

Supplemental Table S2. PCR primers for detection and sequencing and PCR conditions.

target gene	primer name	sequence (5'-3')	PCR conditions (30 cycles)	Size of amplicon (bp)	References
EAEc					
<i>aggR</i>	aggR-F	CAGAATACATCAGTACACTG	94 °C, 30 s / 55 °C, 30 s / 72 °C, 30 s	433	Tsukamoto T. <i>et al.</i> , 1996 ^a
	aggR-R	GAAGCTTACAGCCGATATAT			
<i>aap</i>	aap-F	TGAAAAAATAAGTTTGTATC	94 °C, 30 s / 55 °C, 40 s / 72 °C, 60 s	344	Sarantuya J. <i>et al.</i> , 2004 ^b
	aap-R	AACCCATTCCGTTAGAGC			
<i>aatA</i>	aatA-F	ATGTTACCAGATATAAATATAG	94 °C, 30 s / 55 °C, 40 s / 72 °C, 60 s	1065	Nishi J. <i>et al.</i> , 2003 ^c
	aatA-R	CATTTCCCCTGTATTGGAAATG			
<i>aggA</i>	aggA-1	TTAGTCTTCTATCTAGGG	94 °C, 30 s / 55 °C, 30 s / 72 °C, 30 s	457	Vila J., <i>et al.</i> , 2000 ^d
	aggA-2	AAATTAATCCGGCATGG			
<i>aafA</i>	aafA-1	TGCGATTGCTACTTTATTAT	94 °C, 30 s / 55 °C, 30 s / 72 °C, 30 s	242	Vila J., <i>et al.</i> , 2000 ^d
	aafA-2	ATTGACCGTGATTGGCTTCC			
<i>agg3A</i>	agg3A-F	GTATCATTGCGAGTCTGGTATTCAG	94 °C, 60 s / 55 °C, 60 s / 72 °C, 60 s	462	Bernier C, <i>et al.</i> , 2002 ^e
	agg3A-R	GGGCTGTATAGAGTAACCTCCAG			
<i>hdaA/agg4A</i>	hdaA-F	TCCATTATGTCAGGCTGCAA	94 °C, 30 s / 55 °C, 30 s / 72 °C, 40 s	411	Boisen N., <i>et al.</i> , 2008 ^f
	hdaA-R	GGCGTTAACGTCTGATTTCC			
<i>agg5A</i>	aaf5-F	TATCATTGCGAGTCTGGTATTCA	94 °C, 30 s / 55 °C, 60 s / 72 °C, 60 s	501	Prager R, <i>et al.</i> , 2014 ^g
	aaf5-R	TAATTTAAGCTGAAGAATCCAGTCAA			
<i>pet</i>	pet-1	ACTGGGGGACTCATTGCTGT	94 °C, 30 s / 55 °C, 30 s / 72 °C, 30 s	832	Vila J., <i>et al.</i> , 2000 ^d
	pet-2	GCGTTTTTCCGTTCCCTATT			
<i>astA</i>	EAST1-F	GCCATCAACACAGTATATCC	94 °C, 30 s / 55 °C, 30 s / 72 °C, 30 s	106	Yatsuyanagi J, <i>et al.</i> , 1996 ^h
	EAST1-R	GAGTGACGGCTTTGTAGTCC			
<i>sepA</i>	sepA-F	GCAGTGGAATATGATGCGGC	94 °C, 30 s / 59 °C, 90 s / 72 °C, 90 s	794	Boisen N, <i>et al.</i> , 2012 ⁱ
	sepA-R	TTGTTCCAGATCGGAGAAGAACG			
<i>sat</i>	sat-F	TCAGAAGCTCAGCGAATCATTG	94 °C, 30 s / 59 °C, 90 s / 72 °C, 90 s	932	Boisen N, <i>et al.</i> , 2012 ⁱ
	sat-R	CCATTATCACCAGTAAAACGCCACC			
<i>pic</i>	pic-F	ACTGGATCTTAAGGCTCAGGAT	94 °C, 30 s / 59 °C, 90 s / 72 °C, 90 s	572	Boisen N, <i>et al.</i> , 2012 ⁱ
	pic-R	GACTTAATGTCAGTTCAGCG			
<i>aaiC</i>	aaiC-F	TGGTGACTACTTTGATGGACATTGT	94 °C, 50 s / 57 °C, 90 s / 72 °C, 90 s	313	Boisen N, <i>et al.</i> , 2012 ⁱ
	aaiC-R	GACACTCTCTTGGGGTAACGA			
UPEC					
<i>sfa*</i>	sfa1	CTCCGGAGAAGTGGGTGCATCTTAC	94 °C, 60 s / 63 °C, 60 s / 72 °C, 80 s	410	Yamamoto S, <i>et al.</i> , 1995 ^j
	sfa2	CGGAGGAGTAATTACAACTGGCA			
<i>pap*</i>	pap1	GACGGCTGTACTGCAGGTTGTGGCG	94 °C, 60 s / 63 °C, 60 s / 72 °C, 80 s	328	Yamamoto S, <i>et al.</i> , 1995 ^j
	pap2	ATATCCTTTCTGCAGGGATGCAATA			
	pap3	GCAACAGCAACGCTGGTTGCATCAT			
	pap4	AGAGAGGCCACTCTTATACGGACA			
<i>afa*</i>	afa1	CTGGGCGACAAACTGATAACTCTC	94 °C, 60 s / 63 °C, 60 s / 72 °C, 80 s	750	Yamamoto S, <i>et al.</i> , 1995 ^j
	afa2	CATCAAGCTGTTTGTTCGTCGCGCG			
<i>aer*</i>	aer1	TACGGATTGTCATATGCAGACCGT	94 °C, 60 s / 63 °C, 60 s / 72 °C, 80 s	602	Yamamoto S, <i>et al.</i> , 1995 ^j
	aer2	AATATCTTCTCCAGTCCGGAGAAG			
<i>cnf1</i>	cnf1	GGGGGAAGTACAGAAGAATTA	94 °C, 60 s / 55 °C, 60 s / 72 °C, 60 s	1126	Hinenoya A., <i>et al.</i> , 2009 ^k
	cnf2	TTGCCGTCCACTCTCACCAGT			
ESBL					
<i>bla_{CTX-M}</i>	CTX-M-U1	ATGTGCAGYACCAGTAARGTKATGGC [§]	94 °C, 30 s / 60 °C, 30 s / 72 °C, 40 s	593	Boyd DA, <i>et al.</i> , 2004 ^l
	CTX-M-U2	TGGGTRAARTARGTSACCAGAAYCAGCGG [§]			
H antigen					
<i>fliC</i>	F-FLIC1	ATGGCACAAGTCATTAATACCCAAC	95 °C, 30 s / 60 °C, 60 s / 72 °C, 120s	ca. 1.3–2.6 kb	Fields PI, <i>et al.</i> , 1997 ^m
	R-FLIC2	CTAACCTGCGCAGAGACA			

*amplified by multiplex PCR, ^SY stands for pyrimidine, R stands for purine, K stands for G or T, and S stands for G or C

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^bSarantuya J, *et al.* 2004. Typical enteroaggregative *Escherichia coli* is the most prevalent pathotype among *E. coli* strains causing diarrhea in Mongolian children. J Clin Microbiol 42:133-139.

^cNishi J, *et al.* 2003. The export of coat protein from enteroaggregative *Escherichia coli* by a specific ATP-binding cassette transporter system. J Biol Chem 278:45680-45689.

^dVila J, *et al.* Enteroaggregative *Escherichia coli* virulence factors in traveler's diarrhea strains. J Infect Dis 182(6):1780-1783, 2000

^eBernier C, *et al.* Identification of an aggregative adhesion fimbria (AAF) type III-encoding operon in enteroaggregative *Escherichia coli* as a sensitive probe for detecting the AAF-encoding operon family. Infect Immun 70(8):4302-4311, 2002

^fBoisen N, *et al.* New adhesin of enteroaggregative *Escherichia coli* related to the Afa/Dr/AAF family. Infect Immun 76(7):3281-3292, 2008

^gPrager R, *et al.* Two Novel EHEC/EAEc Hybrid Strains Isolated from Human Infections. PLoS One 9:e95379.

^hYatsuyanagi J, *et al.* Enteropathogenic *Escherichia coli* strains harboring enteroaggregative *Escherichia coli* (EAaggEC) heat-stable enterotoxin-1 gene isolated from a food-borne like outbreak. Kansenshogaku Zasshi 70(1):73-79, 1996 (jpn)

ⁱBoisen N, *et al.* 2012. Genomic characterization of enteroaggregative *Escherichia coli* from children in Mali. J Infect Dis 205:431-444.

^jYamamoto S, *et al.* 1995. Detection of urovirulence factors in *Escherichia coli* by multiplex polymerase chain reaction. FEMS Immunol Med Microbiol 12:85-90.

^kHinenoya A, *et al.* Prevalence and characteristics of cytolethal distending toxin-producing *Escherichia coli* from children with diarrhea in Japan. Microbiol Immunol 53(4):206-215, 2009

^lBoyd DA, *et al.* 2004. Complete nucleotide sequence of a 92-kilobase plasmid harboring the CTX-M-15 extended-spectrum β -lactamase involved in an outbreak in long-term-care facilities in Toronto, Canada. Antimicrob Agents Chemother 48:3758-3764.

^mFields PI, *et al.* 1997. Molecular characterization of the gene encoding H antigen in *Escherichia coli* and development of a PCR-restriction fragment length polymorphism test for identification of *E. coli* O157:H7 and O157:NM. J Clin Microbiol 35:1066-1070.