

1 ***New Phytologist* Supporting Information**

2

3 Article title: **The abiotic and biotic drivers of rapid diversification in Andean bellflowers**
4 (**Campanulaceae**)

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10 The following Supporting Information is available for this article:

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12 **Fig. S1** Time-calibrated phylogeny of Neotropical Campanulaceae, with outgroup
13 representatives of Lobelioideae and Campanuloideae collapsed.

14

15 **Fig. S2** Time-calibrated phylogeny of Neotropical Campanulaceae, with the ingroup collapsed
16 and showing outgroup relationships.

17

18 **Fig. S3** Phylogram showing molecular-proportional branch lengths, illustrating substitution rate
19 heterogeneity within clade.

20

21 **Fig. S4** Diversification rate analysis for Neotropical Campanulaceae in BAMM.

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23 **Fig. S5** Posterior distributions of trait-dependent speciation (i), extinction (ii), transition (iii), and
24 net diversification rates (iv) as estimated by BiSSE.

25

26 **Table S1** Taxon sampling including voucher information, country of origin, and GenBank
27 accession numbers for seven plastid loci.

28

29 **Table S2** Divergence time estimation to assess sensitivity to dating method, calibration selection,
30 and branch length heterogeneity.

31

32 **Table S3** Trait codings for the four traits used in our BiSSE analyses.

33

34 **Table S4** Paleoenvironmental-dependent diversification analyses using paleoaltimetry (a) and
35 Cenozoic climate data (b).

36

37 **Table S5** Model comparison for the four BiSSE analyses presented in the main text.

38

39 **Table S6** Correlated evolution of binary traits using BayesTraits.

40

41 **Table S7** Model comparison for the two diversity-dependence analyses presented, with mean
42 parameter estimates for each model.

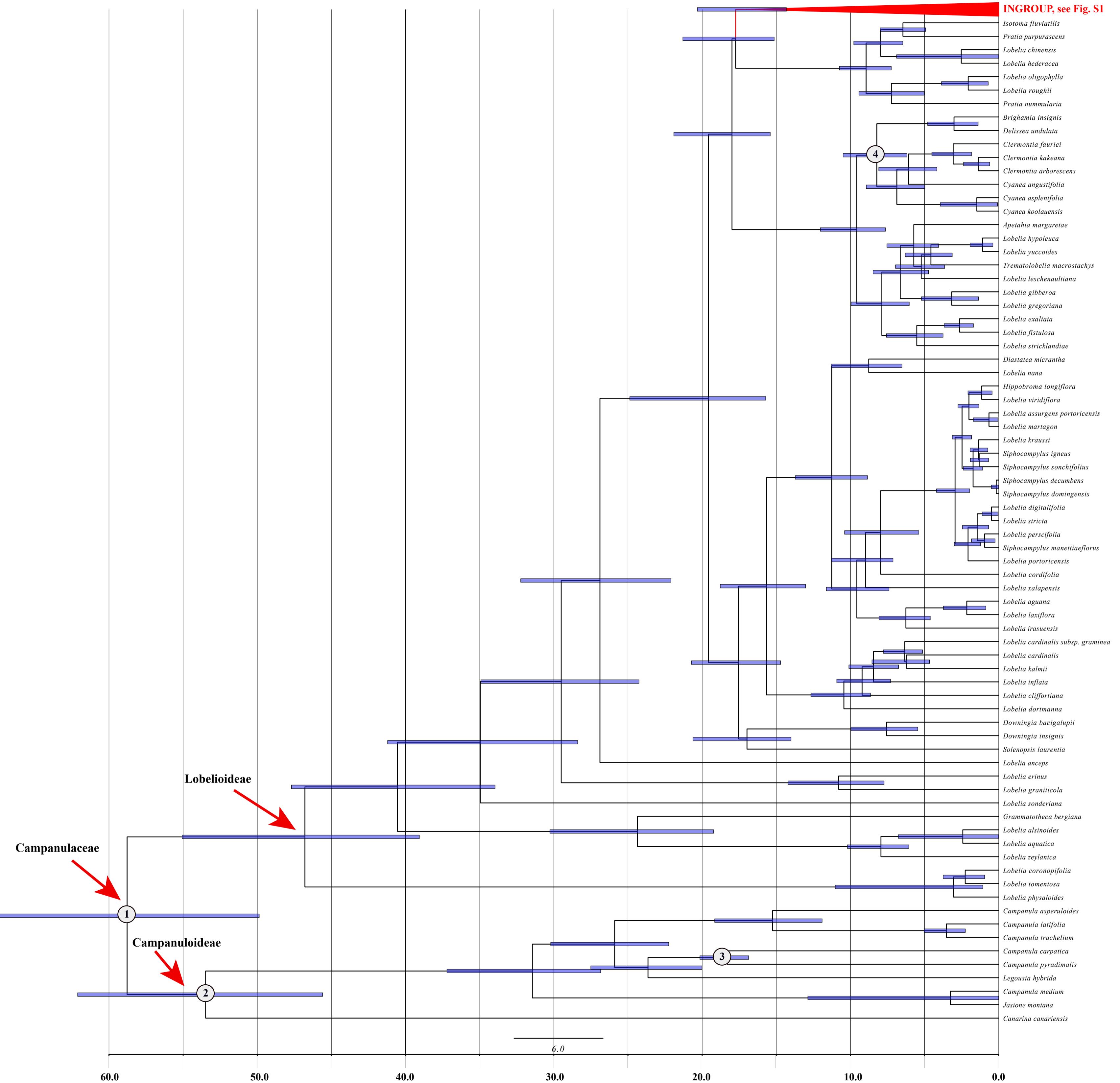
43

44 **Figure S1 (next page). Time-calibrated phylogeny of Neotropical Campanulaceae, with**
45 **outgroup representatives of Lobelioideae and Campanuloideae collapsed.** Phylogeny
46 represents a maximum clade credibility tree resulting from BEAST analysis. Bars at nodes
47 represent the 95% highest posterior density (HPD) of inferred ages. Time series below the
48 chronogram is in Ma. Ingroup relationships shown, with outgroup clades collapsed. Red arrow
49 marks the ingroup, the Neotropical bellflowers. Taxon names are color-coded accorded to
50 current taxonomy: *Burmeistera* (green), *Centropogon* (yellow), *Siphocampylus* (red), *Lysipomia*
51 (purple), and *Lobelia* section *Tupa* (light blue).

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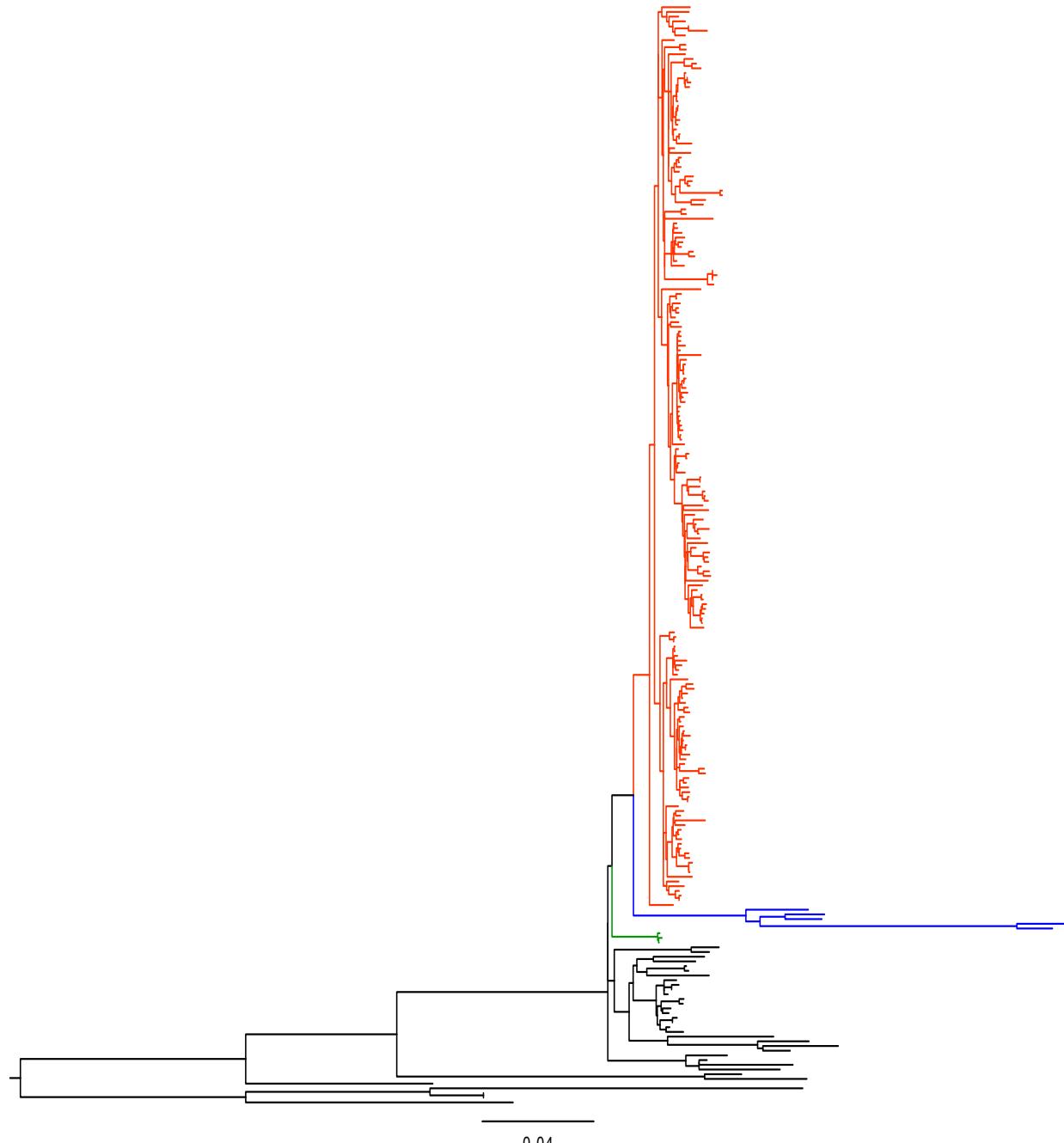


54 **Figure S2 (next page). Time-calibrated phylogeny of Neotropical Campanulaceae, with the**
55 **ingroup collapsed and showing outgroup relationships.** Phylogeny represents a maximum
56 clade credibility tree resulting from BEAST analysis. Bars at nodes represent the 95% highest
57 posterior density (HPD) of inferred ages. Time series below the chronogram is in Ma. Outgroup
58 relationships, with ingroup collapsed (in red, at top). Red arrows denote major taxonomic
59 groups: Campanulaceae, Campanuloideae, and Lobelioideae. Numbers circled at nodes
60 correspond to the four calibration points used to infer ages: 1) secondary age constraint for
61 crown group Campanulaceae; 2) secondary age constraint for crown group Campanuloideae; 3)
62 the fossil, *Campanula paleopyramidalis* at most recent common ancestor of *C. pyramidalis*
63 Gilib. and *C. carpatica* Jacq.; and 4) the geological age constraint for the oldest island of the
64 Hawaiian Ridge. See methods in main text for additional details.
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67



68 **Figure S3 Phylogram showing molecular-proportional branch lengths, illustrating**
69 **substitution rate heterogeneity within clade.** Branches colored in red correspond to the
70 centropogonid clade, in blue to *Lysipomia*, and in green to *Lobelia* section *Tupa*; black branches
71 represent species in the outgroup. Note that the branch lengths within the genus *Lysipomia* are
72 much longer than in other parts of the tree, reflecting an elevated substitution rate within this
73 genus, which may result from its minute habit or the extreme high-elevation habitat to which it is
74 endemic. The scale at bottom represents substitutions/site.

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78 **Figure S4. Diversification rate analysis for Neotropical Campanulaceae in BAMM.** A. 95%
 79 credible set of shift configurations from a BAMM analysis. Colors of branches denote
 80 directionality and strength of rate change, cooler and warmer colors designate slower and faster
 81 rates, respectively. Significant rate changes are designated by a circle along the branch at which
 82 this shift takes place. Note that among the 95% credible set of shift configurations, the majority
 83 (~67% of the shift configurations) indicate a significant increase in diversification rate at or near
 84 the origin of the centropogonid clade. Bi–iii. Diversification rates (in events/Ma/lineage) through
 85 time for the Neotropical bellflower clade (i), Neotropical bellflowers minus the centropogonid
 86 clade (ii), and the centropogonid clade alone (iii). Shading in B denotes confidence interval in
 87 rate reconstructions.

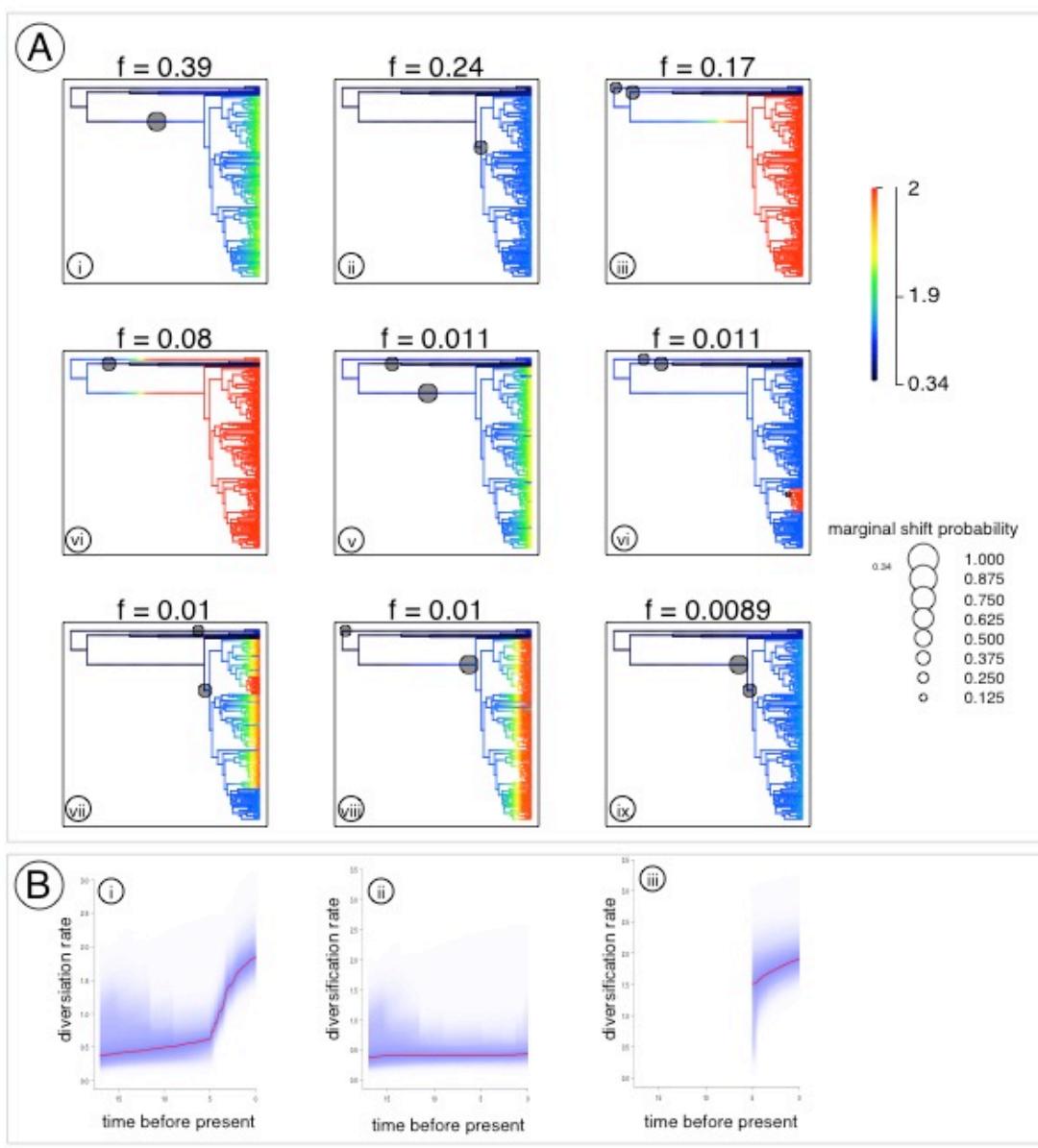
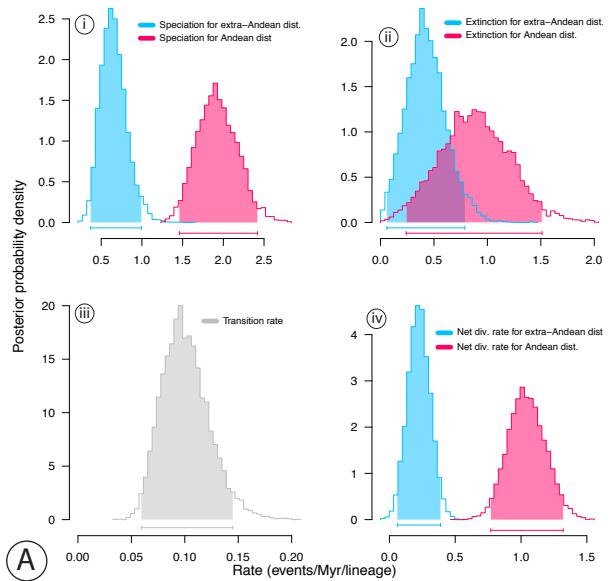
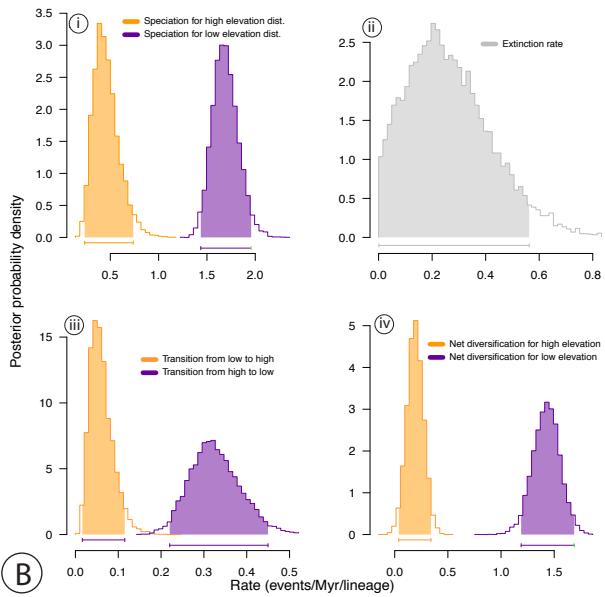


Figure S5. Posterior distributions of trait-dependent speciation (i), extinction (ii), transition (iii), and net diversification rates (iv) as estimated by BiSSE. Histogram colors correspond to Fig. 1 in the main text for each trait, except when the parameter is indistinguishable between the two traits (in which case it is colored gray). A. Andean vs. extra-Andean parameter estimates. B. High vs. low elevation parameter estimates. C. Dry capsule vs. fleshy berry parameter estimates. D. Vertebrate vs. invertebrate pollination parameter estimates.

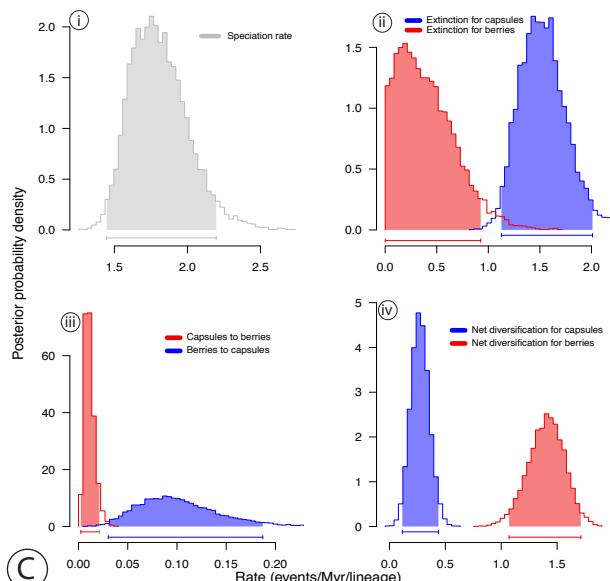
Andean v. Extra-Andean Distribution



High vs. Low Elevation



Dry vs. Fleshy Fruit



Invertebrate vs. Vertebrate Pollination

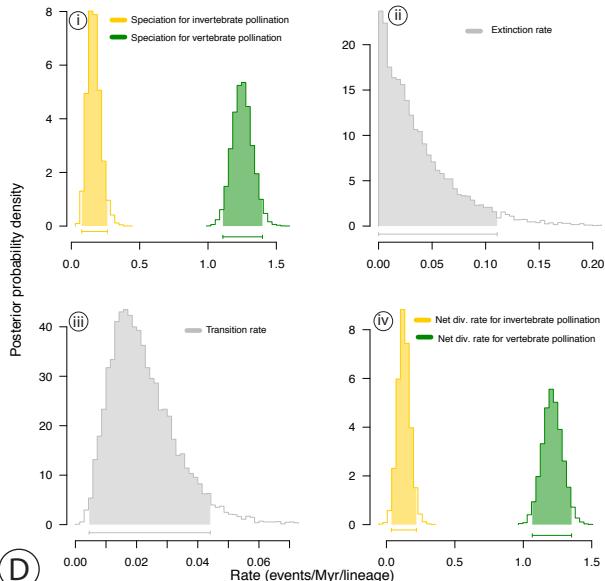


Table S1. Taxon sampling including voucher information, country of origin, and GenBank accession numbers for seven plastid loci.

Taxon	Voucher	Origin	rpl32-trnL	rpl32-ndhF	rps16-trnK	trnG-trnG-trnS	rbcL	trnL-trnF	ndhF
INGROUP									
Burmeistera									
<i>Burmeistera almedae</i> Wilbur	L. Lagomarsino 86 (GH)	Costa Rica	KP014195	KP014848	KP014511	KU670709	-	-	-
<i>Burmeistera</i> aff. <i>bullatifolia</i> J. Garzón & F. Gonzalez	C. D. Bacon 264A (GB)	Colombia	KP014167	KP014861	KP014501	KU670707	-	-	-
<i>Burmeistera</i> aff. <i>bullatifolia</i> J. Garzón & F. Gonzalez	C. D. Bacon 287 (GB)	Colombia	KP014168	KP014862	KP014502	KP014770	-	-	-
<i>Burmeistera ceratocarpa</i> Zahlb.	M. L. Bristol 993 (GH)	Colombia	KP014172	KP014852	KP014343	-	-	-	-
<i>Burmeistera chiriquiensis</i> Wilbur	D. Santamaría 8996 (INB)	Costa Rica	KP014192	KP014858	KP014521	KP014754	-	-	-
<i>Burmeistera crebra</i> McVaugh	L. Lagomarsino 70 (GH)	Costa Rica	KP014234	KP014870	KP014507	KP014758	-	-	-
<i>Burmeistera cyclostigmata</i> J. D. Smith	L. Lagomarsino 51 (GH); Andersson & Nilsson 2451	Costa Rica	KP014181	KP014876	KP014514	KP014707	DQ356147	DQ356213	DQ356097
<i>Burmeistera dendrophila</i> E. Wimm	L. Lagomarsino 255 (GH)	Panama	KP014163	KP014855	KP014524	KP014750	-	-	-
<i>Burmeistera domingensis</i> Jeppesen	N. Muchhalá 102; Harling and Andersson 23154 (GB)	Ecuador	KP014189	KP014860	KP014532	KP014699	EF174633	DQ356210	-
<i>Burmeistera fuchsoides</i> J. Garzón & F. Gonzalez	C. D. Bacon 258 (GB)	Colombia	KP014186	KP014865	KP014503	KU670692	-	-	-
<i>Burmeistera mcvaughii</i> Wilbur	L. Lagomarsino 257 (GH)	Panama	KP014166	KP014853	KP014526	KP014768	-	-	-
<i>Burmeistera mcvaughii</i> Wilbur	L. Lagomarsino 98 (GH)	Panama	KP014233	KP014854	KP014527	KP014769	-	-	-

<i>Burmeistera microphylla</i> J. D. Smith	L. Lagomarsino 34 (GH)	Costa Rica	KP014187	KP014850	KP014350	KP014720	-	-	-
<i>Burmeistera</i> aff. <i>minutiflora</i> J. Garzón & F. Gonzalez	C. D. Bacon 262 (GB)	Colombia	KP014235	KP014869	KP014508	KP014712	-	-	-
<i>Burmeistera morii</i> Wilbur	L. Lagomarsino 273 (GH)	Panama	KP014165	KP014864	KP014525	KP014771	-	-	-
<i>Burmeistera multiflora</i> Zahlb.	N. Muchhala 114	Ecuador	KP014160	KP014879	KP014530	KP014583	-	-	-
<i>Burmeistera obtusifolia</i> E. Wimm	L. Lagomarsino 61 (GH)	Costa Rica	KP014197	KP014969	KP014510	KU670700	-	-	-
<i>Burmeistera obtusifolia</i> E. Wimm	L. Lagomarsino 163 (GH)	Panama	KP014196	KP014851	KP014513	KP014701	-	-	-
<i>Burmeistera parviflora</i> E. Wimm	L. Lagomarsino 53 (GH)	Costa Rica	KP014236	KP014859	KP014498	KP014752	-	-	-
<i>Burmeistera racemiflora</i> Lammers	M. Madison 7145 (AAU)	Ecuador	KP014170	KP014845	KP014504	KP014760	-	-	-
<i>Burmeistera refracta</i> E. Wimm.	L. Andersson 2464 (GB)	Ecuador	KP014175	KP014871	KP014493	KP014713	-	-	-
<i>Burmeistera refracta</i> E. Wimm.	N. Muchhala 110	Ecuador	KP014176	KP014880	KP014494	KP014711	-	-	-
<i>Burmeistera sodiroana</i> Zahlb.	N. Muchhala 115	Ecuador	KP014239	KP014867	KP014528	KU67071	-	-	-
<i>Burmeistera succulenta</i> H. Karst. & Triana	C. D. Bacon 297 (GB)	Colombia	KP014240	KP014866	KP014529	KU670697	-	-	-
<i>Burmeistera tenuiflora</i> J.D. Smith	L. Lagomarsino 48 (GH)	Costa Rica	KP014194	KP014923	-	KP014757	-	-	-
<i>Burmeistera toroensis</i> Wilbur	L. Lagomarsino 92 (GH)	Panama	KP014241	KP014868	KP014497	KP014709	-	-	-
<i>Burmeistera truncata</i> Zahlbr.	J. Luteyn 14362	Ecuador	-	-	KU670737	-	-	-	-
<i>Burmeistera utleyi</i> Wilbur	L. Lagomarsino 253 (GH)	Panama	KP014164	KP014856	KP014523	KP014751	-	-	-
<i>Burmeistera variabilis</i> (Gleason) E. Wimm.	C. D. Bacon 298 (GB)	Colombia	KP014169	KP014846	KP014505	KP014759	KU670751	-	-
<i>Burmeistera vulgaris</i> E. Wimm.	D. Santamaría S-980 (GB)	Costa Rica	KP014174	KP014872	KP014495	KP014714	-	-	-
<i>Burmeistera zurquiensis</i> Wilbur	L. Lagomarsino 26 (GH)	Costa Rica	KP014193	KP014849	KP014512	KP014702	KU670805	-	-
<i>Burmeistera</i> sp.	C. D. Bacon 283 (GB)	Colombia	KP014171	KP014844	KP014506	KP014638	-	-	-

<i>Burmeistera</i> sp. nov.	D. Santamaría 6640 (INB)	Costa Rica	KP014237	KP014857	KP014522	KP014753	-	-	-	
<i>Centropogon</i>										
<i>Centropogon aequatorialis</i> E. Wimm.	N. Muchhala 131	Ecuador	KP014206	KP014957	KP014531	KP014767	-	-	-	
<i>Centropogon argutus</i> E. Wimm.	L. Lagomarsino 329	Peru	KU670740	KU670689	KU670726	KU670711	KU670756	-	-	
<i>Centropogon asclepiadeus</i> (Willd.) E. Wimm.	R. E. Weaver, Jr. 1556 (GH)	Colombia	KP014228	KP014927	KP014560	KU670712	-	-	-	
<i>Centropogon azuayensis</i> Jeppesen	A. Antonelli 576 (GB)	Ecuador	KP014231	KP014931	KP014558	KU670719	-	-	-	
<i>Centropogon baezanus</i> Jeppesen	N. Muchhala 162	Ecuador	KP014141	KP014836	KP014408	KP014624	-	-	-	
<i>Centropogon bangii</i> Zahlb.	S. Beck 20763 (OSH)	Bolivia	KP014122	KP014826	KP014409	KP014630	-	-	-	
<i>Centropogon brittonianus</i> Zahlb.	L. Lagomarsino 196 (GH)	Bolivia	KP014283	KP015004	KP014477	KP014686	KU670757	-	-	
<i>Centropogon coccineus</i> (Hook.) Regel ex B.D. Jacks.	D. Santamaría 8218 (INB)	Costa Rica	KP014304	KP014955	KP014465	KP014763	KU670771	-	-	
<i>Centropogon comosus</i> Gleason	P. Nuñez 8162 (AAU)	Ecuador	KP014229	KP014937	KP014546	KP014739	-	-	-	
<i>Centropogon congestus</i> Gleason	L. Lagomarsino 40 (GH)	Costa Rica	KP014127	KP014822	KP014406	KP014595	-	-	-	
<i>Centropogon cordifolius</i> Benth.	R. Kriebel 5559 (NY)	Guatemala	KP014253	KP014945	KP014457	KP014773	-	-	-	
<i>Centropogon cornutus</i> (L.) Druce	L. Lagomarsino 93 (GH); Jacobs et al. 4955	Panama	KP014121	KP014810	KP014345	KP014593	DQ356158	DQ356226	DQ356106	
<i>Centropogon costaricae</i> (Vatke) E. Wimm.	L. Lagomarsino 24A (GH)	Costa Rica	KP014306	KP014949	KP014470	KP014742	KU670774	-	-	
<i>Centropogon dianae</i> Lammers	L. Lagomarsino 415 (GH)	Peru	KP014267	KP014988	KP014444	-	KU670769	-	-	
<i>Centropogon dissectus</i> E. Wimm.	J. Luteyn 5678 (GB)	Ecuador	KP014212	KP014992	KP014561	KP014722	EF141026	DQ356215	-	
<i>Centropogon dombeyanus</i> E. Wimm.	L. Lagomarsino 332 (GH)	Peru	-	KP014986	KP014488	KP014663	-	-	-	
Dillon	<i>Centropogon eilersii</i> Lammers & M.O.	L. Lagomarsino 411 (GH)	Peru	KU670739	KU670685	KU670729	KU670715	-	-	-

<i>Centropogon erythraeus</i> Drake	L. Andersson 2529 (GB)	Ecuador	KP014227	KP014936	KP014559	KP014740	-	-	-
<i>Centropogon featherstonei</i> Gleason	L. Lagomarsino 318 (GH)	Peru	KP014219	KP014933	KP014548	KP014737	KU670758	-	-
<i>Centropogon ferrugineus</i> (L.f) Gleason	D. Santamaría S-902 (GB)	Costa Rica	KP014216	KP014920	KP014543	KP014729	-	-	-
<i>Centropogon ferrugineus</i> (L.f) Gleason	L. Lagomarsino 22 (GH)	Costa Rica	KP014217	KP014925	KP014544	KP014730	KU670759	-	-
<i>Centropogon ferrugineus</i> (L.f) Gleason	L. Lagomarsino 103 (GH)	Panama	KP014203	KP014919	KP014538	KP014696	-	-	-
<i>Centropogon foetidus</i> (Kunth) E. Wimm.	C. D. Bacon 288A (GB)	Colombia	KP014200	KP014971	KP014563	KP014774	-	-	-
<i>Centropogon gamosepalus</i> Zahlb.	N. Muchhala 429; Harling and Andersson 24342	Ecuador	KP014130	KP014823	KP014404	KP014627	DQ356157	DQ356225	DQ356105
<i>Centropogon glabrifilis</i> (E. Wimm.) Jeppsen	L. Andersson 2423 (GB)	Ecuador	KP014222	KP014916	KP014542	KP014728	KU670760	-	-
<i>Centropogon gloriosus</i> (Britton) Zahlb.	L. Lagomarsino 199 (GH)	Bolivia	KP014294	KP014979	KP014451	KP014670	KU670761	-	-
<i>Centropogon grandidentatus</i> (Schltrd.) Zahlbr.	UCBG 81.0952	Cultivated	KU670747	KU670688	KU670721	KU670702	-	-	-
<i>Centropogon granulosus</i> subsp. <i>granulosus</i> Stein (ined.) 1	Andersson 2477	Ecuador	KP014136	KU670682	KP014407	KU670701	DQ356152	DQ356220	DQ356101
<i>Centropogon granulosus</i> subsp. <i>granulosus</i> Stein (ined.) 2	L. Lagomarsino 330 (GH)	Peru	KP014142	KP014841	KP014413	KP014614	-	-	-
<i>Centropogon granulosus</i> subsp. <i>lugens</i> Stein (ined)	C. D. Bacon 303 (GB)	Colombia	KP014123	KP014840	KP014416	KP014618	-	-	-
<i>Centropogon granulosus</i> subsp. <i>nutans</i> Stein (ined.)	D. Santamaría S-962 (GB)	Costa Rica	KP014145	KP014830	KP014394	KP014623	-	-	-
<i>Centropogon granulosus</i> subsp. <i>tortilis</i> Stein (ined.)	C. Persson 829 (GB)	Panama	KP014126	KP014821	KP014411	KP014605	-	-	-
<i>Centropogon granulosus</i> C. Presl. 1	C. D. Bacon 306 (GB)	Colombia	KU670749	-	KU670725	KU670703	-	-	-
<i>Centropogon granulosus</i> C. Presl. 2	L. Lagomarsino 56 (GH)	Costa Rica	KP014147	KP014833	KP014395	KP014625	-	-	-
<i>Centropogon granulosus</i> C. Presl. 3	L. Lagomarsino 81 (GH)	Costa Rica	KP014144	KP014829	KP014393	KP014622	-	-	-

<i>Centropogon granulosus</i> C. Presl. 4	L. Lagomarsino 301 (GH)	Panama	KP014140	-	KP014396	KP014616	-	-	-
<i>Centropogon gutierrezii</i> (Planch & Oerst.) E. Wimm.	L. Lagomarsino 19 (GH)	Costa Rica	KP014301	KP014950	KP014466	KP014745	-	-	-
<i>Centropogon hirtus</i> (Cav.) C. Presl.	L. Lagomarsino 392	Peru	KU670743	KU670680	KU670727	KU670708	KU670762	-	-
<i>Centropogon incanus</i> (Britton) Zahlb.	S. Acha 207 (MO)	Bolivia	KP014279	KP014998	KP014480	KP014659	-	-	-
<i>Centropogon incanus</i> (Britton) Zahlbr..	L. Lagomarsino 223 (GH)	Bolivia	KP014285	KP014997	KP014476	KP014662	-	-	-
<i>Centropogon isabellinus</i> E. Wimm.	L. Lagomarsino 406 (GH)	Peru	KP014280	KP014983	KP014445	KP014679	-	-	-
<i>Centropogon leucocarpus</i> McVaugh	L. Lagomarsino 280 (GH)	Panama	KP014162	KP014951	KP014462	KP014743	KU670763	-	-
<i>Centropogon llanganatensis</i> Jeppsen	N. Muchhala 139	Ecuador	KP014226	KP014926	KP014556	KP014727	-	-	-
<i>Centropogon luteus</i> E. Wimm.	L. Andersson 2411 (GB)	Ecuador	KP014214	KP014922	KP014555	KP014731	DQ356151	DQ356219	DQ356100
<i>Centropogon luteynii</i> Wilbur	L. Lagomarsino 259 (GH)	Panama	KP014161	KP014911	KP014432	KP014747	KU670764	-	-
<i>Centropogon macbridei</i> Gleason	L. Lagomarsino 337 (GH)	Peru	KP014208	KP014917	KP014540	KP014732	-	-	-
<i>Centropogon macrocarpus</i> Zahlb.	L. Valenzuela 858 (OSH)	Peru	KP014289	-	KP014460	-	-	-	-
<i>Centropogon macrophyllus</i> (G. Don.) E. Wimm	L. Lagomarsino 345 (GH)	Peru	KP014120	KP014824	KP014405	KP014629	-	-	-
<i>Centropogon magnificus</i> Zahlb.	A. Fuentes 10044 (LPB)	Bolivia	KP014288	KP014995	KP014475	KP014658	-	-	-
<i>Centropogon mandonis</i> Zahlb.	A. Timmermann 1 (AAU)	Bolivia	KP014282	KP014999	KP014478	KP014687	-	-	-
<i>Centropogon mandonis</i> Zahlb.	L. Lagomarsino 243 (GH)	Bolivia	KP014295	KP014977	KP014452	KP014666	-	-	-
<i>Centropogon medusa</i> E. Wimm.	N. Muchhala 174	Ecuador	KP014223	KP014930	KP014549	KP014692	KU670765	-	-
<i>Centropogon minimus</i> McVaugh	C. D. Bacon 264B (GB)	Colombia	KP014254	KP014947	KP014458	KP014698	-	-	-
<i>Centropogon nervosus</i> Wimm	L. Lagomarsino 407 (GH)	Peru	KP014202	KP014915	KP014539	KP014724	-	-	-
<i>Centropogon nigricans</i> Zahlb.	J. Ramos 7097 (OSH)	Ecuador	KP014209	-	KP014565	-	-	-	-

<i>Centropogon nigricans</i> Zahlb.	N. Muchhala 118	Ecuador	KP014210	KP015007	KP014566	KP014723	-	-	-
<i>Centropogon palmanus</i> (J. D. Smith) E. Wimm.	L. Lagomarsino 193a (GH)	Costa Rica	KP014303	KP014843	KP014746	KP014746	-	-	-
<i>Centropogon panamensis</i> Wilbur	L. Lagomarsino 308 (GH)	Panama	KP014131	KP014828	KP014403	KP014613	-	-	-
<i>Centropogon perlongus</i> Gleason	L. Lagomarsino 409 (GH)	Peru	KP014265	KP014989	KP014447	KP014681	-	-	-
<i>Centropogon peruvianus</i> (E. Wimm.) McVaugh	L. Lagomarsino 386 (GH)	Peru	KU670744	KU670683	KU670723	KU670695	KU670766	-	-
<i>Centropogon pichinchensis</i> Zahlb.	J. Luteyn 14284 (OSH)	Ecuador	KP014220	-	KP014550	-	-	-	-
<i>Centropogon preslii</i> E. Wimm.	L. Andersson 2503 (GB)	Ecuador	KP014225	KP014929	KP014553	KP014734	-	-	-
<i>Centropogon pulcher</i> Zahlb.	L. Lagomarsino 344 (GH)	Peru	KP014143	KP014842	KP014415	KP014615	KU670767	-	-
<i>Centropogon reticulatus</i> Drake	M. Merello 1099 (MO)	Peru	KP014185	KP014921	KP014557	KP014718	-	-	-
<i>Centropogon rex</i> E. Wimm.	C. D. Bacon 288 (GB)	Colombia	KP014201	KP014972	KP014564	KU670693	KU670768	-	-
<i>Centropogon roseus</i> Rusby	L. Lagomarsino 372 (GH)	Peru	KP014119	KP014827	KP014412	KP014628	KU670808	-	-
<i>Centropogon salviaeformis</i> Zahlb.	N. Muchhala 173	Ecuador	KP014230	KP014932	KP014554	KP014726	-	-	-
<i>Centropogon simulans</i> Lammers	L. Lagomarsino 402 (GH)	Peru	KP014281	KP014974	KP014461	KP014646	KU670770	-	-
<i>Centropogon smithii</i> E. Wimm.	L. Lagomarsino 68 (GH)	Costa Rica	KP014211	KP015008	KP014562	KP014772	-	-	-
<i>Centropogon sodiroanus</i> Zahlb.	N. Muchhala 106	Ecuador	KP014204	KP015011	KP014568	KU670706	-	-	-
<i>Centropogon solanifolius</i> Benth. 1	C. D. Bacon 261 (GB)	Colombia	KP014134	KP014839	KP014402	KU670696	-	-	-
<i>Centropogon solanifolius</i> Benth. 2	L. Lagomarsino 25 (GH)	Costa Rica	KP014132	KP014832	KP014398	KP014626	-	-	-
<i>Centropogon solanifolius</i> Benth. 3	D. Santamaría S-904 (GB)	Costa Rica	KP014133	KP014834	KP014397	KP014621	-	-	-
<i>Centropogon subandinus</i> Zahlb.	N. Muchhala 475	Ecuador	KP014207	KP014913	KP014569	KP014756	-	-	-

<i>Centropogon talamancensis</i> Wilbur	L. Lagomarsino 140 (GH)	Costa Rica	KP014302	KP014953	KP014467	KP014748	-	-	-
<i>Centropogon tessmannii</i> E. Wimm.	N. Muchhala 168	Ecuador	KP014150	KP014882	KP014344	KP014700	KU670773	-	-
<i>Centropogon trachyanthus</i> E. Wimm.	N. Muchhala 138	Ecuador	KP014137	KP014838	KP014401	KP014606	-	-	-
<i>Centropogon trichodes</i> E. Wimm.	L. Andersson 2546 (GB)	Ecuador	KP014213	KP014970	KP014535	KP014766	KU670780	DQ356217	DQ356098
<i>Centropogon umbrosus</i> E. Wimm	L. Lagomarsino 349 (GH)	Peru	KP014125	KP014825	-	KP014631	-	-	-
<i>Centropogon unduavensis</i> (Britton) Zahlb.	J. Solomon 15418 (LPB)	Bolivia	KP014290	KP014978	KP014453	-	-	-	-
<i>Centropogon urubambae</i> E. Wimm	L. Lagomarsino 347 (GH)	Peru	KP014148	KP014831	KP014414	KP014617	-	-	-
<i>Centropogon valerioi</i> Standley	D. Santamaría S-977 (GB)	Costa Rica	KP014215	KP014924	KP014545	KP014733	-	-	-
<i>Centropogon vargasii</i> B. A. Stein (ined.)	L. Lagomarsino 356 (GH)	Peru	KU670745	KU670681	KU670732		KU670772	-	-
<i>Centropogon viriduliflorus</i> Wimm	L. Lagomarsino 370 (GH)	Peru	KP014268	KP015006	KP014487	KP014665	-	-	-
<i>Centropogon viriduliflorus</i> Wimm	L. Valenzuela 712 (OSH)	Peru	KP014266	-	KP014448	-	-	-	-
<i>Centropogon weberbaueri</i> Zahlb.	L. Lagomarsino 317(GH)	Peru	KP014218	KP014934	KP014547	KP014736	-	-	-
<i>Centropogon cf. weberbaueri</i> Zahlb.	L. Lagomarsino 351 (GH)	Peru	KP014232	KP014935	KP014552	KP014735	-	-	-
<i>Centropogon yarumalensis</i> E. Wimm.	C. D. Bacon 263 (GB)	Colombia	KP014221	KP014918	KP014541	KP014725	KU670775	-	-
<i>Centropogon yungasensis</i> Britton	L. Lagomarsino 392 (GH)	Peru	KP014149	KP014818	KP014391	KP014619	KU670776	-	-
<i>Centropogon</i> sp. nov. (Lagom. & Santam. ined.)	L. Lagomarsino 354 (GH)	Peru	KP014269	KP014987	KP014489	KP014664	-	-	-
<i>Centropogon</i> sp.	A. Antonelli 611	Ecuador	KU670748	KU670687	KU670722	KU670705	-	-	-
<i>Lobelia</i>									
<i>Lobelia bridgesii</i> Hook & Arn.	M. Gustafsson 1019 (AAU)	Cultivated	KP014077	KP014800	KP014338	KP014584	-	-	-
<i>Lobelia excelsa</i> Bonpland	L. Lagomarsino 426 (GH); Lammers et al. 6357	Cultivated	KP014078	KP014801	KP014341	KP014586	DQ356146	DQ356212	-

<i>Lobelia polypylla</i> Hook. & Arn.	Lammers et al. 6331 (GB)	Cultivated	-	-	-	-	DQ356123	DQ356177	-
<i>Lobelia tupa</i> L.	Cultivated, University of Aarhus; Lammers and Rodriguez 6329 (GB)	Cultivated	KP014076	KP014802	KP014339	KP014585	KU670784	DQ356211	-
<i>Lysipomia</i>									
<i>Lysipomia cuspidata</i> McVaugh	L. Andersson 2559 (GB)	Ecuador	KP014057	-	KP014318	KP014579	DQ356133	DQ356198	-
<i>Lysipomia muscoides</i> Hook f.	L. Lagomarsino 325 (GH)	Peru	KP014051	-	KP014312	KP014575	-	-	-
<i>Lysipomia pumila</i> (Wedd.) E. Wimm.	L. Lagomarsino 238 (GH)	Bolivia	KP014056	KP014791	KP014317	KP014580	KU670783	-	-
<i>Lysipomia sphagnophila</i> Griseb. Ex Wedd.	L. Andersson 2444 (GB)	Ecuador	KP014052	-	KP014313	KP014576	DQ356132	DQ356197	-
<i>Lysipomia vitreola</i> McVaugh	A. Antonelli 581 (GB)	Ecuador	KP014058	KP014792	KP014319	KU670704	-	-	-
<i>Siphocampylus</i> (excluding section <i>Hemisiphocampylus</i>)									
<i>Siphocampylus actinothrix</i> E. Wimm.	L. Lagomarsino 403 (GH)	Peru	KP014086	KP014812	KP014367	KP014643	-	-	-
<i>Siphocampylus</i> aff. <i>actinothrix</i> E. Wimm.	L. Lagomarsino 361 (GH)	Peru	KP014086	KP014812	KP014367	KP014643	-	-	-
<i>Siphocampylus affinis</i> (Mirb.) McVaugh	B. Lojtnant 11426 (GB)	Ecuador	KP014270	-	KP014456	KP014661	DQ356155	DQ356223	DQ356104
<i>Siphocampylus ambivalens</i> Lammers	J. Wood 9986	Bolivia	KP014151	KP014806	KP014421	KP014650	-	-	-
<i>Siphocampylus andinus</i> Britton	L. Lagomarsino 226 (GH)	Bolivia	KP014260	KP014984	KP014500	KP014654	KU670782	-	-
<i>Siphocampylus angustiflorus</i> Schlechtend.	L. Lagomarsino 417 (GH)	Peru	KP014097	KP014887	KP014379	KP014633	KU670785	-	-
<i>Siphocampylus antioquianus</i> E. Wimm.	C. D. Bacon 242 (GB)	Colombia	KP014090	KP014943	KP014364	KU670710	-	-	-
<i>Siphocampylus aureus</i> Rusby	J. Wood 14956	Bolivia	KP014105	KP014898	KP014383	KU670718	KU670786	-	-
<i>Siphocampylus ayersiae</i> Lammers	L. Lagomarsino 212 (GH)	Bolivia	KP014297	KP014965	KP014437	KP014648	-	-	-
<i>Siphocampylus betulaefolius</i> (Cham.) G.	S. Godoy 3000 (UEC)	Brazil	KP014153	KP014904	KP014420	KP014694	KU670788	-	-

Don.

<i>Siphocampylus bilabiatus</i> Zahlb.	A. Araujo-Murakami 4230 (MO)	Bolivia	KP014256	KP015016	KP014423	KP014677	KU670789	-	-
<i>Siphocampylus boliviensis</i> Zahlb.	L. Lagomarsino 239 (GH)	Bolivia	KP014252	KP014960	KP014483	KP014673	-	-	-
<i>Siphocampylus brevicalyx</i> E. Wimm.	N. Muchhala 480; Lojtnant et al 11826	Ecuador	KP014245	KP014901	KP014433	KP014715	KU670790	DQ356224	-
<i>Siphocampylus chloroleucus</i> E. Wimm	Asplund 13060 (S)	Ecuador	KP014098	KP014903	KP014377	KP014611	-	-	-
<i>Siphocampylus citrinus</i> E. Wimm	L. Lagomarsino 368 (GH)	Peru	KP014115	KP014896	KP014374	KP014604	KU670792	-	-
<i>Siphocampylus clotho</i> E. Wimm.	L. Lagomarsino 367 (GH)	Peru	KU670742	KU670686	KU670728	KU670714	KU670791	-	-
<i>Siphocampylus convolvulaceus</i> (Cham.) G. Don.	F. M. Souza 104 (UEC)	Brazil	KP014159	KP014907	KP014481	KP014645	-	-	-
<i>Siphocampylus correoides</i> Zahlb.	A. Timmermann 3 (AAU)	Bolivia	KP014298	KP014967	KP014436	KP014657	-	-	-
<i>Siphocampylus corymbifer</i> Pohl	A. Timmermann 9 (AAU)	Bolivia	KP014155	KP014941	KP014347	-	-	-	-
<i>Siphocampylus corymbifer</i> Pohl	A. Glaziou s.n.	Bolivia	KP014156	KP014940	KP014348	-	-	-	-
<i>Siphocampylus corymbifer</i> Pohl	L. Lagomarsino 216 (GH)	Bolivia	KP014157	KP014942	KP014346	KP014693	KU670796	-	-
<i>Siphocampylus corynoides</i> E. Wimm	L. Lagomarsino 353 (GH)	Peru	KP014117	KP014910	KP014361	KP014610	-	-	-
<i>Siphocampylus dependens</i> (Ruiz & Pavon) G. Don	L. Lagomarsino 316 (GH)	Peru	KP014261	KP015000	KP014485	KP014690	-	-	-
<i>Siphocampylus elfriedii</i> E. Wimm	L. Lagomarsino 387 (GH)	Peru	KP014084	KP014813	KP014371	KP014641	KU670793	-	-
<i>Siphocampylus fiebrigii</i> E. Wimm	S. Beck 31446 (LPB)	Bolivia	KP014100	KP014900	KP014382	KP014599	-	-	-
<i>Siphocampylus flagelliformis</i> Zahlb.	E. Garcia 2006 (GB)	Bolivia	KP014096	KP014883	KP014378	KP014612	-	-	-
<i>Siphocampylus flagelliformis</i> Zahlb.	L. Lagomarsino 220 (GH)	Bolivia	KP014099	KP014888	KP014380	KP014632	KU670794	-	-
<i>Siphocampylus fulgens</i> Hort.	Hatschbach 44995 (GB)	Ecuador	KP014081	KP015014	KP014434	KP014719	EF141032	DQ356216	-
<i>Siphocampylus giganteus</i> (Cav.) G. Don.	L. Andersson 2432 (GB)	Ecuador	-	KP015002	KP014533	KP014761	DQ356154	DQ356222	DQ356103

<i>Siphocampylus giganteus</i> (Cav.) G. Don.	N. Muchhal 101	Ecuador	KP014205	KP015001	KP014534	KU670699	EF174621	-	-
<i>Siphocampylus imbricatus</i> (Cham.) G. Don.	P. Nuñez 8182 (AAU)	Brazil	-	KU670679	KP014329	KU670698	-	-	-
<i>Siphocampylus jelskii</i> Zahlb.	L. Lagomarsino 321 (GH)	Peru	KP014199	KP014956	KP014567	KP014764	KU670795	-	-
<i>Siphocampylus krauseanus</i> E. Wimm.	L. Lagomarsino 418	Peru	-	KU670678	KU670736	KU670713	KU670797	-	-
<i>Siphocampylus longipedunculatus</i> Pohl	A. Antonelli 299 (GB)	Brazil	KP014238	KP014905	KP014471	KP014697	-	-	-
<i>Siphocampylus lycoides</i> (Cham.) G. Don.	R. Romero 3805 (UEC)	Brazil	KP014247	KP014808	KP014352	KP014710	KU670798	-	-
<i>Siphocampylus macropodus</i> (Billb.) G. Don.	A. Antonelli 298 (GB)	Brazil	KP014107	KP014890	KP014365	KP014639	DQ356153	DQ356221	DQ356102
<i>Siphocampylus matthiae</i> A. DC.	L. Lagomarsino 313 (GH)	Peru	KP014190	KP014968	KP014536	KP014765	KU670807	-	-
<i>Siphocampylus membraceus</i> Britton	A. Gentry 44,526 (MO)	Bolivia	KP014094	KP014885	KP014388	-	-	-	-
<i>Siphocampylus nematosepalus</i> (Donn. Smith) E. Wimm	L. Lagomarsino 164 (GH)	Costa Rica	KP014300	KP014952	KP014463	KP014741	KU670800	-	-
<i>Siphocampylus nemoralis</i> Griseb.	D. Spooner 6605	Bolivia	KP014112	KP014944	KP014426	-	-	-	-
<i>Siphocampylus neurotrichus</i> E. Wimm	M. Nee 42251 (NY)	Bolivia	KP014108	KP014889	KP014381	KP014603	-	-	-
<i>Siphocampylus nitidus</i> Pohl	J. Pirani 689 (UEC)	Brazil	-	-	KP014349	-	-	-	-
<i>Siphocampylus oblongifolius</i> Rusby	J. L. Clark 6687 (OSH)	Bolivia	KP014093	KP014884	KP014358	KP014636	-	-	-
<i>Siphocampylus oblongifolius</i> Rusby	S. Beck 30201 (LPB)	Bolivia	KP014095	KP014886	KP014389	-	-	-	-
<i>Siphocampylus obovatus</i> (G. Don) E. Wimm	L. Lagomarsino 350 (GH)	Peru	KP014116	-	KP014360	KP014609	-	-	-
<i>Siphocampylus orbignianus</i> A. DC.	L. Lagomarsino 217 (GH)	Bolivia	KP014152	KP014807	KP014422	KP014655	-	-	-
<i>Siphocampylus puberulus</i> E. Wimm	Vargas 226 (LPB)	Bolivia	KP014109	KP014964	KP014427	-	-	-	-
<i>Siphocampylus pubescens</i> Benth.	B. Ollgaard 74216 (GH)	Ecuador	KP014080	-	KP014419	-	-	-	-
<i>Siphocampylus</i> aff. <i>rictus</i> Lammers	L. Lagomarsino 341 (GH)	Peru	KP014082	KP014811	KP014363	KP014640	-	-	-

<i>Siphocampylus rosmarinifolius</i> G. Don.	L. Lagomarsino 331 (GH)	Peru	KP014113	KP014909	KP014362	KP014634	KU670801	-	-
<i>Siphocampylus rusbyanus</i> Britton	A. Fuentes 4760 (MO)	Bolivia	KP014264	KP015012	KP014431	KP014689	KU670802	-	-
<i>Siphocampylus scandens</i> (HBK) G. Don.	L. Andersson 2515 (GB)	Ecuador	KP014248	KP014973	KP014356	KP014592	DQ356150	DQ356218	DQ356099
<i>Siphocampylus sceptrum</i> Decaisne apud Linden	L. Dorr 20513	Venezuela	KP014242	KP015009	KP014392	KP014597	-	-	-
<i>Siphocampylus smilax</i> Lammers	L. Lagomarsino 250 (GH)	Bolivia	KP014079	KP014908	KP014342	KP014649	KU670803	-	-
<i>Siphocampylus sparsipilus</i> E. Wimm	L. Lagomarsino 197 (GH)	Bolivia	KP014278	KP014939	KP014492	KP014652	KU670804	-	-
<i>Siphocampylus tunarensis</i> Zahlb.	L. Lagomarsino 232 (GH)	Bolivia	KP014271	KP014975	KP014435	KP014674	KU670752	-	-
<i>Siphocampylus tunicatus</i> Zahlb.	L. Lagomarsino 235 (GH)	Bolivia	KP014272	KP014991	KP014443	KP014676	KU670754	-	-
<i>Siphocampylus tupaeformis</i> Zahlb.	A. Timmermann 7 (LPB)	Bolivia	KP014111	KP014962	KP014429	KP014708	-	-	-
<i>Siphocampylus umbellatus</i> (HBK) G. Don.	L. Lagomarsino 193 (GH)	Bolivia	KP014275	KP014994	KP014442	KP014672	-	-	-
<i>Siphocampylus vatkeanus</i> Zahlb.	N. Paniagua 5774 (MO)	Bolivia	KP014088	KP014816	KP014369	KP014608	KU670753	-	-
<i>Siphocampylus veteranus</i> Wimm	L. Lagomarsino 388 (GH)	Peru	KP014087	KP014815	KP014373	KP014644	KU670781	-	-
<i>Siphocampylus virgatus</i> A. DC.	L. Lagomarsino 343 (GH)	Peru	KP014091	KP014958	KP014357	KP014607	KU670755	-	-
<i>Siphocampylus werdermannii</i> E. Wimm	L. Lagomarsino 245 (GH)	Bolivia	KP014106	KP014893	KP014376	KP014602	-	-	-
<i>Siphocampylus westinianus</i> (Billb.) Pohl	A. Antonelli 308 (GB)	Brazil	KP014154	KP014906	KP014472	KP014587	-	-	-
<i>Siphocampylus</i> sp.	L. Lagomarsino 333 (GH)	Peru	KP014262	KP015010	KP014486	KP014691	-	-	-
<i>Siphocampylus</i> sp. nov. 1 (Lagom. & Santam. ined.)	L. Lagomarsino 241 (GH)	Bolivia	KP014274	KP014990	KP014440	KP014675	-	-	-
<i>Siphocampylus</i> sp. nov. 2 (Lagom. & Santam. ined.)	L. Lagomarsino 400 (GH)	Peru	KP014085	KP014814	KP014372	KP014642	-	-	-

OUTGROUP

<i>Diastatea micrantha</i> (Kunth) McVaugh	L. Lagomarsino 44; Lojtnant and Molau 15184	Costa Rica	KP014066	KP014803	KP014327	KP014581	DQ356138	DQ356203	DQ356095
<i>Downingia</i>									
<i>Downingia bacigalupii</i> Weiler	Mason 14384(S)	California	-	-	-	-	EF141031	DQ356183	-
<i>Downingia insignis</i> Greene	Tiehm 12143 (S)	California	-	-	-	-	EF141030	DQ356185	-
<i>Grammatotheca</i>									
<i>Grammatotheca bergiana</i> (Cham.) C. Presl	Bean & Viviers 2628 (GB)	South Africa	-	-	-	-	DQ356116	DQ356168	DQ356088
<i>Hippobroma</i>									
<i>Hippobroma longiflora</i> (L.) G. Don	L. Lagomarsino 346; Andersson and Nilsson 2492 (GB)	Peru	KP014072	-	KP014337	KP014782	KU670778	DQ356206	DQ356096
<i>Isotoma</i>									
<i>Isotoma fluviatilis</i> F. Muell. Ex Bentham	Cultivated, University of Aarhus, no voucher	Clutivated	KP014054	KP014790	KP014321	KP014783	DQ356161	DQ356230	DQ356108
<i>Jasione</i>									
<i>Jasione montana</i> L.	F.I.Sales & C.Hedge (EGHB); Andersson 2562 (GB)	9898 unknown	-	-	-	-	EU713354	DQ356174	-
<i>Legousia</i>									
<i>Legousia hybrid</i> Delarb.	Corneliuson s.n.; (GB 152920)	Sweden	-	-	-	-	DQ356163	DQ356234	DQ356111
<i>Lobelia</i>									
<i>Lobelia aguana</i> E. Wimm.	Koch 82120 (NY)	Mexico	-	-	-	-	DQ356122	DQ356176	-
<i>Lobelia alsinoides</i> Lam.	Larsen 34131 (AAU)	Thailand	KP014050	KP014785	KP014309	KP014572	-	-	-
<i>Lobelia anceps</i> L.	Skottsberg 4536 (GB)	Chile	-	-	-	-	DQ356124	DQ356184	-
<i>Lobelia aquatica</i> Cham.	Jacobs et al. 4080(GB)	Guyana	-	-	-	-	EF141029	DQ356182	-
<i>Lobelia assurgens</i> var. <i>portoricensis</i> Urb.	Trejo 28 (GH)	Puerto Rico	-	-	KU670730	-	-	-	-
<i>Lobelia cardinalis</i> L.	L. Lagomarsino 423 (GH); T. Lammers 8710 (F)	USA	KP014060	KP014794	KP014314	KU670717	AY655144	DQ356231	AF130187 (Vincent 4377 (GB))

<i>Lobelia cardinalis</i> subsp. <i>graminea</i> (Lam.) McVaugh	R. Kriebel 5570 (NY)	Guatemala	KP014061	KP014795	KP014315	-	-	-	-
<i>Lobelia chinensis</i> Hance	G.Kokubugata 11397; Hashimoto 409 (GB)	Japan	-	-	-	-	AB645974	DQ356228	-
<i>Lobelia cliffortiana</i> L.	R. A. Howard 15704 (NY)	Puerto Rico	-	KU670691	KU670733	-	-	-	-
<i>Lobelia cordifolia</i> Hook. & Arn.	Mexia 8780 (S)	Mexico	-	-	-	-	-	DQ356204	-
<i>Lobelia coronopifolia</i> L.	Dahlstrand 1084(GB)	South Africa	-	-	-	-	EF141025	DQ356181	-
<i>Lobelia digitalifolia</i> (Griseb.) Urb.	R. A. Howard 19407 (NY)	Guadeloupe	-	-	KP014324	-	-	-	-
<i>Lobelia dortmanna</i> L.	Andersson 2561 (GB)	Sweden	-	-	-	-	DQ356162	DQ356232	DQ356109
<i>Lobelia erinus</i> L.	Andersson 2570 (GB)	Cultivated	-	-	-	-	HM850129	DQ356233	DQ356110
<i>Lobelia exaltata</i> Pohl	Antonelli 335 (GB)	Brazil	-	-	-	-	DQ356135	DQ356200	DQ356093
<i>Lobelia fistulosa</i> Raf.	Antonelli & Andersson 279 (GB)	Brazil	-	-	-	-	DQ356136	DQ356201	DQ356094
<i>Lobelia gibberoa</i> Hemsl.	Jaasund s.n.; (GB 1.1.67)	Kenya	-	-	-	-	DQ356127	DQ356192	-
<i>Lobelia graniticola</i> E. Wimm.	Thulin & Mhoro 3210 (C)	Tanzania	-	-	-	-	DQ356129	DQ356194	-
<i>Lobelia gregoriana</i> Baker f.	Hedberg 1608 (S)	Kenya	-	-	-	-	DQ356187	DQ356187	-
<i>Lobelia hederacea</i> Cham.	A. Schinini 12141 (AAU)	Argentina	KP014053	KP014797	KP014323	KP014578	-	-	-
<i>Lobelia hypoleuca</i> Hillebr.	Selling & Skottsberg 3194 (GB)	Hawaii	-	-	-	-	DQ356126	DQ356191	-
<i>Lobelia inflata</i> L.	H. Schmidt 1531 (MO)	USA	KP014062	-	KP014320	-	-	-	-
<i>Lobelia irasuensis</i> Planh. & Oerst.	L. Lagomarsino 5 (GH); Burger & Burger 8151 (GB)	Costa Rica	-	KU670690	KU670720	-	KU670750	DQ356175	-
<i>Lobelia kalmii</i> L.	L. Lagomarsino 424 (GH); D. Boufford 7292	USA	KP014059	KP014793	KP014316	KP014577	DQ356166 (Boufford 7292)	EF126736	DQ356114
<i>Lobelia kraussii</i> Graham	Hill 25677(NY)	Dominica	-	-	-	-	EF141024	DQ356179	-

<i>Lobelia laxiflora</i> Kunth.	L. Lagomarsino 289 (GH); D. Santamaría S-985	Panama	KP014065	KP014799	KP014326	KP014777	DQ356143	DQ356209	-
<i>Lobelia leschenaultiana</i> Skottsb.	Skottsberg s.n.; (GB) 11.XII.1926)	Sri Lanka	-	-	-	-	DQ356131	DQ356196	-
<i>Lobelia martagon</i> Hitchc.	W. Judd 5375 (NY)	Jamaica	KP014073	-	KP014335	-	KU670806	DQ356205	-
<i>Lobelia nana</i> Kunth.	L. Lagomarsino 228 (GH)	Bolivia	KP014063	KP014796	KP014328	KP014775	-	-	-
<i>Lobelia oligophylla</i> (Wedd.) Lammers	C. M. Taylor 11210 (NY); (Harling et al. 6807)	Argentina	KP014055	-	KP014322	KP014784	KU670787	DQ356227	-
<i>Lobelia persicifolia</i> Cav.	Whitefand 3782 (GH)	Guadeloupe	-	-	KU670731	-	-	-	-
<i>Lobelia physaloides</i> A. Cunn.	Cameron s.n. (AAU)	New Zealand	-	-	KP014311	KP014574	-	-	-
<i>Lobelia portoricensis</i> (Vatke) Urb.	Boom 10069 (NY)	Puerto Rico	-	-	-	-	DQ356142	DQ356208	-
<i>Lobelia roughii</i> Hook f.	Skottsberg s.n. (GB)	New Zealand	-	-	-	-	DQ356165	EF126737	DQ356113
<i>Lobelia sonderiana</i> (Kuntze) Lammers	Volk 1095 (S)	Namibia	-	-	-	-	DQ356130	DQ356195	-
<i>Lobelia stricklandiae</i> Gilliland	Fisher & Thweicher 291 (S)	Zimbabwe	-	-	-	-	-	DQ356186	-
<i>Lobelia stricta</i> M. Martens & Galeotti	Wilbur et al. 7849 (NY)	Dominica	-	-	KU670738	-	DQ356141	DQ356207	-
<i>Lobelia tomentosa</i> L. f.	Dahlstrand 2600(GB)	South Africa	-	-	-	-	EF141028	DQ356180	-
<i>Lobelia viridiflora</i> McVaugh	Weaver 910 (GH)	Jamaica	-	-	KU670735	-	-	-	-
<i>Lobelia xalapensis</i> Kunth	Santamaría S-1009 (GB)	Costa Rica	-	-	-	-	DQ356144	DQ356210	-
<i>Lobelia yuccoides</i> Hillebr.	Cranwell et al. 2909 (GB)	Hawaii	-	-	-	-	DQ356125	DQ356190	-
<i>Lobelia zeylanica</i> L.	D. Boufford 24898 (GH)	Taiwan	-	-	KP014310	KP014573	-	-	-
Pratia									
<i>Pratia nummularia</i> (Lam.) A. Braun & Asch.	Andersson 2571 (GB)	Cultivated	-	-	-	-	DQ356164	DQ356235	DQ356112
<i>Pratia purpurascens</i> E. Wimm.	Egerod s.n. (GB)	Australia	-	-	-	-	DQ356160	DQ356229	DQ356107

Siphocampylus* section *Hemisiphocampylus

<i>Siphocampylus decumbens</i> (Rich.) Juss & A. DC.	P. Acevedo-Rodriguez 14172 (NY)	Dominican Republic	KP014067	-	KP014330	-	-	-	-
<i>Siphocampylus domingensis</i> A. DC.	A. Veloz 2001 (JBSD)	Dominican Republic	KP014068	KP014787	KP014331	KP014582	-	-	-
<i>Siphocampylus igneus</i> Urb.	J. R. Abbott 21003 (FLAS)	Dominican Republic	KP014069	KP014788	KP014332	KP014779	-	-	-
<i>Siphocampylus manettiaeflorus</i> Hook.	Cultivated, Marbury Garden, no voucher	Bot.	Cultivated	KP014074	KP014786	KP014334	KP014780	KU670799	-
<i>Siphocampylus sonchifolius</i> (Sw.) McVaugh	T. Clase 3048 (JBSD)	Dominican Republic	KP014070	KP014789	KP014333	KP014778	-	-	-
<i>Solenopsis</i>									
<i>Solenopsis laurentia</i> (L.) C. Presl.	Dept.Syst.Bot. 381 (GB)	Cultivated	-	-	-	-	DQ356134	DQ356199	-
<i>Trematolobelia</i>									
<i>Trematolobelia macrostachys</i> (Hook. & Arn.) Zahlbr.	Fagerlind 6872 (S)	Hawaii	-	-	-	-	DQ356137	DQ356202	-
<i>Triodanis</i>									
<i>Triodanis perfoliata</i> (L.) Nieuwl.	L. Lagomarsino 425 (GH)	USA	KP014049	-	KP014308	-	-	-	-

Herbarium acronyms follow the Index Herbariorum. Taxa are organized alphabetically within the ingroup and outgroup categories

Table S2. Divergence time estimation to assess sensitivity to dating method, calibration selection, and branch length heterogeneity.

	Campanulaceae	Campanuloideae	Lobelioideae	Neotropical Lobelioideae	<i>Lobelia</i> section <i>Tupa</i>	<i>Lysipomia</i>	centropogonids
treePL dates from RAxML bootstrap trees							
All priors	58.19 (55.89–61.62)	56.13 (55.87–57.0)	54.98 (51.71–56.00)	20.15 (18.06–22.04)	1.07 (0.36–1.35)	13.05 (11.13–14.36)	5.12 (4.24–5.83)
Three priors (fossil and secondary ages)	57.15 (55.90–61.76)	55.93 (55.87–56.0)	55.03 (51.72–56.0)	20.24 (18.09–22.04)	0.86 (0.34–1.33)	13.08 (11.13–14.37)	5.32 (4.22–5.86)
Two priors (fossil and root age)	66.89 (66.88–67.0)	65.41 (60.64–66.95)	64.65 (56.13–66.94)	24.73 (20.74–25.40)	1.01 (0.15–1.20)	15.21 (13.01–16.38)	6.22 (4.76–6.67)
One prior (root age only)	66.69 (66.88–67.0)	65.34 (60.65–66.96)	63.66 (56.13–66.93)	23.58 (21.19–25.25)	1.01 (0.147–1.20)	15.3 (12.85–16.38)	6.24 (4.76–6.68)
All priors, <i>Lysipomia</i> removed	60.17 (55.98–62.09)	56 (55.87–56.0)	53.42 (50.96–56.12)	17.33 (13.97–18.31)	0.34 (0.31–1.31)	N/A	4.62 (4.14–5.45)
BEAST dates, all with expanded outgroup							
All priors, exponential relaxed clock	57.1 (44–69.55)	46.57 (36.28–56.39)	49.8 (35.88–65.20)	16.26 (9.44–22.51)	0.9 (0.18–2.63)	8.2 (4.70–13.06)	5.94 (3.32–9.54)
All priors, lognormal relaxed clock*	58.78 (49.87–68.04)	53.48 (45.60–62.11)	46.79 (39.07–55.07)	17.74 (14.32–20.32)	1.35 (0.42–2.78)	11.69 (9.00–15.24)	4.98 (3.95–6.13)
Three priors (fossil and secondary ages), lognormal relaxed clock	59.2 (46.81–72.03)	48.55 (38.38–59.31)	51.78 (37.87–66.55)	16.07 (10.54–22.32)	0.95 (0.22–2.75)	8.97 (5.08–13.83)	6.55 (3.74–9.95)
Two priors (fossil and root age), lognormal relaxed clock	62.47 (48.31–76.38)	53.23 (39.67–68.62)	51.52 (34.62–69.17)	17.78 (9.69–24.49)	0.97 (0.30–2.90)	9.24 (5.03–13.89)	6.71 (3.87–10.46)
One prior (root age only), lognormal relaxed clock	53.4 (35.61–69.51)	20.33 (9.69–34.34)	43.62 (19.68–57.10)	12.11 (6.27–18.80)	0.68 (0.10–2.17)	6.37 (2.84–9.89)	4.77 (2.25–7.67)
All priors, <i>Lysipomia</i> removed	58.77 (47.0–71.94)	49.16 (37.82–59.07)	50.05 (35.34–66.99)	11.8 (6.93–17.63)	0.85 (0.15–0.86)	N/A	6.11 (3.78–9.47)

Analysis in boldface denoted with an asterisk is the one presented in the main text and used in subsequent diversification analyses.

Age estimates reported for major clades relevant to the study reported in million years (Ma). Root node = crown group Campanulaceae.

Table S3. Trait codings for the four traits used in our BiSSE analyses.

	Andean Distribution	Elevation	Fruit	Pollinator
<i>Burmeistera almedae</i> Wilbur	0	1	1	1
<i>Burmeistera aff. bullatifolia</i> J. Garzón & F. Gonzalez	1	1	1	1
<i>Burmeistera aff. bullatifolia</i> J. Garzón & F. Gonzalez	1	1	1	1
<i>Burmeistera ceratocarpa</i> Zahlb.	1	0	1	1
<i>Burmeistera chiriquiensis</i> Wilbur	0	1	1	1
<i>Burmeistera crebra</i> McVaugh	0	1	1	1
<i>Burmeistera cyclostigmata</i> J. D. Smith	0	1	1	1
<i>Burmeistera dendrophila</i> E. Wimm	0	1	1	1
<i>Burmeistera domingensis</i> Jeppesen	1	0	1	1
<i>Burmeistera fuchsoides</i> J. Garzón & F. Gonzalez	1	1	1	1
<i>Burmeistera mcvaughii</i> Wilbur	0	1	1	1
<i>Burmeistera mcvaughii</i> Wilbur	0	1	1	1
<i>Burmeistera microphylla</i> J. D. Smith	0	0	1	1
<i>Burmeistera aff. minutiflora</i> J. Garzón & F. Gonzalez	1	1	1	0
<i>Burmeistera morii</i> Wilbur	0	0	1	1
<i>Burmeistera multiflora</i> Zahlb.	1	0	1	1
<i>Burmeistera obtusifolia</i> E. Wimm	0	1	1	1
<i>Burmeistera obtusifolia</i> E. Wimm	0	1	1	0
<i>Burmeistera racemiflora</i> Lammers	1	0	1	1
<i>Burmeistera refracta</i> E. Wimm.	1	0	1	1
<i>Burmeistera refracta</i> E. Wimm.	1	0	1	1
<i>Burmeistera sodiroana</i> Zahlb.	1	1	1	1
<i>Burmeistera succulenta</i> H. Karst. & Triana	1	0	1	1
<i>Burmeistera tenuiflora</i> J.D. Smith	0	0	1	1
<i>Burmeistera toroensis</i> Wilbur	0	0	1	1
<i>Burmeistera truncata</i> Zahlbr.	1	0	1	1
<i>Burmeistera utleyi</i> Wilbur	0	1	1	1
<i>Burmeistera variabilis</i> (Gleason) E. Wimm.	1	0	1	1
<i>Burmeistera vulgaris</i> E. Wimm.	0	0	1	1
<i>Burmeistera zurquiensis</i> Wilbur	0	1	1	1
<i>Burmeistera</i> sp.	1	1	1	1
<i>Burmeistera</i> sp. nov.	0	1	1	0

<i>Centropogon aequatorialis</i> E. Wimm.	1	1	1	1
<i>Centropogon argutus</i> E. Wimm.	1	1	1	1
<i>Centropogon asclepiadeus</i> (Willd.) E. Wimm.	1	1	1	1
<i>Centropogon azuayensis</i> Jeppesen	1	1	1	1
<i>Centropogon baezanus</i> Jeppesen	1	1	1	1
<i>Centropogon bangii</i> Zahlb.	1	0	1	1
<i>Centropogon brittonianus</i> Zahlb.	1	1	1	1
<i>Centropogon coccineus</i> (Hook.) Regel ex B.D. Jacks.	0	0	1	1
<i>Centropogon comosus</i> Gleason	1	1	1	1
<i>Centropogon congestus</i> Gleason	0	0	1	1
<i>Centropogon cordifolius</i> Benth.	0	0	1	1
<i>Centropogon cornutus</i> (L.) Druce	0	0	1	1
<i>Centropogon costaricae</i> (Vatke) E. Wimm.	0	1	1	1
<i>Centropogon dianae</i> Lammers	1	1	1	1
<i>Centropogon dissectus</i> E. Wimm.	1	1	1	1
<i>Centropogon dombeyanus</i> E. Wimm	1	1	1	1
<i>Centropogon eilersii</i> Lammers & M.O. Dillon	1	1	1	1
<i>Centropogon erythraeus</i> Drake	1	1	1	1
<i>Centropogon featherstonei</i> Gleason	1	1	1	1
<i>Centropogon ferrugineus</i> (L.f) Gleason	0	1	1	1
<i>Centropogon ferrugineus</i> (L.f) Gleason	0	1	1	1
<i>Centropogon ferrugineus</i> (L.f) Gleason	0	1	1	1
<i>Centropogon foetidus</i> (Kunth) E. Wimm.	1	1	1	1
<i>Centropogon gamosepalus</i> Zahlb.	1	0	1	1
<i>Centropogon glabrifilis</i> (E. Wimm.) Jeppesen	1	1	1	1
<i>Centropogon gloriosus</i> (Britton) Zahlb.	1	1	1	1
<i>Centropogon grandidentatus</i> (Schltdl.) Zahlbr.	0	0	1	1
<i>Centropogon granulosus</i> subsp. <i>granulosus</i> Stein (ined.) 1	1	0	1	1
<i>Centropogon granulosus</i> subsp. <i>granulosus</i> Stein (ined.) 2	1	0	1	1
<i>Centropogon granulosus</i> subsp. <i>lugens</i> Stein (ined)	1	0	1	1
<i>Centropogon granulosus</i> subsp. <i>nutans</i> Stein (ined.)	0	0	1	1
<i>Centropogon granulosus</i> subsp. <i>tortilis</i> Stein (ined.)	0	0	1	1
<i>Centropogon granulosus</i> C. Presl. 1	1	0	1	1
<i>Centropogon granulosus</i> C. Presl. 2	0	0	1	1

<i>Centropogon granulosus</i> C. Presl. 3	0	0	1	1
<i>Centropogon granulosus</i> C. Presl. 4	0	0	1	1
<i>Centropogon gutierrezii</i> (Planch & Oerst.) E. Wimm.	0	1	1	1
<i>Centropogon hirtus</i> (Cav.) C. Presl.	1	1	1	1
<i>Centropogon incanus</i> (Britton) Zahlb.	1	1	1	1
<i>Centropogon incanus</i> (Britton) Zahlbr..	1	1	1	1
<i>Centropogon isabellinus</i> E. Wimm.	1	1	1	1
<i>Centropogon leucocarpus</i> McVaugh	0	1	1	1
<i>Centropogon llanganatensis</i> Jeppsen	1	1	1	1
<i>Centropogon luteus</i> E. Wimm.	1	1	1	1
<i>Centropogon luteynii</i> Wilbur	0	1	1	1
<i>Centropogon macbridei</i> Gleason	1	1	1	1
<i>Centropogon macrocarpus</i> Zahlb.	1	1	1	1
<i>Centropogon macrophyllus</i> (G. Don.) E. Wimm	1	0	1	1
<i>Centropogon magnificus</i> Zahlb.	1	1	1	1
<i>Centropogon mandonis</i> Zahlb.	1	1	1	1
<i>Centropogon mandonis</i> Zahlb.	1	1	1	1
<i>Centropogon medusa</i> E. Wimm.	1	1	1	1
<i>Centropogon minimus</i> McVaugh	1	1	1	1
<i>Centropogon nervosus</i> Wimm	1	1	1	1
<i>Centropogon nigricans</i> Zahlb.	1	1	1	1
<i>Centropogon nigricans</i> Zahlb.	1	1	1	1
<i>Centropogon palmanus</i> (J. D. Smith) E. Wimm.	0	0	1	1
<i>Centropogon panamensis</i> Wilbur	0	0	1	1
<i>Centropogon perlóngus</i> Gleason	1	1	1	1
<i>Centropogon peruvianus</i> (E. Wimm.) McVaugh	1	1	1	1
<i>Centropogon pichinchensis</i> Zahlb.	1	1	1	1
<i>Centropogon preslii</i> E. Wimm.	1	1	1	1
<i>Centropogon pulcher</i> Zahlb.	1	0	1	1
<i>Centropogon reticulatus</i> Drake	1	1	1	1
<i>Centropogon rex</i> E. Wimm.	1	1	1	1
<i>Centropogon roseus</i> Rusby	1	0	1	1
<i>Centropogon salviaeformis</i> Zahlb.	1	1	1	1
<i>Centropogon simulans</i> Lammers	1	1	1	1
<i>Centropogon smithii</i> E. Wimm.	0	1	1	1
<i>Centropogon sodiroanus</i> Zahlb.	1	1	1	1

<i>Centropogon solanifolius</i> Benth. 1	1	1	1	1
<i>Centropogon solanifolius</i> Benth. 2	0	1	1	1
<i>Centropogon solanifolius</i> Benth. 3	0	1	1	1
<i>Centropogon subandinus</i> Zahlb.	1	1	1	1
<i>Centropogon talamancensis</i> Wilbur	0	1	1	1
<i>Centropogon tessmannii</i> E. Wimm.	1	0	1	1
<i>Centropogon trachyanthus</i> E. Wimm.	1	1	1	1
<i>Centropogon trichodes</i> E. Wimm.	1	1	1	1
<i>Centropogon umbrosus</i> E. Wimm	1	1	1	1
<i>Centropogon unduavensis</i> (Britton) Zahlb.	1	1	1	1
<i>Centropogon urubambae</i> E. Wimm	1	0	1	1
<i>Centropogon valerii</i> Standley	0	1	1	1
<i>Centropogon vargasii</i> B. A. Stein (ined.)	1	1	1	1
<i>Centropogon viriduliflorus</i> Wimm	1	1	1	1
<i>Centropogon viriduliflorus</i> Wimm	1	1	1	1
<i>Centropogon weberbaueri</i> Zahlb.	1	1	1	1
<i>Centropogon cf. weberbaueri</i> Zahlb.	1	1	1	1
<i>Centropogon yarumalensis</i> E. Wimm.	1	1	1	1
<i>Centropogon yungasensis</i> Britton	1	1	1	1
<i>Centropogon</i> sp. nov. (Lagom. & Santam. ined.)	1	1	1	1
<i>Centropogon</i> sp.	1	1	1	1
<i>Lobelia bridgesii</i> Hook & Arn.	1	0	0	1
<i>Lobelia excelsa</i> Bonpland	1	0	0	1
<i>Lobelia polyphylla</i> Hook. & Arn.	1	0	0	1
<i>Lobelia tupa</i> L.	1	0	0	1
<i>Lysipomia cuspidata</i> McVaugh	1	1	0	0
<i>Lysipomia muscoides</i> Hook f.	1	1	0	0
<i>Lysipomia pumila</i> (Wedd.) E. Wimm.	1	1	0	0
<i>Lysipomia sphagnophila</i> Griseb. Ex Wedd.	1	1	0	0
<i>Lysipomia vitreola</i> McVaugh	1	1	0	0
<i>Siphocampylus actinothrix</i> E. Wimm.	1	1	0	1
<i>Siphocampylus</i> aff. <i>actinothrix</i> E. Wimm.	1	1	0	1
<i>Siphocampylus affinis</i> (Mirb.) McVaugh	1	1	0	1
<i>Siphocampylus ambivalens</i> Lammers	1	1	0	1
<i>Siphocampylus andinus</i> Britton	1	1	0	1
<i>Siphocampylus angustiflorus</i> Schlechtend.	1	1	0	1

<i>Siphocampylus antioquianus</i> E. Wimm.	1	1	0	1
<i>Siphocampylus aureus</i> Rusby	1	0	0	1
<i>Siphocampylus ayersiae</i> Lammers	1	1	0	1
<i>Siphocampylus betulaefolius</i> (Cham.) G. Don.	0	0	0	1
<i>Siphocampylus bilabiatus</i> Zahlb.	1	0	0	1
<i>Siphocampylus boliviensis</i> Zahlb.	1	1	0	1
<i>Siphocampylus brevicalyx</i> E. Wimm.	1	1	0	1
<i>Siphocampylus chloroleucus</i> E. Wimm	1	1	0	1
<i>Siphocampylus citrinus</i> E. Wimm	1	1	0	1
<i>Siphocampylus clotho</i> E. Wimm.	1	1	0	1
<i>Siphocampylus convolvulaceus</i> (Cham.) G. Don.	0	0	0	1
<i>Siphocampylus correoides</i> Zahlb.	1	1	0	1
<i>Siphocampylus corymbifer</i> Pohl	1	0	0	1
<i>Siphocampylus corymbifer</i> Pohl	1	0	0	1
<i>Siphocampylus corynoides</i> E. Wimm	1	1	0	1
<i>Siphocampylus dependens</i> (Ruiz & Pavon) G. Don	1	1	0	1
<i>Siphocampylus elfriedii</i> E. Wimm	1	1	0	1
<i>Siphocampylus fiebrigii</i> E. Wimm	1	0	0	1
<i>Siphocampylus flagelliformis</i> Zahlb.	1	1	0	1
<i>Siphocampylus flagelliformis</i> Zahlb.	1	1	0	1
<i>Siphocampylus fulgens</i> Hort.	0	0	0	1
<i>Siphocampylus giganteus</i> (Cav.) G. Don.	1	1	0	1
<i>Siphocampylus giganteus</i> (Cav.) G. Don.	1	1	0	1
<i>Siphocampylus imbricatus</i> (Cham.) G. Don.	0	0	0	1
<i>Siphocampylus jelskii</i> Zahlb.	1	1	0	1
<i>Siphocampylus krauseanus</i> E. Wimm.	1	1	0	1
<i>Siphocampylus longipedunculatus</i> Pohl	0	0	0	1
<i>Siphocampylus lycioides</i> (Cham.) G. Don.	0	0	0	1
<i>Siphocampylus macropodus</i> (Billb.) G. Don.	0	0	0	1
<i>Siphocampylus matthiae</i> A. DC.	1	1	0	1
<i>Siphocampylus membraceus</i> Britton	1	0	0	1
<i>Siphocampylus nematosepalus</i> (Donn. Smith) E. Wimm	0	1	0	1
<i>Siphocampylus nemoralis</i> Griseb.	1	1	0	1
<i>Siphocampylus neurotrichus</i> E. Wimm	1	0	0	1

<i>Siphocampylus nitidus</i> Pohl	0	0	0	1
<i>Siphocampylus oblongifolius</i> Rusby	1	0	0	1
<i>Siphocampylus oblongifolius</i> Rusby	1	0	0	1
<i>Siphocampylus obovatus</i> (G. Don) E. Wimm	1	1	0	1
<i>Siphocampylus orbignianus</i> A. DC.	1	0	0	1
<i>Siphocampylus puberulus</i> E. Wimm	1	1	0	1
<i>Siphocampylus pubescens</i> Benth.	1	1	0	1
<i>Siphocampylus aff. rictus</i> Lammers	1	1	0	1
<i>Siphocampylus rosmarinifolius</i> G. Don.	1	1	0	1
<i>Siphocampylus rusbyanus</i> Britton	1	1	0	1
<i>Siphocampylus scandens</i> (HBK) G. Don.	1	1	0	1
<i>Siphocampylus sceptrum</i> Decaisne apud Linden	1	1	0	1
<i>Siphocampylus smilax</i> Lammers	1	0	0	1
<i>Siphocampylus sparsipilus</i> E. Wimm	1	1	0	1
<i>Siphocampylus tunarensis</i> Zahlb.	1	1	0	1
<i>Siphocampylus tunicatus</i> Zahlb.	1	1	0	1
<i>Siphocampylus tupaeformis</i> Zahlb.	1	1	0	1
<i>Siphocampylus umbellatus</i> (HBK) G. Don.	1	1	0	1
<i>Siphocampylus vatkeanus</i> Zahlb.	1	1	0	1
<i>Siphocampylus veteranus</i> Wimm	1	1	0	1
<i>Siphocampylus virgatus</i> A. DC.	1	0	0	1
<i>Siphocampylus werdermannii</i> E. Wimm	1	1	0	1
<i>Siphocampylus westinianus</i> (Billb.) Pohl	0	0	0	1
<i>Siphocampylus</i> sp.	1	1	0	1
<i>Siphocampylus</i> sp. nov. 1 (Lagom. & Santam. ined.)	1	1	0	1
<i>Siphocampylus</i> sp. nov. 2 (Lagom. & Santam. ined.)	1	1	0	1

Character States are as follows: Andean Distribution— 0= extra-Andean, 1= Andean; Elevation— 0= Low elevation (<1900m), 1= High elevation (>1900 m); Fruit— 0= Capsule, 1= Berry; Pollinator— 0= Invertebrate, 1= Vertebrate

Table S4. Paleoenvironmental-dependent diversification analyses using paleoaltimetry (a) and Cenozoic climate data (b).

a) Paleoaltitude-dependence diversification models

Models	Mode of dependency	NP	logL	AICc	ΔAIC	P (LRT)	λ	α	μ	β
λ constant (no μ) (Yule)	-	1	-313.082	628.184	89.408	<0.001	1.016	-	-	-
λ and μ constant	-	2	-276.699	557.458	18.682	<0.001	2.915	-		-
λ Alti. and no μ	Exponential	2	-267.358	538.776	0	Best	0.0010	0.00249	-	-
λ Alti. and μ constant	Exponential	3	-266.863	539.848	1.072	0.3197	0.0004	0.00258	0.0184	-
λ constant and μ Alti.	Exponential	3	-271.228	548.578	9.802	0.999	2.3894	-	6.1267	-0.00026
λ Alti. and μ Alti.	Exponential	4	-266.523	541.250	2.474	0.4339	0.0016	0.00213	0.0611	-0.03312
λ Alti. and no μ	Linear	2	-281.176	566.413	27.637	0.999	2.1972	-0.00077	-	-
λ Alti. and μ constant	Linear	3	-268.544	543.211	4.435	0.999	2.3254	-0.00094	0.9437	-
λ constant and μ Alti.	Linear	3	-271.499	549.120	10.344	0.999	2.1654	-	4.1010	-0.00065
λ Alti. and μ Alti.	Linear	4	-270.284	548.773	9.997	0.999	2.7205	0.00011	1.7161	-0.00013

b) Temperature-dependence diversification models

Models	Mode of dependency	NP	logL	AICc	Δ AIC	P (LRT)	λ	α	μ	β
λ constant (no μ) (Yule)	-	1	-313.082	628.184	92.14	<0.001	1.016	-	-	-
λ and μ constant	-	2	-276.699	557.458	21.414	<0.001	2.915	-		-
λ Temp. and no μ	Exponential	2	-268.613	541.286	5.242	0.999	2.7463	-0.43320	-	-
λ Temp. and μ constant	Exponential	3	-265.353	536.829	0.785	0.999	2.5085	-0.14360	1.0536	-
λ constant and μ Temp.	Exponential	3	-265.248	536.619	0.575	0.999	2.2038	-	1.2030	0.10578
λ Temp. and μ Temp.	Exponential	4	-264.955	538.114	2.070	0.9128	2.2776	-0.05523	1.0695	0.07642
λ Temp. and no μ	Linear	2	-272.381	548.824	12.779	0.999	1.6588	-0.23915	-	-
λ Temp. and μ constant	Linear	3	-265.748	537.618	1.573	0.999	2.1378	-0.26233	0.6189	-
λ constant and μ Temp.	Linear	3	-264.961	536.044	0	Best	2.1238	-	0.9730	0.20179
λ Temp. and μ Temp.	Linear	4	-264.575	537.355	1.310	0.3796	1.7996	0.28453	0.7834	0.47501

Mean parameter estimates presented for each model. Best-fitting model, as determined via a combination of the Akaike Information Criterion and likelihood ratio tests (see main text) highlighted in red and boldface. In our best-fit paleoaltitude-dependent model, speciation is positively exponentially correlated to Andean orogeny over time; adding extinction as a parameter did not improve model fit. Likewise, extinction is positively linearly correlated to temperature variation over time; allowing speciation to vary with temperature did not improve model fit. λ = speciation rate (in events/Ma/lineage); μ = extinction rate (in events/Ma/lineage); α = rate of variation of the speciation according to the relevant paleoenvironmental variable; β = rate of variation of the extinction according to the paleoenvironmental variable; NP: number of parameters in each model.

Table S5. Model comparison for the four BiSSE analyses presented in the main text.

Andes (0: extra-Andean; 1: Andean)											
Model	N P	LogL	AICc	ΔAIC	P (LRT)	λ ₀	λ ₁	μ ₀	μ ₁	q ₀₁	q ₁₀
Null model (λ ₀ =λ ₁ ; μ ₀ =μ ₁ ; q ₀₁ =q ₁₀)	3	-361.737	729.597	15.332	0.99	2.932	2.932	2.592	2.592	0.084	0.084
λ ₀ ≠λ ₁ ; μ ₀ =μ ₁ ; q ₀₁ =q ₁₀	4	-354.666	717.537	3.271	0.99	1.908	2.541	1.818	1.818	0.089	0.089
λ ₀ =λ ₁ ; μ ₀ ≠μ ₁ ; q ₀₁ =q ₁₀	4	-357.126	722.457	8.192	0.99	2.713	2.713	2.621	2.164	0.102	0.102
λ ₀ =λ ₁ ; μ ₀ =μ ₁ ; q ₀₁ ≠q ₁₀	4	-359.261	726.726	12.460	0.99	2.924	2.924	2.583	2.583	0.273	0.077
λ₀≠λ₁; μ₀≠μ₁; q₀₁=q₁₀	5	-351.978	714.266	0	Best	1.267	2.975	1.152	2.275	2.275	2.275
λ ₀ ≠λ ₁ ; μ ₀ =μ ₁ ; q ₀₁ ≠q ₁₀	5	-352.912	716.133	1.867	0.99	1.690	2.477	1.636	1.636	0.043	0.109
λ ₀ =λ ₁ ; μ ₀ ≠μ ₁ ; q ₀₁ ≠q ₁₀	5	-354.944	720.197	5.931	0.99	2.348	2.348	2.305	1.510	0.031	0.123
λ ₀ ≠λ ₁ ; μ ₀ ≠μ ₁ ; q ₀₁ ≠q ₁₀	6	-351.892	716.219	1.953	0.678	1.292	2.938	1.184	2.224	0.081	0.088
Elevation (0: low; 1: high)											
Model	N P	LogL	AICc	ΔAIC	P (LRT)	λ ₀	λ ₁	μ ₀	μ ₁	q ₀₁	q ₁₀
Null model (λ ₀ =λ ₁ ; μ ₀ =μ ₁ ; q ₀₁ =q ₁₀)	3	-370.173	746.468	16.468	0.99	2.994	2.994	2.656	2.656	0.175	0.175
λ ₀ ≠λ ₁ ; μ ₀ =μ ₁ ; q ₀₁ =q ₁₀	4	-365.927	740.060	10.060	0.99	2.554	2.973	2.465	2.465	0.209	0.209
λ ₀ =λ ₁ ; μ ₀ ≠μ ₁ ; q ₀₁ =q ₁₀	4	-367.173	742.551	12.551	0.99	2.944	2.944	2.822	2.480	0.206	0.206
λ ₀ =λ ₁ ; μ ₀ =μ ₁ ; q ₀₁ ≠q ₁₀	4	-367.360	742.924	12.924	0.99	2.989	2.989	2.650	2.650	0.353	0.119
λ ₀ ≠λ ₁ ; μ ₀ ≠μ ₁ ; q ₀₁ =q ₁₀	5	-360.316	730.942	0.942	0.99	0.963	3.566	0.740	2.876	2.876	2.876
λ₀≠λ₁; μ₀=μ₁; q₀₁≠q₁₀	5	-359.845	730.000	0	Best	1.418	2.427	1.349	1.349	0.034	0.274
λ ₀ =λ ₁ ; μ ₀ ≠μ ₁ ; q ₀₁ ≠q ₁₀	5	-362.785	735.880	5.880	0.99	2.244	2.244	2.193	1.177	0.024	0.300

$\lambda_0 \neq \lambda_1; \mu_0 \neq \mu_1; q_{01} \neq q_{10}$	6	-358.759	729.952	-0.048	0.141	1.079	2.956	0.965	2.037	0.064	0.227
* the lowest AIC is the model with 6 parameters, but it has a small ΔAIC as compared to simplest models											
Fruit type (0: capsule; 1: berry)											
Model	N P	LogL	AICc	ΔAIC	P (LRT)	λ_0	λ_1	μ_0	μ_1	q_{01}	q_{10}
Null model $(\lambda_0 = \lambda_1; \mu_0 = \mu_1; q_{01} = q_{10})$	3	-319.636	645.395	22.804	0.99	2.815	2.815	2.469	2.469	0.037	0.037
$\lambda_0 \neq \lambda_1; \mu_0 = \mu_1; q_{01} = q_{10}$	4	-310.059	628.324	5.733	0.99	1.954	2.666	1.692	1.692	0.024	0.024
$\lambda_0 = \lambda_1; \mu_0 \neq \mu_1; q_{01} = q_{10}$	4	-310.192	628.588	5.998	0.99	2.280	2.280	2.044	1.186	0.018	0.018
$\lambda_0 = \lambda_1; \mu_0 = \mu_1; q_{01} \neq q_{10}$	4	-319.270	646.745	24.155	0.99	2.814	2.814	2.467	2.467	0.030	0.052
$\lambda_0 \neq \lambda_1; \mu_0 \neq \mu_1; q_{01} = q_{10}$	5	-309.859	630.028	7.437	0.99	2.055	2.498	1.804	1.460	1.460	1.460
$\lambda_0 \neq \lambda_1; \mu_0 = \mu_1; q_{01} \neq q_{10}$	5	-307.565	625.438	2.848	0.99	1.937	2.715	1.701	1.701	0.014	0.067
$\lambda_0 = \lambda_1; \mu_0 \neq \mu_1; q_{01} \neq q_{10}$	5	-306.141	622.591	0	Best	2.220	2.220	2.018	1.017	0.009	0.081
$\lambda_0 \neq \lambda_1; \mu_0 \neq \mu_1; q_{01} \neq q_{10}$	6	-305.982	624.400	1.809	0.573	2.364	2.111	2.170	0.875	0.008	0.085
Pollinator class (0: invertebrate; 1: vertebrate)											
Model	N P	LogL	AICc	ΔAIC	P (LRT)	λ_0	λ_1	μ_0	μ_1	q_{01}	q_{10}
Null model $(\lambda_0 = \lambda_1; \mu_0 = \mu_1; q_{01} = q_{10})$	3	-290.715	587.553	37.753	0.99	2.847	2.847	2.501	2.501	0.013	0.013
$\lambda_0 \neq \lambda_1; \mu_0 = \mu_1; q_{01} = q_{10}$	4	-270.798	549.800	0	Best	0.998	1.943	0.970	0.970	0.011	0.011
$\lambda_0 = \lambda_1; \mu_0 \neq \mu_1; q_{01} = q_{10}$	4	-272.780	553.766	3.966	0.99	2.051	2.051	2.045	1.144	0.007	0.007
$\lambda_0 = \lambda_1; \mu_0 = \mu_1; q_{01} \neq q_{10}$	4	-290.087	588.379	38.579	0.99	2.844	2.844	2.498	2.498	0.001	0.015
$\lambda_0 \neq \lambda_1; \mu_0 \neq \mu_1; q_{01} = q_{10}$	5	-269.357	549.023	-0.777	0.089	0.351	2.093	0.255	1.155	1.155	1.155

$\lambda_0 \neq \lambda_1; \mu_0 = \mu_1; q_{01} \neq q_{10}$	5	-270.113	550.534	0.734	0.242	0.990	1.952	0.968	0.968	0.007	0.021
$\lambda_0 = \lambda_1; \mu_0 \neq \mu_1; q_{01} \neq q_{10}$	5	-271.390	553.089	3.289	0.99	1.992	1.992	1.998	1.037	0.004	0.022
$\lambda_0 \neq \lambda_1; \mu_0 \neq \mu_1; q_{01} \neq q_{10}$	6	-269.330	551.094	1.294	0.23	0.377	2.083	0.288	1.140	0.016	0.019
				<i>* the lowest AIC is the model with 5 parameters, but it has a small ΔAIC as compared to simplest models</i>							

Best-fitting model, as determined via a combination of the Aikake Information Criterion and likelihood ratio tests (LRT), is in red and bold text. The best model is first identified with the lowest AICc and then compared to other models with ΔAIC and LRT. The best-fitting models show that there is higher net diversification in the Andes (vs. extra-Andes), at high elevation (vs. low elevation), in berry-producing lineages (vs. capsular lineages), and in vertebrate-pollinated lineages (vs. invertebrate-pollinated lineages). Our second and third best models in each scenario also support these conclusions. λ = speciation rate (in events/Ma/lineage); μ = extinction rate (in events/Ma/lineage); q = transition rate between states; NP, number of parameter in each model.

Table S6. Correlated evolution of binary traits using BayesTraits.

Maximum Likelihood					
Pair of Characters	LH (Dependent)	LH (Independent)	χ^2 statistic	df	p-value
Pollination: Fruit	-56.583684	-60.354574	7.54178	4	0.109881
Pollination: Elevation	-104.072122	-107.581998	7.019752	4	0.1348481
Pollination: Andean	-88.546479	-92.452286	7.811614	4	0.09872793
Fruit: Elevation	-130.009089	-133.796692	7.575206	4	0.1084384
Fruit: Andean	-118.2341643	-122.1769823	7.8856359	4	0.09585848
Elevation: Andean	-159.696796	-168.894404	18.395216	4	0.001032828*

Bayesian Inference			
Pair of Characters	Harmonic Mean (Dependent)	Harmonic Mean (Independent)	Bayes Factor
Pollination: Fruit	-66.279838	-64.657858	3.24396
Pollination: Elevation	-111.382757	-111.797966	-0.830418
Pollination: Andean	-98.023314	-97.954698	0.137232
Fruit: Elevation	-138.288508	-138.146294	0.284428
Fruit: Andean	-127.542706	-126.737234	1.610944
Elevation: Andean	-229.079744	-172.277752	113.603984*

Results of likelihood ratio test (above) and Bayes factor of harmonic means from MCMC (below) for the correlated evolution of 6 pairs of binary characters. Tests were performed by comparing fit of a dependent v. independent model of trait evolution in BayesTraits. Asterisks mark traits whose evolution was found to be significantly correlated. LH: log-likelihood; df: degrees of freedom.

Table S7. Model comparison for the two diversity-dependence analyses presented, with mean parameter estimates for each model.

Diversity Dependence							
Model	Mode of dependency	NP	logL	AICc	λ	μ	K
λ depends on diversity (no extinction)	Exponential variation	2	-1056.369	2108.677	0.05975	-	31190.11
λ depends on diversity and μ constant	Exponential variation	3	-958.0664	1910.01	0.07683	3.041e-05	5873.402

Both models show that the group has not reached its ecological carrying capacity, suggesting that there currently is no limit to species diversity. λ = speciation rate (in events/Ma/lineage); μ = extinction rate (in events/Ma/lineage); K = carrying capacity; NP, number of parameter in each model.