14-3-3σ loss leads to activation of the epithelial to mesenchymal transition due to the stabilization of c-Jun.

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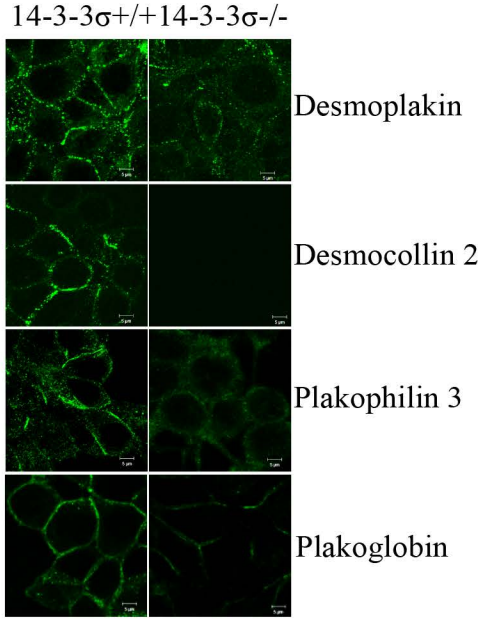
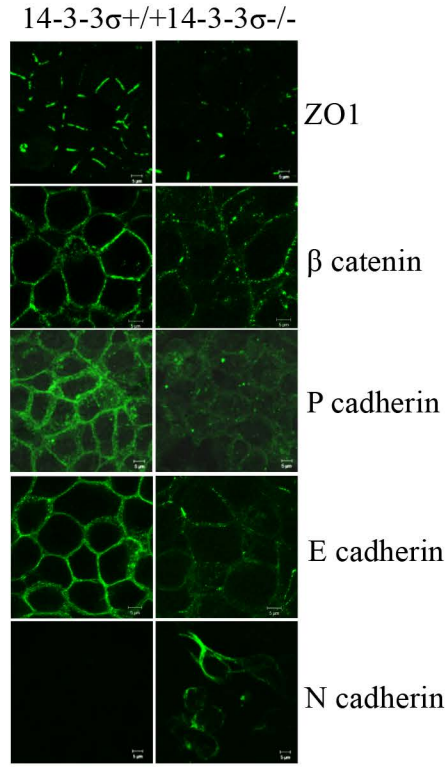
Supplemental materials

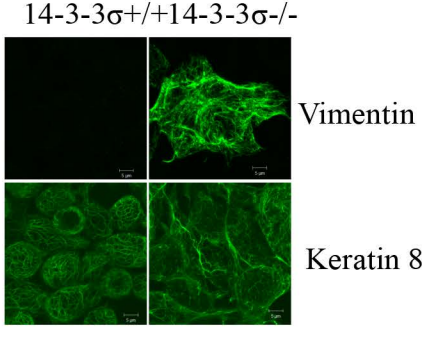
A


14-3-3σ+/+14-3-3σ-/-
Desmoplakin
Desmocollin 2
Plakophilin 3
Plakoglobin

B


14-3-3σ+/+14-3-3σ-/-
ZO1
β catenin
P cadherin
E cadherin
N cadherin

C


14-3-3σ+/+14-3-3σ-/-
Vimentin
Keratin 8

Supplemental figure legend

Supplemental figure S1. Loss of 14-3-3σ does not lead to change in the localization of the adhesion proteins. *A*-*C*, 14-3-3σ-/- and 14-3-3σ+/+ cells were fixed and stained with the indicated antibodies followed by confocal microscopy. Micrographs for desmosomal proteins (*A*), adherens junction proteins (*B*) and intermediate filament proteins (*C*) are shown in the figure. Magnification is X63 with 2X optical zoom and bars indicate 5μm.

Supplemental table 1. Oligonucleotide sequences for generation of site directed mutants of c-Jun.

Point mutant	Sequence
c JUN S58A FWD	gcgccaagaacGCGgacctcctca
c JUN S58A REV	tgaggaggtcCGCgttcttggcgc
c JUN S267A FWD	gcatcgctgccGCCaagtgccgaaa
c JUN S267A REV	tttcggcacttGGCggcagcgatgc

Supplemental table 2. List of antibodies used for Western blot experiments along with the appropriate dilution.

Antibody	Supplier	Dilution
Plakophilin3	Zymed	1:2000
Desmoglein2	Zymed	1:500
Desmocollin2/3	Zymed	1:500
DP-200	Abexome	1:1000
Plakoglobin	Abcam	1:500
Vimentin	Sigma	1:5000
N cadherin	BD Biosciences	1:100
Cytokeratin 8	Sigma	1:5000
Cytokeratin 18	Sigma	1:5000
Snail	Santa Cruz	1:100
Slug	Santa Cruz	1:100
Zeb1	Santa Cruz	1:100
c-Jun	Abcam	1:100
ZO1	Zymed	1:100
E cadherin	BD Biosciences	1:500
β catenin	Abcam	1:500
Laminin A	Abcam	1:2000
α tubulin	Abcam	1:1500
Flag	Sigma	1:2000
GST	Sigma	1:1000
Ubiquitin	Millipore	1:1000
anti-Myc	Santa Cruz	1:2000
β-actin	Sigma	1:5000
14-3-3σ	Hybridoma supernatant CS112	1:10
HA	12CA5 hybridoma supernatant	1:10

Supplemental table 3. Oligonucleotide sequences for quantitative real time PCR.

Gene	Sequence
E cadherin 5’	GCAGGTCTCCTCTTGGCTCTG
E cadherin 3’	TGTGCCCACTTTGAATCGG
Desmoplakin I 5’	GCTGATTAATGATTTACAG
Desmoplakin I 3’	CACCAGAAGGCTCTCTCTTTC
Desmoplakin II 5’	AAGGTTGAGGGTTCTACTGC
Desmoplakin II 3’	TTGTCTTGCTCCAGGACTT
Desmocollin 2 5’	GTTTTACTCAGCCCCGTCTTG
Desmocollin 2 3’	GCCCATCTTCTTCTTGTCGT
Desmoglein 2 5’	TACGCCCTGCTGCTTCTCC
Desmoglein 2 3’	TCTCCCTCCCGAAGAGCCACG
ZO1 5’	CAAGAGCACAGCAATGGAGGA
ZO1 3’	TCCCCACTCTGAAAATGAGGA
β catenin 5’	AGTGCTGAAGGTGCTATCTGT
β catenin 3’	GAACAAGAGTCCCAAGGAGAC
Plakoglobin 5’	CTACGGCAACCAGGAGAGC
Plakoglobin 3’	GGGACACACGGATAGCACCT
Slug RT5’	AGACCCCCA TGCCA TTGAAG
Slug RT3’	GGCCAGCCCAGAAAAAGTTG
Snail RT5’	TAGCGAGTGGTTCTTCTGCG
Snail RT3’	AGGGCTGCTGGAAGGTAAAC
Twist 1 RT5’	AGCTGAGCAAGATTCAGACCC
Twist 1 RT3’	GCAGCTTGCCATCTTGGAGT
Zeb 1 RT5’	AGGA TGACCTGCCAACAGAC
Zeb 1 RT3’	CTTCAGGCCCCAGGATTTCTT