

# Supplementary Material

for

## Ammonia as an in-situ sanitizer: influence of virus genome type on inactivation.

Loïc Decrey<sup>a</sup>, Shinobu Kazama<sup>a,b</sup>, and Tamar Kohn<sup>a\*</sup>

<sup>a</sup>*Laboratory of Environmental Chemistry, School of Architecture, Civil and Environmental Engineering (ENAC), École Polytechnique Fédérale de Lausanne (EPFL), CH-1015 Lausanne, Switzerland*

<sup>b</sup>*New Industry Creation Hatchery Center (NICHe), Tohoku University Hatchery Square 303, 6-6-04, Aoba, Aramaki, Aoba-ku, Sendai, Miyagi, 980-0879, Japan*

\*Corresponding author

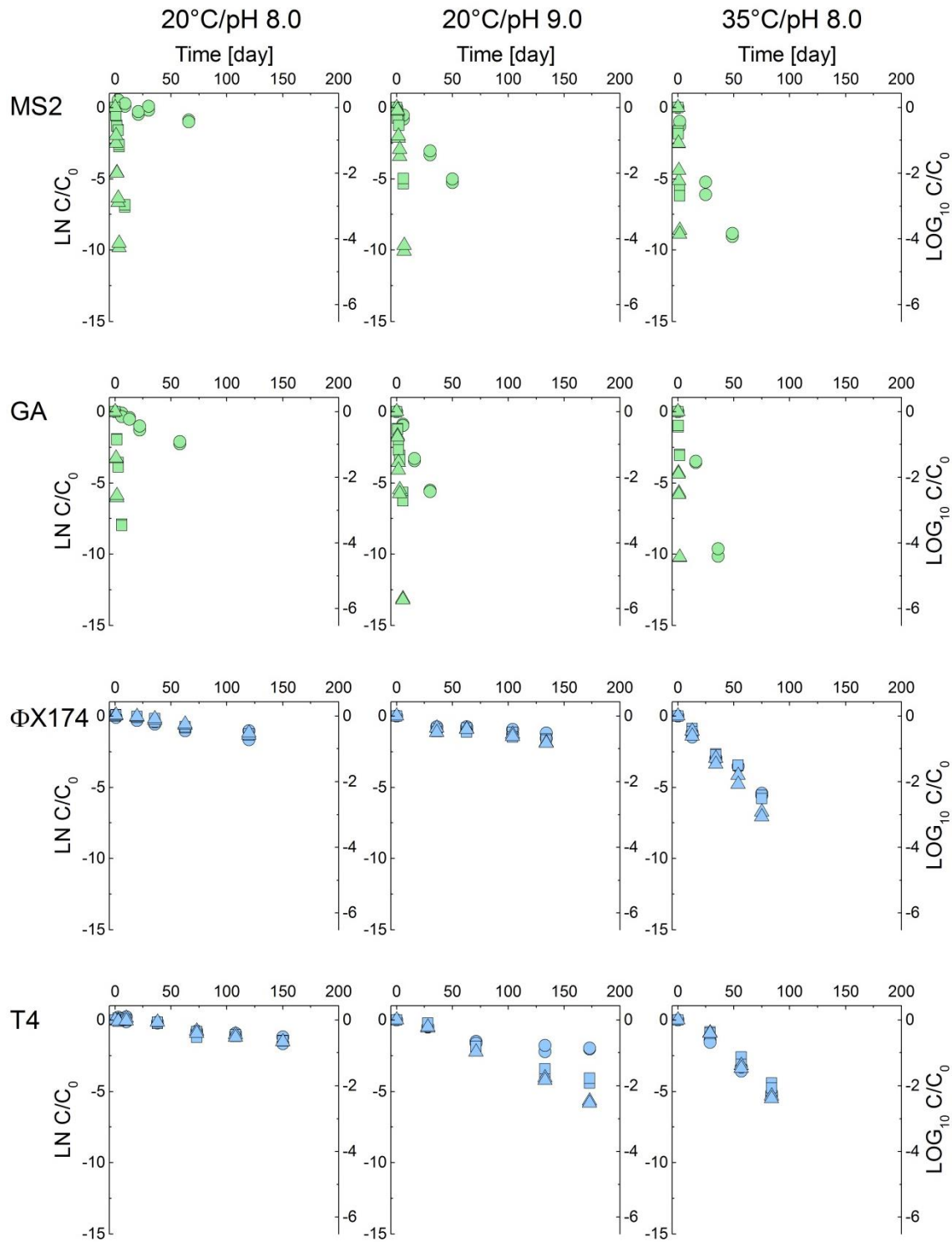
phone: +41 (0)21 693 0891; fax: +41 (0)21 693 8070;

e-mail: tamar.kohn@epfl.ch

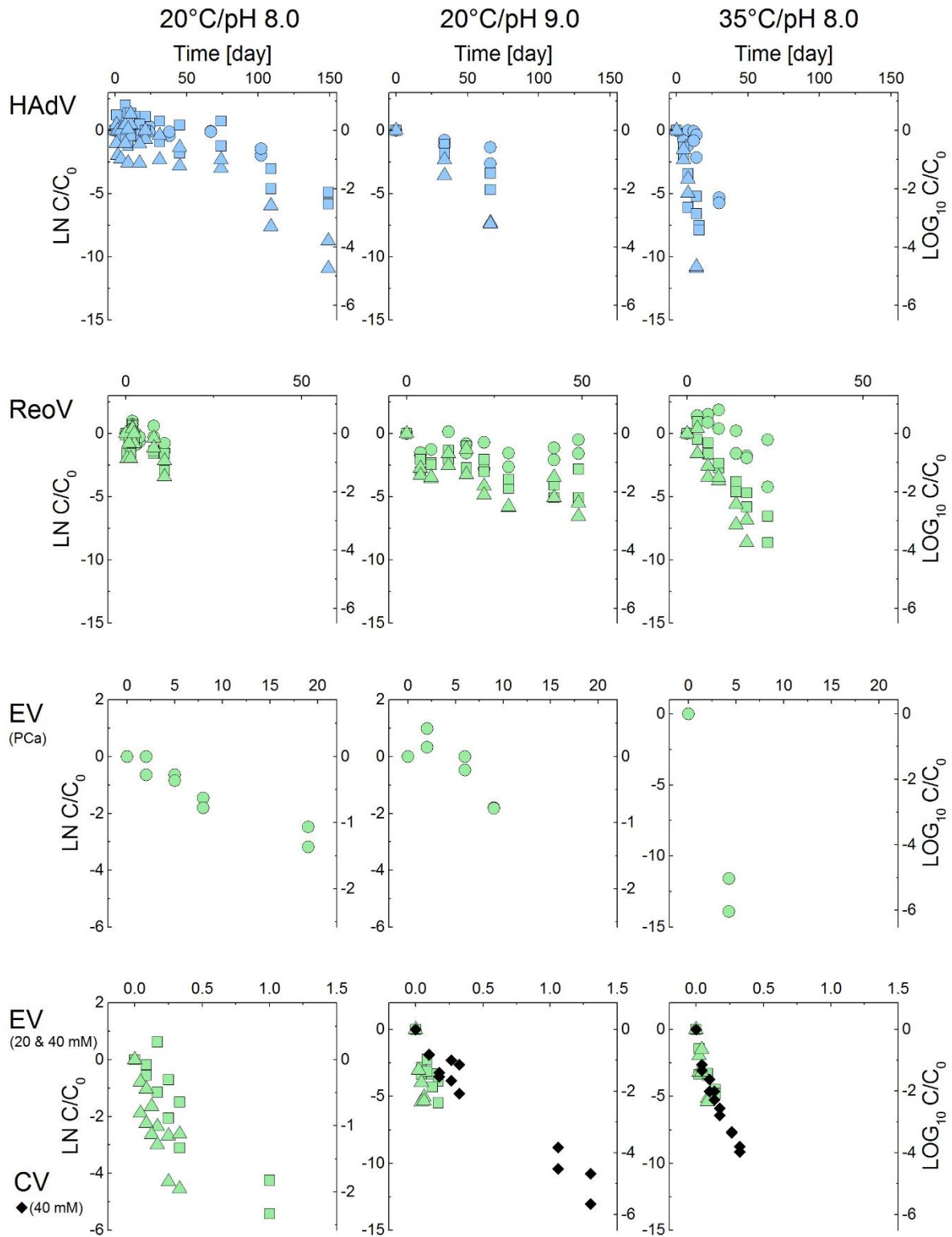
Number of pages : 9

Number of figures : 3

Number of tables : 5



**FIG S1** Kinetics of phage inactivation. Circles, squares and triangles represent PCa and AmCa with an approximate  $\{NH_3(aq)\}$  of 0, 20 and 40 mmol L<sup>-1</sup> respectively. The actual, measured  $\{NH_3(aq)\}$  is listed in Table S1.



**FIG S2 Kinetics of mammalian virus inactivation.** Circles, squares and triangles represent PCa and AmCa with an approximate  $\{\text{NH}_3(\text{aq})\}$  of 0, 20 and 40  $\text{mmol L}^{-1}$  respectively. The actual, measured  $\{\text{NH}_3(\text{aq})\}$  is listed in Table S1.

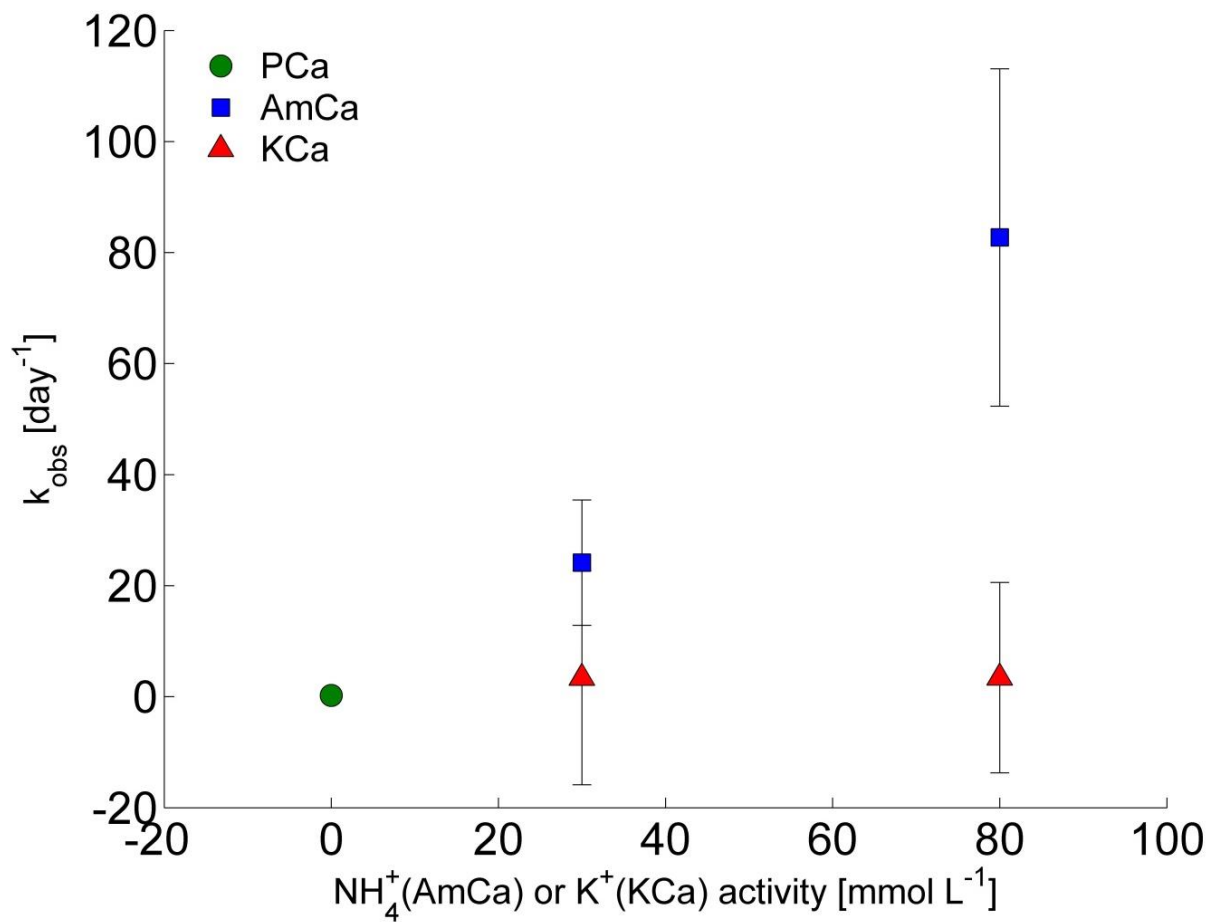


FIG S3. Influence of  $\text{NH}_4^+$  and  $\text{K}^+$  activity on EV inactivation at pH 9/20°C. The influence of  $\text{NH}_4^+$  and  $\text{K}^+$  activity were examined in AmCa and KCa buffer, respectively. The error bars depict the 95% confidence intervals associated with  $k_{\text{obs}}$ .

TABLE S1 Composition of working solutions.

T		pH <sup>a</sup>	EC [mS cm <sup>-1</sup> ]	Buffer composition (added substances)							{NH <sub>3</sub> (aq)} <sup>e</sup> [mmol L <sup>-1</sup> ]	
				Na <sub>2</sub> CO <sub>3</sub> <sup>c</sup> [mmol L <sup>-1</sup> ]	NaHCO <sub>3</sub> <sup>c</sup> [mmol L <sup>-1</sup> ]	NH <sub>4</sub> Cl <sup>b</sup> [mmol L <sup>-1</sup> ]	NaH <sub>2</sub> PO <sub>4</sub> *2H <sub>2</sub> O <sup>c</sup> [mmol L <sup>-1</sup> ]	HCl <sup>c</sup> [mmol L <sup>-1</sup> ]	NaOH <sup>c</sup> [mmol L <sup>-1</sup> ]	NaCl <sup>c</sup> [mmol L <sup>-1</sup> ]		KCl <sup>c</sup> [mmol L <sup>-1</sup> ]
20°C	PCa	7.86	10.4	50.0			60.0		5.7			0.0
	PCaH	7.87	127.0	50.0			60.0		12.0	1770.0		0.0
	AmCa 20	8.03	91.6	50.0		920.4		26.3				22.7
	AmCa 40	7.95	174.2	50.0		1976.8			4.9			37.6
20°C	PCa	8.86		50.0			45.0		1.0			0.0
	AmCa 20 <sup>d</sup>	8.93		50.0		54.9		23.5				10.8
	AmCa 40 <sup>d</sup>	8.94		50.0		147.0		1.2				28.4
	KCa 30 <sup>d</sup>	9.00		50.0				44.3			44.0	0.0
	KCa 80 <sup>d</sup>	9.00		50.0				43.8			113.0	0.0
35°C	PCa	7.92		50.0			60.0		7.0			0.0
	AmCa 20	7.98		50.0		295.1		28.1				19.5
	AmCa 40	8.03		50.0		652.9		5.6				44.3
50°C	PCa	8.00		50.0			60.0		8.8			0.0
60°C	PCa	8.00		50.0			60.0		10.6			0.0
20°C		10.00	0.45	4.5	5.5					10.0		0.0
20°C		11.00	0.57	9.8	0.2					10.0		0.0
20°C		11.50	0.65				10.0		15.1	10.0		0.0
20°C		12.00	0.95				10.0		24.0	10.0		0.0

<sup>a</sup> Measured at experimental temperature.

<sup>b</sup> Total ammonia measured by IC (see experimental method).

<sup>c</sup> Concentration determined from the amount of the compound added at the beginning of the experiment. The concentration was calculated with PHREEQC (Pitzer database), see material and methods.

<sup>d</sup> Correspond respectively to NH<sub>4</sub><sup>+</sup> activity of 30 and 80 mM and K<sup>+</sup> activity of 30 and 80 mM.

<sup>e</sup> NH<sub>3</sub> activities determined from the buffer composition (see material and method).

**TABLE S2 Kinetic parameters associated with phage inactivation:  $k_{obs}$  and coefficient of determination ( $R^2$ ).**

Phage	pH	T		$k_{obs}$ [day <sup>-1</sup> ]	IC 95%	$R^2$
MS2	8	20°C	PCa	0.02	0.01	0.75
			PCaH	0.11	0.01	0.98
			AmCa 20	0.78	0.05	0.99
			AmCa 40	2.50	0.23	0.99
	9	20°C	PCa	0.10	0.01	0.99
			AmCa 20	0.86	0.11	0.98
			AmCa 40	1.47	0.14	0.99
	8	35°C	PCa	0.18	0.03	0.98
			AmCa 20	3.18	0.32	0.99
			AmCa 40	5.32	0.58	0.99
	8	50°C	PCa	5.51	0.34	0.99
			60°C	PCa	294.01	42.69
	10	20°C		0.44	0.06	0.98
	11	20°C		5.16	0.23	0.99
	11.5	20°C		37.76	4.12	0.98
12	20°C		11623	5384	0.99	
GA	8	20°C	PCa	0.04	0.01	0.96
			PCaH	0.14	0.01	0.99
			AmCa 20	1.35	0.13	0.99
			AmCa 40	3.33	0.08	0.99
	9	20°C	PCa	0.19	0.02	0.99
			AmCa 20	1.02	0.11	0.98
			AmCa 40	2.30	0.14	0.99
	8	35°C	PCa	0.28	0.04	0.99
			AmCa 20	1.84	0.19	0.99
			AmCa 40	6.10	0.22	0.99
PhiX174	8	20°C	PCa	0.0114	0.0029	0.88
			PCaH	0.0115	0.0012	0.98
			AmCa 20	0.0117	0.0016	0.96
			AmCa 40	0.0110	0.0013	0.97
	9	20°C	PCa	0.0088	0.0034	0.81
			AmCa 20	0.0108	0.0044	0.79
			AmCa 40	0.0124	0.0044	0.84
	8	35°C	PCa	0.0686	0.0094	0.97
			AmCa 20	0.0726	0.0078	0.98
			AmCa 40	0.0893	0.0085	0.99
	8	50°C	PCa	1.1280	0.1548	0.97
			60°C	PCa	265.62	26.25
	11	20°C		0.0101	0.0068	0.67
	12	20°C		0.0669	0.0128	0.93
	T4	8	20°C	PCa	0.0104	0.0018
PCaH				0.0079	0.0023	0.83
AmCa 20				0.0106	0.0018	0.93
AmCa 40				0.0108	0.0014	0.96
9		20°C	PCa	0.0119	0.0038	0.86
			AmCa 20	0.0263	0.0030	0.98
			AmCa 40	0.0336	0.0021	0.99
8		35°C	PCa	0.0628	0.0049	0.99
			AmCa 20	0.0560	0.0097	0.97
			AmCa 40	0.0663	0.0118	0.97
8		50°C	PCa	0.8331	0.0883	0.98
8		60°C	PCa	196.93	35.54	0.95
10		20°C		0.0315	0.0067	0.93
11		20°C		0.9625	0.1295	0.99
11.5		20°C		22198	14815	0.98

**TABLE S3 Kinetic parameters associated with mammalian virus inactivation:  $k_{obs}$  and coefficient of determination ( $R^2$ ).**

Virus	pH	T		$k_{obs}$ [day <sup>-1</sup> ]	IC 95%	$R^2$	
HAdV	8	20°C	PCa	0.018	0.008	0.62	
			PCaH	0.003 *	0.007	0.04	
			AmCa 20	0.037	0.009	0.71	
			AmCa 40	0.060	0.011	0.83	
	9	20°C	PCa	0.030	0.019	0.80	
			AmCa 20	0.061	0.025	0.91	
			AmCa 40	0.111	0.026	0.97	
	8	35°C	PCa	0.185	0.056	0.81	
			AmCa 20	0.468	0.120	0.90	
			AmCa 40	0.786	0.205	0.94	
	ReoV	8	20°C	PCa	0.040 *	0.123	0.04
				PCaH	0.048 *	0.124	0.05
AmCa 20				0.186	0.103	0.52	
AmCa 40				0.174	0.131	0.37	
9		20°C	PCa	0.015 *	0.028	0.09	
			AmCa 20	0.069	0.031	0.59	
			AmCa 40	0.089	0.040	0.58	
8		35°C	PCa	0.154	0.085	0.56	
			AmCa 20	0.352	0.041	0.97	
			AmCa 40	0.470	0.067	0.96	
EV		8	20°C	PCa	0.15	0.03	0.92
				PCaH	0.25	0.06	0.91
	AmCa 20			4.97	1.51	0.84	
	AmCa 40			10.24	4.08	0.71	
	9	20°C	PCa	0.22	0.15	0.66	
			AmCa 20	24.15	11.28	0.75	
			AmCa 40	82.72	30.38	0.87	
	8	35°C	PCa	3.00	0.87	0.98	
			AmCa 20	27.73	13.98	0.79	
			AmCa 40	57.78	24.31	0.84	
	CV	9	20°C	AmCa 40	8.40	1.19	0.95
		8	35°C	AmCa 40	25.31	3.89	0.94

\* not significantly different from zero.

**TABLE S4 Kinetic parameters associated with virus and phage inactivation:  $k_{\text{NH}_3}$  and coefficient of determination ( $R^2$ ).**

Phage/Virus	pH	T	$k_{\text{NH}_3}^b$ [day <sup>-1</sup> M <sup>-1</sup> ]	IC 95%	$R^2$
MS2	8	20°C	63.4	38.3	0.89
	9	20°C	46.7	16.6	0.96
	8	35°C	114.8	82.7	0.97
GA	8	20°C	85.2	35.2	0.95
	9	20°C	74.3	1.4	0.99
	8	35°C	133.3	110.8	0.96
PhiX174	8	20°C	-0.009 *	0.031	0.20
	9	20°C	0.122	0.041	0.96
	8	35°C	0.480 *	1.690	0.93
T4	8	20°C	0.009	0.001	0.99
	9	20°C	0.729	0.423	0.90
	8	35°C	0.090 *	0.930	0.16
HAdV	8	20°C	1.1	0.4	0.97
	9	20°C	2.9	0.0	1.00
	8	35°C	13.6	2.1	0.99
ReoV	8	20°C	3.8	3.4	0.79
	9	20°C	2.4	1.8	0.85
	8	35°C	7.0	6.6	0.95
EV	8	20°C	264.0	66.3	0.98
	9	20°C	2944.3	519.7	0.99
	8	35°C	1236.1	72.8	0.99
f2 <sup>a</sup>	8	20°C	35.3	5.4	0.99
Poliovirus <sup>a</sup>	8	20°C	235.0	91.3	0.97

<sup>a</sup> Determined from Burge et al. (1983) data.

<sup>b</sup> According to Eq.2 (see material and methods).

\* Not significantly different from zero.



**TABLE S5 Comparison of DNA, RNA and protein cleavage rate constants.**

<b>Spontaneous DNA hydrolysis (Gates, 2009)</b>				
<b>A</b>	<b>Spontaneous hydrolysis of phosphodiester backbone (Schroeder, 2006)</b>			
			[day <sup>-1</sup> ]	
	pH 7.0	25°C	8.64E-11	
<b>B</b>	<b>Hydrolytic deamination of DNA base --&gt; mutagenesis, no evidence of cleavage (Gates, 2009)</b>			
			[day <sup>-1</sup> ]	
	dsDNA	pH 7.4	37°C	3.79E-08
	ssDNA	pH 7.4	37°C	9.49E-06
<b>C Hydrolysis of phosphodiester backbone through abasic site (C1 and C2 are the rate limiting steps)</b>				
<i>C1.a Depurination/depyrimidation (Lindhal, 1972 &amp; 1973) --&gt; acid-catalyzed reaction</i>				
			[day <sup>-1</sup> ]	
	pyrimidine (CT)	pH 7.4	37°C	1.30E-07
	purine (AG)	pH 7.4	37°C	2.59E-06
<i>C1.b Depurination (An, 2014) --&gt; acid-catalyzed reaction</i>				
			[day <sup>-1</sup> ]	
	Adenine	pH 7.1	37°C	1.90E-05
	Guanine	pH 7.1	37°C	2.16E-05
<i>C2 Self catalyzed (site specific) DNA depurination (Amosova, 2005)</i>				
			[day <sup>-1</sup> ]	
		pH 7.0	37°C	1.01E-02
<i>C3 Chain breakage at abasic site (alkaline-catalyzed reaction)(Gates, 2009)(Küpfer, 2007)</i>				
			[day <sup>-1</sup> ]	
		pH 7.4	37°C	1.20E-02-1.25E-01
	in 0.1 M NaOH			2.61E+02
<b>Spontaneous RNA hydrolysis (Li, 1999)</b>				
			[day <sup>-1</sup> ]	
		pH 7.0	37°C	1.73E-04
		pH 11.0	37°C	1.48E+00
<b>Spontaneous peptide bond hydrolysis (Smith, 1998)</b>				
			[day <sup>-1</sup> ]	
		pH 7.0	37°C	4.02E-06
		pH 11.0	37°C	1.49E-04
<b>Protein denaturation</b>				
<i>Ricin denaturation (Levy, 1950)</i>				
			[day <sup>-1</sup> ]	
		pH 11.0	37°C	5.97E-01
<i>Penicillin Acylase denaturation (Guranda, 2004)</i>				
			[day <sup>-1</sup> ]	
		pH 11.0	37°C	4.67E+03

(1–12)

## References

1. **Burge WD, Cramer WN, Kawata K.** 1983. Effect of Heat on Virus Inactivation by Ammonia. *Appl Environ Microbiol* **46**:446–451.
2. **Schroeder GK, Lad C, Wyman P, Williams NH, Wolfenden R.** 2006. The time required for water attack at the phosphorus atom of simple phosphodiester and of DNA. *Proc Natl Acad Sci U S A* **103**:4052–4055.
3. **Gates KS.** 2009. An overview of chemical processes that damage cellular DNA: spontaneous hydrolysis, alkylation, and reactions with radicals. *Chem Res Toxicol* **22**:1747–60.
4. **Lindahl T, Nyberg B.** 1972. Rate of depurination of native deoxyribonucleic acid. *Biochemistry* **11**:3610–3618.
5. **Lindahl T, Karlström O.** 1973. Heat-induced depyrimidination of deoxyribonucleic acid in neutral solution. *Biochemistry* **12**:5151–5154.
6. **An R, Jia Y, Wan B, Zhang Y, Dong P, Li J, Liang X.** 2014. Non-enzymatic depurination of nucleic acids: factors and mechanisms. *PLoS One* **9**:e115950.
7. **Amosova O, Coulter R, Fresco JR.** 2006. Self-catalyzed site-specific depurination of guanine residues within gene sequences. *Proc Natl Acad Sci U S A* **103**:4392–4397.
8. **Küpfer PA, Leumann CJ.** 2007. The chemical stability of abasic RNA compared to abasic DNA. *Nucleic Acids Res* **35**:58–68.
9. **Li Y, Breaker RR.** 1999. Kinetics of RNA Degradation by Specific Base Catalysis of Transesterification Involving the 2'-Hydroxyl Group. *J Am Chem Soc* **121**:5364–5372.
10. **Smith RM, Hansen DE.** 1998. The pH-Rate Profile for the Hydrolysis of a Peptide Bond. *J Am Chem Soc* **120**:8910–8913.
11. **Levy M, Benaglia AE.** 1950. The influence of temperature and pH upon the rate of denaturation of ricin. *J Biol Chem* **186**:829–47.
12. **Guranda DT, Volovik TS, Švedas VK.** 2004. pH stability of penicillin acylase from *Escherichia coli*. *Biochem* **69**:1386–1390.