064%)	1112/1235(90%)	7/1235(0.056%)	717/993(72%)	36/993(3%)

Supplemental Table 4: Oligonucleotides used in this study

no.	primer Fwd name	primer Fwd seq 5'-3'	primer Rev name	primer Rev seq 5'-3'	Figure	
1	TaPKS_RT_F1	GTTGGAGCCACGAAGCTAG	TaPKS_RT_R1	GACGCCGTAGCTCATCAAGC	Figure 3	
2	TaHyd_RT_F1	GACGAATTCTACCCATTAATCC	TaHyd_RT_R1	GGACACACCGGTGCCCTG		
3	TaP450_RT_F1	GGCAAATTGAAGCTGCTCAAC	TaP450_RT_R1	TTCGGCACCTTCACATGTG		
4	ACT2_RT_F	CAATCATGTTTGAGACCTCAATG ¹	ACT2_RT_R	ACCAGAATCCAACACGATACCTG ¹	Figure 4	
5	PKS_VIGS_F1	AACCACCACCACCGTGGTTACTGCACCGAGACC ²	PKS_VIGS_R1	AAGGAAGTTTAACTTCTTTGCTGCTGCTG ²		
6	PKS_VIGS_F2	AACCACCACCACCGTCACACTGGTAGCAGACACC ²	PKS_VIGS_R2	AAGGAAGTTTAAAGCTGCGTACATAGCTTAACG ²		
7	Hyd_VIGS_F1	AACCACCACCACCGTGTGAGGACCATGGCTTC ²	Hyd_VIGS_R1	AAGGAAGTTTAACTCATCACGAATCTTACC ²		
8	TaPKS_RT_F2	CCATCTTAAACGGTGTCTC	TaPKS_RT_R2	CTGCAGAAGCCAGAAATG		
9	TaHyd_RT_F2	ATGACAGCATCATGGACATC	TaHyd_RT_R2	CACTGGTGGGAACATGG		
10	CDC48_RT_F	GTCTCTGGCTGGTAAAAC ³	CDC48_RT_R	AGCAGCTCAGGTCCCTTGATAC ³		
11	MLOC_59804_F1	CCATAATTGTTTAGTCTGCTGC	MLOC_59804_R1	AATCAAATAACATTTGGGTGAG		Figure 5, Supp. Figure 6
12	MLOC_59804_F2	CAGTCCATAATGTTTAGTCTGCTGC	MLOC_59804_R2	CTACAAAGTGAAGGTGTGTGC		
13	MLOC_59804_F3	CGTTGACCCACATTTTCAG	MLOC_59804_R3	GGAGTTGAGCAGTTCATCC		
14	MLOC_13397_F1	TCGTCAACATATCCCGTAG	MLOC_13397_R1	ACTCCAGACGTGATCATAACC		
15	MLOC_13397_F2	GGTTGGTCATGGTGCATC	MLOC_13397_R2	GCTCTTATAAGCTTCTTGCTG		
16	MLOC_13397_F3	CCATGGTGATCCAGATG	MLOC_13397_R3	CAATGTTCCATTGCAATACC		
17	MLOC_12151_F1	GAAAAACATTCTGTACTGGTTG	MLOC_12151_R1	CAGTATTACTAAGTCTTCTGTAAC		
18	MLOC_12151_F2	AACATTCTGTACTGGTTGG	MLOC_12151_R2	GAGTCGTGCATCATGTCG		
19	MLOC_12151_F3	GGAAGGATGTTAAAGCTGCTC	MLOC_12151_R3	GAACATGGCCCAGGTAATAAG		
20	MLOC_59804_RT_F	TGTCCCAAGAAGAGTACCCCTGAC	MLOC_59804_RT_R	GCCCGTGAGACCACATATTATCT	Figure 5	
21	MLOC_13397_RT_F	CCAACATAATATAGATCTGACAGC	MLOC_13397_RT_R	GGTGACGGGTGACCTACG		
22	MLOC_12151_RT_F	TCCTGTACTGGTTGGGACC	MLOC_12151_RT_R	TTGGAACATGTCTGTCTG		
23	HvActin RT_F	GCCGTGCTTTCCCTCTATG ⁴	HvActin RT_R	GCTTCTCTTGATGTCCCTTA ⁵		
24	Hyd_RF_F	TCCGCGGGTAAAACCTGTACTTCCAGGGT CCTGCAACAAGACTTACCCTC ⁵	Hyd_RF_F_R	GTGGTGGTCTCGAGTGCGGCCGCAAGCTTTTA GAAACAGTTGTTTCATCATGGATC ⁵	Figure 6	

¹Wheat actin gene [TC234027] (Tenea, G.N., et al., Reference genes for gene expression studies in wheat flag leaves grown under different farming conditions. BMC Res Notes, 2011. 4: p. 373.)

²Sequences complementary to the target gene cDNA sequence are underlined. Ligation independent cloning (LIC) adaptor sequences are shown in bold.

³Wheat cell division cycle 48 [Unigene Ta.46201]

⁴Barley actin gene [GenBank: AY145451] (Kapazoglou, A., et al., Epigenetic chromatin modifiers in barley: IV. The study of barley polycomb group (PcG) genes during seed development and in response to external ABA. BMC Plant Biol, 2010. 10: p. 73.)

⁵Sequences complementary to the target gene cDNA sequence are underlined. Restriction free (RF) sequences adaptors from pET28.TEV are shown in bold.

Supplemental File 1: Gene Sequences of β -diketone biosynthesis and those annotated in the genome of the Zavitan accession.

β -diketone genes from *T. aestivum* cultivar Bethlehem (BL):

>TaDMP

ATGGCAGGCAGCTCACCGAAGGTTAGTGAGATCCGGTGTGCGCAGCGTGCGGAAGGCTCCGCGGCAATGCTGGCTATCGGAACAGCAAATCCGGCGAACAAGGTGTCCCAAGAAGAGTACCCTGACTATTATTTCCGCGTTACCAAGAGCGAGCACCTTACTGACCATAAAGACACATTCAAGATAATATGTGGTCTAACGGGCACGGAGAATCGTTTTCTTCTACCACACGGATGAACTGCTCAACTCCCACCTGTCTTGCTGGACAACACGTCACCGTCCCCTGAGGCTCGGCATGATATCGTGGCCAAGGCTGCTCCAGAGCTTGCGGCAGCAGCAGCAAAGAAGGCCATCGCAAAGTGGGGCCGTCCGGCCAGTGACATCACCCACCTTATCGTCAGCACCAACTCTGACGCTGGCGCCCAAGCGCTGACGTACGCTTGGTTTTCACTCCTTGGCCTTCGCGCCGATGTCTGTCTGCTACCATGCTCCATCTTAACGGCTGCTTCGCCGGCTGCTCCGCCTTGCCTAGCCAAGGACCTTGCTGAGAATAACTGTGGGGCACGCGTCTCGTAGCTTGGCTGGAGCTCGCCATTTCTGGCTTCTGCAGCCCCGGCGAAGGGGACTGCTTAGACACCCTCATCACTCATGCGTTGTTTGGTGATGGGGCAGGCGCGGTAATCGTCGGCGCCGACCCCATGCACCCCATCGAGAATCCTCTGTTTGGAGATGGTGCTCGTCTCTCAGACCCTCGTGCCGAGCACTGAGAATGTGCTCACCCCTGAACTTGGGGAGCAATGGCACTCATGGGAAAGTTTACACCAAACGCCCACACTGGTAGCAGACACCATGGAACCTTGTCTTCTGGAAGCGTTTGGTCCACTTGAGATGGACTTCCAATGGAATGACCTCTTCTGGGCAGTGCACCCTGGCAGCCGTGGGATCTTGGACCAACTTGACAAGCACTCCAGTTGGAGCCCACGAAGCTAGCGGCAGCAGAAGTGTCTGACAGAAGTTTGGGAACATGTTTAGCGCCACCCTGATCTTCGTGCTTGATGAGCTACAGCGTCGAATGGAGGAGGAAGGAGAGCAAGCTGAGTGGGGGGCCATGGTGGGATTTGGACCAGGCTTCACTATCGAGACCATGGTGTCCATGCAACCCGGCGCTCTCAAGAAAAAT**TAG**

> TaDMP_protein

MAGSSPKVSEIRCAQRAEGSAAMLAIQTANPANKVVSQEEYPDYYFRVTKSEHLTDHKDTFKIICGLTGTENRFFYHTDELLNSHPVLLDNTSPSREARHDIVAKAAPELAAAAAKKAIKWGRPASDIITHLIVSTNSDAGAPSADVRLVSLGLRADVCRMLHLNGCFAGCSALRLAKDLAENNCGARVLVACVELAISGFCSPEGDCLDTLIHALFGDGAGAVIVGADPMHPIENPLFEMVSVSQTLPVSTENVLTLNLGSNGTHGKVYTKLPTLVADTMEPCLLEAFGPLEMDFQWNDLFWAVHPGSRGILDQLDKTLQLEPTKLAASRTVVQKFGNMFSAIVIFVLDELQRRMEEEEGEQAEWGAMVGFPGFTIETMVLHATGALKKK*

>TaDMH

ATGCCTGCAAACAAGACTTACCCCTCCCATAAGAATGCCAACGGTGAGGTGGACGATGAATTCTACCCATTAATCCGCAAGTACAAGGATGGCCGGATCGAGCGGTTTCATGAGCTCATTCGTGCCGGCGTCGGAGGACCCGGCCGACAGTTCGTGGTGTGGTGACGAGGGACGTCGTCATCGACCAGGGCACCGGCGTGTCCGTGCGCCTGTTCTTCCCTGTCCAGGCTGCCAGGCTGGCACGAGGCTCCCCCTTGTGTGTACGTCCATGGTGGTTCCTTTTGCACGGAGAGTGCCTTCTCCCGGACGTACCACCGTTACGCCACTTCCCTCGCCGCCAGCGCTGGGGCGCTCATCGTGTCCGTGGAGTACCGTCTGGCGCCGGAATATCCCGTGCCAACGTCTACGATGACACATGGCCCGCTGCGGTGGGTGGCGTCCCTTGTCCGACCTTGGCTCGCCAAGTACGCAGACCCTAGCCGCACATTCCTCGCCGGCGACAGCGCCGGCGGCAACATCGTGTACCACACGGCCGTGCGCGCCACACGTGATGACAGCATCATGGACATCCAGGGGTTGGTTCATGGTGCATCCATTCTTCTGGGGCCCCGAGCGTCTCCCGGCGGAGAAGGTTTTGGACGGCGACCCATGTTCCCGCCAGTGTGGGTGGATAAGCTGTGGCCCTTCGTGACGGCGGCGGGGCTGGCAACGATGATCCTCGGATCAATCCTCCGGACGAGGAGATCGCGTTGCTAACTGGCAGGCG

GGTGCTTGTGGCCATTGCAGAGAAGGACACCCTGCGCGACCGGGGGCGCCAGTTTGTGTGCAGCATGCGC
GGGTGTGGGTGGGTGATGGCAGCCTCACCCTGGTGGAGTCGGAGGGTGAGGACCATGGTTTTCCATCTGT
ATGCTCCCCTACGTGCGACCAGCAAGAAGCTTATGAAGAGCATCGTGCAGTTCATAAACCATCGCGCCAC
CTTGCCGTCACCGGCCATGGTGTATCCCAGGAGGCTCGGCCGAAACTATGCTAGGCGTCCCTAGTAGGCCA
TTTAAGGACATATTTGGCTACGGGATGCGCATGAAACGTTGGAGTGGCACGAGTTTGGGGCTCAAAGTTG
GTCGTGCAAAAGCATCGACGACGAGCTATGGGTACGTTTGAAGCAAGCTCGTACCTTCGGAGACCCTGT
TTCAGCACCAACTTCGGTAAGATTCGCGATGAGGAACTGTTTC**TAG**

>TaDMH_protein

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AALRWVASLSDPWLAKYADPSRTFLAGDSAGNIVYHTAVRATRDDSIMDIQGLVMVHPFFWGPRLPAE
KVLGDGAMFPPVWVDKLPFVTAGGAGNDDPRINPPDEEIALLTGRRVLVAIAEKDTLRDRGRQFVCSMR
GCGWVDGSLTVVESEGEDHGFHLYAPLRATSKKLMKSIQVFINHRATLPSAMVIPGGSAAETMLGVPSRP
FKDIFGYGMRMKRWSGTSFGLKVGRAKASTTSYGLRLKQARTFGDPVSAPTSVRFAMRNC*

>TaDMC

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ATTCTGCTTTGGGTCCTGCCCGACTGCCAAAGGATGCTCGTTGCCGGGAGAGTCAAGGATCTGGACACG
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GAACATGCAGCAAGCCGAGATCGACATGAGGTACGACTCCGATGACATAGCAATGCGTGTATAGCACGA
GTGATGCTGGGCAAGAACTACAGGGAGGCCTGGGAAGTGTTCATGGCAGGAAGGGAGCAACTGAAGCTCG
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CTGGCAACTCGACAAGCTTGTGAGAAGCAAGATTACAGAGATCATAAAGGCGCGGCTTGCTAGCAGTGTC
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TGAGCACGGAGGAGATGGTTCGGCGAGTGCAGGACCTTCTTCATGGCTGGGTACGAAACCAGCGCCAACCT
CATTACCTGGGCCATGTTCTGCTCGCCAGCCACCCACGGTGGCAGGAGATGGTCAGGGACGAGGTGCTC
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TCTGGGAGACATTGAGGCTCTATGGCCCATATCATTCCTGCAGAGGAAGACAGCCTCAGACACAATCCT
CACACATGTGAAGGTGCCGAAAGGAACGATGATAACGATACCTCTGGTGATGTTGCACCGGGACAAAGAG
GTCTGGGGACCCGACGCCGACAAGTTTAAACCAATGAGGTCCAGAATGGCTTCGCGAGAGCCGCCAAGC
ATTCACATGCACTGCTGGCCTTCTCGTATGGGCTTAGGGTCTGTGTGCGGACAGAACTTGGCCATGGTGG
GGTGCAGATCGTCATAGCGACGATGCTCAAAGTTTCTCCTTCTCCCTGTCCCCACTTATGTGCACAAG
CCGAGCAATTCGTCACATTGACGCCCAAGTACGGGCTCCCTCTCATCGTGAGGAACCTGCAGCTGACTA
GG**TGA**

>TaDMC_protein

MAANVVQALAAVLTLLVITRALWYLLWRPYAVARWFEQQGIRGPPYKFLVGSPLDCQRMVLVAGRVDLDT
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CKRRRLEAAPQIHPFQFPREDQVHVSNNVGVHTADDGTVAHSNAGENMQQAEIDMRYSDDDIAMRVIAR
VMLGKNYREAVEVFMAGREQLKLAAYAFADPPVPFGRYLPTRNRRTWQLDKLVRSKITEIKARLASSV
YGDDLLGQMLWLQSRGAGANAENLSTEEMVGE CRTFFMAGYETSANLITWAMFLLASHPRWQEMVRDEVV
QEFPAHKPPLGDGLGKLLNMLLWETLRLYGPISFLQRKTASDTILTHVKVPKGTMITIPLVMLHRDKE
VWGPDADKFNPMRFQNGFARAAKHSHALLAFSYGPRVCVGNFAMVEVQIIVIATMLKSFSFSLSPITYVHK
PSNFVTLTPKYGLPLIVRNQLTR*

Zavitan genes:

Nucleotide and protein sequence of 22 annotated genes in the Zavitan *W1* physical map interval. Gene/protein nomenclature is similar to Figure 2C.

>NB-Arc-like_1

ATGGCGACGTTTCGTGATCCAGCACAAACACAGCACTAGCTAAGCAAGAGTCCGTGGTTAGCACTAATAAGC
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 TCATCAAACCAACACCAGGTACGCACACTACGCAGCACACACGTTAACCTCAGAGTCCCATTACATTGATG
 GCTTCCCTGGCCCCGAAATACCGTGCGGCTCCTTGTCTGGTGGCGTCGTTCTGCAGCTGAGCTCCATCA
 TGCCGGCCGCGGGCTGGCAGGATGCTTGCAGGGGACGTACTGAAGTGTGAAGGCGGGTTGGTATTCAA
 GAGAGATCCCTCCATTAGCGGGCGGGCATGTGCGTTCGTAGCCGACATACTCAAGTGCGGCGGGGTTG
 GTATTCAAGGAAGATCCCTCCGTTAGCGGGCGGGCAGTGCCTCGTAGCCAACGTACTCAAGTGCGGCG
 CCGGGTTTCGAATTCGAAGGAAGATCCATCCATTAACGGCGGTGGCGGGTGCCTCGTAGCCGCCGTACTCA
 GTGCGGGCGGGGTTTCAATTCAAGAAAGATCCCTCCATGAGCGGGCGGGCGGGATGCGTTCGTAGCCGCC
 TTACTCAAGTGCGGTGTGGGTTTCAATTCAAGGAAGATCCCTCCATTCGCGGGCGGGCGGGAGTGTGTTG
 TAACCAACGTAGTCAAGTGCAGCGGGGTTTCAATTCAAGAAAGACAACCCCCCTACCGGGCGGGCGGAA
 GTTCCCATTAAATGGTGCATGCGTCTGTCGTCGTCGTTGATTGGCTATTTTCTTTGTTCTTCAATCC
 TCTGTGTATGGTTTGTGATAACAGTGACTATTTTATTCAAGGCTGGGGCGCCCCCTCCCGTCGATCGC
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 GGAAGATAGGCACGGTGTACGGCACGGACAAAGTTGCCAAGAGTTGTATTGGACAGGCCCGCAAACCTGG
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 AAAGACCATCCCATGTCACGTGAGAGCCTGGTACGACTTTGGGTTTTCAGAAGGCCCTTGTGGTGAAGT
 GCATGGACACAACAGAGATGCTAGCCGAGGAAAATATCAGGGAATTGGTCTGCCGCAATATGCTTGAAGT
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>NB-Arc-like_1_protein

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 VVANVLKCGAGFEFKEDPSINGGGCVVAVLKCGAGFEFKKDPSSGGGGCVVAALLKCGAGFEFKEDPSIRGGGE
 CVVTNVVKCDAGFEFKDNPPTGGGKFLPMVHASVVVVVDWLFPLFLQSSVYGLSITVTILFKAGAPPSRRSLSYAI
 VSSEQLAVAPQTPCKSVDDLMAIEPMMGMQRPNVLRFMLAQEEAAMKQRFAYKVDVPTVVVRAVMKIGSILNVEAA
 GASIYKLSDEVTRKELPVKVYQISEPLIMSGSVIGKIGTVYGTDKVAKSCIGQARKLAYHVEDWITVSQTYTVEVL
 MPLAGGTDKTDMEQEVYFHIHDALQNFQGSIVSTTWNEHVAMISSPTGRLRLEQLSQPDAFDLLCRRDFTETTATC
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 KTCEM*

>NB-Arc-like_2

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 CCATTAGCGGGCGGGCATGTGCGTTCGTAGCCGACATACTCAAGTGCGGTGGCGGGTTGGTATTCAAGGAAGATCCC
 TCCGTTAGCGGGCGGGCAGTGCCTCGTAGCCAACGTACTCAAGTGCGGCGGGGTTTCAATTCAAGGAAGATCC

ATCCATTAATGGCGGTGGCGGGTGCCTCGTAGCCGCCATAGTCAAGTGCGGCGCCGGGTTTCAATTCAAGAAAGATC
 CCTCCATAAGCGGGCGGGCGGATGCGTCTGAGCCGCTTACTCAAGTGCGGTGTGGGTTTCAATTCAAGGAAGAT
 CCCTCCATTTCGCGGGCGGGCGGAGTGTGTTGTAACCAACGTAGTCAAGTGCAGCGCCGGGTTTCAATTCAAGAAAGA
 CAACCCCCCTACCGGGCGGAGCAAGATAACAGTGACTATTTTTATTCAAGGCTGGGGCGCCCCCTCCCGTTCGATCGC
 TCAGCTATGCCAATGTTTTGCTCTGAGCAGCTTGCAGTTGCACCCCAAACACCTTGCAAAAGTGTAGATCACCCCTCGT
 GGCAATCCAGCCAATGATGGGCATGCAAAGGCCCAATATCATCCGTTTTATGCTAGCCCAGGAGGAAGCCGGATATC
 ATGA

>NB-Arc-like_2_protein

MASLARNTVLLLLVLVGVVLQLSSIMPAAAAGRMLAEDVLKCEGGLVFKRDPSISGGGMCVVADILKCGGGLVFKEDP
 SVSGGGECVVANVLKCGAGFEFKEDPSINGGGGCVVAAIVKCGAGFEFKDPSISGGGGCVVAALLKCGAGFEFKED
 PSIRGGGECVVTNVKCDAGFEFKDNPPTGGSKITVTILFKAGAPSRRLSYANVSSEQLAVAPQTPCKSVDPHPR
 GNPANDGHAKAQYHPFHASPGGSRIS*

>DUF4220

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 GCTGTGGCCGCGCCACCTGCTCTCTGTTGGTGCAGGCATTAGGAGTAGGTTATGTCTTCTACAAGTATGTGCTG
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>DUF4220_protein

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 LRNSKLD SIRKFLDEVEKPRVVKEAEREERPYPLPAQWGEEDKLDSEEVLQGAHLLPICMGQFVDYKFLPSRLQI
 QANCRFKRKGCLYELIEMQLSLMHDVLYTKAAVFTWYGC SIRAI SLVATISAFFLFQSSIGKNDFSRGDI IVTYIL
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 EVAKVWELKHCKDLWKKLHRSRPVSYGTELVMRHVTRMVEECHGQEDVMRSYSQCALSRWQVTFEDPTSRAGID
 FDDKILAWYFATKMMFFQFHVEPRQLEPVVEAIDTLESEYMI FLLLERPYMLPSPVPRPVYANAEEAYQKLNHLRCVGS
 FIKRITRMAELDKRREELEKTKRDRWHARELKKIREELETIREEVQVLERGKENLLSVRLHPPALVRGAEIVERLIY
 NDARDLDNMAVGTVLLGVWVEMLCYAAHCSRDSHARQLNNGGEFITIVWLLSTAI FNGNYIR*

>PKS-1.1

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 GATGAACTGCTCAACTCCCACCCTGTCTTGCTGGACAACACGTCACCGTCCCCTGAGGCTCGGCATGATATCGTGGC
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>PKS-1.1_protein

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 HATGALKKN*

>PKS-1.2

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>PKS-1.2_protein

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>P450-1.1

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>P450-1.1_protein

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 QLKLAAYAFADPPVPGFRYLPTRRNRRTWQLDKLVRSKITEIIKARLASSVYGDDLLGQMLWLQRSAGANAETLST
 EEMVGEERTFFMAGYETSANLITWAMFLLASHPRWQEMVRDEVVQEFPAHKPPLGDGLGKLLKLLNMLLWETLRLYGP
 LSFLQRKTASDITLTHVKVPKGMTITPLVMLHRDKEVWGPDADEFNPMRFQNGFARAAKHSHALLAFSYGPRVCVG
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>Hyd-1.1

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 CTAG

>Hyd-1.1_protein

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>Hyd-1.2 (pseudogene)

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>Hyd-1.2 (pseudogene)_protein

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 AKYTDPSRTFLAGDSAGGNIVYHTAVRATHDDSIMDIQGLVMVHPFFWGPRLPAEKVLDGDAMFPPVWVDKLPWFV
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 RPARSL*

>HlyIII-1.1

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>HlyIII-1.1_protein

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>P450-1.2 (pseudogene)

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>P450-1.2 (pseudogene)_protein

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 D*

>Hyd-1.3

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>Hyd-1.3_protein

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 PLKQARTFGDPVSAPTSVRFRVNRNCF*

>HlyIII-1.2 (partial)

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>HlyIII-1.2 (partial_protein)

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 VVYYTFLCDPFSRSLYLGSIITICGAAAVAVSLLPVFQAPDLRWARAALFACMGASGIVPIVHKMLMFHSRPEAVLTT
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>PKS-2.1

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>PKS-2.1_protein

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 DPIHPVENPLFEMVSVSQTFVPSTEHVLTLLNLGSHGTHGKVVYTKLPTLVADTMEPCLSEAFGRLEMDFKWNDFWAV
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 HATSALKEN*

>P450-2.1

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>P450-2.1_protein

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YAFADPPVPGLWYLPTRRNRRRAWYLDKLVKHKI SQIIEARLASGVYEDDLLGQMLQLQLQTCSSGSETLSTEEMVG
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>P450-3.1

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>P450-3.1_protein

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CCDKVENGLLPFFRRMASTGTPFDVQELMSRFMFDLAAMPFFGVDPGLLSSEMLQDR LHMDAAVALDTFMEVGF SLL
MTPSSYWKLMRWN IGP ERK LKVARTV LREFAAEMMERRKMNTCFVGNED EQEDGDI LSCFLNDPDYADDDLLRAMI
IGYMF AARDTVGTTLTWIF YKLAQNP NIVSNIRKELSPIASRKEAVGVDAMLI FEPKETRSLVYLKAVLYETLR LYP
AAPLECKTVVADDIMPSGHEVHAGDTIIISIHSMGRMEGVWGEDCLHYK PDRWLLVGGNNMRYVPSHKFLAFNSGPR
MCLGKDIAIMQKT V IASTLWNFDVEVMDGQTIQPKQACIQQMKNGLIVK LKKREM*

>PKS-2.2 (partial)

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>PKS-2.2 (partial)_protein

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>P450-2.2

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>P450-2.2_protein

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 IHPAFNQEIKSMSAITLECTQQTMERWRNRQOAEIDMMHDSDEIAMSVIARVMLGKCYKEAWEVFIAGKEQLKLAT
 YAFADPPVPGLWYLPTRNRRAWYLDKLVKHKISQIEEARLASGVYEDDLLGQMLQLQLQTCSSGSTETLSTEEMVG
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>P450-3.2 (pseudogene)

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 TCTTGGCCAACCTCCACAACCTGCACGAGTATTTAACCTTGTCTTGGCGGATCAGGGCACAACCTCAGGGCGCAT
 GGCCACCCGGGACCGGGTTGCGGTTCTTTGTACATGCGACCCTACAAATGTCCGACACATTTTACGACCAACTA
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 GAACT**GATGTCAAGTTTTATGTTTGACCTGGCTGCTATGCCTTTCTTTGGCGTGGATCCTGGCCTCCTGTCTCGGA**
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TGACACCAAGTTCTTATTGGAAGTTGATGAGGTGGCTAAACATTGGCCCTGAGAGAAAGCTCAAAGTGGCGCGCACG
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>P450-3.2 (pseudogene) _protein

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 CCDKVENGLLPFLGEWRALALHLTKN*

>PKS-3

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>PKS-3_protein

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 DAVHPIERPLFEMVSASQTTIPATDGLVLTMLQTEAGLDGHI FTKELIPLAAQHIEQCLMDAFQQLGIMNGGTNWNLD
 FVVVHPGTHGIMDHIQRALRLDPGKLAASRTVLSEYGNMLGATLIFVLDEQRRQMEEDGETGEWGVMMGFPGPFTVE
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>Hyd-2

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>Hyd-2_protein

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 VLSLSDPWLSDYADLGRFTLVGDSAGGNIVYNTAVRAASGGSHIHIEGLVIVHPYFWGVERLSSSEAVWDGIAMFAP
 EDVDRLWPFITAGRLGNDPLVNPVDEEIASLTCRRVLVAVAEKDTLRDRGRRLAARMRDCCSWADDENAVTLIESK
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>Wax ester synthase (WES)

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>Wax ester synthase (WES) _protein

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>Ank_PGG

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>Ank_PGG_protein

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