Supporting Information

Fabrication of Mediatorless/Membraneless Glucose/Oxygen Based Biofuel Cell using

Biocatalysts Including Glucose Oxidase and Laccase Enzymes

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Figure S1. Cyclic voltammograms (CVs) of (a) $GA/[[GOx/PEI]_2/CNT]$ operated at N₂-state and airstate. The concentrations of glucose provided at air-state were 0, 1, and 2 mM glucose, (b) Lac/CNT, (c) Lac/PEI/Lac/CNT and (d) GA/[Lac/PEI/Lac/CNT] operated at N₂-state and air-state. Insets of (b)~(d) indicate redox peak of cupric ions within Lac molecule that was shifted to positive direction and appeared around 0.6 V vs. Ag/AgCl at O₂-state. For the CV tests, 0.01 M PBS (pH 7.4) was used as electrolyte and potential scan rate was 5 mV·s⁻¹.



Figure S2. Laviron's plots for (a) $GA/[[GOx/PEI]_2/CNT]$ as anode and (b) Lac/CNT, (c) Lac/PEI/Lac/CNT and (d) GA/[Lac/PEI/Lac/CNT] as cathode. For the CV tests, 0.1 M PBS (pH 7.4) was served as electrolyte and potential scan rate was 5 mV·s⁻¹.



Figure S3. Michaelis-Menten plots (a) $GA/[[GOx/PEI]_2/CNT]$ as anode and (b) Lac/CNT, Lac/PEI/Lac/CNT and GA/[Lac/PEI/Lac/CNT] as cathode. In the tests of the anodic catalyst, 0.1 M PBS (pH 7.4) was used as an electrolyte at N₂ state and 0~2 mM glucose was supplied, while In the tests of the cathodic catalysts, 0.1 M PBS (pH 7.4) was used as an electrolyte at N₂ state and 0~10 mM ABTS was supplied. In all the tests, potential scan rate used was 5 mV·s⁻¹.



Figure S4. Stability evaluations by measurements of catalytic activities of (a) GA/[GOx/PEI]₂/CNT] and (b) Lac/CNT, Lac/PEI/Lac/CNT, GA/[Lac/PEI/Lac/CNT] catalysts for four weeks.



Figure S5. Stability evaluations by measurements of membraneless mediatorless EBC with $GA/[GOx/PEI]_2/CNT]$ as anode catalyst and GA/[Lac/PEI/Lac/CNT] as cathode catalysts for four weeks. In this experiment, 0.1 M PBS (pH 3) was used as an electrolyte, 40 mM glucose solution was supplied with flowrate 0.5 cc min⁻¹, and the distance between electrode was 1 mm. For cathode, 100 cc·min⁻¹ O₂ gas was supplied.