

## **Supplemental Material**

# **Metabolism of *myo*-inositol by *Legionella pneumophila* promotes infection of amoeba and macrophages**

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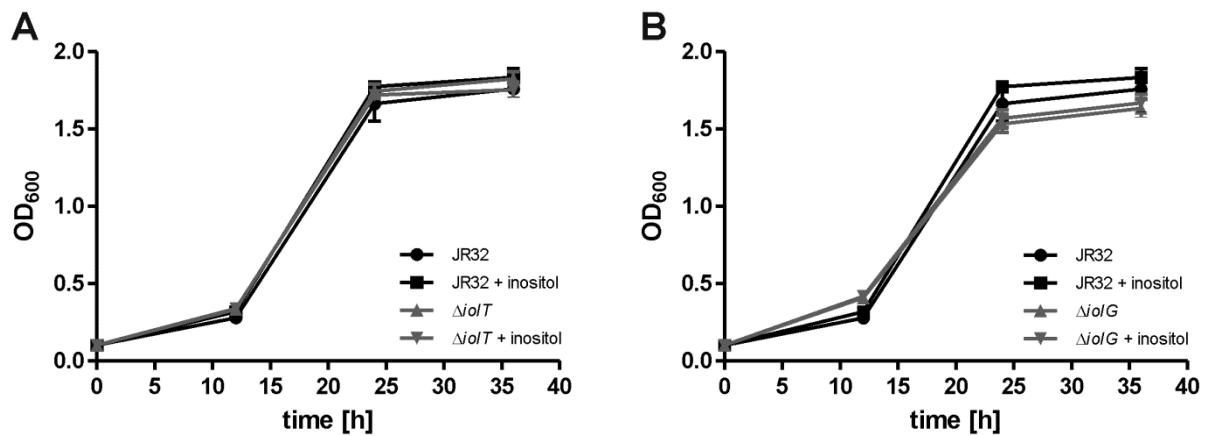
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Running title: Inositol metabolism by *L. pneumophila*

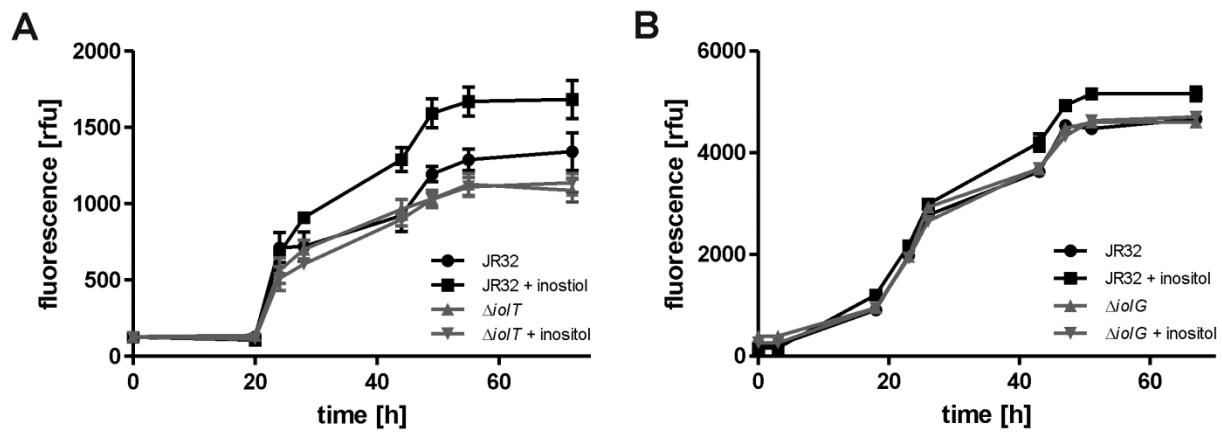
**Key words:** amoeba, bipartite metabolism, Legionnaires' disease, macrophage, nutrition, pathogen vacuole, phytate, transport, type IV secretion

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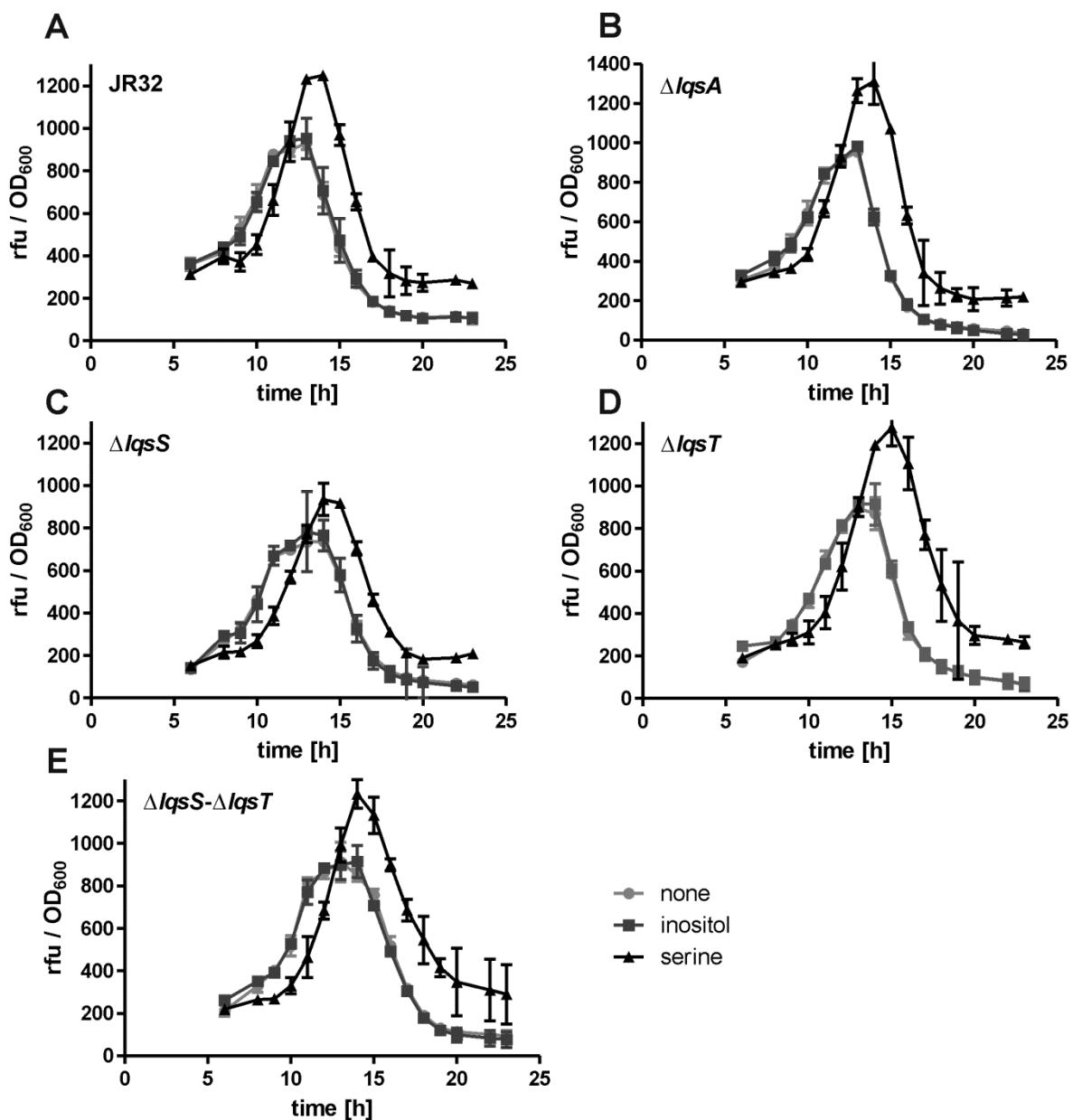
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**FIG S1** Growth of the *L. pneumophila* mutant strains  $\Delta iolT$  and  $\Delta iolG$  in CDM. Extracellular growth of *L. pneumophila* JR32,  $\Delta iolT$  or  $\Delta iolG$  in CDM with and without 10 mM inositol was assessed. Optical density at 600 nm was determined at the time points indicated.



**FIG S2** Inositol promotes intracellular growth of *L. pneumophila* at 4 h post infection. *A. castellanii* was infected (MOI 20) with *L. pneumophila* JR32, (A)  $\Delta\text{iolT}$  or (B)  $\Delta\text{iolG}$  harboring plasmid pNT28 (constitutive GFP production). 20 mM inositol was added 4 h post infection, and replication was determined by fluorescence. Mean and SD of triplicates are shown (Student's *t*-test; A, > 44 h:  $p < 0.01$ ; B, > 26 h:  $p < 0.05$ ). Data are representative of three independent experiments.



**FIG S3** Expression of  $P_{iol}$  is not regulated by  $lqs$  genes. Exponentially growing cultures of (A) *L. pneumophila* JR32, (B)  $\Delta lqsA$ , (C)  $\Delta lqsS$ , (D)  $\Delta lqsT$  or (E)  $\Delta lqsS-\Delta lqsT$  harboring plasmid pCM007 (unstable GFP (ASV) under control of  $P_{iol}$ ) were diluted to a starting OD<sub>600</sub> of 0.1 in AYE broth. The bacteria were grown at 37°C with 10 mM inositol, 6 mM serine or without additional nutrients (none). Optical density at 600 nm and GFP fluorescence was measured every hour for 24 h, and results were plotted with RFU as function of OD<sub>600</sub> over time. Mean and SD of triplicates are shown (Student's *t*-test; A-E [serine *versus* none], > 14 h:  $p < 0.05$ ). Data are representative of three independent experiments.

**TABLE S1** Oligonucleotides used in this study.

Oligonucleotide	Sequence (5' – 3'), restriction sites underlined	Description
iol-660-fo-Seq	TACTGGGGAGTAGTTGTGTGC	seq. primer of <i>iolT</i>
iol-1347-fo-Seq	TCAGTGGTACTTGATCGCA	seq. primer of <i>iolT</i>
iol-2051-fo-Seq	CATCTCGAACAAATTATTACAGCG	seq. primer of <i>iolT</i>
iol-2744-fo-Seq	GATGGACTACAAGCCTGCG	seq. primer of <i>iolT</i>
iol-3459-fo-Seq	TATGCGATTGATTGTTGGG	seq. primer of <i>iolT</i>
iol-7042-re-Seq	GAAGTCGTAAGCAATTCCGC	seq. primer of <i>iolT</i>
iol-6351-re-Seq	CCCAATCCCTCTGCGTACT	seq. primer of <i>iolT</i>
iol-5655-re-Seq	AAAGCCCATTGGAAGCTGT	seq. primer of <i>iolT</i>
iol-4942-re-Seq	TTTTGTTGGCATAGGCG	seq. primer of <i>iolT</i>
iol-4241-re-Seq	AAAATAAATTTCGTTTCCGTTTC	seq. primer of <i>iolT</i>
<i>iolG</i> -1400up-fo	ATGAACAAAGGAATGAAC	seq. primer of <i>iolG</i>
<i>iolG</i> -511-fo	ATATTATGGACATGTCT	seq. primer of <i>iolG</i>
<i>iolG</i> -451-fo	CGAATTACTCTCGTGAT	seq. primer of <i>iolG</i>
<i>iolG</i> -301-re	TATCTCTCTTCATTCAA	seq. primer of <i>iolG</i>
<i>iolG</i> -1400down-re	GTCTAACTCAAGCATATTAGA	seq. primer of <i>iolG</i>
<i>iolG</i> -850bp-fo	CCGCTGGATAATGATTAA	seq. primer of <i>iolG</i>
<i>iolG</i> -850bp-re	GATTTGCCTGTAATAGA	seq. primer of <i>iolG</i>
<i>iolG</i> -LB-XbaI-fo	GGCCAG <u>TCTAGA</u> ATAATTACAGCAGGAATTG	800 bp 5'-flanking region of <i>iolG</i>
<i>iolG</i> -LB-SalI-re	TTTCAT <u>GTCGACT</u> CCATAATTAAATTAAATTG	800 bp 5'-flanking region of <i>iolG</i>
<i>iolG</i> -RB-SalI-fo	AGTTGAG <u>TCGACT</u> ATGAAACAGTTCCCGG	800 bp 3'-flanking region of <i>iolG</i>
<i>iolG</i> -RB-XbaI-re	CGAG <u>CATCTAGA</u> TCTCCTAATTAAACAAC	800 bp 5'-flanking region of <i>iolG</i>
<i>iolT</i> -400bp-SacI-fo	GACGAT <u>GAGCT</u> TTTGTGTTCAAACGGCAGA	P <sub><i>iolT</i></sub> 400 bp for pCM007
<i>iolT</i> -400bp-XbaI-re	TCC <u>CTTCTAGA</u> GTAACTATCTGTCCCTAATG	P <sub><i>iolT</i></sub> 400 bp for pCM007

<i>iolT</i> -BamHI-fo-comp	ACAGAT <u>GGATCC</u> CATGAACAAGGGAATGAAC	<i>iolT</i> for complementation
<i>iolT</i> -SalI-re-comp	TCTTT <u>GTCGACT</u> TAATCCATAATTAAATTAAATTTC GC	<i>iolT</i> for complementation
<i>iolG</i> -BamHI-fo-comp	TATGG <u>AGGATCC</u> CATGAAAAAGAAAATATGTCGAATT GGG	<i>iolG</i> for complementation
<i>iolG</i> -SalI-re-comp	TTCAT <u>AGTCGACT</u> CAACTCAATACTACTGGCAG	<i>iolG</i> for complementation
<i>iolT</i> -LB-XbaI-fo	AATAT <u>GTCTAG</u> ACAATTGTCTTGCGAGCAGA	500 bp 5'-flanking region of <i>iolT</i>
<i>iolT</i> -LB-SalI-re	CTGTT <u>CGTCGACG</u> ATGTTATCACGGCCAGGTT	500 bp 5'-flanking region of <i>iolT</i>
<i>iolT</i> -RB-SalI-fo	TCGAAT <u>GTGACAGT</u> AGGTGCGGGAAAGAATTG	500 bp 3'-flanking region of <i>iolT</i>
<i>iolT</i> -RB-XbaI-re	TCAAGG <u>TCTAGA</u> TATTGAGGTTTGCGCCATC	500 bp 3'-flanking region of <i>iolT</i>