

Inhibition of DNA methylation promotes tumor sensitivity to netrin-1 interference.

Mérodie Grandin¹, Pauline Mathot¹, Guillaume Devailly¹, Yannick Bidet², Akram Ghantous³, Clementine Favrot¹, Benjamin Gibert¹, Nicolas Gadot⁴, Isabelle Puisieux⁵, Zdenko Herceg³, Jean-Guy Delcros¹, Agnès Bernet¹, Patrick Mehlen^{1§}, Robert Dante^{1§}

Appendix Figure S1 is related to Figure 3.

Appendix Figure S2 is related to Figure 3.

Appendix Figure S3 is related to Figures 3 and 4.

Appendix Figure S4 is related to Figures 3 and 4.

Appendix Figure S5 is related to Figure 5.

Appendix Figure S6 is related to Figure 5.

Appendix Table S1 is related to Experimental Procedures

Appendix Figure S1 - The netrin-1 blocking antibody net1-mAb triggered apoptotic cell death in breast cancer cell lines upon 5-azacytidine treatment.

A, B Gene expression was measured by Q-RT-PCR after 72 hr in MDA-MB-231 (A) and HMLER (B) cells treated daily with 2 μ M of 5-azacytidine (Aza). *PBGD* expression level was used as internal control. Error bars, s.e.m of at least 3 independent experiments, two-tailed unpaired Student's t test, ns = not significant.

C, F MDA-MB-231 and HMLER cell lines were treated with Aza, 2 μ M for 72h, and/or net1-mAb (10 μ g/mL, 48h) and/or recombinant netrin-1 (5 μ g/mL, 48h). Caspase-3 activities (C and E) and cellular mortality (D and F) were determined in 3 independent experiments. **** P<0.0001. ANOVA1. Error bars = s.e.m

Appendix Figure S2 – Tumor growth inhibition induced by treatment combining DAC and therapeutic netrin-1 antibody, *in vivo*.

A. When tumors reached 100 mm³, mice were injected subcutaneously with decitabine (0.5mg/kg) or PBS and/or intraperitoneally with NET1-mAb-T (10 mg/kg) or a Control IgG (Iso-mAb (10 mg/kg). Tumor volumes were measured twice a week. Statistical significance of differences between PBS-groups and DAC + NET1-mAb-T-groups was determined by two-way ANOVA and post-hoc Tukey-test. (n = 6 mice per group). **** P<0.0001.

B. Activated caspase-3, median number of cells per mm² were measured using antibody against cleaved caspase-3 and the DNA fragmentation median number of cells per mm² were determined by TUNEL assay (B); mean values from 4 tumors per group. **** P<0.0001, ANOVA1

C. Representative tumor sections corresponding to the histograms (B), bars represent 50 μ m.

Appendix Figure S3 – Angiogenesis and cell proliferation in patient-derived xenografts treated net1-mAb and/ or decitabine (DAC).

A, D Paraffin embedded tumor xenografts were stained using CD31 for angiogenesis and Ki-67 for cell proliferation in MDA-MB-231 (A, B) and PDX HBC 146 (C, D).

(A and C) Vessel numbers in tumors were measured by CD31 immunohistochemistry staining and expressed as median number of vessels per mm²

(A and C) Mean proliferation rate assessed by Ki- 67 staining; proliferative cells were divided by non-proliferative cells. Error bars = s.e.m. ANOVA1; *** P<0.001, n = not significant, ANOVA1

(B and D) Representative sections of CD31 and KI67 staining for angiogenesis and proliferation count.

Appendix Figure S4 - Combined treatment led to a reduced number of metastasis formation, and was not associated with changes in angiogenesis or proliferation.

A, B. MDA-MB-231 cells previously treated with control IgG1, DAC and/or net1-mAb alone or in combination were grafted on the chorio-allantoic membrane (CAM) of 10 days old chicken embryos. Error bars = s.e.m, Kruskal–Wallis test.

Appendix Figure S5 - Genes upregulated by decitabine treatment in MDA-MB231 and HMLER cell lines.

A. Venn diagram showing the overlap of genes upregulated by decitabine treatment in MDA-MB-231 and HMLER cell lines.

B. Gene ontology and KEGG pathways identified from of common DAC-upregulated genes between MDA-MB-231 and HMLER cell lines.

Appendix Figure S6 – Expression of genes involved in the YAP signaling pathway and effect of *IRF7* depletion on *DAPK1*, *UNC5B*, and *NTN1* expression.

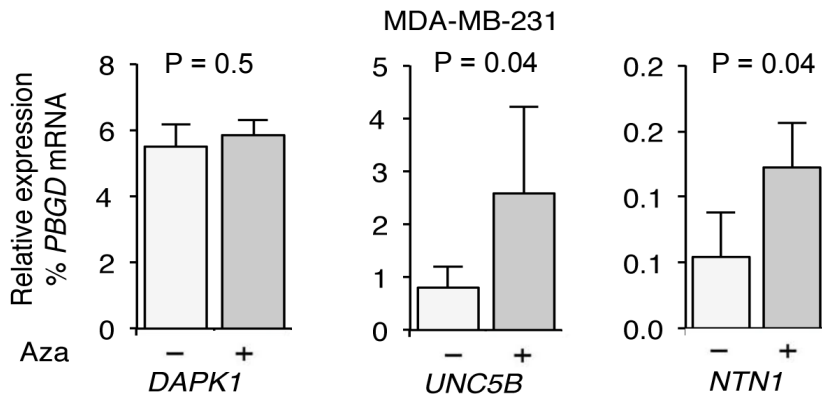
A, B Q-RT-PCR assays of the expression of genes linked to the netrin-1 dependence receptor pathway, *DAPK1*, *UNC5B*, and *NTN1*, and genes linked to the YAP signaling pathway, *YAP*, *TAZ*, and *CTGF* in MDA-MB-231 (A) and HMLER (B) cells.

Error bars = s.e.m. *** P<0.001, ns = not significant. ANOVA1

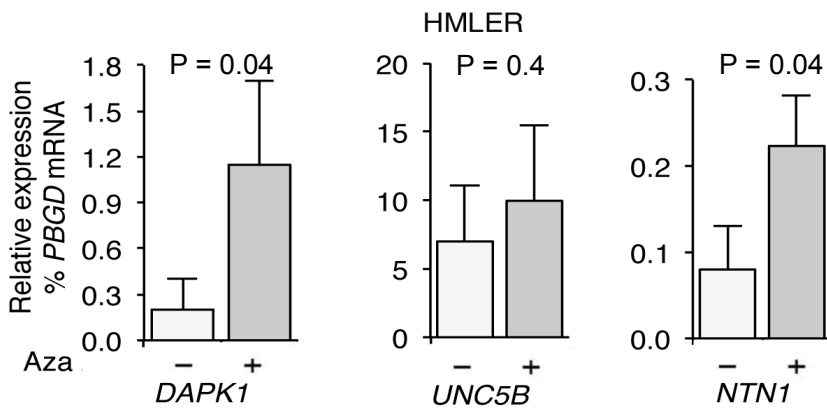
C, D Q-RT-PCR assays of the expression of *DAPK1*, *UNC5B*, *NTN1*, and *IRF7* upon treatment combining DAC treatment and transient transfection with a siRNA targeting *iRF7* in MDA-MB-231 (C) and HMLER (D) cells. **** P<0.0001, ns = not significant.

ANOVA1.

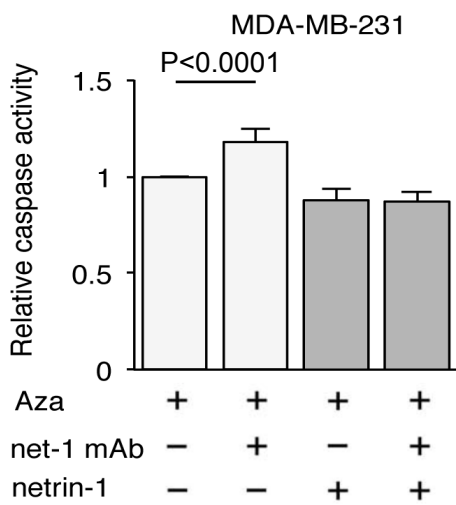
A



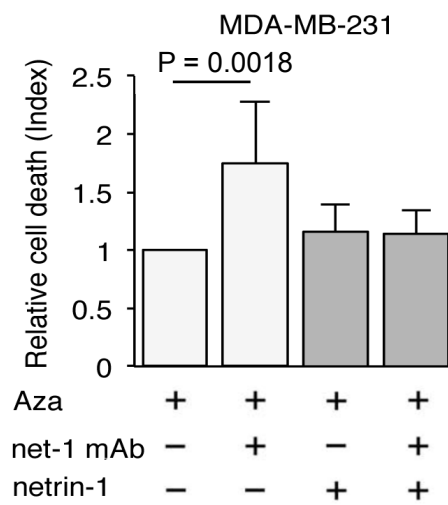
B



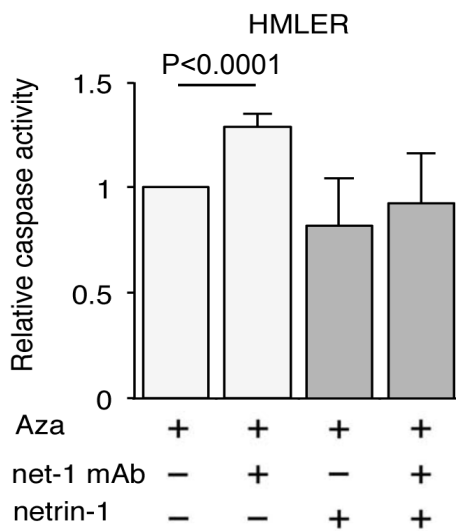
C



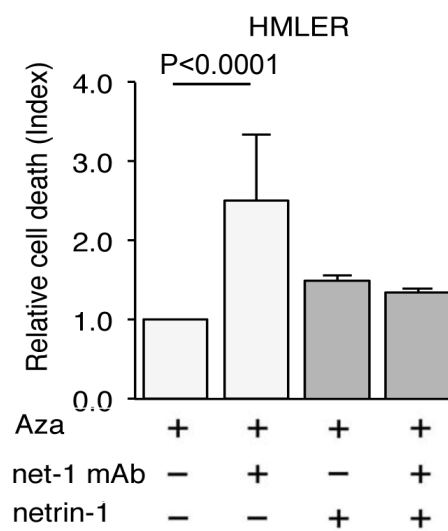
D

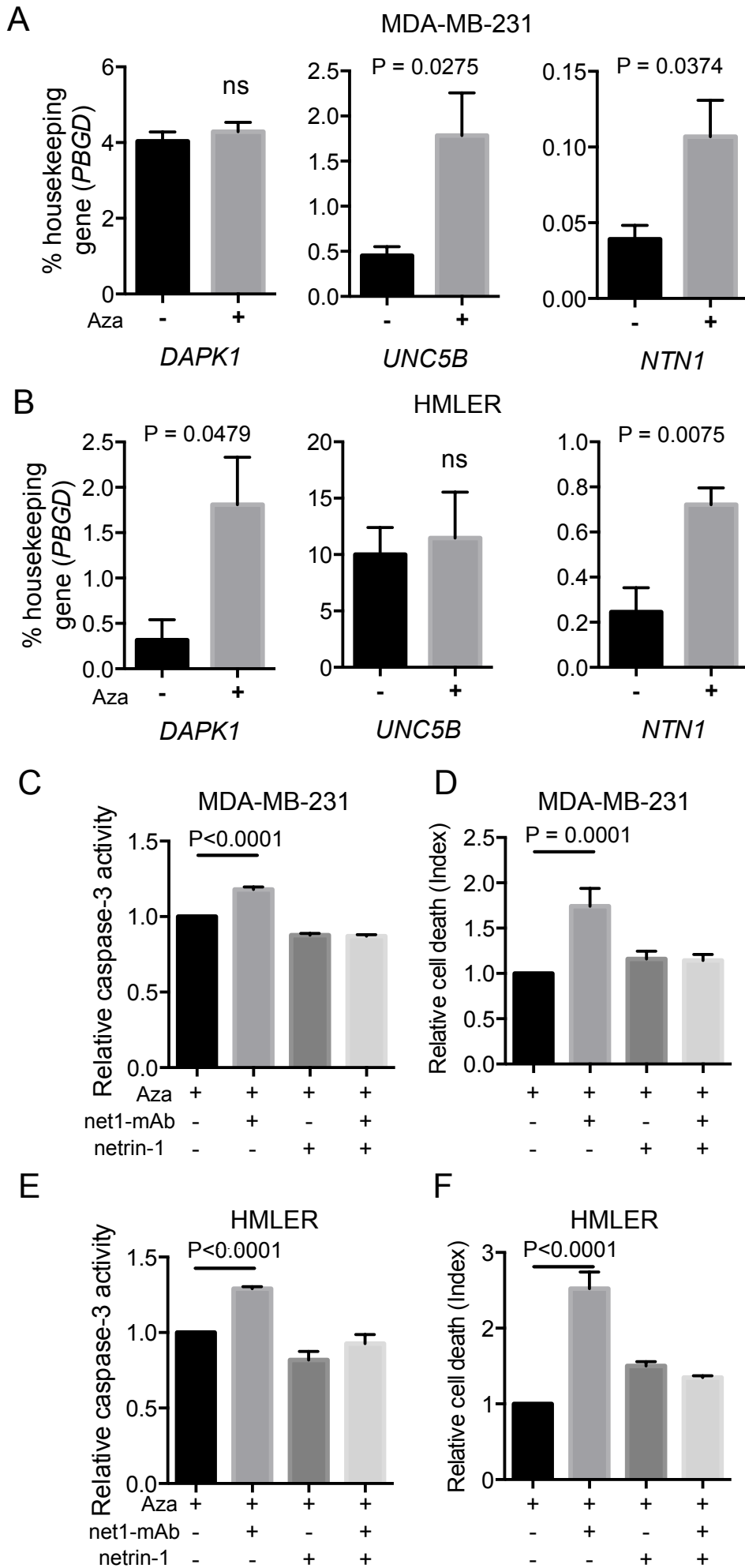


E

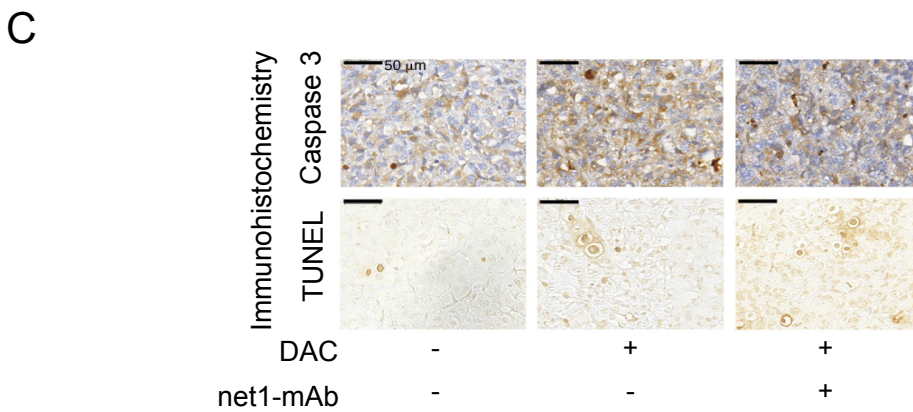
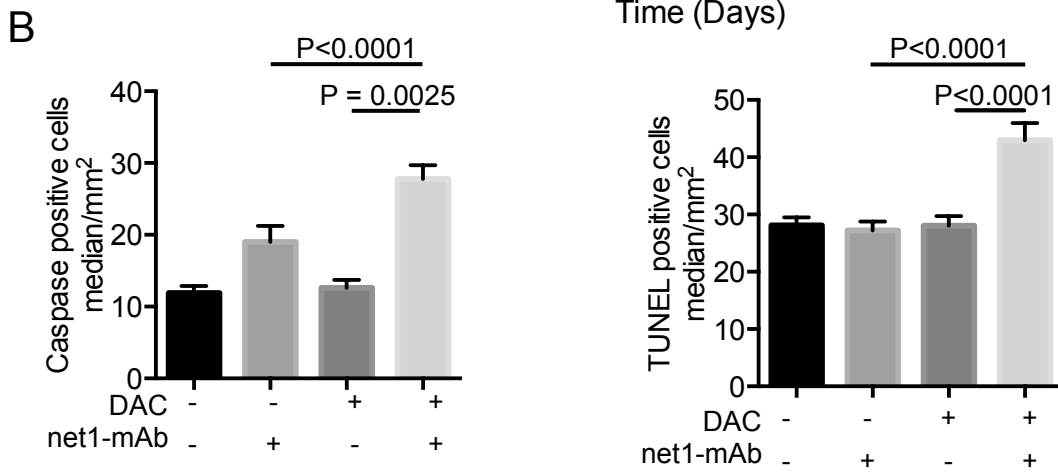
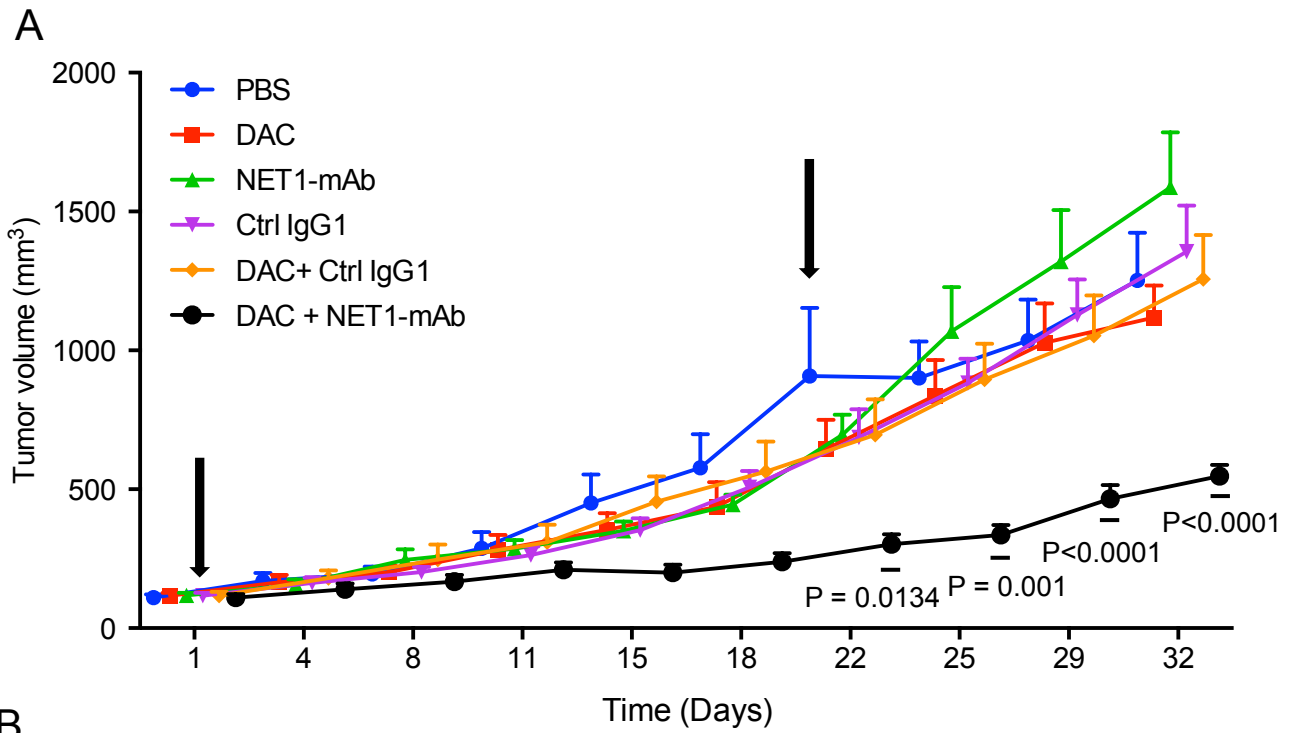


F

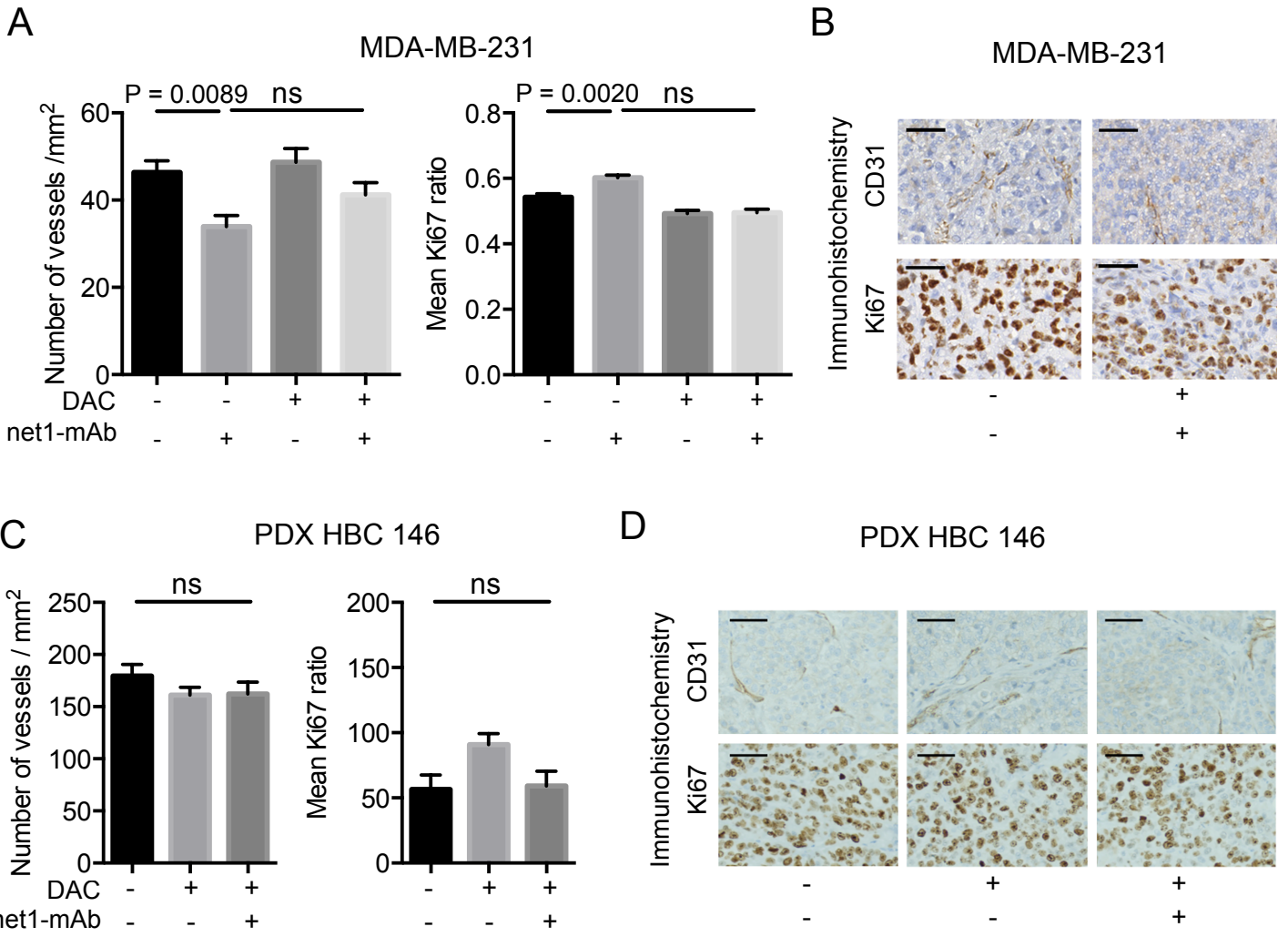




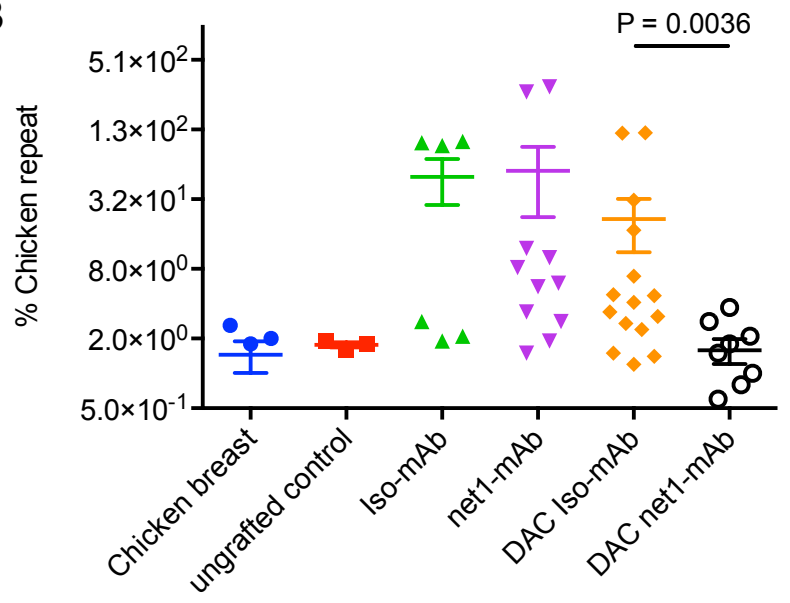
Appendix Figure S1. Grandin et al.



Appendix Figure S2. Grandin et al.

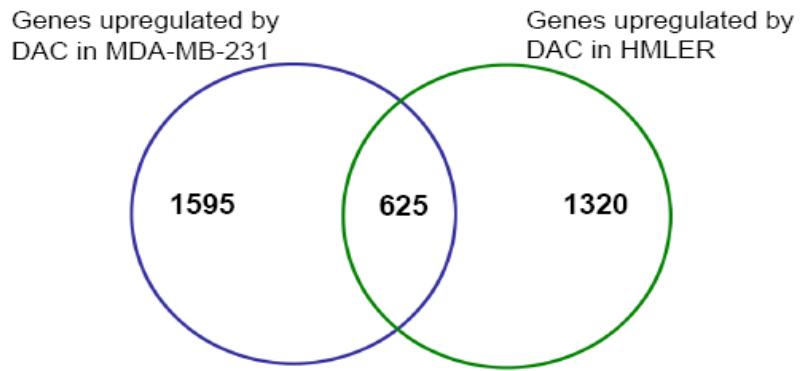


Appendix Figure S3. Grandin et al.

A**B**

Appendix Figure S4. Grandin et al.

A



B

Set	Rank	Name	p value	ajusted p value
KEGG pathways	1	Osteoclast differentiation	2.40E-07	1.42E-05
	2	MAPK signaling pathway	7.50E-06	8.85E-05
	3	Rheumatoid arthritis	5.86E-06	8.85E-05
	4	Cytokine-cytokine receptor interaction	6.85E-06	8.85E-05
	5	Pathways in cancer	5.18E-06	8.85E-05
	6	NOD-like receptor signaling pathway	9.69E-06	9.53E-05
	7	African trypanosomiasis	2.54E-05	2.00E-04
	8	Leishmaniasis	2.80E-05	2.00E-04
	9	Type I diabetes mellitus	5.81E-05	4.00E-04
	10	Malaria	1.00E-04	6.00E-04
Gene Ontology	1	Response to molecule of bacterial origin	1.06E-06	0.0014
	2	Response to lipid	1.84E-05	0.0046
	3	Response to bacterium	2.76E-05	0.0046
	4	Regulation of cell proliferation	2.43E-05	0.0046
	5	Immune response	3.27E-05	0.0048
	6	Cell proliferation	4.70E-05	0.0053
	7	Cytokine activity	0.0003	0.0286

Appendix Figure S5. Grandin et al.

Appendix Table S1 - List of primers

Q-RT-PCR Primers	Forward (5' to 3')	Reverse (5' to 3')
<i>PBGD</i>	GAGTGATTTCGCGTGGGTACC	GGCTCCGATGGTGAAGCC
<i>GAPDH</i>	CGGAGTCAACGGATTTGGTCGTAT	AGCCTTCTCCATGGTGGTGAAGAC
<i>DAPK1</i>	GATAGAAATGTCCCCAAACCTCG	TCTTCTTGATCCTTGACCAGAA
<i>UNC5B</i>	CCCGCCACACAGATCTACTT	CAGTAATCCTCCAGCCCAA
<i>NTN1</i>	TGCAAGAAGGACTATGCCGTC	GCTCGTGCCCTGCTTATACAC
<i>MBD2</i>	CCCACAACGAATGAATGAACAGC	TGAAGACCTTTGGGTAGTTCCA
<i>UNC5A</i>	ATCACCAAGGACACAAGTTTTGC	GGCTGGAAATTATCTTCTGCCGAA
<i>UNC5C</i>	GCAAATTGCTGGCTAAATATCAGGAA	GCTCCACTGTGTTCAAGGCTAAATCTT
<i>DCC</i>	AGCCAATGGGAAAATTACTGCTTAC	AGGTTGAGATCCATGATTTGATGAG
<i>YAP</i>	CGCTCTTCAACGCCGTCA	AGTACTGGCCTGTCCGGAGT
<i>TAZ</i>	CCAGTGCCTCAGAGGTCCA	ATCTGCTGCTGGTGTGGTG
<i>CTGF</i>	CCAATGACAACGCCTCCTG	TGGTGCAGCCAGAAAGCTC
<i>IRF7</i>	ACTGTGACACCCCATCTTC	GCTGCTATCCAGGGAAGACACA
Pyrosequencing		
Amplification primers		
<i>NTN1</i>	BIO-TTATGGTTATTTATAAGTTTATGGA	TAACCCAATCCTACAACAC
<i>DAPK1</i>	BIO-TTTTTTGGATTGTGGAAATGTATAA	AACCCTAAACTACTACCTCTCCTCC
<i>GAPDH unmodified</i>	CTCTTGCTACTCTGCTCTGG	GCTAAGTTTAGCCTGCCTGG
<i>GAPDH modified</i>	GTATTTGTTGATGGGTTAAGG	ATAAAAACAAATCCCCTACCC
Sequencing primers		
NTN1-a	TAACCCAATCCTACAACAC	
NTN1-b	CAATCCTACAACAC	
DAPK1-a	AACCCTTAAATCAA	
DAPK1-b	TTCTCTATAATTTA	
DNA methylation analysis		
<i>NTN1</i>	ACACTGACGACATGGTTCTACATGAGTTGTGGTGAGTTTTTATTTT TACGGTAGCAGAGACTTGGTCTAACAACATCCTTTAATCATCTTACAAC	
<i>DAPK1</i>	ACACTGACGACATGGTTCTACAGAGGTTTTTAGTGGATATGGGATT TACGGTAGCAGAGACTTGGTCTTCCACCTCCAAAATTCAAATAATT	
Human DNA analysis in Chicken lungs		
Alu	CACCTGTAATCCCAGCACTTT	CCCAGGCTGGAGTGCAGT
Avian Repeat	ATAGAATGGCCTGGGTTGAAAAG	AAGTTTTTCACACAGAGGGTGGT
siRNA		
IRF7	CAGCCUCUAUGACGACAUC	