Supplementary Material legends

Supplementary Figure S1. List of the *CRX* human mutations leading to the production of a premature stop codon at amino acid 185 compared to the human wild-type and feline mutant sequence.

Supplementary Figure S2. Retinal regions imaged by SD-OCT. Cross sectional retinal images were captured for measurement of retinal layer thicknesses at the following regions (as indicated on the fundus image by an asterisk): the *area centralis*; 4 optic nerve diameters from the optic nerve edge superiorly, inferiorly, temporally and nasally.

Supplementary Figure S3. Sites of retinal sampling for qRT-PCR. A. For the 2-week-old kittens, due to the small globe size, retina was collected from two regions as indicated (central and peripheral retinal regions). B. In the 6, 12 and 20 weeks of age kittens retinal biopsies were collected from the indicated five regions (*area centralis* – *AC*; inferior mid-peripheral – Inf MP; inferior far-peripheral – Inf FP; superior mid-peripheral – Sup MP; and superior far-periphery – Sup FP).

Supplementary Figure S4. TR, REC+, ONL and IR layer thicknesses on SD-OCT images, four optic nerve distance from the optic nerve rim itself A. Superiorly, B. Inferiorly, C. Temporally and D. Nasally of $Crx^{Rdy/+}$ kittens normalized to control WT kittens at 4, 6, 8, 10, 12, 15, 20 weeks-old and 6 months of age.

The ONL and REC+ thicknesses showed thinning with age in $Crx^{Rdy/+}$ kittens compared to WT. TR was not significant thinned until 6 months of age inferiorly. Conversely, the IR became thicker in $Crx^{Rdy/+}$ kittens compared to the wild-type kittens in all regions from 10 weeks of age.

Supplementary Figure S5. Immunolabeling of the retina for S cones and inner retinal cells.

Frozen sections of central retinal labeled with PNA combined with S-opsin, and with GFAP, and PKCalpha.

The $Crx^{Rdy/+}$ retina showed a lack of S-opsin staining cells. Cone nuclei (PNA positive) became mislocalized to the subretinal space but did not stain for S-opsin (indicated by white arrowheads in the bottom panel – high magnification view). There was marked Müller cell activation as indicated by GFAP upregulation at 12 and 20 weeks of age. Rod bipolar cells were labeled by PKCalpha and showed dendrite retraction in the $Crx^{Rdy/+}$ retina.

Key: OS= Photoreceptor Outer segment, IS= Photoreceptor Inner segment, ONL= Outer Nuclear Layer, OPL= Outer Plexiform Layer, INL= Inner Nuclear Layer, IPL= Inner Plexiform Layer, GCL/NFL= Ganglion Cell Layer/Nerve Fiber Layer; White arrow head= Mislocalized photoreceptor nuclei.

Supplementary Figure S6. Western blot for Crx protein in retinal nuclear and cytoplasmic extracts from 2 week old kittens.

Note that the truncated mutant Crx protein was exclusively detected in the nuclear extract from the $Crx^{Rdy/+}$ kitten and was at a higher level than the wild-type protein (immunolabeled with anti-Crx antibody 119b1).

Beta-actin was used as protein loading control.

Supplementary Figure S7. Dual-Luciferase assays for CRX transactivation activity on mouse *Crx-Luc* reporter.

Crx auto-activation ability of WT or mutant Crx protein on its own promoter Crx (containing 2 binding sites within 500-bp upstream region of the mouse Crx gene) was tested using HEK293 cells transfected by plasmids containing the 500bp mouse Crx promoter-luciferase reporter (mCrx-Luc) and the indicated Crx protein expression vector. Comparing to pcDNA3.1hisc control, only pCAGIG-feline Crx WT significantly activated the mCrx-Luc reporter. pCAGIG-feline Crx^{Rdy} mutant did not show any transactivation compared to the control vector.

P-values indicate as followed: ***P*<0.01 and ****P*<0.001.

Supplementary Table S1. List of antibodies used for IHC – their origins and dilutions.

Supplementary Table S2. Primer sequences for qRT-PCR assays.

CRX	1	135:	PLGISDSYSPPLPGPSGSPTTAVATVSIWSPASESPLPEAQRAGLVASGPS <mark>L</mark> TSAPYAMTYAPASA	200299
CRX	pI138	d1(1):	PLGTQIPTVPLCPAPQAPQPRQWPLCPSGAQPQSPLCLRRSGLGWWPQGRL-	
CRX	pS141	d1(2):	PLGISDPTVPLCPAPQAPQPRQWPLCPSGAQPQSPLCLRRSGLGWWPQGRL-	
CRX	pS143	d2i1(3):	PLGISDSYAPLCPAPQAPQPRQWPLCPSGAQPQSPLCLRRSGLGWWPQGRL-	
CRX	pP153	d1(4,5):	PLGISDSYSPPLPGPSGSQPRQWPLCPSGAQPQSPLCLRRSGLGWWPQGRL-	
CRX	_ pE168	d1(6):	PLGISDSYSPPLPGPSGSPTTAVATVSIWSPASSPLCLRRSGLGWWPQGRL-	
CRX	pP170	d1(7):	PLGISDSYSPPLPGPSGSPTTAVATVSIWSPASESPCLRRSGLGWWPQGRL-	
CRX	pA174	d1(8):	PLGISDSYSPPLPGPSGSPTTAVATVSIWSPASESPLPERSGLGWWPQGRL-	
CRX	pA177	d1(9):	PLGISDSYSPPLPGPSGSPTTAVATVSIWSPASESPLPEAQRLGWWPQGRL-	
CRX	pA181	dl(10):	PLGISDSYSPPLPGPSGSPTTAVATVSIWSPASESPLPEAQRAGLVPQGRL-	
fCrx	pA18	2d1 (11):	PLGISDSYSPPLPGPSVSPTSAVATVSIWSPASESPLPEAQRAGLVAAGPL-	

- 1. Nichols LL, 2nd, Alur RP, Boobalan E, et al. Two novel CRX mutant proteins causing autosomal dominant Leber congenital amaurosis interact differently with NRL. Hum Mutat. 2010;31:E1472-1483.
- 2. Zou X, Yao F, Liang X, et al. De novo mutations in the cone-rod homeobox gene associated with leber congenital amaurosis in Chinese patients. *Ophthalmic Genet*. 2015;36:21-26.pl138d1.
- 3. Stone EM. Leber congenital amaurosis a model for efficient genetic testing of heterogeneous disorders: LXIV Edward Jackson Memorial Lecture. Am J Ophthalmol. 2007;144:791-811.
- 4. Wang P, Guo X, Zhang Q. Further evidence of autosomal-dominant Leber congenital amaurosis caused by heterozygous CRX mutation. Graefes Arch Clin Exp Ophthalmol. 2007;245:1401-1402.
- 5. Ziviello C, Simonelli F, Testa F, et al. Molecular genetics of autosomal dominant retinitis pigmentosa (ADRP): a comprehensive study of 43 Italian families. J Med Genet. 2005;42:e47.
- Freund CL, Gregory-Evans CY, Furukawa T, et al. Cone-rod dystrophy due to mutation in a novel photoreceptor-specific homeobox gene (CRX) essential for maintenance of the photoreceptor. Cell. 1997;91:543-553.
- 7. Perrault I, Hanein S, Gerber S, et al. Evidence of autosomal dominant Leber congenital amaurosis (LCA) underlain by a CRX heterozygous null allele. J Med Genet. 2003;40:e90.

8. Nakamura M, Ito S, Miyake Y. Novel de novo mutation in CRX gene in a Japanese patient with leber congenital amaurosis. Am J Ophthalmol. 2002;134:465-467.

- 9. Koenekoop RK, Loyer M, Dembinska O, Beneish R. Visual improvement in Leber congenital amaurosis and the CRX genotype. Ophthalmic Genet 2002;23:49-59.
- 10. Zhang Q, Li S, Guo X, et al. Screening for CRX gene mutations in Chinese patients with Leber congenital amaurosis and mutational phenotype. *Ophthalmic Genet* 2001;22:89-96.
- 11. Menotti-Raymond M, Deckman KH, David V, Myrkalo J, O'Brien SJ, Narfstrom K. Mutation discovered in a feline model of human congenital retinal blinding disease. Invest Ophthalmol Vis Sci. 2010;51:2852-2859.



Supplementary Figure S2



Supplementary Figure S3



Supplementary Figure S4



Supplementary Figure S5



Supplementary Figure S6



Supplementary Figure S7

Supplementary Table S1. List of antibodies used for IHC – their origins and dilutions.

Antibody – Source	Type	Primary	Secondary	Secondary
	турс	Dilution	Antibody – Source	Dilution
hCAR	Polyclonal	1:10,000	Alexa Fluor 488 Goat anti-	1:500
(Human cone arrestin)	rabbit		rabbit IgG	
Dr. Cheryl Craft; LUMIJ,			Life technologies,	
University of Southern California,			Carlsbad, CA, USA	
Los Angeles, CA, USA				
PNA	Biotinylated	1:500	Alexa Fluor 488	1:500
(Biotinylated Peanut Agglutinin)	Lectin		Streptavadin	
Vector Labs Inc., Burlin-game, CA,			Life technologies,	
USA			Carlsbad, CA, USA	
ML-opsin	Polyclonal	1:1,000	Alexa Fluor 568 or 594	1:500
(Anti-Opsin, Red/Green; Medium/	rabbit		Goat anti-rabbit IgG	
Long wavelength cone opsin)			Life technologies,	
Millipore Corp., Billerica, MA, USA			Carlsbad, CA, USA	
S-opsin	Polyclonal	1:1,000	Alexa Fluor 568 or 594	1:500
(Anti-Opsin, Blue; Short wavelength	rabbit		Goat anti-rabbit IgG	
cone opsin)			Life technologies,	
Millipore Corp., Billerica, MA, USA			Carlsbad, CA, USA	
RetP1	Monoclonal	1:2	Alexa Fluor 594 Rabbit	1:500
(Rhodopsin Ab-1)	mouse		anti-mouse IgG	
Thermo Scientific, Rockford, IL,			Life technologies,	
USA			Carlsbad, CA, USA	
GFAP	Monoclonal	1:300	Alexa Fluor 594 Rabbit	1:500
(Anti-Glial Fibrillary Acidic Protein)	mouse		anti-mouse IgG	
Cell Signaling Technology Inc.,			Life technologies,	
Danvers, MA, USA			Carlsbad, CA, USA	
PKCa	Monoclonal	1:500	Alexa Fluor 594 Rabbit	1:500
(Protein Kinase C-alpha)	mouse		anti-mouse IgG	
BD Biosciences, San Jose, CA, USA			Life technologies,	
			Carlsbad, CA, USA	

Supplementary Table S2. Primer sequences for qRT-PCR assays.

Primer name	Forward primer	Reverse primer	Amplicon size (bp)	Annealing temperature (°C)
Crx Total	5' AAGACTCAGTACCCGGATGTGTA 3'	5' GGGGCTGTAGGAGTCTGAGAT 3'	223	60
Arr3	5' CGTTGTCCTGTATTCCCTAGAC 3'	5' GCTAGAGGCCAGATTAGTATCAC 3'	190	60
Rho	5' GGTGCCCTACGCCAGCGTG 3'	5' CAGTGGGTTCTTGCCACAG 3'	190	60
Tuba1b	5' GCTCTATTGCCTGGAACACG 3'	5' CATCTTCCTTGCCCGTGATG 3'	230	60
GAPDH	5' GGTCTTCACCACCATGGAGA 3'	5' TGGACTGTGGTCATGAGTCC 3'	237	60