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Table S1
Enterobacteriaceae tailed phage MCP matches in bacterial genomes

Cluster	Prototype phage	Prototype phage host	Bacterial genera / authentic phage with >50% MCP matches outside the <i>Enterobacteriaceae</i> ¹
Lytic 1	T1	<i>Escherichia</i>	None / none
Lytic 2	T4	<i>Escherichia</i>	None / <i>Acinetobacter</i> phage Acj9-75%
Lytic 3	Vi01	<i>Salmonella</i>	None / <i>Delftia</i> phage øW-14-54%
Lytic 4	T5	<i>Escherichia</i>	None / <i>Vibrio</i> phage pVp-1-60%
Supercluster	T7		
Lytic 5	T7	<i>Escherichia</i>	None / <i>Stenotrophomonas</i> phage IME15 82%
Lytic 6	SP6	<i>Salmonella</i>	None / none
Lytic 7	KP34	<i>Klebsiella</i>	None / <i>Vibrio</i> phage VP93-74%
Lytic 8	LIMEzero	<i>Pantoea</i>	None / <i>Ralstonia</i> phage RSB3-51%
Lytic 9	øKT	<i>Escherichia</i>	None / none
Lytic 10	GAP227	<i>Cronobacter</i>	None / <i>Aeromonas</i> phage øAS7-82%
Lytic 11	N4	<i>Escherichia</i>	<i>Mezorhizobium</i> -75% / <i>Delftia</i> phage RG-2014-62%
Lytic 12	9NA	<i>Salmonella</i>	None / none
Lytic 13	Chi	<i>Salmonella</i>	None / none
Lytic 14	øEco32	<i>Escherichia</i>	None / none
Lytic 15	Felix-O1	<i>Salmonella</i>	None / none
Supercluster	SETP3		
Lytic 16	SETP3	<i>Salmonella</i>	<i>Acinetobacter</i> -64% / none
	K1-dep(1)	<i>Escherichia</i>	<i>Sinorhizobium</i> -61% / <i>Synechococcus</i> phage S-CBS4-56%
Lytic 17	SO-1	<i>Sodalis</i>	None / none
Lytic 18	ECO1230-10	<i>Escherichia</i>	<i>Marinobacter</i> -60% / <i>Pseudomonas</i> phage PPpW-3-68%
Lytic 19	Gj1	<i>Escherichia</i>	<i>Aeromonas</i> -60% / <i>Shewanella</i> phage Spp001-56%
Lytic 20	PY100	<i>Yersinia</i>	None / none
Supercluster	rV5		
Lytic 21	Ø92	<i>Escherichia</i>	None / none
Lytic 22	rV5	<i>Escherichia</i>	None / none
Lytic 23	SPN3US	<i>Salmonella</i>	None / none
Lytic 24	RaK2	<i>Klebsiella</i>	None / none
Lytic 25	øR1-37	<i>Yersinia</i>	None / none
Lytic 26	E1	<i>Salmonella</i>	None / <i>Vibrio</i> phage pYD38-A-79%
Lytic 27	EMCL-117	<i>Escherichia</i>	<i>Burkholderia</i> -52% / <i>Pseudomonas</i> phage DL60-67% ^c
Lytic 28	KF-1	<i>Edwardsiella</i>	None / <i>Vibrio</i> phage VPMS1-69%
Lytic 29	MSW-3	<i>Edwardsiella</i>	None / <i>Iodobacter</i> phage øPLPE-68%
Lytic 30	Ea35-70	<i>Erwinia</i>	None / none
Lytic 31	øEaH1	<i>Erwinia</i>	None / none
Lytic 32	9g	<i>Escherichia</i>	None / none
Lytic 33	IME-EC2	<i>Escherichia</i>	None / none
Lytic 34	Ss1	<i>Cronobacter</i>	None / none
Lytic 35	CVT22	<i>Citrobacter</i>	None / i phage CW02-63%
Supercluster	Lambda		
Temperate 1	Lambda	<i>Escherichia</i>	None / none
Temperate 2 & 3	ø80 & N15	<i>Escherichia</i>	None / none
	PY54	<i>Yersinia</i>	<i>Vibrio</i> -76% / none
Temperate 4	HK97	<i>Escherichia</i>	<i>Pseudomonas</i> 78% / none
	mEp235	<i>Escherichia</i>	<i>Pseudomonas</i> -80% / none
Temperate 5	ES18	<i>Salmonella</i>	<i>Pseudomonas</i> -69% / <i>Pseudomonas</i> phage ø297-69%
Temperate 6	Gifsy-2	<i>Salmonella</i>	<i>Photobacterium</i> -57% / <i>Pseudomonas</i> phage F10-55%
Temperate 7	BP-4795	<i>Escherichia</i>	None, none
Temperate 8	SfV	<i>Shigella</i>	<i>Aggregatibacter</i> -64% / none
	SfI	<i>Shigella</i>	<i>Vibrio</i> -84% / none
Temperate 9	P22	<i>Salmonella</i>	None / none
	Sf6	<i>Shigella</i>	None / none
Temperate 10	APSE-1	<i>Hamiltonella</i>	None / none
Temperate 11	933W	<i>Escherichia</i>	<i>Photobacterium</i> -63%, none

Table S1 cont.

Cluster	Prototype phage	Prototype phage host	Bacterial genera / phage with >50% MCP matches ¹ outside <i>Enterobacteriaceae</i>
Temperate 12	HK639	<i>Escherichia</i>	<i>Castellaniella</i> -55% / none
Temperate 13	øES15	<i>Cronobacter</i>	None / none
Temperate 14	HS2*	<i>Sodalis</i>	<i>Acinetobacter</i> -77% / none
Temperate 15	ENT47670	<i>Cronobacter</i>	<i>Vibrio</i> -77% / none
Temperate 16	ZF40	<i>Pectobacterium</i>	None / none
Temperate 17	øEt88	<i>Erwinia</i>	None / <i>Xanthomonas</i> phage DIBBI-53%
Temperate 18	ε15	<i>Salmonella</i>	<i>Pseudomonas</i> -65% / none
Temperate 19	P1	<i>Escherichia</i>	None / none
Supercluster	P2		
Temperate 20	P2	<i>Escherichia</i>	<i>Burkholderia</i> -55% / <i>Burkholderia</i> phage-55%
Temperate 21	ESSI-2	<i>Cronobacter</i>	<i>Vibrio</i> -59% / <i>Vibrio</i> phage-53%
Temperate 22	Mu	<i>Escherichia</i>	<i>Marinomonas</i> -62% / <i>Pseudomonas</i> phage DMS3-51%
Temperate 23	SSU5	<i>Salmonella</i>	<i>Acinetobacter</i> -60% / none
Temperate 24	PEp14	<i>Erwinia</i>	<i>Rhizobium</i> -66% / <i>Sinorhizobium</i> phage PBC5 phage-66%
Temperate 25	GF-2	<i>Edwardsiella</i>	None / none
Temperate 26	SEN34	<i>Salmonella</i>	<i>Pseudomonas</i> -59% / none

- Percent MCP identity as reported by BLASTp
 - Red text highlights prophages in other bacterial families with MCPs >75% identical to MCPs of a temperate *Enterobacteriaceae* phage cluster.
 - Blue text highlights prophages in other bacterial families with MCPs between 50% and 70% identical to MCPs of a lytic *Enterobacteriaceae* phage cluster.
- Several phages in the EMCL-117-like phage group, for example phage øVader, have recently been reported to grow lytically on both *E. coli* and *Pseudomonas aeruginosa* (Malki *et al.*, 2015).

Table S2
P2 Supercluster Phages and Prophages**A** *Enterobacteriaceae* P2-like phages in this study

Cluster	Phage	Host species	Accession number	Genome orientation	Genome sequence publication
Temperate 20	P2	<i>Escherichia coli</i>	AF063097	P2	numerous publications
	W ϕ	<i>Escherichia coli</i>	AY135739	P2	–
	L-413C	<i>Yersinia pestis</i>	AY251033	P2	Virology 372:85
	ϕ AA91-ss	<i>Escherichia coli</i>	KF322032	P2	JVirol 87:13665
	pro147	<i>Escherichia coli</i>	KR073660	P2	–
	pro483	<i>Escherichia coli</i>	KR073661	P2	–
	186	<i>Escherichia coli</i>	U32222	P2	numerous publications
	PsP3	<i>Salmonella enterica</i>	NC_005340	P2	–
	FSL-SP-004	<i>Salmonella enterica</i>	KC139521	P2	BMCgenomics 14:481
	ENT90	<i>Erwinia amylovora</i>	HQ110084	P2	–
	SopE ϕ	<i>Salmonella enterica</i>	AY319521	P2	JBact 185:5182
	Fels-2	<i>Salmonella enterica</i>	NC_010463	P2	Nature 413:852
	RE-2010 (EL ϕ S)	<i>Salmonella enterica</i>	HM770079	P2	ApplEnvironMicro 78:1785
	SEN1	<i>Salmonella enterica</i>	KT630644	P2	–
	SEN4	<i>Salmonella enterica</i>	KT630645	P2	–
	SEN5	<i>Salmonella enterica</i>	KT630646	P2	–
	SEN8	<i>Salmonella enterica</i>	KT630647	P2	–
	P88	<i>Escherichia coli</i>	NC_026014	P2	–
Temperate 21	ESSI-2	<i>Cronobacter sakazakii</i>	HQ110083	HP1	Arc hVirol 156:721

Table S2
P2 Supercluster Phages and Prophages

B P2-like Enterobacteriaceae prophage panel used in this study

Prophage	Host species and strain	MCP locus_tag
AgonaP2-A	<i>Salmonella enterica</i> (Agona73.H.09)	SA73_2208
CedP2-1	<i>Cedecea davisae</i> DSM 4568	HMPREF0201_03602
CronoP2-B	<i>Cronobacter sakazakii</i> SP291	CSSP291_11880
DickP2-B	<i>Dickeya zeae</i> EC1	W909_12715
Eco541P2-A	<i>Escherichia coli</i> 541-15	EC54115_23648
EcoC3P2-A*	<i>Escherichia coli</i> 2-011-08_S4_C3	AD24_4183
EcoP2-CVM-A	<i>Escherichia coli</i> CVM N36225PS	PL46_04800
EcoP2-E22-A	<i>Escherichia coli</i> E22	E6E22_1442
EcoP2-KTE10-A	<i>Escherichia coli</i> KTE10	WCM_01332
EcoP2-KTE234-A	<i>Escherichia coli</i> KTE234	A193_03379
EdwardP2-A	<i>Edwardsiella tarda</i> ATCC 15947	ET1_14_00560
EdwardP2-B	<i>Edwardsiella piscicida</i> C07-87	ETAC_02100
EntP2-A	<i>Enterobacter cloacae</i> ATCC 13047	ECL_02851
ErwinP2-A	<i>Erwinia billingiae</i> Eb661	Ebc_01060
HafP2-A	<i>Hafnia alvei</i> FB1	AT03_17965
KlebP2-A	<i>Klebsiella pneumoniae</i> UHKPC57	H237_1271
MorgP2-A	<i>Morganella morganii</i> KT	MU9_2040
PantP2-A	<i>Pantoea rwandensis</i> ND04	LH22_RS05065
PectP2-C	<i>Pectobacterium carotovorum</i> PC1	PC1_2643
ProdP2-A	<i>Pantoea rodasi</i> ND03	QU24_21505
ProvP2-B	<i>Providencia rustigianii</i> DSM 4541	PROVRUST_04967
RahnP2-B	<i>Rahnella aquatilis</i> HX2	Q7S_07135
RauoIP2-A	<i>Raoultella ornithinolytica</i> TNT	DF41_23110
SalP2-C	<i>Salmonella enterica</i> Typhimurim YUHS 05-78	SE14_01921
SerrP2-C	<i>Serratia odorifer</i> DSM 4582	HMPREF0768_1209
ThompP2-A	<i>Salmonella enterica</i> (Thompson CFSAN000736)	SEETH391_21320
XenoP2-A	<i>Xenorhabdus doucetiae</i> FRM16	XDD1_2854
YersP2-A	<i>Yersinia pseudotuberculosis</i> YP111	YPK_0899
YersP2-B	<i>Yersinia enterocolitica</i> Y11	Y11_25391
YokP2-A	<i>Yokenella regensburgeri</i> ATCC 43003	HMPREF0880_02763

Table S2
P2 Supercluster Phages and Prophages

C P2-like phages outside the *Enterobacteriaceae*

Phage	Host species	Host Family	Class	MCP locus_tag	Genome sequence publication
BEK	<i>Burkholderia pseudomallei</i>	Burkholderiaceae	Beta-Proteobacteria	CP008753	–
KL3	<i>Burkholderia cepacia</i>	Burkholderiaceae	Beta-Proteobacteria	GU911304	BMCGenomics 11:599
KS14	<i>Burkholderia cenocepacia</i>	Burkholderiaceae	Beta-Proteobacteria	NC_015273	BMCGenomics 11:599
KS5	<i>Burkholderia cenocepacia</i>	Burkholderiaceae	Beta-Proteobacteria	NC_015265	BMCGenomics 11:599
ø52237	<i>Burkholderia pseudomallei</i>	Burkholderiaceae	Beta-Proteobacteria	NC_007145	–
øE12-2	<i>Burkholderia pseudomallei</i>	Burkholderiaceae	Beta-Proteobacteria	NC_009236	–
øE202	<i>Burkholderia thailandensis</i>	Burkholderiaceae	Beta-Proteobacteria	CP000623	–
øRSA1	<i>Ralstonia solanacearum</i>	Ralstoniaceae	Beta-Proteobacteria	AB276040	JBact 190:143
øX216	<i>Burkholderia pseudomallei</i>	Burkholderiaceae	Beta-Proteobacteria	JX681814	BMC Microbiol 12:289
RSY1	<i>Ralstonia solanacearum</i>	Ralstoniaceae	Beta-Proteobacteria	AB981169	–
ST79	<i>Burkholderia pseudomallei</i>	Burkholderiaceae	Beta-Proteobacteria	NC_021343	–
535AP1	<i>Mannheimia haemolytica</i>	Pasteurellae	Gamma-Proteobacteria	KP137432	BMCGenomics15:175
587AP1	<i>Mannheimia haemolytica</i>	Pasteurellae	Gamma-Proteobacteria	KP137434	BMCGenomics15:175
1127AP1	<i>Mannheimia haemolytica</i>	Pasteurellae	Gamma-Proteobacteria	KP137436	BMCGenomics15:175
1152AP1	<i>Mannheimia haemolytica</i>	Pasteurellae	Gamma-Proteobacteria	NC_021778	BMCGenomics15:175
2256AP1	<i>Mannheimia haemolytica</i>	Pasteurellae	Gamma-Proteobacteria	KP137438	BMCGenomics15:175
BAA410	<i>Mannheimia haemolytica</i>	Pasteurellae	Gamma-Proteobacteria	DQ426905	Virology 350:79
F108	<i>Pasteurella multocida</i>	Pasteurellae	Gamma-Proteobacteria	DQ114220	AEM 72:3154
HP1	<i>Haemophilus influenzae</i>	Pasteurellae	Gamma-Proteobacteria	NC_001697	NucleicAcidRes 24:685
HP2	<i>Haemophilus influenzae</i>	Pasteurellae	Gamma-Proteobacteria	AY027935	JBact 184:6893
K139	<i>Vibrio cholerae</i>	Vibrionaceae	Gamma-Proteobacteria	NC_003313	JBact 184:6592
Kappa	<i>Vibrio cholerae</i>	Vibrionaceae	Gamma-Proteobacteria	NC_010275	–
øCTX	<i>Vibrio cholerae</i>	Vibrionaceae	Gamma-Proteobacteria	AB008550	MolMicro 31:399
ø3	<i>Pseudomonas aeruginosa</i>	Pseudomonadaceae	Gamma-Proteobacteria	KT887559	–
øMHaA1-PHL101	<i>Pseudomonas aeruginosa</i>	Pseudomonadaceae	Gamma-Proteobacteria	NC_008201	Virology 350:79
øO18P	<i>Mannheimia haemolytica</i>	Pasteurellae	Gamma-Proteobacteria	NC_009542	Virology 373:25
øAeromonas media	<i>Aeromonas media</i>	Aeromonadaceae	Gamma-Proteobacteria	NC_009542	–
PV94	<i>Vibrio vulnificus</i>	Vibrionaceae	Gamma-Proteobacteria	HG803181	PLoS One 9:e94707
Smp131	<i>Stenotrophomonas maltophilia</i>	Xanthomonaceae	Gamma-Proteobacteria	JO809663	BMC Microbiol 14:17
VPUSM 8	<i>Vibrio cholerae</i>	Vibrionaceae	Gamma-Proteobacteria	NC_022747	–

Table S2
P2 Supercluster Phages and Prophages

D P2-like prophages outside the *Enterobacteriaceae*

Prophage	Host species and strain	Host Family	Class	MCP locus_tag
BrevuP2-A	<i>Brevundimonas diminuta</i> ATCC-11568	Caudobacteriaceae	Alpha-Proteobacteria	BDIM_14580
NovoP2-A	<i>Novosphingobium pentarothovans</i> US6-1	Sphingomonadaceae	Alpha-Proteobacteria	J159_04365
ZymoP2-A	<i>Zymomonas mobilis</i> ZM4	Sphingomonadaceae	Alpha-Proteobacteria	ZZM4_0034
AchromoP2-A	<i>Achromobacter</i> sp. LC458	Alcaligenaceae	Beta-Proteobacteria	YH64_11480
BurkP2-A	<i>Burkholderia thailandensis</i> MSMB121	Burkholderiaceae	Beta-Proteobacteria	BTL_1970
BurkP2-B	<i>Burkholderia cenocepacia</i> MCO-3	Burkholderiaceae	Beta-Proteobacteria	Bcenm03_0205
Collip2-A	<i>Collimonas fungivorans</i> Ter331	Collimonadaceae	Beta-Proteobacteria	CFU_0996
ComaP2-A	<i>Comamonas testosteroni</i> TK102	Comamonadaceae	Beta-Proteobacteria	O987_RS22905
CupP2-A	<i>Cupriavidus taiwanensis</i> LMG19424	Cupriavidaceae	Beta-Proteobacteria	RALTA_A0929
KingP2-A	<i>Kingella oralis</i> ATCC 51147	Burkholderiaceae	Beta-Proteobacteria	GCWU000324_01254
NeissP2-A	<i>Neisseria meningitidis</i> alpha710	Neisseriaceae	Beta-Proteobacteria	NMMB_1121
PolarP2-A	<i>Polaromonas naphthalenivorans</i> CJ2	Comamonadaceae	Beta-Proteobacteria	Prap_3291
DesulfP2-A	<i>Desulfivibrio desulfuricans</i> ATCC 27774	Desulfivibrionaceae	Delta-Proteobacteria	Ddes_0241
DesulfP2-B	<i>Desulfivibrio vulgaris</i> DP4	Desulfivibrionaceae	Delta-Proteobacteria	Dvul_2765
AcinP2-A	<i>Acinetobacter baumannii</i> AYE	Pseudomonadales; Moraxellaceae	Gamma-Proteobacteria	ABAYE0571
AcinP2-B	<i>Acinetobacter ursingii</i> NIPH 706	Pseudomonadales; Moraxellaceae	Gamma-Proteobacteria	F943_01500
AeroP2-A	<i>Aeromonas hydrophila</i> AL06-06	Aeromonadae	Gamma-Proteobacteria	RY45_19020
AeroP2-B	<i>Aeromonas salmonicida</i> 2009-144K3	Aeromonadae	Gamma-Proteobacteria	NX85_17375
AzoP2-A	<i>Azotobacter vinelandii</i> CA	Pseudomonadales; Pseudomonadaceae	Gamma-Proteobacteria	AvCA_37590
CardP2-A	<i>Cardiobacterium hominis</i> ATCC-15926	Cardiobacteriaceae	Gamma-Proteobacteria	HMPREF0198_2128
ChromoP2-A	<i>Chromohalobacter salexigens</i> DSM 3043	Halomonadaceae	Gamma-Proteobacteria	Csal_1397
EndoP2-A	<i>Endozoicomonas montiporae</i> LMG 24815	Halellaceae	Gamma-Proteobacteria	GZ77_07230
Gallip2-A	<i>Gallibacterium anatis</i> UMN179	Pasteurellae	Gamma-Proteobacteria	UMN179_00236
HaliP2-A	<i>Halella chejuensis</i> KCCTC 2396	Halellaceae	Gamma-Proteobacteria	HCH_00394
HaliP2-B	<i>Halomonas</i> sp. PEN3	Halomonadaceae	Gamma-Proteobacteria	Q671_08135
MannP2-A	<i>Mannheimia haemolytica</i> PKL10	Pasteurellae	Gamma-Proteobacteria	AK33_08830
MarmoP2-A	<i>Mairimonas</i> sp. MMYL1	Oceanospirillaceae	Gamma-Proteobacteria	Mmwy11_0585
MarinP2-A	<i>Marinobacter nanhaiticus</i> D15-8W	Alteromonadales; Alteromonadaceae	Gamma-Proteobacteria	J057_04576
MortP2-A	<i>Mortella viscosa</i> MM/S1	Alteromonadales; Alteromonadaceae	Gamma-Proteobacteria	MVIS_2005
Oleip2-A	<i>Oleispira antarctica</i> RB-8	Oceanospirillaceae	Gamma-Proteobacteria	OLEAN_C08680
PasP2-A	<i>Pasteurella multocida</i> P1062	Pasteurellae	Gamma-Proteobacteria	P1062_021455
PseudoalteroP2-A	<i>Pseudoalteromonas arctica</i> A 37-1-2	Pseudoalteromonadaceae; Alteromonadales	Gamma-Proteobacteria	PARC_05148
PseudoalteroP2-B	<i>Pseudoalteromonas atlantica</i> A 37-1-2	Pseudoalteromonadaceae; Alteromonadales	Gamma-Proteobacteria	Patl_0697
PseudoP2	<i>Pseudomonas aeruginosa</i> BL13	Pseudomonadales; Pseudomonadaceae	Gamma-Proteobacteria	Q067_06185
SheP2-B	<i>Shewanella baltica</i> OS195	Shewanellaceae; Alteromonadales	Gamma-Proteobacteria	Sbal195_2914
StentioP2-A	<i>Stenotrophomonas maltophilia</i> D457	Xanthomonadaceae	Gamma-Proteobacteria	SMD_3273
Vibriop2-B	<i>Vibrio cholerae</i> AM-19226	Vibrionaceae	Gamma-Proteobacteria	A33_2125
Vibriop2-A	<i>Vibrio fischeri</i> ES114	Vibrionaceae	Gamma-Proteobacteria	VF2012
XanthoP2-A	<i>Xanthomonas campestris</i>	Xanthomonadaceae	Gamma-Proteobacteria	XCCEB1459_1200

Table S3
P22-like and P2-like prophages in Salmonella genome sequences

<i>Salmonella enterica</i> subsp. <i>enterica</i>	O Group	Serovar	Number of genome sequences	P22 MCP	CUS-3 MCP	Sf6 MCP	Percentage of genomes with P22-like prophage	P22 Tailspike type††	Number of tailspike genes†	P2-like Fels-2 MCP >50%	P2-like P88 MCP >50%	P2-like ESSI-2 MCP >45%
	O:1,3,19	Serftenberg	10	0	0	0	0.0%		0	5	0	2
	O:2	Paratyphi A	139	130	0	0	93.5%	la	130	107	32	124
	O:3,10	Anatum	5	2	0	0	40.0%	III / IIb	1 / 1	1	0	1
	O:3,10	Give	2	1	0	0	50.0%	la	1	1	0	2
	O:3,10	Muenster	4	2	0	0	50.0%	III / Vb	1 / 1	8	0	0
	O:3,10	Nchanga	2	2	0	0	100.0%	III	2	0	0	2
	O:3,10	Uganda	1	1	0	0	100.0%	III	1	0	0	0
	O:3,10	Weltvredden	2	1	0	0	50.0%	III	1	2	0	1
	O:3,10	London	1	0	0	0	0.0%		0	1	0	1
	O:3,10	Meleagridis	1	0	0	0	0.0%		0	0	0	1
	O:4	Abony	1	1	0	0	100.0%	la	1	0	0	0
	O:4	Agona	114	11	1	0	10.5%	la / lb	8 / 4	111	1	83
	O:4	Brandenburg	1	1	0	0	100.0%	la	1	1	0	1
	O:4	Bredeney	1	0	0	0	0.0%		0	0	0	0
	O:4	Chester	1	0	0	0	0.0%		0	1	0	0
	O:4	Derby	2	0	0	0	0.0%		0	0	0	0
	O:4	Heidelberg	103	103	0	0	100.0%	la	103	38	0	102
	O:4	Indiana	1	0	0	0	0.0%		0	1	0	0
	O:4	Paratyphi B	10	1	0	0	10.0%	la	1	4	2	3
	O:4	Saintpaul	6	1	0	0	16.7%	la	1	1	0	2
	O:4	Schwarzengrund	2	0	0	2	100.0%	XI	1	1	0	1
	O:4	Sloterdijk	1	0	0	0	0.0%		0	0	0	0
	O:4	Stanley	1	0	0	0	0.0%		0	1	0	1
	O:4	Stanleyville	1	0	0	0	0.0%		0	0	0	0
	O:4	Typhimurium	445	388	0	0	87.2%	la / lb / Vc	381 / 1 / 1	30	3	26
	O:6,7	Bareilly	103	12	0	0	11.7%	Vb	12	6	0	10
	O:6,7	Braenderup	2	0	0	0	0.0%		0	4	0	0
	O:6,7	Choleraesuis	5	5	0	0	100.0%	Va	5	0	0	0
	O:6,7	Hartford	1	0	0	0	0.0%		0	1	0	0
	O:6,7	Infantis	5	1	0	0	20.0%	Vb	1	2	0	5
	O:6,7	Livingstone	1	1	0	0	100.0%	Vc	1	1	0	0
	O:6,7	Lubbock	2	2	0	0	100.0%	Va	2	5	0	2
	O:6,7	Mbandaka	1	0	0	0	0.0%	Vc	1	1	0	0
	O:6,7/O:54*	Montevideo	55	4	0	0	7.3%	Vb / la	3 / 1	51	0	4
	O:6,7	Norwich	1	0	0	0	0.0%		0	1	0	0

Table S3
P22-like and P2-like prophages in Salmonella genome sequences

<i>Salmonella enterica</i> subsp. <i>enterica</i>	O Group	Serovar	Number of Salmonella genome sequences	P22 MCP	CUS-3 MCP	Sf6 MCP	Percentage of genomes with P22- like prophage	P22 Taispike type††	Number of taispike genest	Fels-2 MCP >50%	P88 MCP >50%	ESSI-2 MCP >45%
	O:6.7	Ohio	1	1	0	0	100.0%	Vc	1	0	0	0
	O:6.7	Oranienburg	3	0	0	0	0.0%		0	3	0	0
	O:6.7	Paratyphi C	1	0	1	0	100.0%	Vb	1	0	0	1
	O:6.7	Rissen	1	1	0	0	100.0%	Vc	1	2	0	0
	O:6.7	Tennessee	3	2	0	0	66.7%	Va / Vc	1 / 1	1	0	2
	O:6.7	Thompson	2	0	0	0	0.0%		0	0	0	0
	O:6.7	Virchow	3	0	0	0	0.0%		0	1	0	1
	O:8	Albany	1	1	0	0	100.0%	Vlia	1	0	0	0
	O:8	Bovismorbificans	2	0	0	0	0.0%		0	0	0	1
	O:8	Hadar	5	2	1	0	60.0%	Vlic	3	0	0	0
	O:8	Kentucky	15	0	0	0	0.0%		0	0	0	0
	O:8	Litchfield	1	0	0	0	0.0%		0	0	0	0
	O:8	Manhattan	2	0	0	0	0.0%		0	1	0	0
	O:8	Muenchen	4	0	0	0	0.0%		0	3	0	2
	O:8	Newport	47	5	3	2	21.3%	Vlia / Vlib / Vlid	7 / 3 / 6	23	2	10
	O:8	Tallahassee	1	0	1	0	100.0%	Vlic	1	0	0	0
	O:9	Berta	3	0	0	0	0.0%		0	3	0	0
	O:9	Dublin	10	10	0	0	100.0%	la	10	10	0	0
	O:9	Eastbourne	1	0	0	0	0.0%		0	1	0	0
	O:9	Enteritidis	318	1	0	0	0.3%	III	1	208	0	22
	O:9	Galinarum/Pullorum	12	1	0	0	8.3%	la	1	5	0	0
	O:9	Javiana	5	1	0	0	20.0%	la	1	5	0	0
	O:9	Miami	1	0	0	0	0.0%	la	1	0	0	0
	O:9	Napoli	1	0	0	0	0.0%		0	0	1	0
	O:9	Panama	1	0	0	0	0.0%		0	1	0	0
	O:9	Typhi	1755	0	0	0	0.0%		0	3526	68	24
	O:9	9,12:1,v:	1	1	0	0	100.0%	lb	1	0	0	2
	O:9.46	Bairdon	1	0	0	0	0.0%		0	0	0	0
	O:9.46	Ouakam	1	1	0	0	100.0%		0	0	0	0
	O:11	Abaetetuba	1	0	0	0	0.0%		0	0	0	0
	O:11	Rubislaw	2	1	0	0	50.0%	XIV	1	0	2	0
	O:13	Cubana	4	0	0	0	0.0%		0	2	0	0
	O:13	Havana	1	0	0	0	0.0%		0	1	0	1
	O:13	Mishmarhaemek	1	0	0	0	0.0%		0	1	1	0

Table S3
P22-like and P2-like prophages in Salmonella genome sequences

<i>Salmonella enterica</i> subsp. <i>enterica</i>	O Group†	Serovar	Number of genome sequences	P22 MCP	CUS-3 MCP	Sf6 MCP	Percentage of genomes with P22- like prophage	P22 Tailspike type††	Number of tailspike genest	Fels-2 MCP >50%	P88 MCP >50%	ESSI-2 MCP >45%
	O:13	Mississippi	1	1	0	0	100.0%	VIII	1	0	0	0
	O:13	Poona	1	1	0	0	100.0%	VIII	1	0	0	1
	O:13	Worthington	1	0	0	0	0.0%		0	0	0	0
	O:16	Gaminara	2	1	0	0	50.0%	VI	1	2	1	0
	O:18	Cerro	21	21	0	0	100.0%	IV	21	0	0	0
	O:21	Minnesota	2	1	0	0	50.0%	X	1	0	0	0
	O:28	Pomona	1	0	0	0	0.0%		0	0	0	0
	O:30	Aqua	1	1	0	0	100.0%	Ia	1	0	0	0
	O:30	Soerenga	1	0	0	1	100.0%	XII	1	1	0	0
	O:30	Urbana	2	0	0	0	0.0%		0	2	0	1
	O:35	Adelaide	1	0	0	0	0.0%		0	1	0	0
	O:35	Alachua	1	0	0	0	0.0%		0	1	0	0
	O:38	Inverness	2	0	0	0	0.0%		0	1	0	0
	O:39	Namur	1	0	0	0	0.0%		0	0	0	0
	O:39	Wandsworth	1	1	0	0	100.0%	XIII	1	0	1	0
	O:40	Johannesburg	1	1	0	0	100.0%	IX	1	1	0	1
<i>Salmonella enterica</i> subsp. <i>arizonae</i>	O:62		1	1	0	0	100.0%	XV	1	0	0	0
<i>Salmonella enterica</i> subsp. <i>diarizonae</i>			2	0	0	0	0.0%		0	1	0	0
<i>Salmonella enterica</i> subsp. <i>houstenae</i>			4	1	0	0	25.0%	XVI / V	1 / 1	0	0	2
<i>Salmonella enterica</i> subsp. <i>indica</i>			2	0	0	0	0.0%		0	2	0	0
<i>Salmonella enterica</i> subsp. <i>salamae</i>			2	1	0	0	50.0%	XVII	1	0	0	0
<i>Salmonella bongori</i>			3	0	0	0	0.0%		0	2	1	1
Total			3298	729	7	5			743**	4197	115	446
			Total P22-like MCPs	741					Total P2-like prophages	4758		
			Total P22-like prophages #	744								

Table S3 footnotes

The table was completed on July 31, 2015. GenBank entries with only "WP_XXXXX" accession numbers are not included since the strain names and nucleic acid context cannot be determined.

† O antigen groups are from Grimont, P., Weill, F. 2007. Antigenic Formulae of the *Salmonella* Serovars, (9th ed.) Paris: WHO Collaborating Center for Reference and Research on *Salmonella*, Institut Pasteur. (Available at: http://www.pasteur.fr/sante/clre/cadrechr/salmoms/WKLM_En.pdf)

†† Where several types of P22-like tailspikes are present in different prophages they are separated by slashes (/). Red text highlights the anomalous tailspikes discussed in the text.

* O:6,7 antigen is expressed in Montevideo serotype cells when plasmid controlling O:54 antigen is not present.

** The total numbers of MCPs and tailspikes are not the same. This is because MCP or tailspike genes are occasionally missing from the translated annotation due to their falling between contigs, having been deleted from the prophage or not being correctly annotated. In addition several Newport prophages have two tailspike genes.

741 MCP genes and 3 tailspike genes whose associated MCP genes are missing or not annotated.

Table S4
P22-like phages and prophages used this study

A	P22-like authentic phages			
Phage	Host	Accession number	genome orientation	MCP type
22	<i>Salmonella enterica</i> (Typhimurium)	KR296686	P22	P22
25	<i>Salmonella enterica</i> (Typhimurium)	KR296687	P22	P22
34	<i>Salmonella enterica</i> (Typhimurium)	KR296688	P22	P22
APSE-2	<i>Hamiltonella defensa</i> *	EU794049	APSE-1	Sf6
ASPE-1	<i>Hamiltonella defensa</i> *	AF157835	APSE-1	Sf6
CUS-3	<i>Escherichia coli</i>	CP000711	Sf6	CUS-3
Emek	<i>Salmonella enterica</i> (Haardt)	JQ806763	P22	P22
epsilon34	<i>Salmonella enterica</i> (Anatum)	EU570103	P22	P22
g341	<i>Salmonella enterica</i> (Anatum)	NC_013059	P22	P22
HK620	<i>Escherichia coli</i>	AF335538	Sf6	Sf6
IME10	<i>Escherichia coli</i>	JN600960	P22	P22
L	<i>Salmonella enterica</i> (Typhimurium)	AY795968†	P22	P22
ø20	<i>Salmonella enterica</i> (Typhimurium)	GQ422450	P22	P22
ø75	<i>Salmonella enterica</i> (Typhimurium)	GQ422451	P22	P22
P22	<i>Salmonella enterica</i> (Typhimurium)	BK000583	P22	P22
SE1	<i>Salmonella enterica</i> (Typhimurium)	NC_011802	P22	P22
SEN22	<i>Salmonella enterica</i> (Diarizonae)	KT630648	P22	P22
SETP1	<i>Salmonella enterica</i> (Eneritidis)	EF151184#	-	-
Sf101	<i>Shigella flexneri</i>	KJ832078	Sf6	CUS-3
Sf6	<i>Shigella flexneri</i>	AF547987	Sf6	Sf6
SPC-P1	<i>Salmonella enterica</i> (TyphiC)	CP000857	Sf6	CUS-3
SPN9CC	<i>Salmonella enterica</i> (Typhimurium)	JF900176	P22	P22
ST104	<i>Salmonella enterica</i> (Typhimurium)	AB102868	P22	P22
ST160	<i>Salmonella enterica</i> (Typhimurium)	NC_014900	P22	P22
ST64T	<i>Salmonella enterica</i> (Typhimurium)	AY052766	P22	P22
	* virion-like particles not yet shown to be infectious † our unpublished sequence # partial sequence			

Table S4
P22-like phages and prophages used this study

B	<i>Enterobacteriaceae</i> P22-like prophages			
	Prophage Host	MCP locus_tag	genome orientation	MCP type
	<i>Arsenophonus nasoniae</i> *	ARN_26010	-	Sf6
	<i>Citrobacter</i> sp. BIDMC108	SK40_00812	Sf6	P22
	<i>Cronobacter sakazakii</i> 696	BN128_2797	P22	P22
	<i>Cronobacter malonaticus</i> †	WP_032971372*	-	-
	<i>Enterobacter</i> sp.GN02548*	ABR33_05860	-	CUS-3
	<i>Enterobacter aerogenes</i> UC147	P850_03405	APSE-1	P22
	<i>Erwinia tracheiphila</i> BufGFH	SY86_19110	APSE-1 early right??	P22
	<i>Escherichia coli</i> B7A	EcB7A_4179	Sf6	Sf6
	<i>Escherichia coli</i> CVM N38428PS	PU66_07355	Sf6	CUS-3
	<i>Escherichia coli</i> W	EschWDRAFT_1163	Sf6	P22
	<i>Klebsiella pneumoniae</i> SB3432	KPR_2793	Sf6	CUS-3
	<i>Kluyvera intermedia</i> CAV1151	AB182_19130	P22	P22
	<i>Leclercia adecarboxylata</i> †	WP_039031990*	-	-
	<i>Morganella morganii</i> KT	MU9_2902	P22	P22
	<i>Pantoea</i> sp. PSNIH2	PSNIH2_13380	P22	P22
	<i>Pectobacterium carotovorum</i> BCS2	RC83_13195	P22	P22
	<i>Pectobacterium carotovorum</i> LMG21371	KS44_06225	P22	CUS-3
	<i>Pluralibacter gergoviae</i> DL84A27	ABW09_23385	-	P22
	<i>Proteus mirabilis</i> ATCC7002	DR94_520	Prot1	P22
	<i>Providencia rettgeri</i> DSM 1131	PROVRETT_06010	P22	Sf6
	<i>Providencia rettgeri</i> †	WP_048606460*	-	-
	<i>Salmonella enterica</i> CVM19536	SEEN536_00900	-	Sf6
	<i>Serratia marcescens</i> VGH107	F518_22685	P22	Sf6
	<i>Serratia plymuthica</i> AS9	SerAS9_2688	APSE-1 (ø80 int org)	P22
	<i>Shigella flexneri</i> 2002017	SF xv_0342	P22	P22
	<i>Shimwella blattae</i> DSM 4481	EBL_c21750	P22	Sf6
	<i>Tatumella morbiroisei</i> LMG23360	HA49_08570	P22	P22
	<i>Xenorhabdus bovienii</i> CS03	XBW1_3034	Prot1	P22
	<i>Yersinia aleksiciae</i> 159	ACZ76_14855	P22	Sf6
	<i>Yersinia kristensenii</i> * ATCC33639	DJ57_2548	P22	P22
	* Rather short contig so not included in genome plot Figure 6	* Accession number (Locus_tag not available)	"-" indicates contig too short to determine orientation	
	† Strain names and whole prophage sequence not available			

Table S4
P22-like phages and prophages used this study

C	<i>Salmonella</i> P22-like prophages			
	Prophage Host	MCP locus_tag		
	<i>Salmonella enterica</i> serovar Paratyphi AY09620	YNSPA_2311		
	<i>Salmonella enterica</i> serovar Heidelberg SL476	SeHA_C0403		
	<i>Salmonella enterica</i> serovar Agona 3432613	SEEA2613_05333		
	<i>Salmonella enterica</i> serovar Typhimurium U288	STU288_12845		
	<i>Salmonella enterica</i> serovar Derby CVMN51289	AEX05_09315		
	<i>Salmonella enterica</i> serovar Albany ATCC51960	SEEA1960_00245		
	<i>Salmonella enterica</i> serovar Hadar 05P066	SeH_A0879		
	<i>Salmonella enterica</i> serovar Newport CVM19536	SEEN536_00900		
	<i>Salmonella enterica</i> serovar Choleraesuis SCB-67	SCH_0357		
	<i>Salmonella enterica</i> serovar Bareilly CFSAN00219	CFSAN00219_		
	<i>Salmonella enterica</i> serovar Montevideo 8387	SEEM8387_07975		
	<i>Salmonella enterica</i> serovar Anatum USMARC-1735	AW50_6230		
	<i>Salmonella enterica</i> serovar Cerro CFSAN001588	CFSAN001588_015920		
	<i>Salmonella enterica</i> serovar Dublin SL1438	SEEDSL_005061		
	<i>Salmonella enterica</i> subspecies <i>Arizonae</i> RSK2980	SARI_02647		
D	Paratyphi A P22-like			
	Prophage Host	MCP locus_tag	Isolation location	
	<i>Salmonella enterica</i> serovar Paratyphi A (no strain name reported)	PA045_3010	-	
	<i>Salmonella enterica</i> serovar Paratyphi A AYN09620	YNSPA_2311	Yunnan, China	
	<i>Salmonella enterica</i> serovar Paratyphi A AKU_12601	SSPA2232e	Pakistan	
	<i>Salmonella enterica</i> serovar Paratyphi A CMCC50093	ACN89_12045	Beijing, China	
	<i>Salmonella enterica</i> serovar Paratyphi A GZ9A00052	GZSPA_2297	Guizhou, China	
	<i>Salmonella enterica</i> serovar Paratyphi A JX05-19	JXSPA_2296	Jiangxi, China	
	<i>Salmonella enterica</i> serovar Paratyphi A CMCC50503	LW89_14285	Jaingsu, China	
	<i>Salmonella enterica</i> serovar Paratyphi A (no strain reported)	PA136_4071	-	
	<i>Salmonella enterica</i> serovar Paratyphi A CMCC50973	IT63_14940	Jaingsu, China	

Table S4
P22-like phages and prophages used this study

E	Heidelberg P22-like prophages		
	Prophage Host	MCP locus_tag	Isolation location
	<i>Salmonella enterica</i> serovar Heidelberg 82-2052	SEEH2052_04713	Maine, USA
	<i>Salmonella enterica</i> serovar Heidelberg ABB07-SB3031	TJ50_03995	British Columbia, CA
	<i>Salmonella enterica</i> serovar Heidelberg B182	SU5_01004	France
	<i>Salmonella enterica</i> serovar Heidelberg CVM N45397	AEU76_20940	Minnesota, USA
	<i>Salmonella enterica</i> serovar Heidelberg N26457	SEEH6457_20825	Colorado, USA
	<i>Salmonella enterica</i> serovar Heidelberg SL476	SeHA_C0403	Minnesota, USA
	<i>Salmonella enterica</i> serovar Heidelberg SL486	SeHB_A0358	Georgia, USA
F	Newport P22-like prophages		
	Prophage Host	MCP locus_tag	Isolation location
	<i>Salmonella enterica</i> serovar Newport CVM 19470	SEEN470_21064	Vietnam
	<i>Salmonella enterica</i> serovar Newport CVM 19536	SEEN536_00900	Vietnam
	<i>Salmonella enterica</i> serovar Newport CVM 22425	SEEN425_002235	Arizona, USA
	<i>Salmonella enterica</i> serovar Newport CVM 22462	SEEN462_0001185	Arizona, USA
	<i>Salmonella enterica</i> serovar Newport CVM 4176	SEEN176_15035	California, USA
	<i>Salmonella enterica</i> serovar Newport Henan_3	G208_21144	Henan, China
	<i>Salmonella enterica</i> serovar Newport JS09102	G209_24390	Jiangsu, China
	<i>Salmonella enterica</i> serovar Newport SH111077	G206_16997	Shanghai, China
	<i>Salmonella enterica</i> serovar Newport Shandong_3	G207_20579	Shandong, China
	<i>Salmonella enterica</i> serovar Newport USMARC-S3124.1	SN31241_18260	USA
G	Typhimurium prophages		
	Prophage Host	MCP locus_tag	Isolation location
	<i>Salmonella enterica</i> serovar Typhimurium 34502	SEET4502_15275	–
	<i>Salmonella enterica</i> serovar Typhimurium ABB1162-2	TJ39_05580	British Columbia, CA
	<i>Salmonella enterica</i> serovar Typhimurium CFSAN001921	CFSAN001921_15475	New York, USA
	<i>Salmonella enterica</i> serovar Typhimurium CVM N51254	AEW79_08460	Connecticut, USA
	<i>Salmonella enterica</i> serovar Typhimurium D23580	STMMW_03811	Malawi
	<i>Salmonella enterica</i> serovar Typhimurium DT104	DT104_03541	–
	<i>Salmonella enterica</i> serovar Typhimurium DT104 EWH6	ERS198263_02378	United Kingdom
	<i>Salmonella enterica</i> serovar Typhimurium FORC_015	ZV79_2330	South Korea
	<i>Salmonella enterica</i> serovar Typhimurium L-3553	STL3553_c03780	–
	<i>Salmonella enterica</i> serovar Typhimurium TN061786	SEE_01911	–
	<i>Salmonella enterica</i> serovar Typhimurium U288	STU288_12845	–
	<i>Salmonella enterica</i> serovar Typhimurium var.5 CVM N43832	AEU38_10780	New York, USA

Table S4
P22-like phages and prophages used this study

H	Prophage P22-like tailspikes			
	<i>Salmonella enterica</i> serovar and strain	Tailspike locus_tag	Tailspike type	
	9,12:l,v:- str. 94293	N925_10475	Ib	
	Abony 0014	SEEA0014_11145	Ia	
	Agona 648586-1	SEEA5861_16245	Ib	
	Albany_ATCC 51960(A)	SEEA1960_00180	VIIa	
	Anatum USDA-ARS-USMARC-1735	AW50_6260	Iib	
	AnatumCDC06-0532	AW57_13240	III	
	Aqua NVSL2001	I654_10070	Ia	
	Arizonae serovar 62:z4,z23:- str. RSK2980	SARI_02638	XIV	
	Bareilly CFSAN000183	SEEB0183_09067	Vb	
	Brandenburg CFSAN024765	QS22_03720	Ia	
	Cerro 87	NI35_4331	IV	
	Choleraesuis SC-B67	SCH_0367	Va	
	Dublin SL1438	SEEDSL_005126	Ia	
	Enteritidis 3402	SEEE3402_02210	III	
	Gaminara ATCC BAA-711	SEEGA711_14315	VI	
	Give var. 15 CFSAN004343	CFSAN004343_03735	Ia	
	Hadar ABB1048-1	TJ45_04405	VIIc	
	Heidelberg SL476	SeHA_C0413	Ia	
	Houtenae ATCC BAA-1581	SEHO0A_00503	XV	
	Houtenae serovar 16:z4,z32:-- str. RKS3027	D088_680001	VI	
	Infantis CFSAN024780	QS05_16285	Vb	
	Javiana PRS_2010_0720	SEEJ0720_04071	Ia	
	Johannesburg S5-703	LTSEJOH_2243	VIII	
	Livingstone CKY-S4	VN13_12135	Vc	
	Lubbock 10TTU468	TR76_09235	Va	
	Miami 1923	SEEM1923_08240	Ia	
	Minnesota ATCC 49284	SEEM9284_03916	IX	
	Mississippi A4-633	LTSEMIS_1115	X	
	Montevideo 4441 H	SEEM41H_02992	Vb	
	Montevideo CFSAN004346	CFSAN004346_01470	Ia	
	Muenster 0315	SEEM0315_05630	III	
	Muenster CFSAN004344	CFSAN004344_03295	Vb	
	Nchanga CFSAN001091	CFSAN001091_05799	III	
	Newport CVM 4176	SEEN176_15095	VIIa	
	Newport CVM 4176	SEEN176_15100	VIIId	
	Newport CVM2242	SEEN425_002285	VIIb	
	Ohio CFSAN001079	CFSAN001079_21649	Vc	
	Paratyphi A ATCC 9150	SPA2388	Ia	
	Paratyphi B ATCC 8759	SEEPB759_13629	Ia	
	Poona ATCC BAA-1673	SEEP1673_06839	X	
	Pullorum S06004	I137_12070	Ia	
	Rissen 150	SEER_02967	Vc	
	Rubislaw A4-653	LTSERUB_1231	XIII	

Table S4
P22-like phages and prophages used this study

H cont.	Tailspikes in tree			
	<i>Salmonella enterica</i> serovar and strain	Tailspike locus_tag	Tailspike type	
	Saint Paul SARA26	SES26_19449	Ia	
	Salamae serovar 58:l,z13,z28:z6 str. 00-0163	SES60163_06541	XVI	
	Schwarzengrund SL480	SeSB_A0555	XI	
	Soerenga 695	SEES0695_12821	XII	
	Tallahassee 0012	SEET0012_06891	VIIc	
	Tennessee TXSC_TXSC08-19	SEET0819_19095	Va	
	Typhimurium CFSAN001921	CFSAN001921_15420	Ic	
	Typhimurium D23580	STMMW_03901	Ib	
	Typhimurium FORC_015	ZV79_2330	Vc	
	Typhimurium U228	STU288_12795	Ia	
	Uganda R8-3404	LTSEUGA_0549	III	
	Wandsworth A4-580	LTSEWAN_1004	XII	
	Weltevreden 2007-60-3289-1	SENTW_2521	III	

SUPPLEMENTARY MATERIAL FIGURES

Figure S1

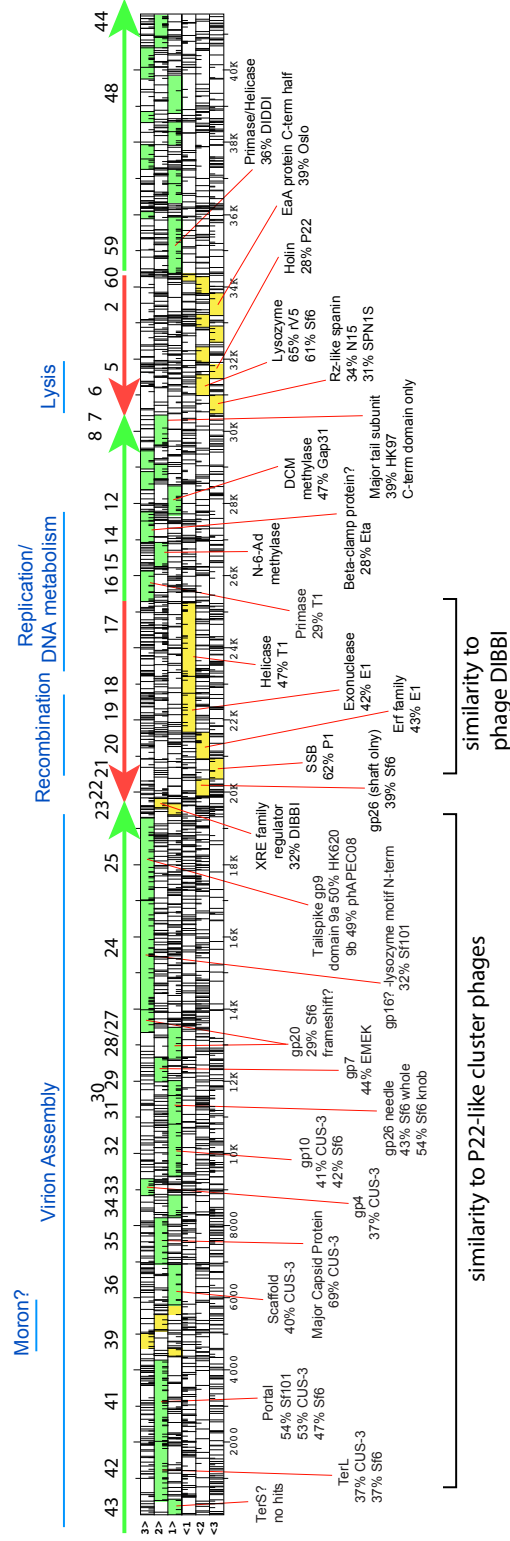


Figure S1. Phage IME-EC2 map and gene database matches.

The phage IME-EC2 genome is shown with the six possible reading frames (indicated at left) with stop codons marked as vertical lines that span the frame rectangle, and AUG codons are indicated by short vertical lines. Open reading frames called as genes are indicated by green and yellow rectangles (transcribed rightward and leftward, respectively). Our gene calls between genes 1 and 5 and between 44 and 59 do not precisely match those of Hua *et al.* (2014); we favor our calls since they leave very little intergenic space as is typical of phage genomes. The opened circular map has been re-oriented from the annotation GenBank accession number KF591601, so that the putative virion assembly genes form the left half of the genome. Above several functional clusters are indicated in blue, selected IME-EC2 locus_tag (gene) numbers of Hua *et al.* (2014) are shown in black, and transcriptional direction (with no operon connotation) is indicated by red and green arrows. Below the ORF map red lines point to the most similar phage proteins found in our analysis. In each graphic annotation the predicted function is given above and similarity to a protein encoded by the indicated phage is given below; no red line indicates there are no good matches in the current sequence database.

In this annotation it is clear that the IME-EC2 virion assembly genes are homologous to and syntenic with those of the P22-like cluster of phages that includes phages Sf6 and CUS-3 (Casjens *et al.*, 2004; King *et al.*, 2007), and “gp#” in the figure refers to the phage P22 gene product names. We note the following additional facts about the IME-EC2 virion assembly genes: (1) genes 43 and 42 overlap by 7 codons, which is typical of terS-terL overlaps in the P22-like phage cluster, suggesting that 43 may encode a TerS protein (TerS proteins are extremely variable, so its lack of homology to known TerS’s is not surprising). (2) Genes 37-40 have no homologues in the P22-like phages (or in other organisms), and we suggest that they are likely a late operon moron (Hendrix *et al.*, 2000; Juhala *et al.*, 2000; Cumby, Davidson, and Maxwell, 2012) that is not involved in virion assembly. (3) Two non-essential genes in this region of the P22-like phages are Orf186 and gene 14. There are syntenic but not recognizably homologous genes, 34 and 30 respectively, in IME-EC2 that could have analogous functions to those poorly understood P22-like phage genes; more experimental work will be required to determine if this is true. Curiously, IME-EC2 genes 21 and 8 are homologous to particular domains of phage Sf6 (a P22-like phage) tail needle shaft, a trimeric coiled-coil (Olia, Casjens, and Cingolani, 2007), and the C-terminal domain of the HK97 tail shaft protein (HK97 has a long non-contractile tail that is similar to the phage lambda tail). The latter is thought not to be

necessary the formation of the actual tail structure in such lambda-like tails; both of these IME-EC2 genes lie in what are likely regions that are expressed early and so may have functions other than virion assembly. (4) The two adjacent IME-EC2 genes 27 and 28 have homology to the 3'- and 5'-sections of P22 gene 20, respectively. This could be the result of an IME-EC2 sequencing frameshift error, or it may indicate a separation of P22 gp20 into two parts so that its function(s) is carried out by two IME-EC2 proteins.

Figure S2

Enterobacteriaceae P2 supercluster MCP dot plot

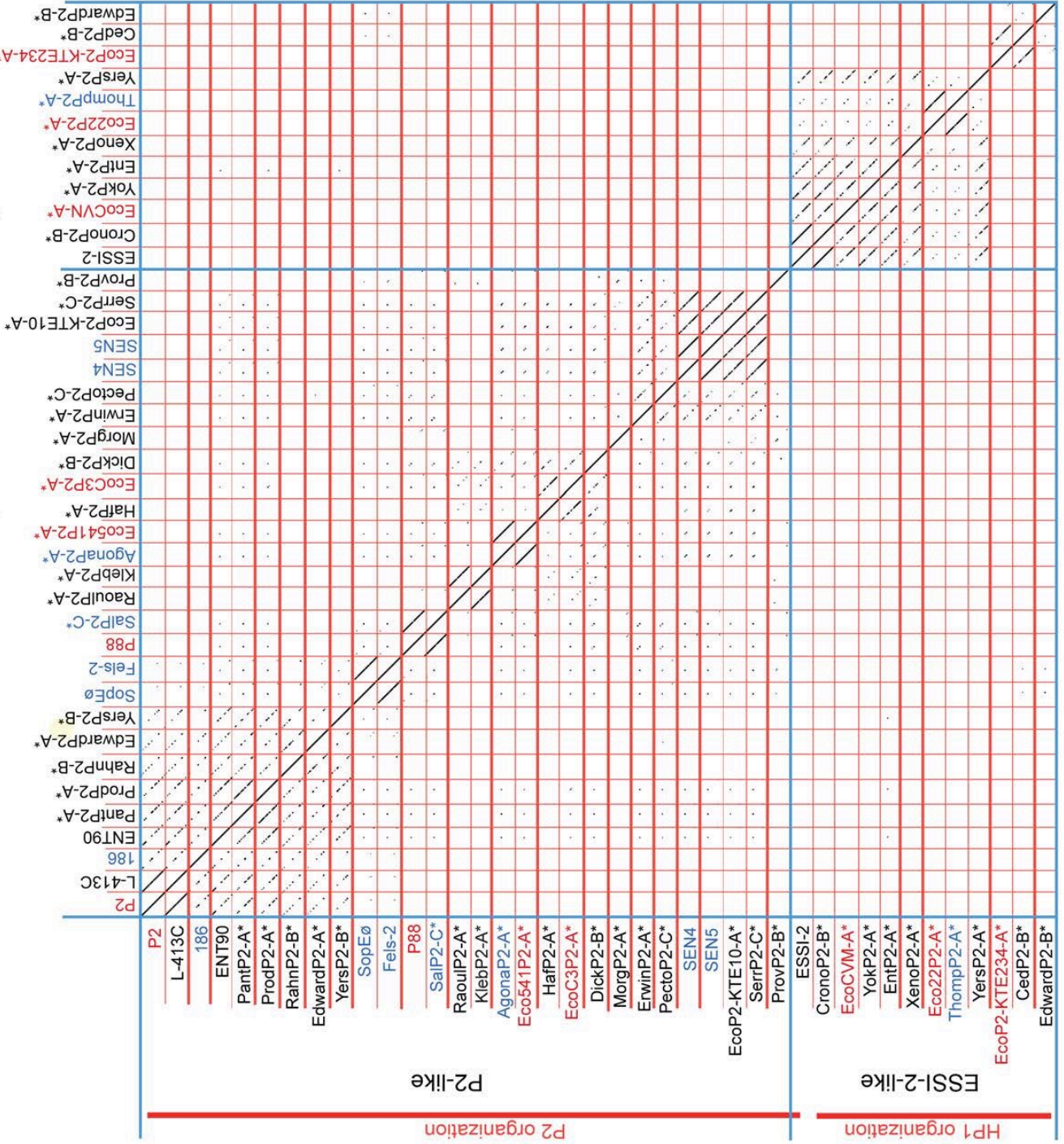


Figure S2. Dot plot analysis of P2-like prophage MCPs from the *Enterobacteriaceae*.

On the left phage and prophage names are shown (red text, *E. coli* host; blue, *Salmonella* host; black, other hosts) and asterisks (*) denote prophage sequences. Blue lines separate the two phage clusters, and thick red lines separate subclusters that are defined in the Figure 2 genome dot plot. Phages and prophages in the plot are listed with host species, genome accession numbers, and MCP locus tags in Table S2A and S2B, respectively. The dot plot was produced by Gepard (Krumstiek, Arnold, and Rattei, 2007) at a word size setting of 9.

Figure S3

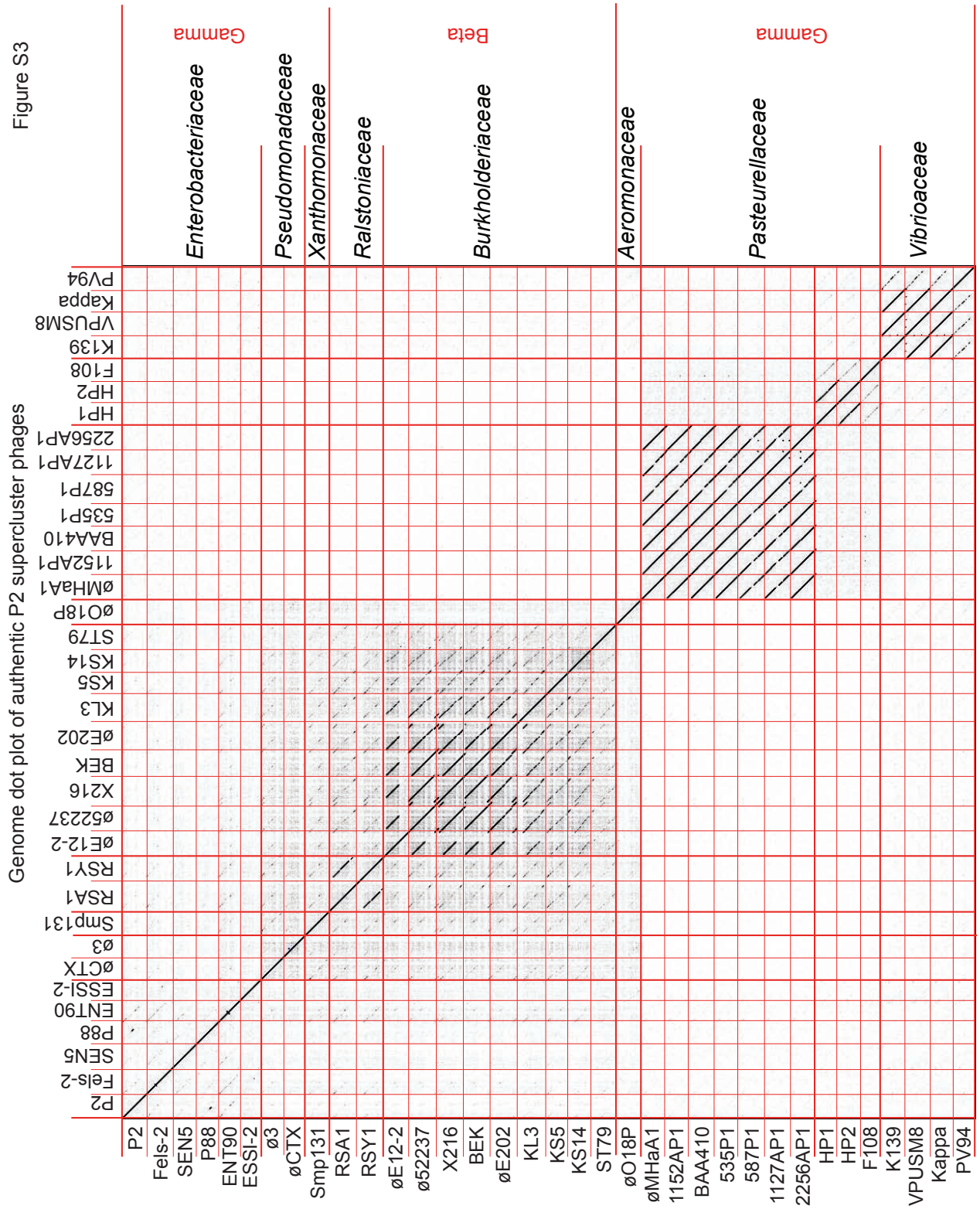


Figure S3. Genome dot plot of authentic P2 supercluster phages.

Phage names are given on the left and top; only representative authentic *Enterobacteriaceae* phage genomes are included. Thin red lines separate individual phage and prophage genomes and thick red line separate the bacterial host taxa shown at the right. Phages in the plot are listed with host species and genome accession numbers in Table S2A and S2C. The dot plot was produced using Gepard (Krumstiek, Arnold, and Rattei, 2007) at a word size setting of 11.

Figure S4

P2-like MCP plot

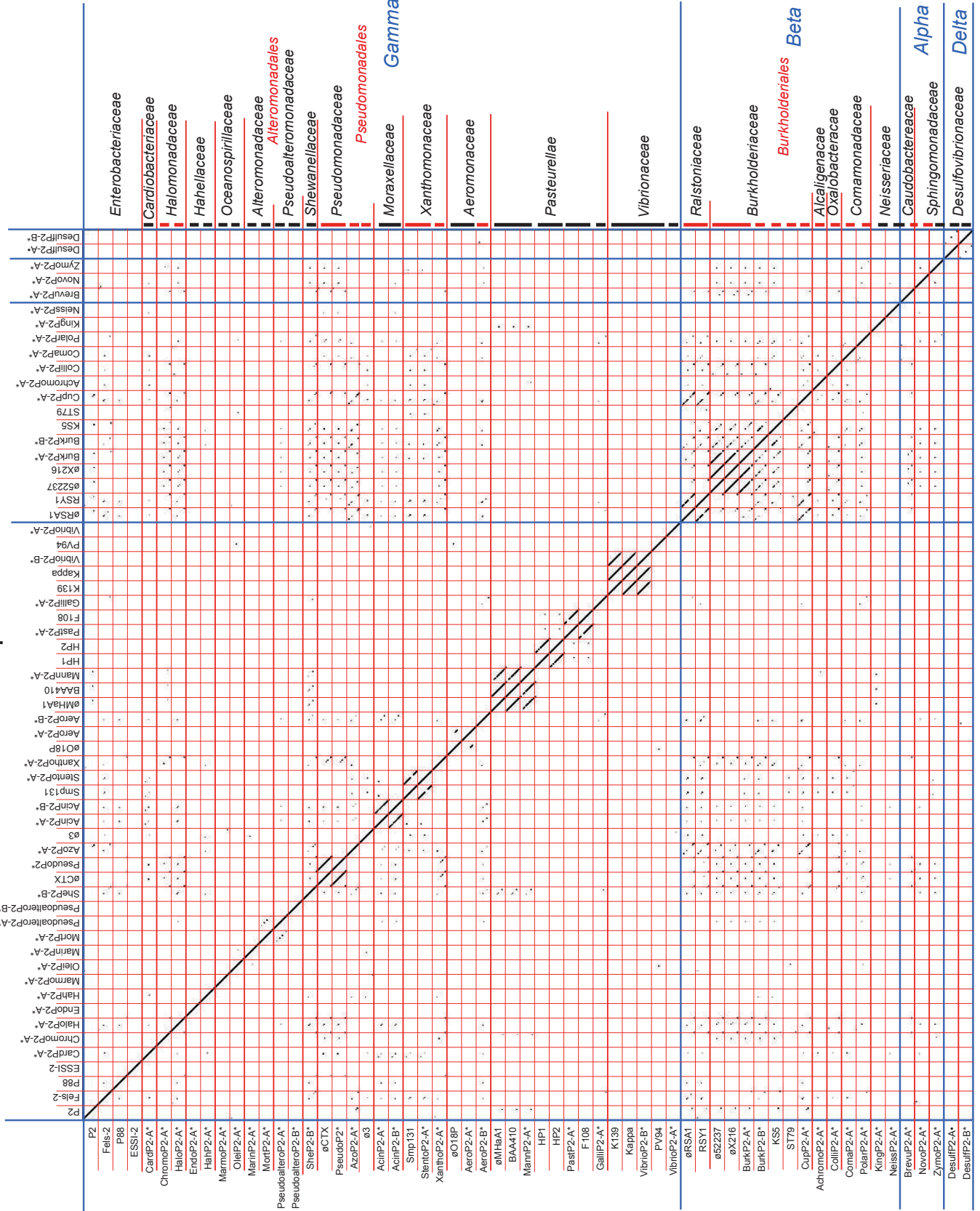


Figure S4. Dot plot analysis of MCPs of P2-like phages and prophages from the Proteobacteria.

Thin red lines separate individual phage and prophage genomes whose names are shown at the top and left; short and longer horizontal red lines separate bacterial host families (black text) and orders (red text), respectively, on the right. Thick blue lines separate Proteobacteria host classes (blue text on the right). Asterisks (*) indicate prophages, and only representative authentic phage genomes are included. Phages and prophages in the plot are listed with host species, genome accession numbers, and MCP locus tags in Table S2. The dot plot was produced using Gepard (Krumnsiek, Arnold, and Rattei, 2007) at a word size setting of 9.

Figure S5

P2-like prophages with distantly related hosts

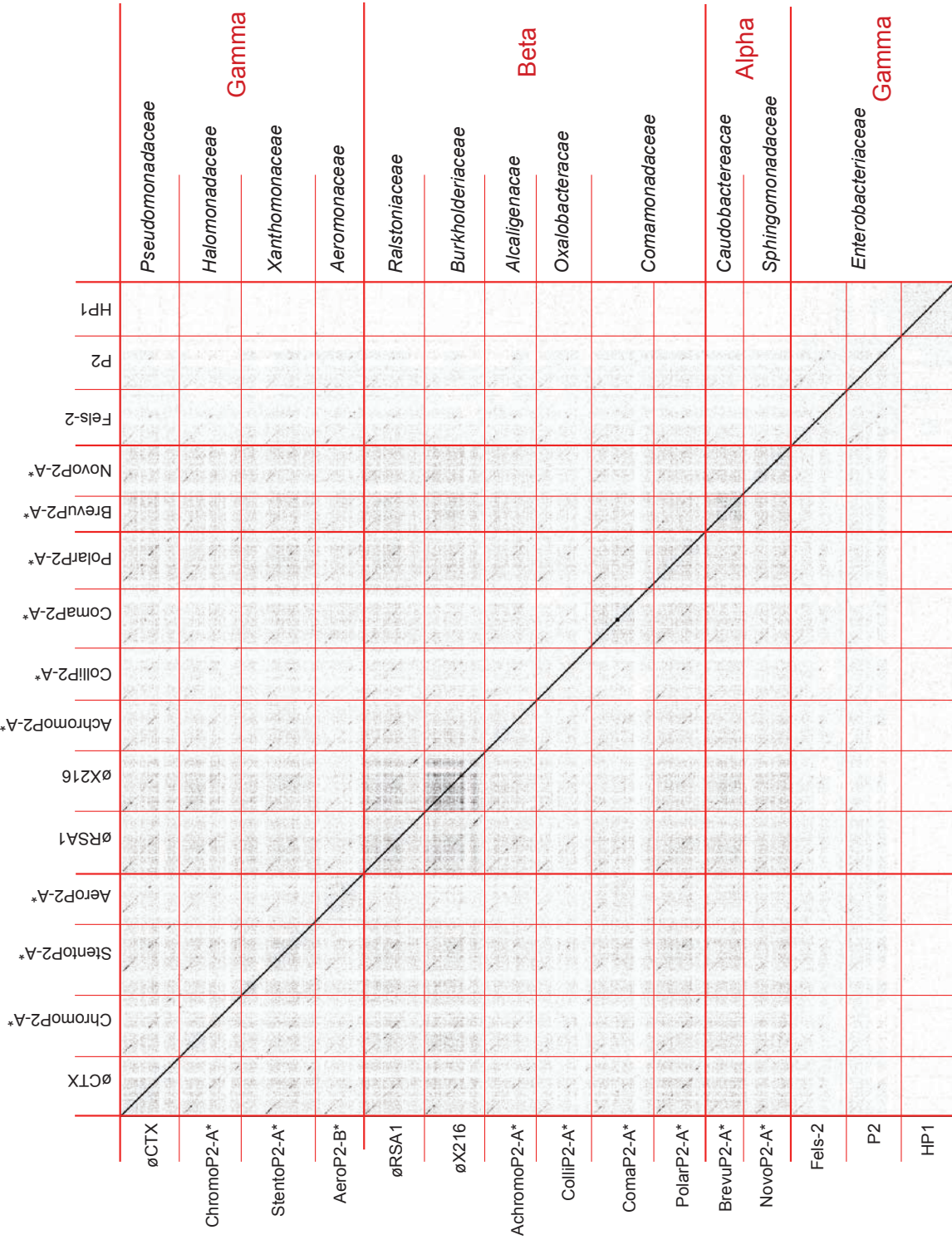


Figure S5. Expanded genome dot plot of P2-like phages and prophages with distantly related hosts.

Red lines separate phage and prophage genomes. Phage and prophage names are given on the left and top; asterisks (*) indicate prophages, and only representative authentic phage genomes are included. Host families (black text) and Proteobacteria classes (red text) are shown at the right. Phages and prophages in the plot are listed with host species, genome accession numbers and MCP locus tags in Table S2. The dot plot was produced using Gepard (Krumstiek, Arnold, and Rattei, 2007) at a word size setting of 10.

Figure S6

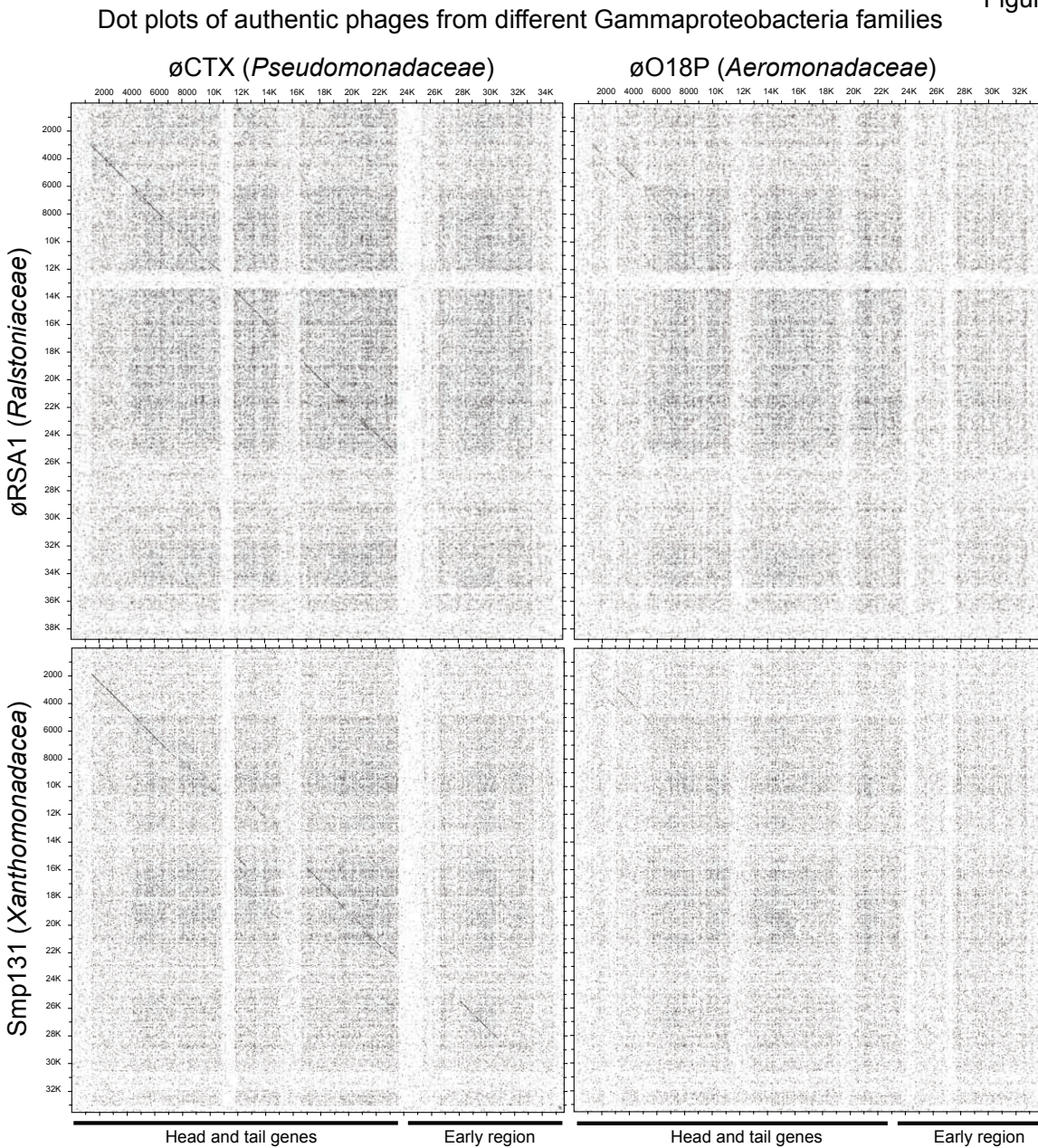


Figure S6. Expanded genome dot plot analysis of authentic P2-like phage examples from different Gammaproteobacteria families.

Phages indicated at the top and left of the plot are listed with host species and genome accession numbers in Table S2C. The dot plots were produced by DNA Strider (Douglas, 1994) with scan window 15 of 23 identities.

Figure S7

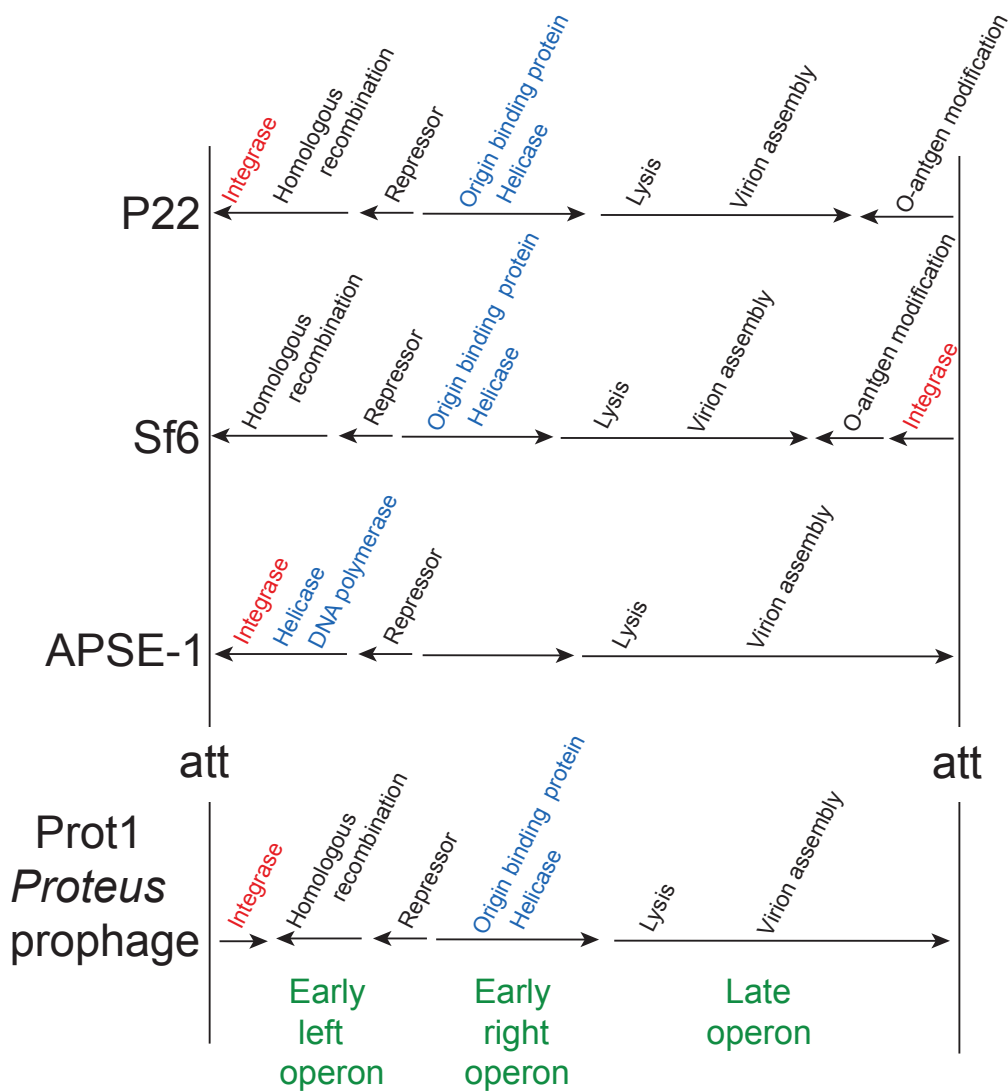


Figure S7. Genome organizational types of the P22-like phages.

Diagrammatic maps of the genomes of four different P22-like phages are shown. Arrows indicate transcriptional direction and the functions of genes in the various regions are shown above each map. The Prot1 prophage is in *Proteus mirabilis* strain ATCC7002 whose MCP locus_tag is DR94_520.

Figure S8

Examples of P22-like prophages

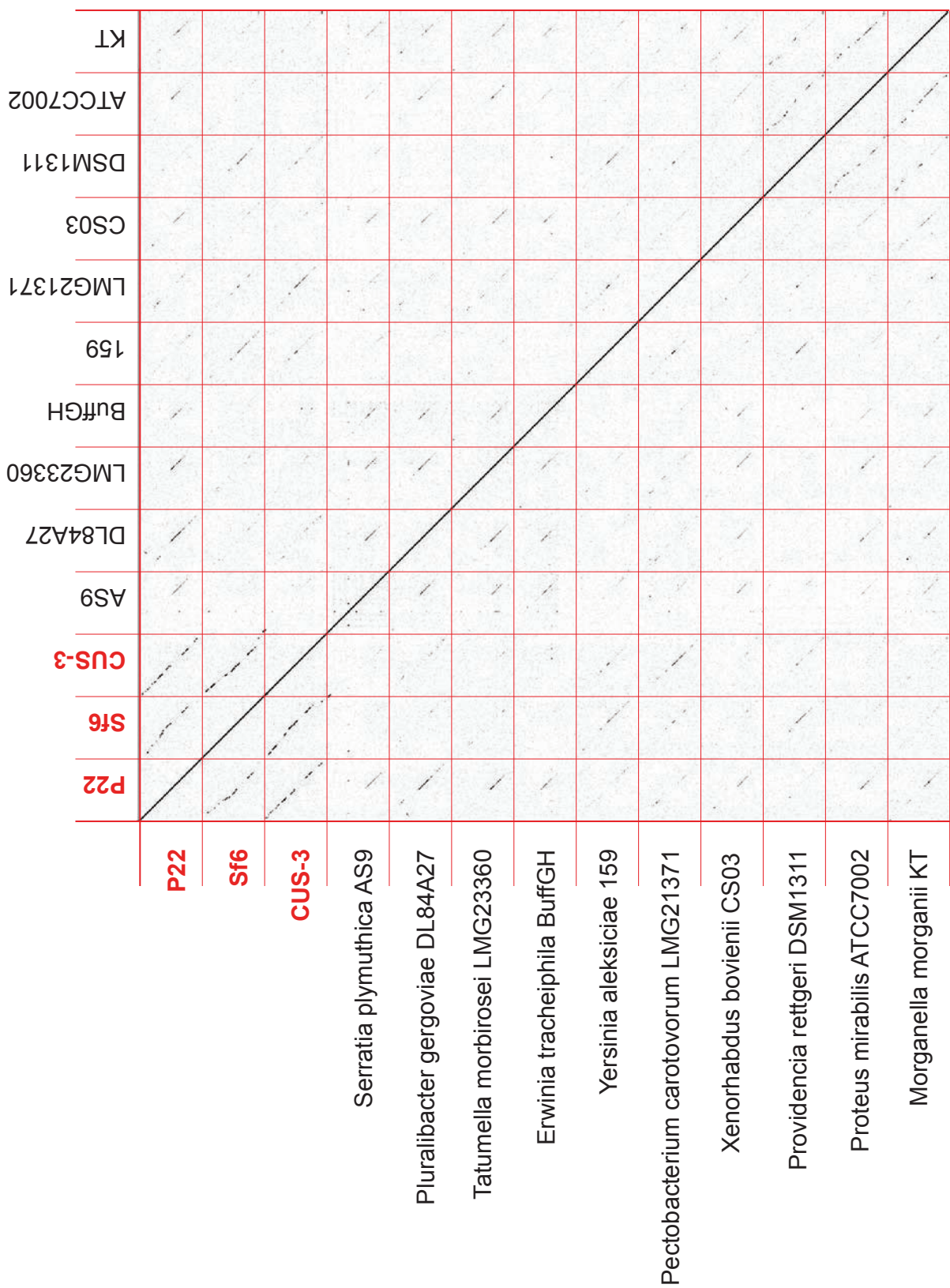


Figure S8. Dot plot analysis of a sample of diverse *Enterobacteriaceae* P22-like prophage genomes.

P22-like prophages in ten different *Enterobacteriaceae* species are shown with three authentic P22-like phages (bold red text) for comparison. The prophages are indicated by the host strain that they inhabit, and their MCP locus_tags are listed in Table S4. The dot plot was produced using Gepard (Krumnsiek, Arnold, and Rattei, 2007) at a word size setting of 10.

P22-like MCP plot

Figure S9

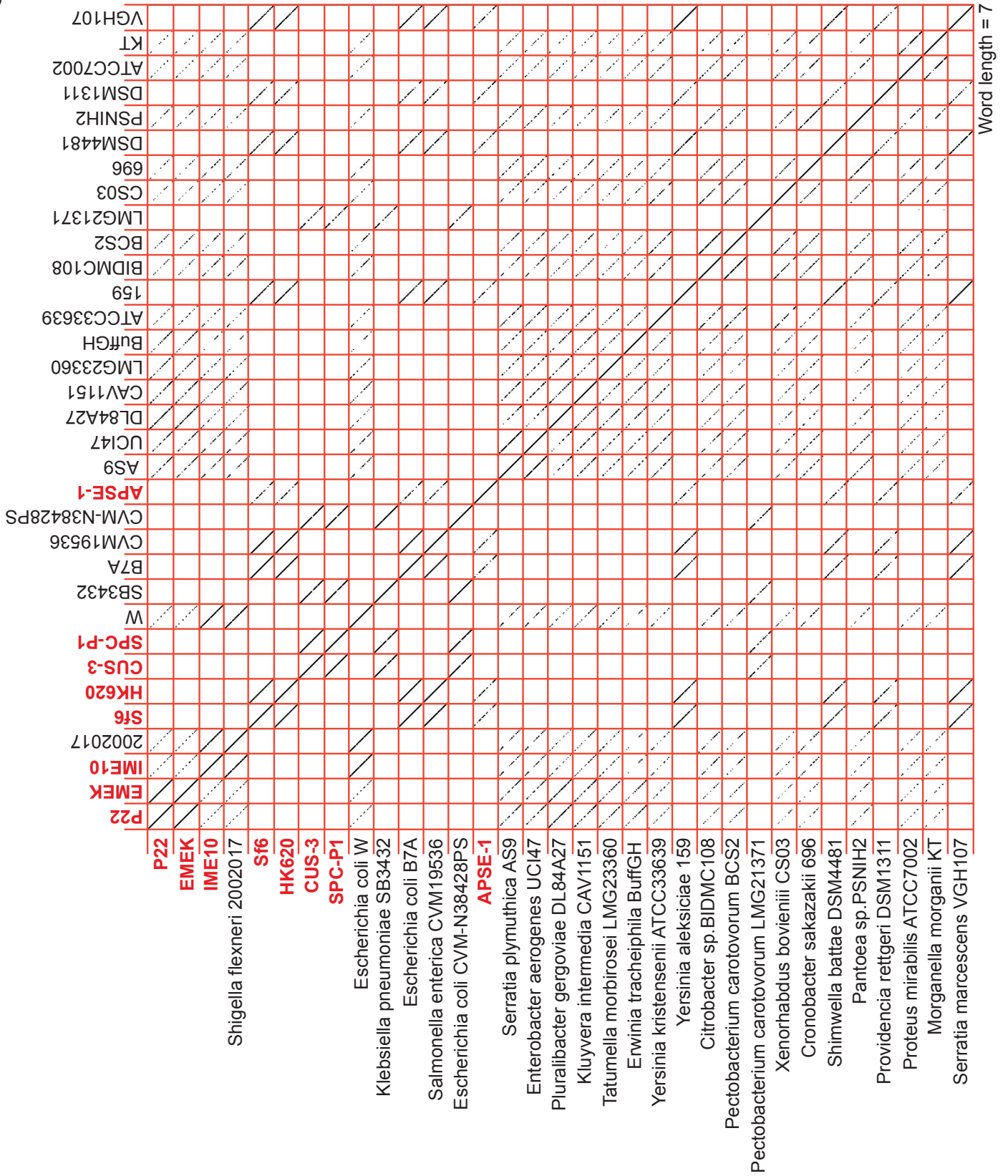
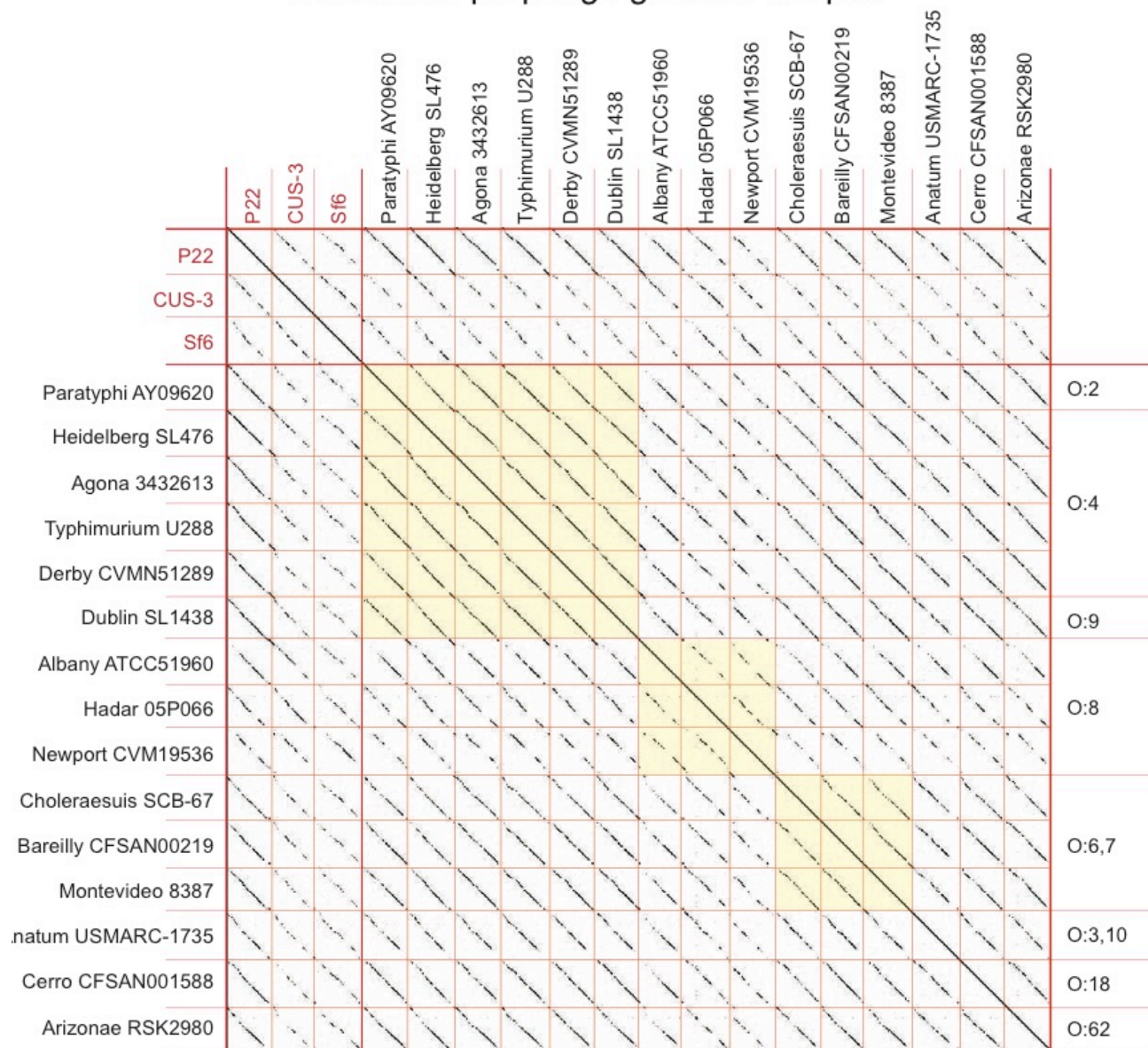


Figure S9. Dot plot analysis of P22-like phage and prophage MCPs.

Red lines separate phage and prophage MCPs. Authentic phage names are in bold red text, and prophage names are indicated by the host species and strain that carries them (listed in Table S4 with their MCP locus_tags). The dot plot was produced by Gepard (Krumnsiek, Arnold, and Rattei, 2007) at a word size setting of 7.

Figure S10

Salmonella prophage genome dot plot**Figure S10. Dot plot analysis of P22-like prophage genomes in *Salmonella enterica*.**

P22-like prophages from fifteen different *Salmonella enterica* serovars are shown with three authentic P22-like phages (red text) shown for comparison. The yellow highlight boxes enclose groups of prophages in serovars with the same or closely related O-antigen structures (see text of report). The prophages are indicated by the host strain that they inhabit, and their MCP locus tags are listed in Table S4. The dot plot was produced using Gepard (Krumsiek, Arnold, and Rattei, 2007) at a word size setting of 11.

Figure S11

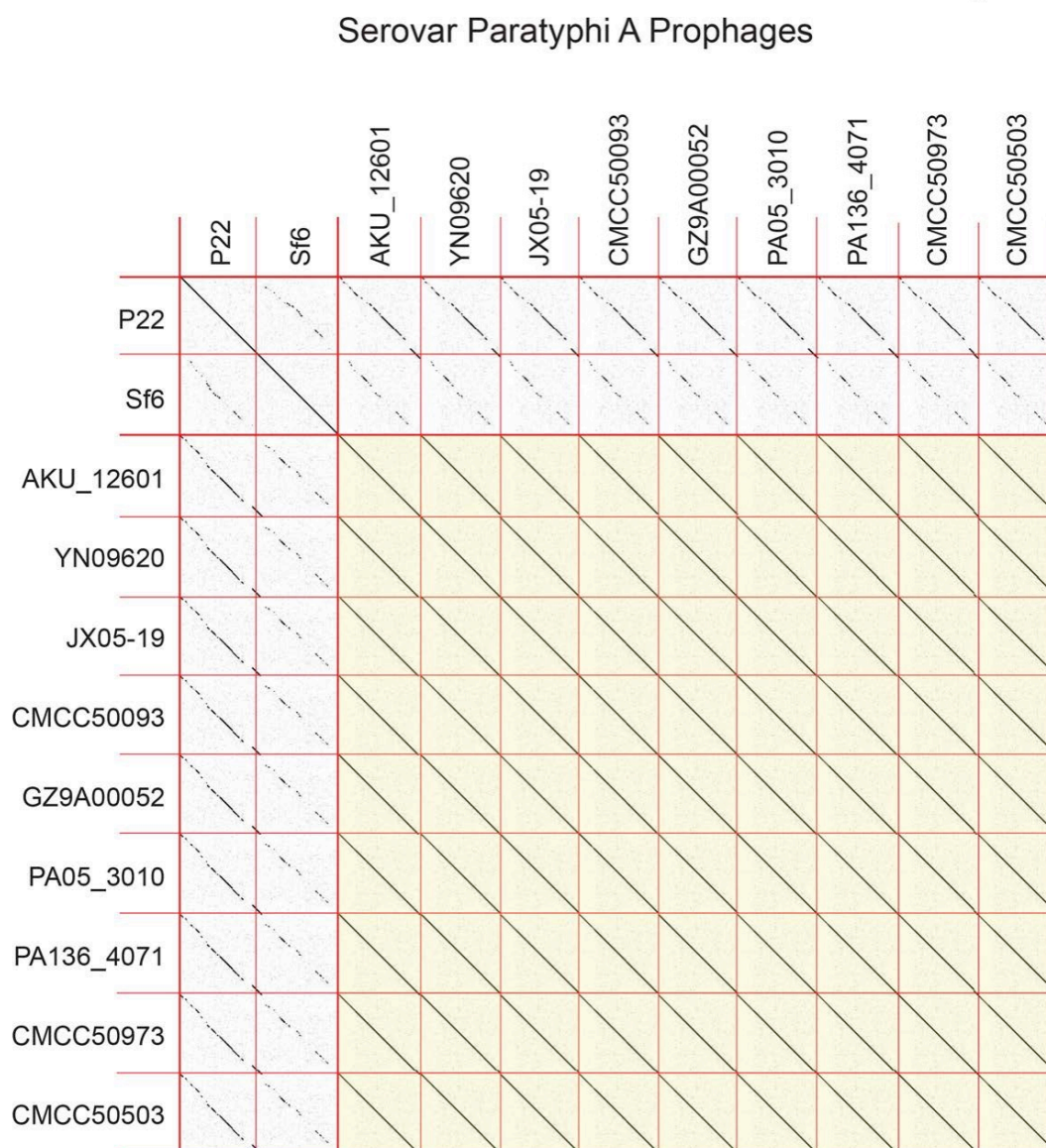


Figure S11. Dot plot analysis of genomes of P22-like prophages in *Salmonella enterica* serovar Paratyphi A isolates.

P22-like prophages from nine randomly chosen Paratyphi A isolates are shown with two authentic P22-like phages (P22 and Sf6) for comparison. The yellow highlight box encloses the nearly identical Paratyphi A prophages. The prophages are indicated by their host strain names (except two whose MCP locus_tags are given since no strain name for the isolate is given in its GenBank entry), and phages and prophages in the plot are listed with host species and MCP locus tags in Table S4D. The dot plot was produced using Gepard (Krumstiek, Arnold, and Rattei, 2007) at a word size setting of 10.

Figure S12

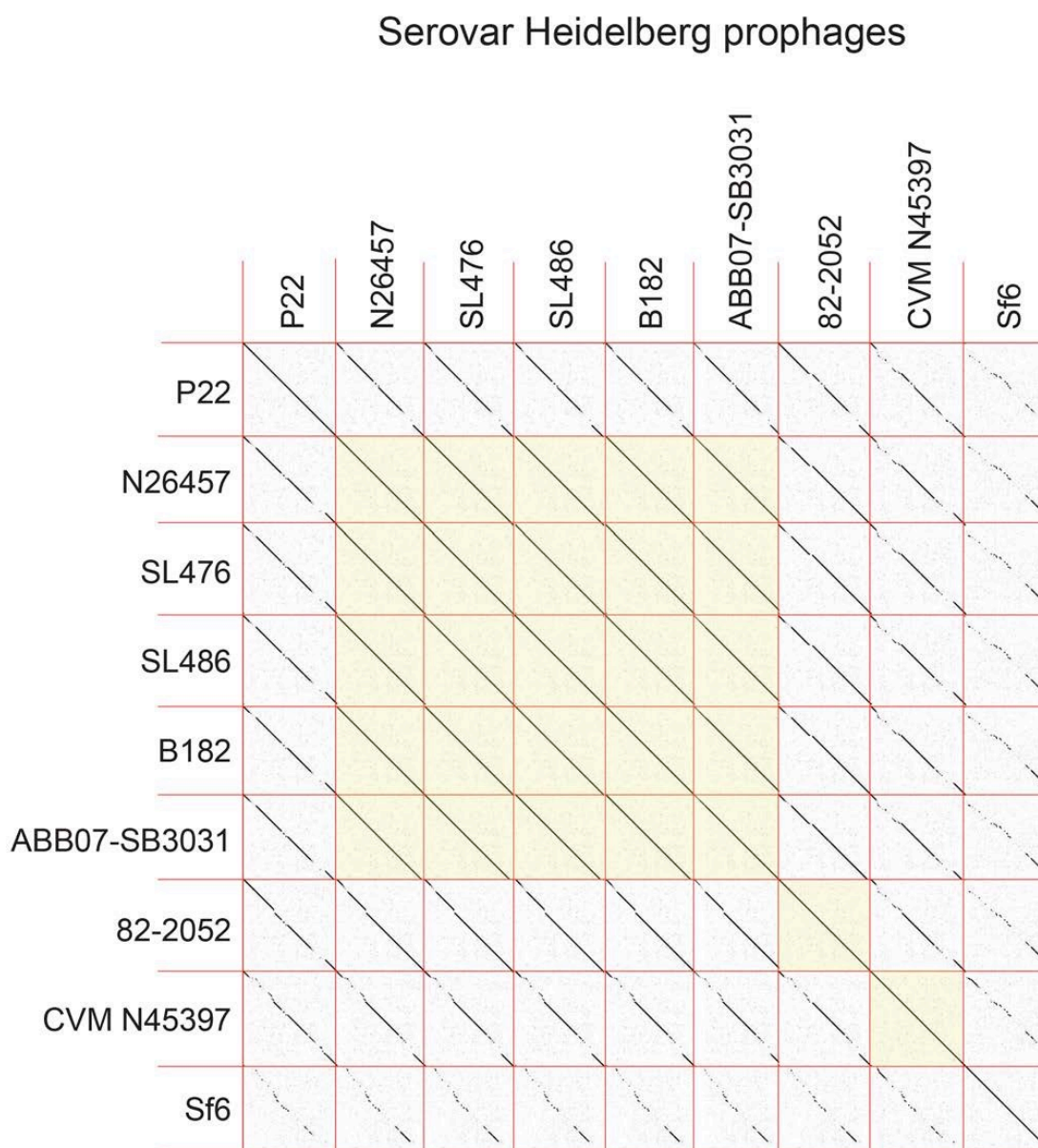


Figure S12. Dot plot analysis of genomes of P22-like prophages in *Salmonella enterica* serovar Heidelberg isolates.

P22-like prophages from seven Heidelberg isolates are shown with two authentic P22-like phages (P22 and Sf6) for comparison. The yellow highlight boxes indicate three different prophage types. The Heidelberg prophages are indicated by their host strain names. Prophages in the plot are listed with host species and MCP locus tags in Table S4E. The dot plot was produced using Gepard (Krumstiek, Arnold, and Rattei, 2007) at a word size setting of 10.

Figure S13

Serovar Typhimurium Prophages

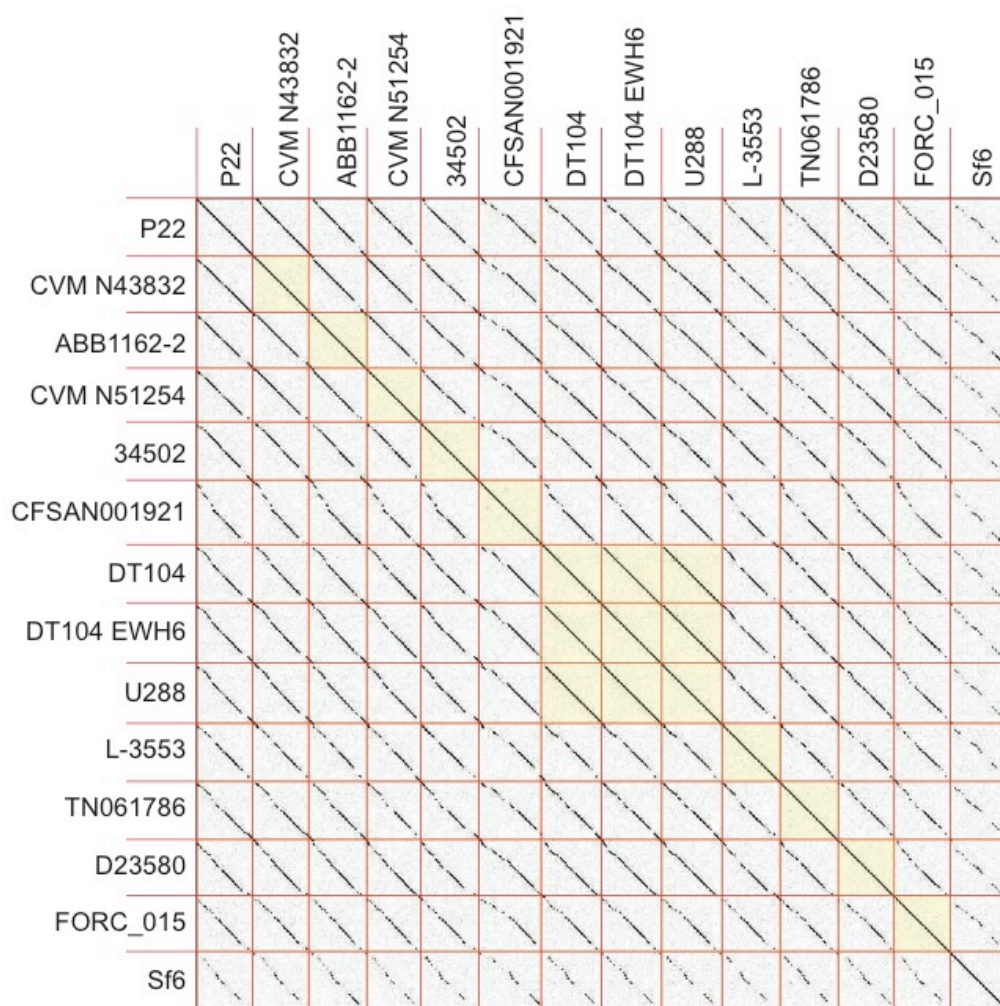


Figure S13. Dot plot analysis of genomes of P22-like prophages in *Salmonella enterica* serovar Typhimurium isolates.

P22-like prophages from twelve Typhimurium isolates are shown with two authentic P22-like phages (P22 and Sf6) for comparison. The Typhimurium sequences are indicated by their host strain names. The yellow highlight boxes indicate significantly different prophage types. Note that the serovar of strain FORC_015 prophage was probably determined incorrectly; it is most likely an O:6,7 strain (see text of report). Phages and prophages in the plot are listed with host species and MCP locus tags in Table S4G. The dot plot was produced using Gepard (Krumstiek, Arnold, and Rattei, 2007) at a word size setting of 10.

Serovar Newport Prophages

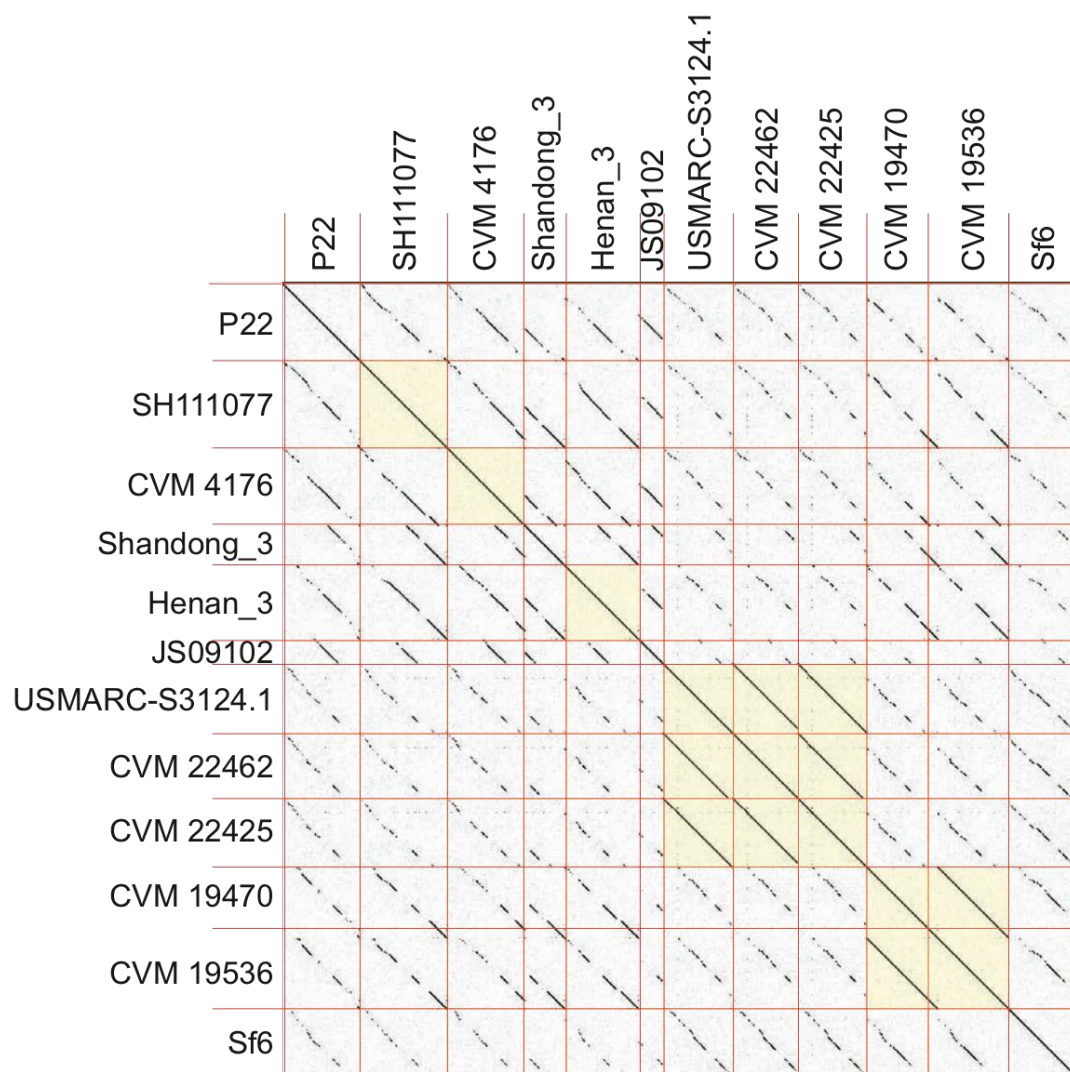


Figure S14. Dot plot analysis of genomes of P22-like prophages in *Salmonella enterica* serovar Newport isolates.

P22-like prophages from ten Newport isolates are shown with two authentic P22-like phages (P22 and Sf6) for comparison. The Newport sequences are indicated by their host strain names. The yellow highlight boxes indicate significantly different prophage types. Two of the ten Newport prophages, in strains Shandong_3 and JS9102, are on short contigs that were not included in the subsequent analysis (see text). Prophages in the plot are listed with host species and MCP locus tags in Table S4F. The dot plot was produced using Gepard (Krumsiek, Arnold, and Rattei, 2007) at a word size setting of 10.

Figure S15

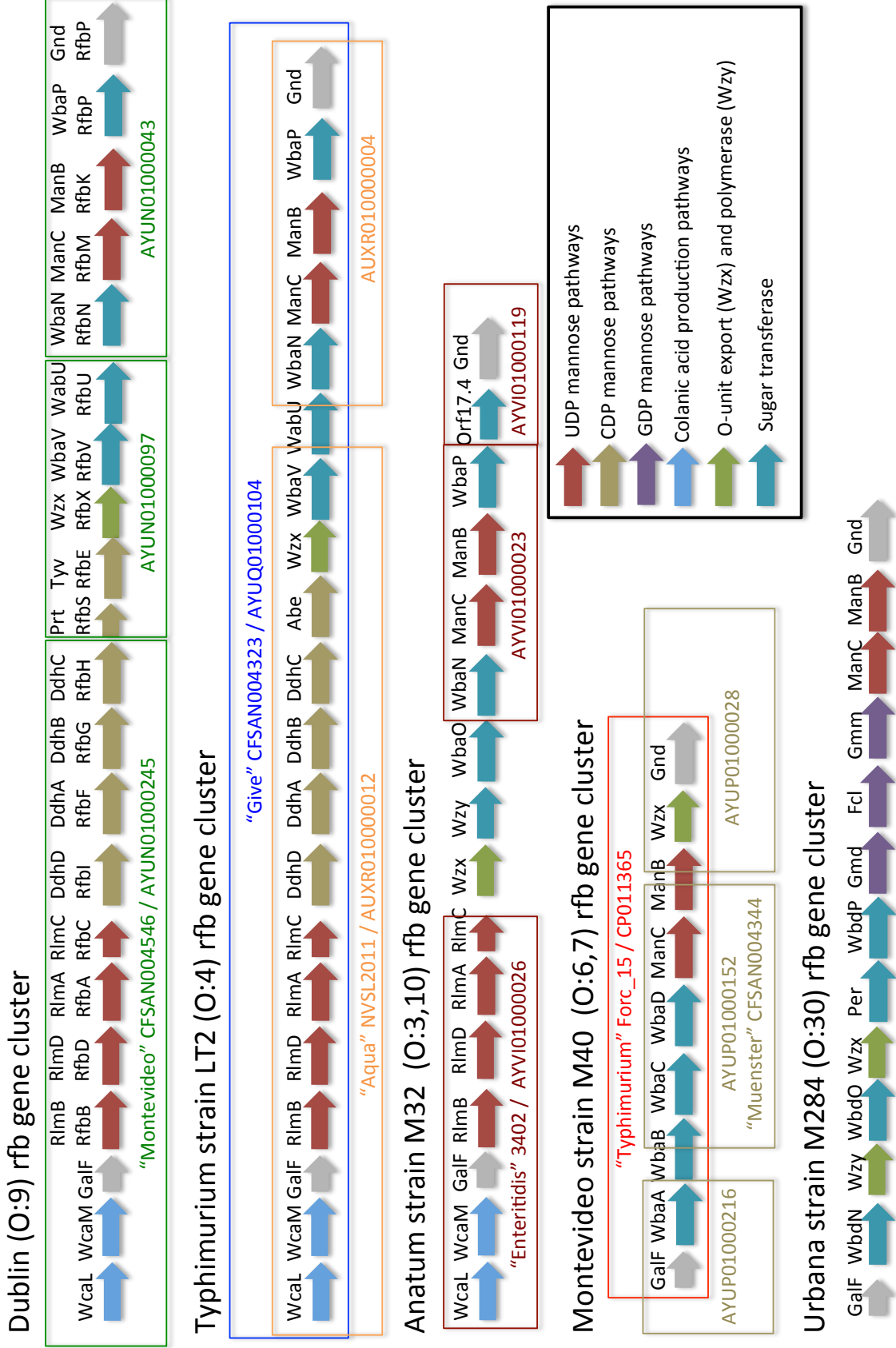


Figure S15. O-antigen synthesis (*rfb* gene) regions of the genomes of the *Salmonella* strains that carry "anomalous" tailspike genes.

Colored arrows represent genes in selected *rfb* regions from the genomes of the indicated *S. enterica* reference strains whose names are given above in large black text; gene names are shown above each map. Colored boxes and similarly colored text indicate the strains and sequence contig accession numbers of the six genomes with "anomalous" tailspike gene discussed in the text of this report. The gene order and content of each of these strains uniquely matches one of the reference regions. The reference *S. enterica rfb* gene regions shown are from the following sources: O:9 serovar Dublin, bp 2019907–1996618 of accession No. LK931502; O:4 serovar Typhimurium strain LT2, X56793 (Wyk and Reeves, 1989; Jiang et al., 1991; Liu et al., 1993; Reeves, 1993); O:3,10 serovar Anatum strain M32, X60665 (Liu et al., 1993) and ATCC BAA-1592, CP007531 (Timme et al., 2013); O:6,7 serovar Montevideo, M84642 (Reeves, 1993); and O:30 serovar Urbana strain M284, AY730593 (Jensen and Reeves, 2001; Samuel et al., 2004).

Figure S16

Novel mosaic boundary inside the tailspike gene

Red = serovar Choleraesuis str. SC-B67 locus_tag="SCH_0367"
 Blue = serovar Ohio str. CFSAN001079 locus_tag="CFSAN001079_21649"

N-terminal head-binding domain

```

1  MSDITANVVVSNRPPIFTESRSFKAVADGKIIYCKIDIDTPVNPANQIPVYIENEDGSHVOIAQPLIINSAGKIYVINGOLVKIVTVOGHSMAIYDAYGSOVDYIANVLKYDDPQFRQELAEFDGSK 125
M+DITANVVVSNRPPIFTESRSFKAVA+GKIYICKIDIDTPVNPANQIPVYIENEDGSHVOIAQPLIINSAGKIYVINGOLVKIVTVOGHSMAIYDAYGSOVDYIANVLKYDDPQFRQELAEFDGSK
1  MTDITANVVVSNRPPIFTESRSFKAVANGKIIYCKIDIDTPVNPANQIPVYIENEDGSHVQIAQPLIINSAGKIYVINGOLVKIVTVOGHSMAIYDAYGSOVDYIANVLKYDDPQFRQELAEFDGSK 125
    
```

New intra-tailspike mosaic boundary



```

126  KVGKDSNYVDP-----LNKLELKFKSFOEMRDDNSNEIGDYALLTGWHTHEHOGYAGVFCVQVDKGLTDDGGTIAVGSYAMKRITGPDATFEGVVENAGSAFDNKAVILAAAATGALIFP 243
KVGKDSNYVDT L KLELKFKSFQ MRDD+SNEIGDYALLTGMH E+QG+GAGVFCVQVDKGLTDDGGTIAVGSYAMKRITGPDATFEGVVENAGSAFDNK YI +A + I P
126  KVGKDSNYVDTLARHDNELAKLELKFKSFOAMRDDNSNEIGDYALLTGMHEEYQGHGAGVFCVQVDKGLTDDGGTIAVGSYAMKRITGPDATFEGVVENAGSAFDNKQYIESAESVKKCILP 250
    140 160 180 200 220 240
244  AGDIYTKFFTLDTYIVRGNSNMLYEINDSEATDFIVHCSRNQEQMDGRIDGHSWEGVNVVYVDPGHRPHFTYFTTNGNMRDCRF-HGGIGSWFDGYSNMFIDSCFEFSGS-----LGGENLINTPKVD 363
A + Y+ L + I + T + WD RI + + VIF+ G H+ FT ++ + G + +GV NW + C F G+ E N P V+
251  AAEXYSTKIILVNGCISGNGGI AVLRLQMSDDVTGGNYIEISAQNWDSRIKNGVIXDLAVYPLSGTVAIHSSFTQYFDIVNVTTISGGYNGMFLGEGVGNMNLARGCTFYGAKNNGCRIETRYNDP=VN 374
    260 280 300 320 340 360
364  PQQIGTWVVIHFKC-FISRSAGVYARTIGLPSVMFRDCIVVYNRD SGLLHYDEDAYPNVEFGVQVTCGCD IDSNDSGVMIRDVYVPI SNHWVSSGR--VLMQPGVVLIRCRDINNVVENSVYF 485
IGT++ F +C F SAG G + PSV+F C +XN GL + + F V + CD DSN S + V D+++NW+ R V+L C D + N +
375  LLMEIGTFFSQACAFRDT SAGPGCVVHPFSVFFEQCTFFFNATVGLYCNSN=-NIMLNFVYVNASNCDFDLSGSIHCLAVLWLDLHNHWIIGGPRDSDGLSDSLLLEDQMDFKITGNEIVE 497
    380 400 420 440 460 480
486  NGTHGISVEVCNFGTISNNNCNDNKNRGISTIQSDAGTSSKLTVSGNVCCTGFLPSLPTAQOEEGHIIEGERIVSYCNVCTGNASSQYVNSASNKQEGMNIIS 586
+G+S+ CN G+I+ N + NKN G+ + S I++ GN T A G HI + N T NA V I S
498  GTDNGLSLYRCNRGSIITGNVLSGNKNSGLFLDSCN SIAATGNLTGNLDW=TRFRQKTGAMLNGEHIVFTGNSLWKN=-TDNAINNVYVDPQVYNSVINS 595
    500 520 540 560 580 595
    
```

Figure S16. New mosaic boundary in the P22-like tailspike gene.

The newly discovered mosaic P22-like phage boundary in the tailspike gene is shown by comparing the amino acid sequences of type Va (*S. enterica* serovar Choleraesuis strain SC-B67) and type Vc (*S. enterica* serovar Ohio strain CFSAN001079) tailspikes as aligned by DNA Strider (Douglas, 1994). The N-terminal head-binding domain is indicated by the yellow box, the green box marks the N-terminal common region of the type V tailspike polysaccharide-binding domain, and the green vertical arrow marks the position of the newly discovered mosaic boundary.

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