

Supplementary Material

Quorum sensing systems differentially regulate the production of phenazine-1-carboxylic acid in the rhizobacterium *Pseudomonas aeruginosa* PA1201

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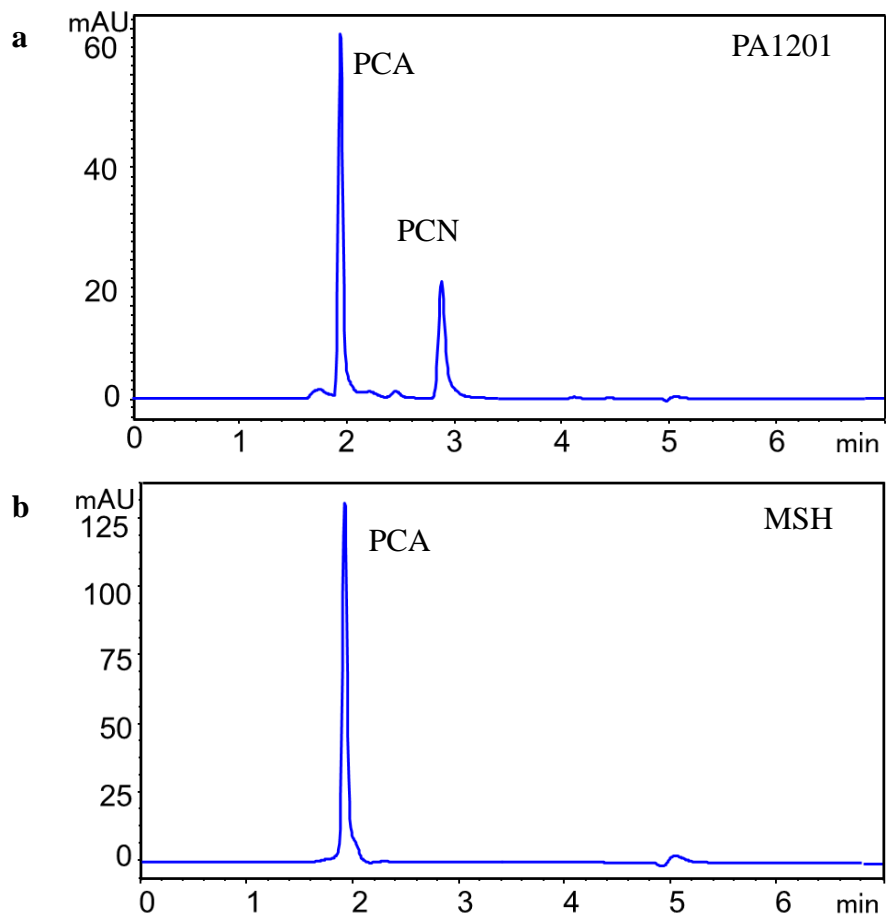
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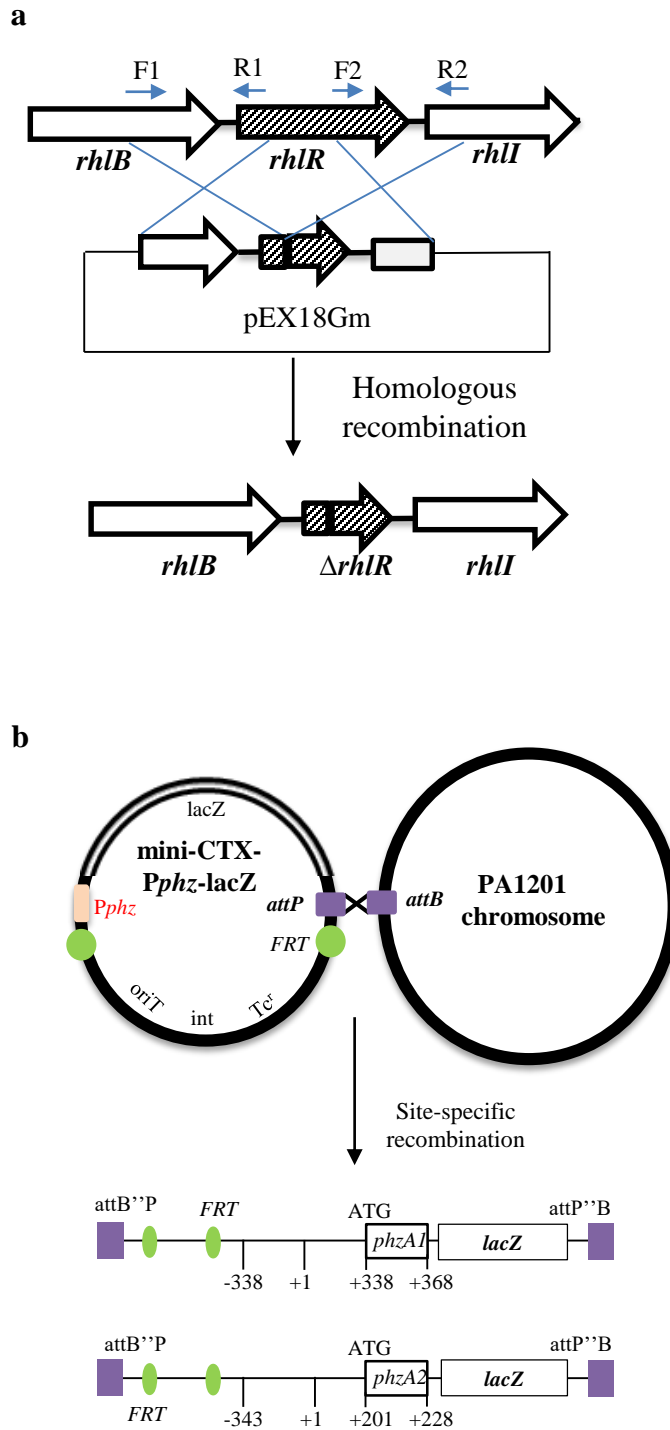
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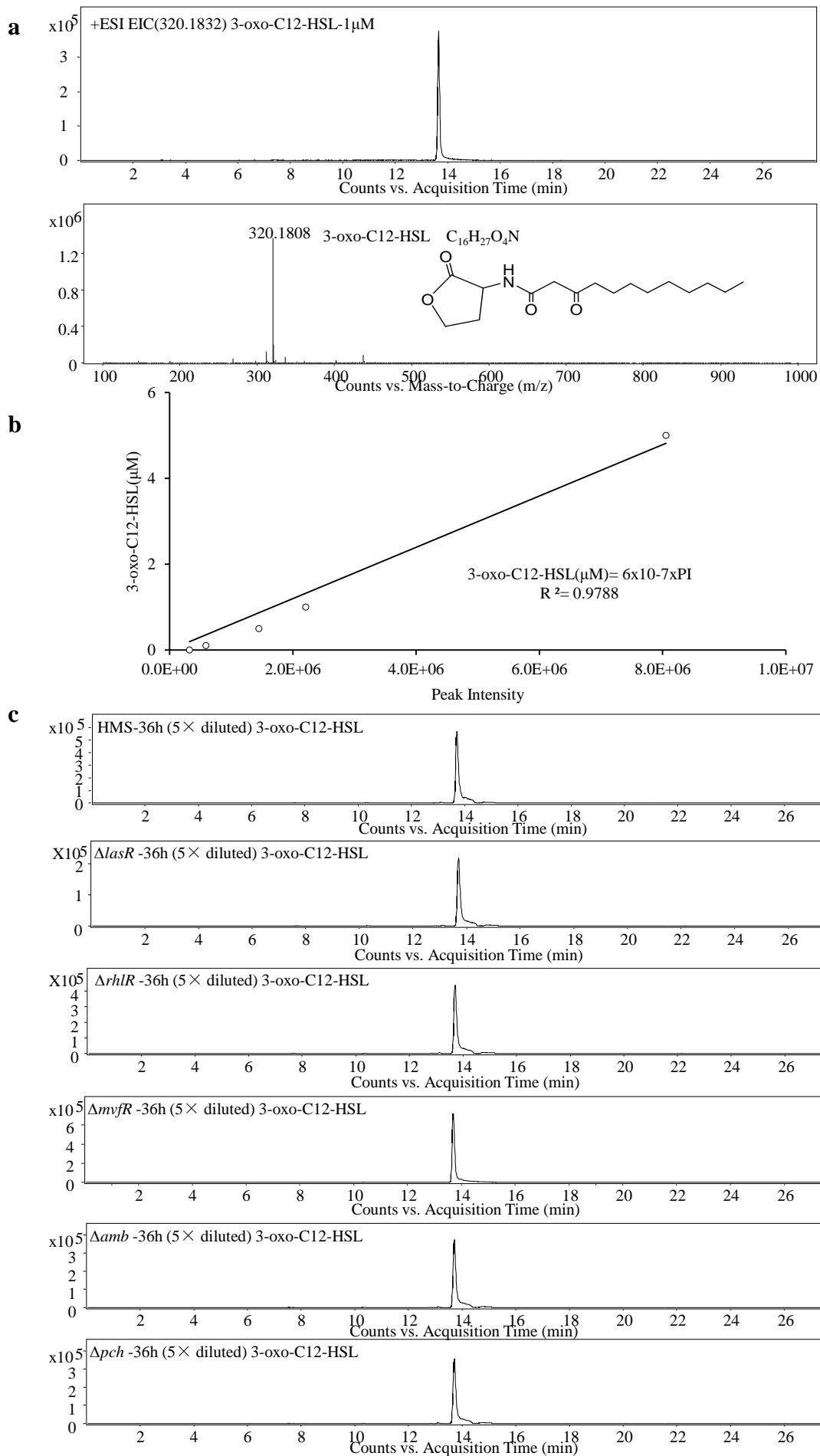
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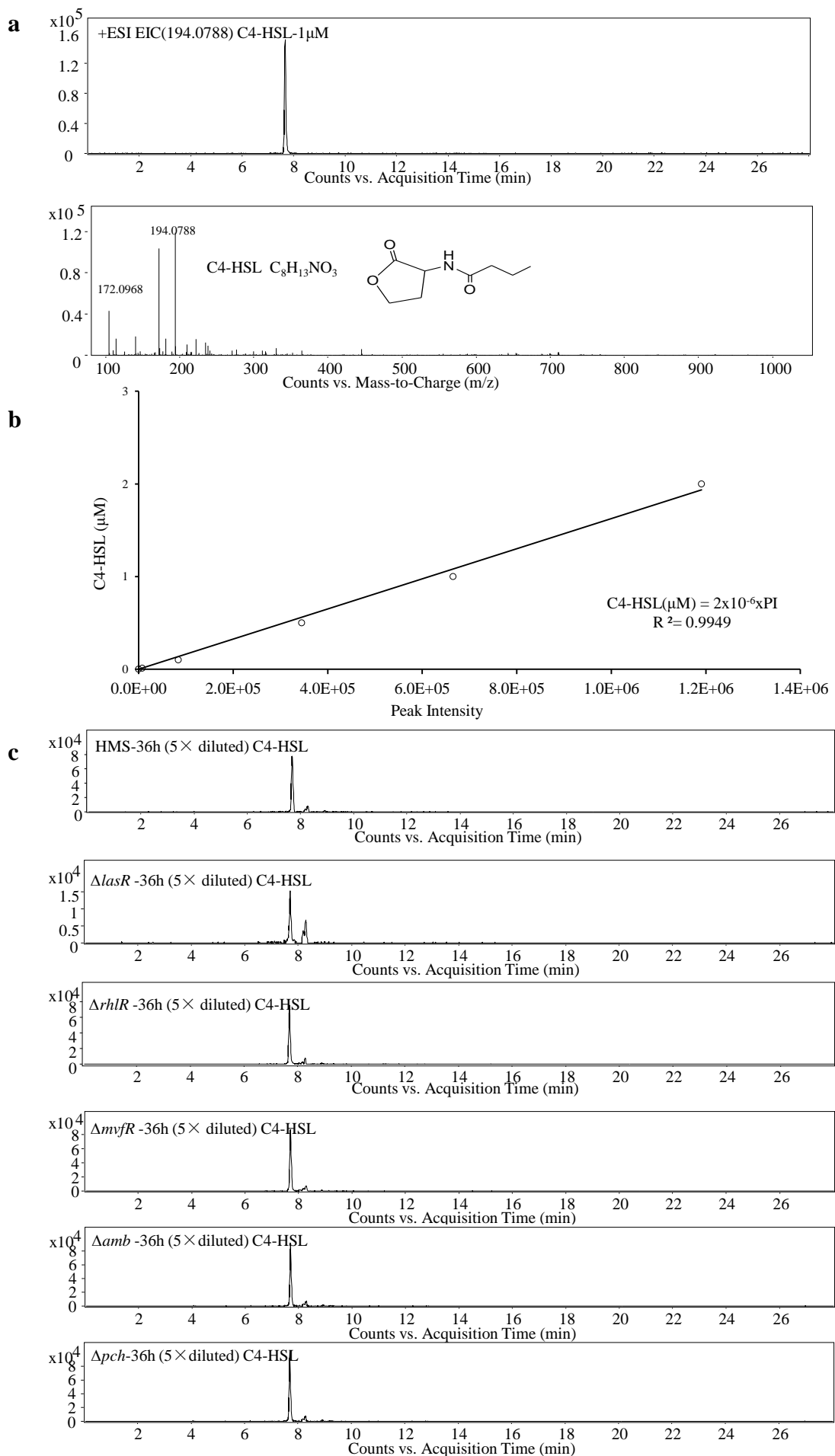
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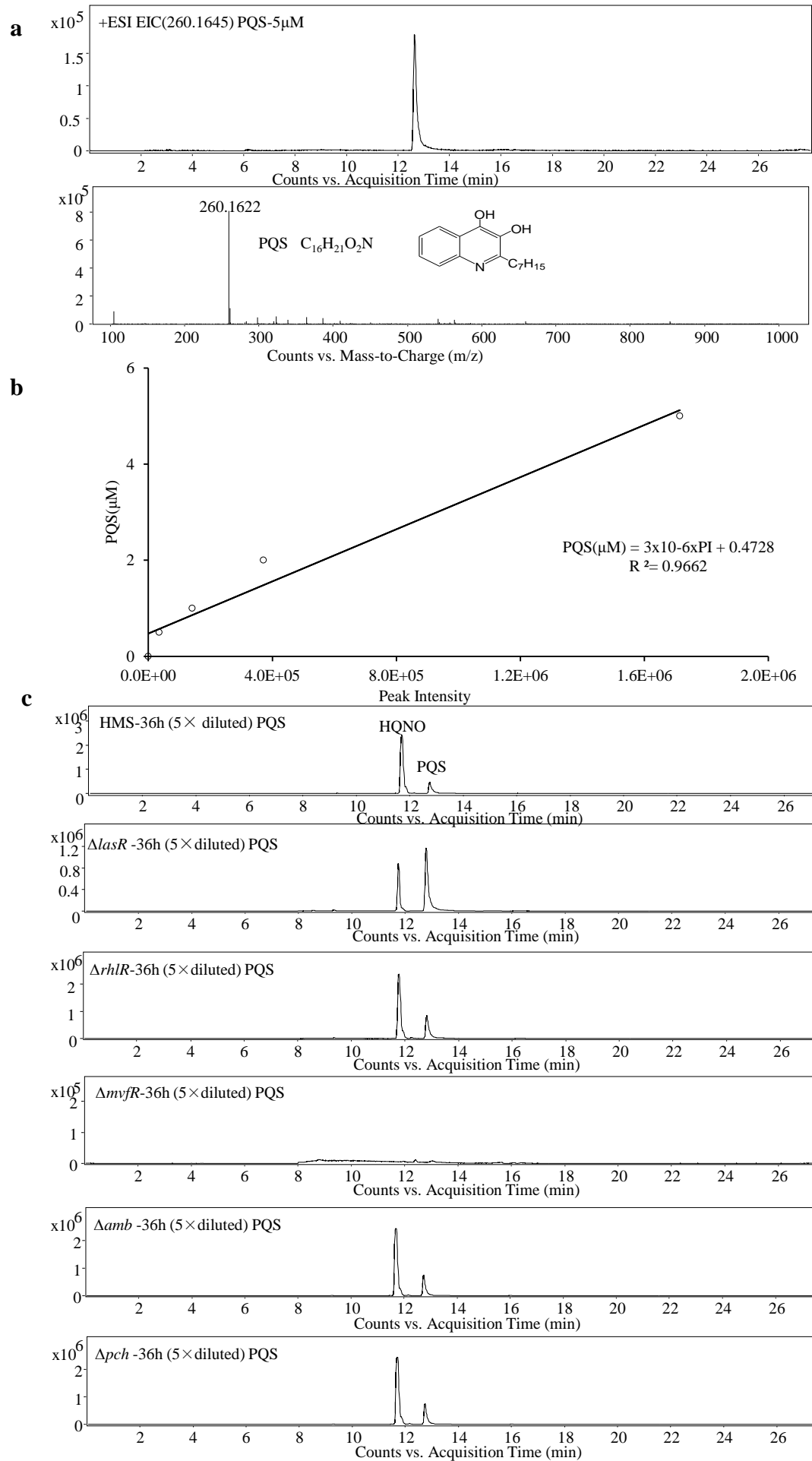
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Supplementary Figure S3 LC-MS analysis of 3-oxo-C12-HSL production in various deletion strains. (a) LC-MS analysis of the commercially available 3-oxo-C12-HSL samples at different concentrations. (b) The standard curve of peak intensity for the specific extracted ion chromatogram (EIC) and 3-oxo-C12-HSL concentrations. (c) EIC chromatograms of extracted 3-oxo-C12-HSL samples.



Supplementary Figure S4 LC-MS analysis of C4-HSL in various PA1201-derived deletion mutation strains. (a) LC-MS analysis of the commercially available C4-HSL. (b) The standard curve of peak intensity for the specific extracted ion chromatogram (EIC) and C4-HSL concentrations. (c) EIC chromatograms of extracted C4-HSL samples



Supplementary Figure S5 LC-MS analysis of PQS in various PA1201-derived deletion mutation strains. (a) LC-MS analysis of the commercially available PQS. (b) The standard curve of peak intensity for the specific extracted ion chromatogram (EIC) and PQS concentrations. (c) EIC chromatograms of extracted PQS samples.

Supplementary Table S1 The strains and plasmids used in this study

Strains	Relevant characteristics	Reference or source
<i>P. aeruginosa</i> strains		
PA1201	<i>P. aeruginosa</i> wild-type strain, Spe ^R	39
MSH	PA1201Δ <i>phzMΔphzSΔphzH</i> The <i>phzM</i> , <i>phzS</i> and <i>phzH</i> in-frame deletion mutant, Spe ^R	40
MSHΔ <i>lasR</i>	<i>lasR</i> in-frame deletion mutant based on MSH, Spe ^R	This study
MSHΔ <i>rhIR</i>	<i>rhIR</i> in-frame deletion mutant based on MSH, Spe ^R	This study
MSHΔ <i>mvfR</i>	<i>mvfR</i> in-frame deletion mutant based on MSH, Spe ^R	This study
MSHΔ <i>amb</i>	<i>ambBCDE</i> in-frame deletion mutant based on MSH, Spe ^R	This study
MSHΔ <i>pch</i>	<i>pchABCDEFG</i> in-frame deletion mutant based on MSH, Spe ^R	This study
MSHΔ <i>phz1</i>	<i>phzA₁B₁C₁D₁E₁F₁G₁</i> in-frame deletion mutant based on MSH, Spe ^R	40
MSHΔ <i>phz1ΔlasR</i>	<i>lasR</i> in-frame deletion mutant based on MSHΔ <i>phz1</i> , Spe ^R	This study
MSHΔ <i>phz1ΔrhIR</i>	<i>rhIR</i> in-frame deletion mutant based on MSHΔ <i>phz1</i> , Spe ^R	This study
MSHΔ <i>phz1ΔmvfR</i>	<i>mvfR</i> in-frame deletion mutant based on MSHΔ <i>phz1</i> , Spe ^R	This study
MSHΔ <i>phz2</i>	<i>phzA₂B₂C₂D₂E₂F₂G₂</i> in-frame deletion mutant based on MSH, Spe ^R	40
MSHΔ <i>phz2ΔlasR</i>	<i>lasR</i> in-frame deletion mutant based on MSHΔ <i>phz2</i> , Spe ^R	This study
MSHΔ <i>phz2ΔrhIR</i>	<i>rhIR</i> in-frame deletion mutant based on MSHΔ <i>phz2</i> , Spe ^R	This study
MSHΔ <i>phz2ΔmvfR</i>	<i>mvfR</i> in-frame deletion mutant based on MSHΔ <i>phz2</i> , Spe ^R	This study
MSH::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSH chromosomal attB site	This study
MSHΔ <i>lasR</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>lasR</i> chromosomal attB site	This study
MSHΔ <i>rhIR</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>rhIR</i> chromosomal attB site	This study
MSHΔ <i>mvfR</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>mvfR</i> chromosomal attB site	This study
MSHΔ <i>amb</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>amb</i> chromosomal attB site	This study
MSHΔ <i>pch</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>pch</i> chromosomal attB site	This study
MSH::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSH chromosomal attB site	This study
MSHΔ <i>lasR</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>lasR</i> chromosomal attB site	This study
MSHΔ <i>rhIR</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>rhIR</i> chromosomal attB site	This study
MSHΔ <i>mvfR</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>mvfR</i> chromosomal attB site	This study
MSHΔ <i>amb</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>amb</i> chromosomal attB site	This study
MSHΔ <i>pch</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>pch</i> chromosomal attB site	This study
MSHΔ <i>phz2</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>phz2</i> chromosomal attB site	This study
MSHΔ <i>phz2ΔlasR</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>phz2ΔlasR</i> chromosomal attB site	This study
MSHΔ <i>phz2ΔrhIR</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>phz2ΔrhIR</i> chromosomal attB site	This study
MSHΔ <i>phz2ΔmvfR</i> ::P <i>phz1</i> -lacZ	Single copy insertion of P <i>phz1</i> -lacZ in the MSHΔ <i>phz2ΔmvfR</i> chromosomal attB site	This study
MSHΔ <i>phz1</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>phz1</i> chromosomal attB site	This study
MSHΔ <i>phz1ΔlasR</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>phz1ΔlasR</i> chromosomal attB site	This study
MSHΔ <i>phz1ΔrhIR</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>phz1ΔrhIR</i> chromosomal attB site	This study
MSHΔ <i>phz1ΔmvfR</i> ::P <i>phz2</i> -lacZ	Single copy insertion of P <i>phz2</i> -lacZ in the MSHΔ <i>phz1ΔmvfR</i> chromosomal attB site	This study

Supplementary Table S1 The strains and plasmids used in this study

Strains	Relevant characteristics	Reference or source
Reporter strains		
<i>C. violaceum</i> CV026	Double mini-Tn5 mutant derived from <i>C. violaceum</i> ATCC 31532, AHL biosensor	42
<i>A. tumefaciens</i> CF11	Harbors two plasmids expressing TraR and the reporter gene lacZ under the tra reporter, respectively	41
<i>E. coli</i> strains		
DH5 α	<i>E. coli</i> F- Φ 80lacZ Δ M15 Δ (lacZYA-argF) U169 recA1 endA1 hsdR17 (rK-, mK+) phoA supE44 λ - thi-1 gyrA96 relA1	Lab collection
pRK2013	A conjugative plasmid, Kan ^R	Lab collection
S17- λ pir	TpR SmR recA, thi, pro, hsdR-M+RP4: 2-Tc:Mu: Kan Tn7 λ pir	Lab collection
Plasmids		
pEX18Gm	Gm ^R ; oriT ⁺ sacB ⁺ , gene replacement vector with MCS This study from pUC18	49
mini-CTX-lacZ	Tet ^R ; integration-proficient vector for chromosomal insertion at the attB site	52
pEX18Gm- Δ lasR	pEX18Gm with the fusion product of the <i>lasR</i> flanking fragments for in-frame deletion of <i>lasR</i> , Gm ^R	This study
pEX18Gm- Δ rhlR	pEX18Gm with the fusion product of the <i>rhlR</i> flanking fragments for in-frame deletion of <i>rhlR</i> , Gm ^R	This study
pEX18Gm- Δ mvfR	pEX18Gm with the fusion product of the <i>mvfR</i> flanking fragments for in-frame deletion of <i>mvfR</i> , Gm ^R	This study
pEX18Gm- Δ amb	pEX18Gm with the fusion product of the <i>ambBCDE</i> flanking fragments for in-frame deletion of <i>amb</i> cluster, Gm ^R	This study
pEX18Gm- Δ pch	pEX18Gm with the fusion product of the <i>pchABCDREFG</i> flanking fragments for in-frame deletion of <i>pch</i> cluster, Gm ^R	40
mini-CTX-Pphz1-lacZ	integration-proficient vector with the promoter region of <i>phzA1</i> , Tet ^R	This study
mini-CTX-Pphz2-lacZ	integration-proficient vector with the promoter region of <i>phzA2</i> , Tet ^R	This study

Supplementary Table S2 Primers used in this study

Application	Oligos and sequence(5'to3')
Deletion of <i>lasR</i>	<i>lasR</i> -F1: <u>GGAATTC</u> CAGCGTGGTGGTGTGCGAGATGTT
	<i>lasR</i> -R1: GTGCCCGACGACCCCTGAAGATGT
	<i>lasR</i> -F2: ggggtcgtcgggcacGCCTGGCGCGAGCATTACGAC
	<i>lasR</i> -R2: <u>CGGGATCC</u> CGCTACGCGGCGGGAGGTCAC
	<i>lasR</i> -con-F: TGGCCTTGGTTGACGGTTTTCTTG
Deletion of <i>rhlR</i>	<i>rhlR</i> -F1: <u>GGAATTC</u> CTGCCGCCGGTACACCCCAAGTTC
	<i>rhlR</i> -R1: CAACCCTATCTGTTATGCCAGCACCG
	<i>rhlR</i> -F2: taacagatagggttTTCGACGCGCCGAACAAGACG
	<i>rhlR</i> -R2: <u>CGGGATCC</u> GTGCGCGAAACGGCTGACGACCT
	<i>rhlR</i> -con-F: GCTCGCGATTGGGGCCTCTGTGT
Deletion of <i>mvfR</i>	<i>mvfR</i> F1: <u>GGAATTC</u> CACGCGCCACCCAATAAAAGGAAT
	<i>mvfR</i> R1: CGGCGGGATGGCGGTGTC
	<i>mvfR</i> F2: accgccatcccgccgCGCGCCGACCAGAGTAGAG
	<i>mvfR</i> R2: <u>CCCAAGCTT</u> GAGGCCGAGGAAACCCGCAAC
	<i>mvfR</i> -con-R AGCCGAGCACGCACTGGTTGAAGC
Deletion of <i>amb</i> cluster	<i>amb</i> F1: <u>GGAATTC</u> CGCGGCGTACTCCACCATCTGTTC
	<i>amb</i> R1: CGCCGCCAGTACCCCGCTCCATT
	<i>amb</i> F2: ggggtactggcggcgCCGGCGAACCTGGCAACCTGA
	<i>amb</i> R2: <u>CCCAAGCTT</u> TGGCCGCCGCGAACGCTTAGAAAAA
	<i>amb</i> -con-F: CTCCTCCGCCATGGGGTGTGCTA
	<i>amb</i> -con-R: CGGCCGTGGACGCTGATGAAGTT
Deletion of <i>pch</i> cluster	<i>pch</i> -F1: <u>CGGAATTC</u> GTTCGAATCGCCTACCAGACCGC
	<i>pch</i> -R1: CAATGGGAAGCCAAGGTGAGCGAC
	<i>pch</i> -F2: cttggettcccattgCCAGCTCACCCATCTCGACCAGGC
	<i>pch</i> -R2: <u>ACGCGTCGAC</u> GAAACCGGCATACCAGTCGTCCCG
	<i>pch</i> -con-F: GCTGGCGCGGACGTTGTAGAGAT
	<i>pch</i> -con-R: TGCCGCCCGCCAATGATAATAAAT
Construction of <i>phz1</i> reporter strain	<i>Pphz1</i> -For: <u>GGAATTC</u> CACATTTCCGTAACCCGAGAAGTACCC
	<i>Pphz1</i> -Rev: <u>CGGGATCC</u> GGGTGTTTCCCTGTACCCTGACC
Construction of <i>phz2</i> reporter strain	<i>Pphz2</i> -For: <u>GGAATTC</u> CTCAACTCCAGCAACAAGGCGGAG
	<i>Pphz2</i> -Rev: <u>CGGGATCC</u> CCCTTCAACCGTTGGTACTCTCG

The sequences with underline are the introduced restriction sites.