

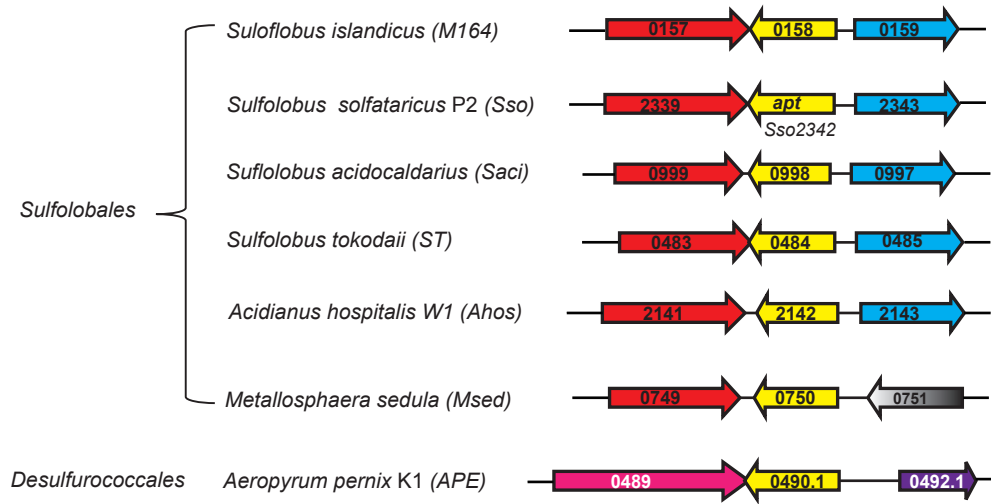
Fig. S1. Comparative and phylogenetic analysis of archaeal *apt* homologues

A. Schematic overview of the *apt* gene that is conserved in some well-studied microorganisms from *Sulfolobales* and *Desulfurococcales*. Homology is indicated by the same colors.

B. Phylogenetic analysis of Apt and Gpt homologues from hyperthermophilic crenarchaea. Phylograms of Apt and Gpt proteins were constructed using the Phylogeny-fr platform (www.phylogeny.fr). For each run, proteins corresponding to hits pertaining to archaea, with BLAST e-value lower than $2E-50$ (for Apt) and $3E-36$ (for Gpt) and covering at least 40% of query sequence, were selected and fed to the “One Click Mode Phylogeny analysis” pipeline available on www.phylogeny.fr.

Fig. S1

A



- Adenine phosphoribosyltransferas
- Peptide chain release factor 1
- 5'-Methylthioadenosine phosphorylase
- Hypothetical protein
- Pyruvate kinase
- Hypothetical protein

B

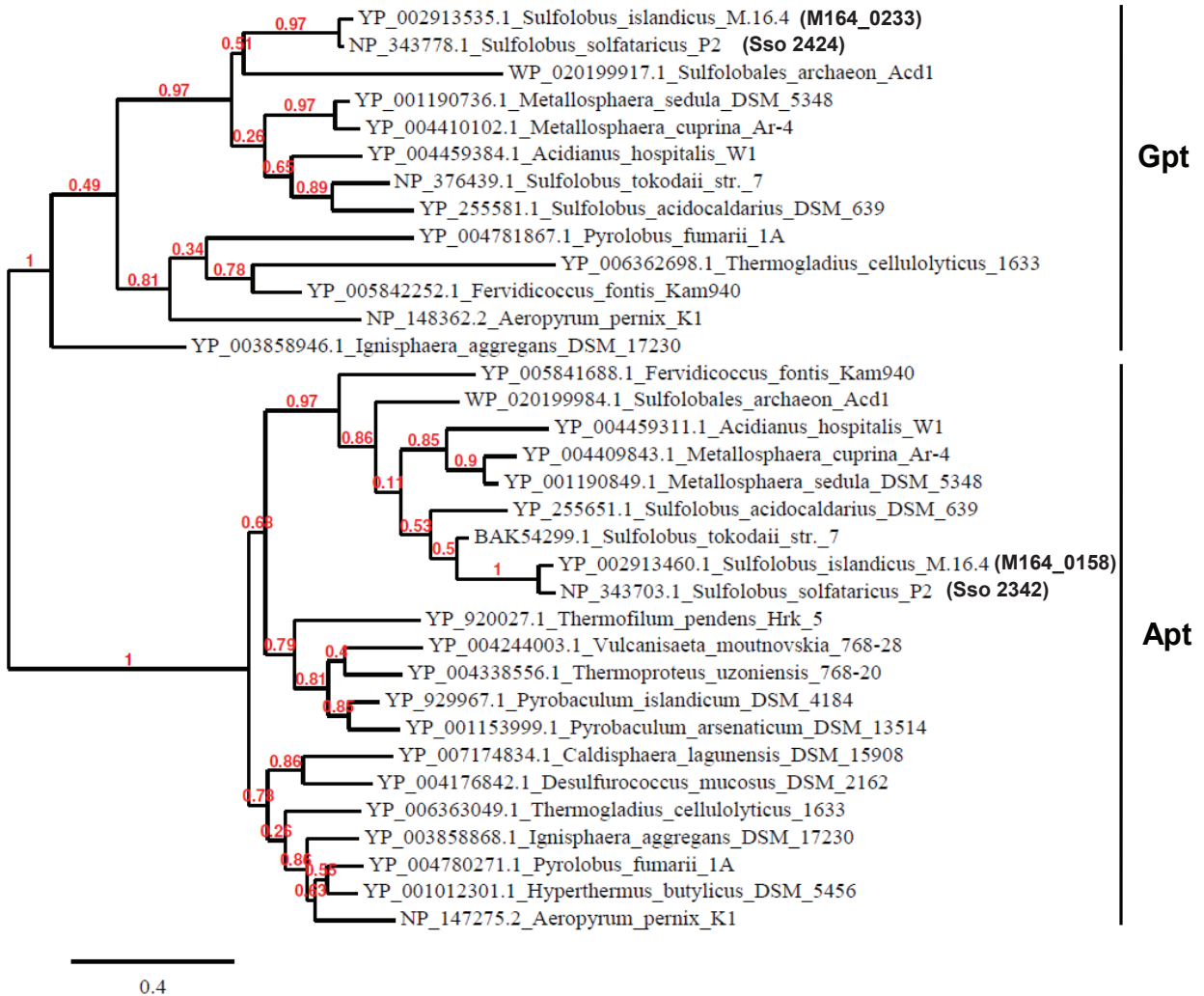


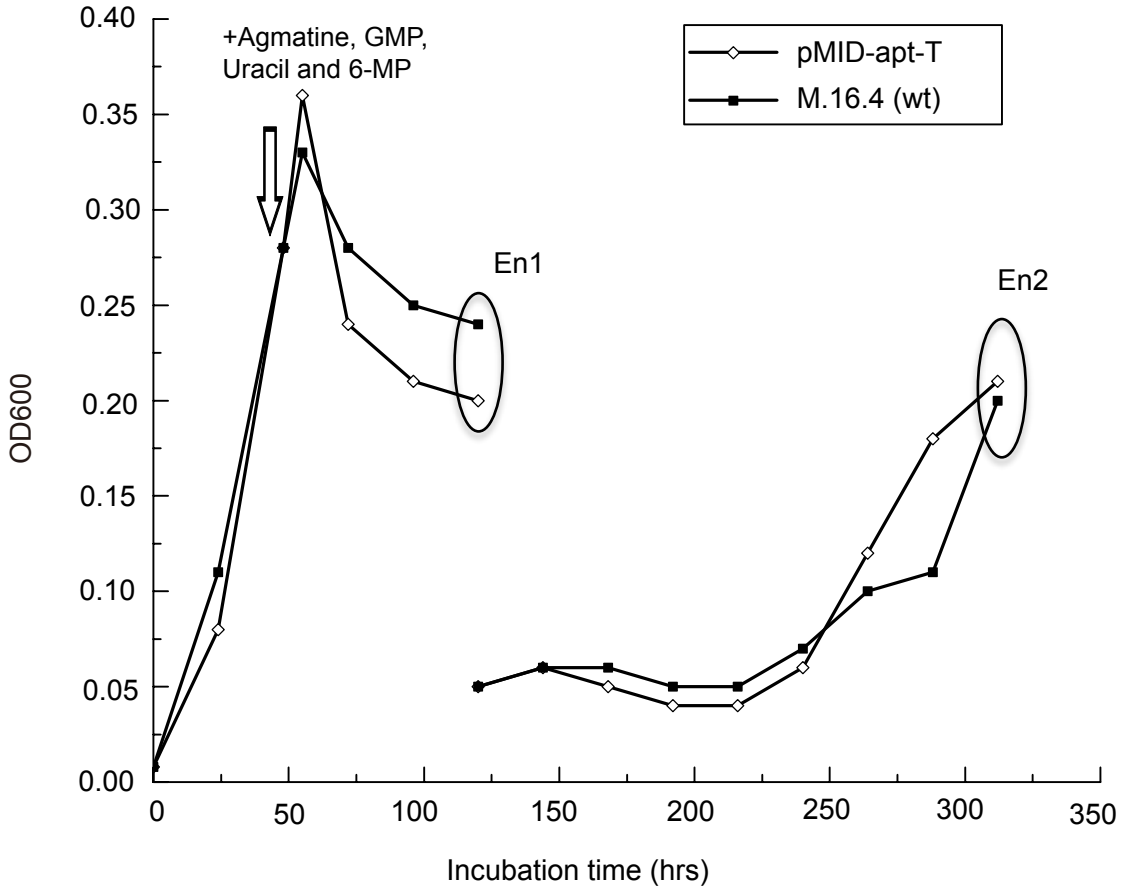
Fig. S2. Generation of Δapt mutant cells via a mutant propagation assay

A. Δapt mutant cells were enriched via a mutant propagation assay as described previously (1), with slight modifications. The transformant pMID-apt-T and a control strain *S. islandicus* M.16.4 were inoculated into DTU liquid medium. When the OD₆₀₀ of cells achieved around 0.26, 6-MP (0.15 mM), agmatine (1 mg/ml) and GMP (0.5 mM) were added. After another three days' incubation, cells were collected (En1), transferred into fresh medium with initial OD₆₀₀=0.05 and then cultivated until obvious growth were observed (~8 days), as indicated by the En2 (A). B. X-gal staining of pMID-apt-T-En2 and M.16.4-En2 cultures. C. Isolation of single 6-MP^R colonies by directly plating pMID-apt-T-En2 on medium plates supplemented with 6-MP, GMP, agmatine and uracil. X-gal staining of colonies indicated that Δapt mutants (White color) were successfully enriched and dominant.

1. **Zhang C, Guo L, Deng L, Wu Y, Liang Y, Huang L, She Q.** 2010. Revealing the essentiality of multiple archaeal pcna genes using a mutant propagation assay based on an improved knockout method. *Microbiology* **156**:3386-3397.

Fig. S2

A



B



C



1 **Fig. S3. Sequences and structural features of the insertion sequence element ISC1205**

2 A. Schematic representation of ISC1205 in the wild-type *S. islandicus* M.16.4 chromosome. The entire
3 insertion sequence element contained a large open reading frame (M164_0453) of 1023 bp, a putative
4 promoter region of 150 bp and terminal inverted repeats (IR, 16 bp). DR means the direct repeat with 100%
5 match.

6 B. Properties of ISC1205 insertions in chromosome of 6-MP^R isolates. The orientation of the ISC1205 in
7 the chromosome was indicated by dotted arrows.

8

9

Fig. S3

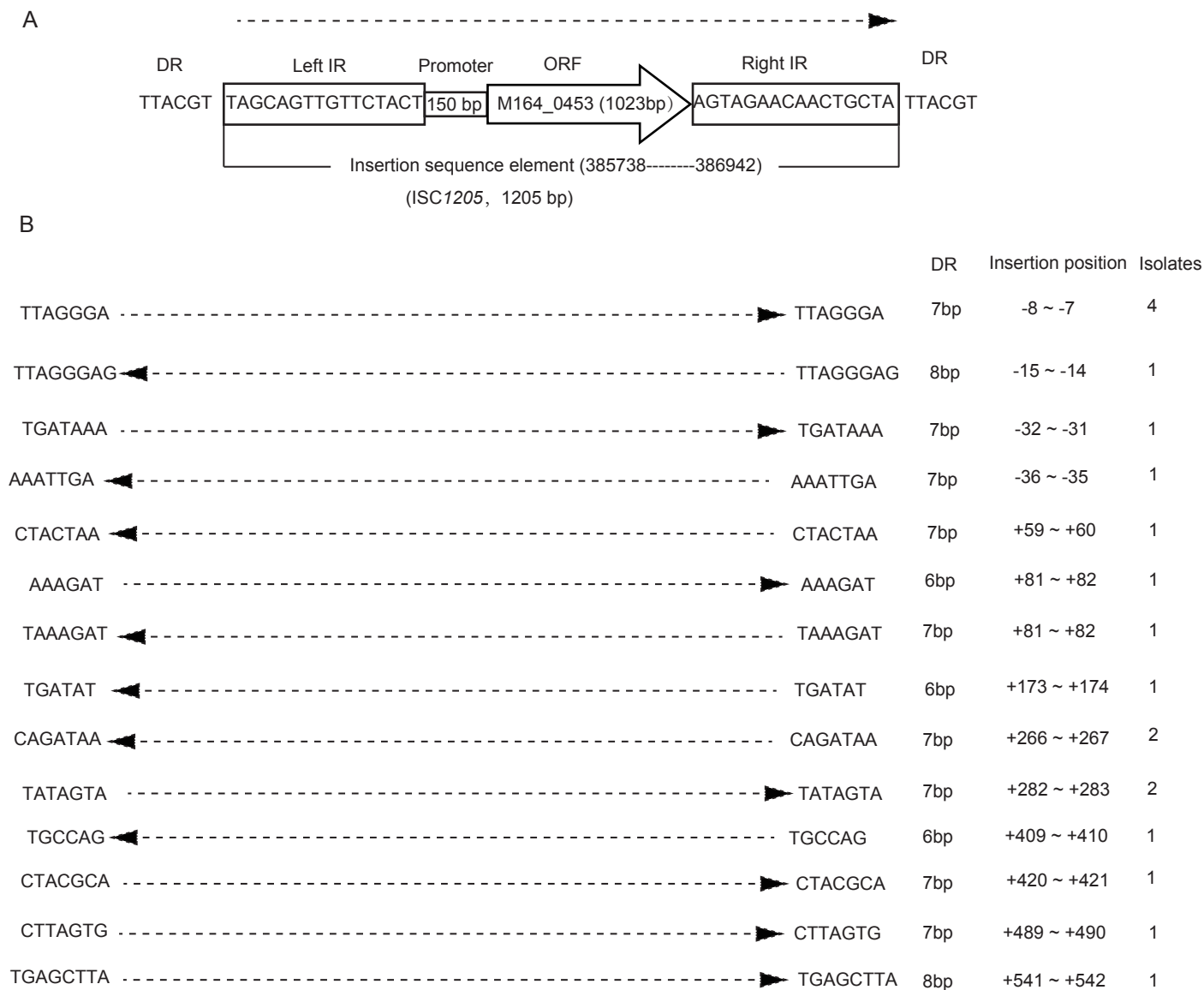


Table S1. MICs of 6-methylpurine (6-MP) on *Sulfolobus* species

<i>Sulfolobus</i> species	MIC of 6-MP (mM)	6-MP-resistant determinant
<i>S. islandicus</i> M.16.4	0.04	<i>M164_0158</i>
<i>S. islandicus</i> REY15A	0.1	<i>SiRe_0139</i>
<i>S. solfataricus</i> P1	0.04	--
<i>S. solfataricus</i> P2	0.04	<i>Sso2342</i>
<i>S. tokodaii</i>	0.02	<i>St0484</i>
<i>S. acidocaldarius</i>	> 0.4	<i>Saci_0998</i>

Sulfolobus cells were inoculated into DT medium (initial OD₆₀₀ = 0.008) supplemented with various concentrations of 6-methylpurine. Cell growth was checked after 10 days' incubation at 75°C.

--: The sequence of 6-MP-resistant determinant in *S. solfataricus* P1 shares 100% identity as *Sso2342*.