

## Appendix

### TssA forms a gp6-like ring attached to the type VI secretion sheath

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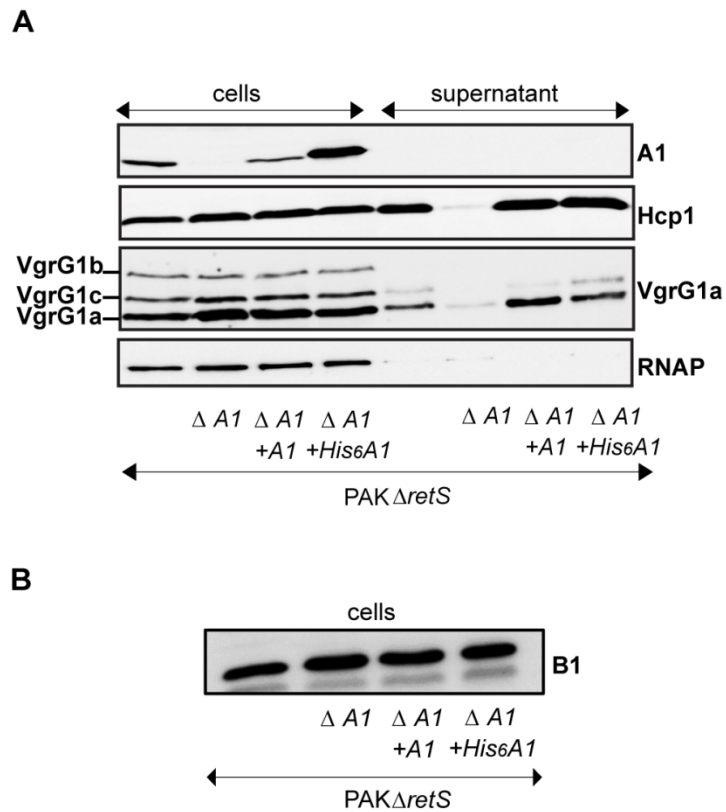
**Appendix Table S1. Strains used in this study**

Strains	Description	References
<i>E. coli</i>		
One-shot top10	F <sup>-</sup> <i>mcrA</i> $\Delta$ ( <i>mrr-hsdRMS-mcrBC</i> ) $\phi$ 80 <i>lacZ</i> $\Delta$ M15 $\Delta$ <i>lacX74</i> <i>recA1</i> <i>araD139</i> $\Delta$ ( <i>ara-leu</i> )	Invitrogen
B834(DE3)	F <sup>-</sup> <i>ompT</i> <i>hsdSB</i> (rB <sup>-</sup> mB <sup>-</sup> ) <i>gal dcm met</i> (DE3)	Laboratory collection
DHM1	<i>cya-854</i> <i>recA1</i> <i>gyrA96</i> (NaI) <i>thi1</i> <i>hsdR17</i> <i>spoT1</i> <i>rfbD1</i> <i>glnV44</i> (AS)	Karimova et al., 1998
C118 $\lambda$ pir	Host strain for pKNG101 replication; D ( <i>ara-leu</i> ) <i>araD</i> <i>DlacX74</i> <i>galE</i> <i>galK-phoA20</i> <i>thi-1</i> <i>rpsE</i> <i>rpoB</i> <i>argE</i> (Am) <i>recA1</i> <i>Rf</i> <sup>*</sup> ( <i>lpir</i> )	Herrero et al., 1990
<i>P. aeruginosa</i>		
PAK $\Delta$ <i>retS</i>	In-frame deletion of <i>retS</i> (PA4856) in <i>P. aeruginosa</i> PAK wild-type strain	Goodman et al., 2004
PAK $\Delta$ <i>retS</i> -pBBR	PAK $\Delta$ <i>retS</i> strain carrying the empty vector pBBR1MCS-4	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i>	In-frame deletion of <i>tssAI</i> (PA0082) in PAK $\Delta$ <i>retS</i>	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> -pBBR	PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> mutant carrying the empty vector pBBR1MCS-4	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> -pBBR- <i>AI</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> mutant carrying the pBBR1MCS-4 vector expressing <i>tssAI</i> (pBBR- <i>AI</i> )	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> -pBBR- <i>His<sub>6</sub>AI</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> mutant carrying the pBBR1MCS-4 vector expressing the N-terminally His <sub>6</sub> tagged <i>tssAI</i> (pBBR- <i>His<sub>6</sub>AI</i> )	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>E1</i>	In-frame deletion of <i>tssE1</i> (PA0087) in PAK $\Delta$ <i>retS</i>	Lossi et al., 2011
PAK $\Delta$ <i>retS</i> $\Delta$ <i>E1</i> -pBBR	PAK $\Delta$ <i>retS</i> $\Delta$ <i>E1</i> mutant carrying the empty vector pBBR1MCS-4	
PAK $\Delta$ <i>retS</i> $\Delta$ <i>E1</i> -pBBR- <i>E1</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>E1</i> mutant carrying the pBBR1MCS-4 vector expressing <i>tssE1</i> (pBBR- <i>E1</i> )	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>B1</i> :: <i>B1-sfGFP</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>B1</i> strain having the gene encoding the C-terminal fusion of the sfGFP with <i>tssB1</i> ( <i>B1-sfGFP</i> ) under a plac promoter integrated into the <i>att</i> site	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> $\Delta$ <i>B1</i> :: <i>B1-sfGFP</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> $\Delta$ <i>B1</i> strain having the gene encoding the <i>B1-sfGFP</i> under a plac promoter integrated into the <i>att</i> site.	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>B1</i> $\Delta$ <i>E1</i> :: <i>B1-sfGFP</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>B1</i> $\Delta$ <i>E1</i> strain having the gene encoding the <i>B1-sfGFP</i> under a plac promoter integrated into the <i>att</i> site	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> $\Delta$ <i>B1</i> :: <i>B1-sfGFP</i> -pBBR- <i>AI</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>AI</i> $\Delta$ <i>B1</i> :: <i>B1-sfGFP</i> carrying the pBBR1MCS-4 vector expressing <i>tssAI</i> (pBBR- <i>AI</i> )	This study
PAK $\Delta$ <i>retS</i> $\Delta$ <i>B1</i> $\Delta$ <i>E1</i> :: <i>B1-sfGFP</i> -pBBR- <i>E1</i>	PAK $\Delta$ <i>retS</i> $\Delta$ <i>B1</i> $\Delta$ <i>E1</i> :: <i>B1-sfGFP</i> carrying the pBBR1MCS-4 vector expressing <i>tssE1</i> (pBBR- <i>E1</i> )	This study

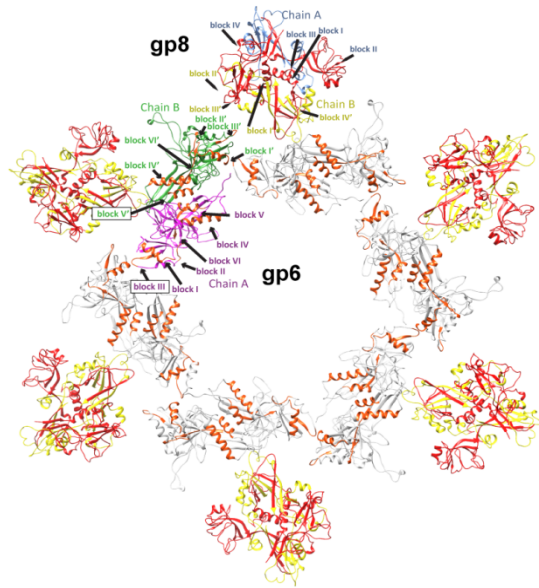
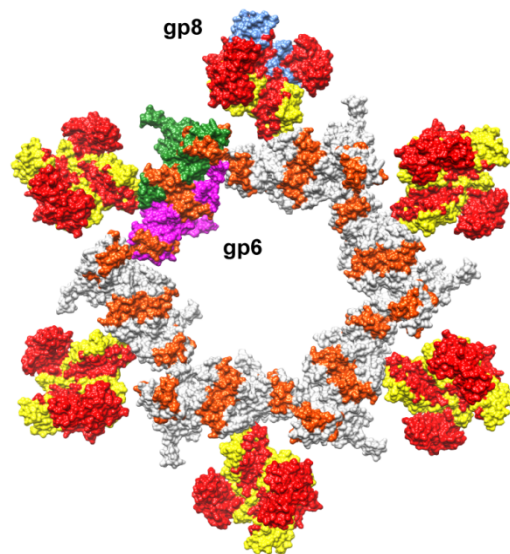
**Appendix Table S2. Plasmids used in this study**

Plasmids	Description	References
pKNG101- $\Delta$ <i>AI</i>	pKNG101 containing the mutator fragment for deletion of <i>tssAI</i> , Str <sup>R</sup>	This study
Mini-CTX- <i>B1-sfGFP</i>	Mini-CTX-1 encoding <i>tssB1-sfGFP</i> under a <i>plac</i> promoter for integration into the <i>att</i> site of the <i>P. aeruginosa</i> chromosome, Tc <sup>R</sup>	This study
pBBR	pBBR1MCS-4, Broad host range vector pBBR1 replicon, Ap <sup>R</sup>	Kovach et al., 1995
pBBR- <i>AI</i>	pBBR1MCS-4 containing <i>tssAI</i>	This study
pBBR- <i>His<sub>6</sub>AI</i>	pBBR1MCS-4 containing <i>His<sub>6</sub>tssAI</i>	This study
pBBR- <i>EI</i>	pBBR1MCS-4 containing <i>tssEI</i>	This study
pET28	Expression vector used for expression of N-terminal His <sub>6</sub> tagged proteins, Km <sup>R</sup>	Novagen
pET26	Expression vector used for expression of untagged proteins, Km <sup>R</sup>	Novagen
pACYC	pACYC-Duet vector, Cm <sup>R</sup>	Novagen
pET- <i>His<sub>6</sub>AI</i>	pET28 expressing TssA1 with an N-terminal His <sub>6</sub> tag (His <sub>6</sub> -TssA1)	This study
pET26- <i>B1C1</i>	pET26 expressing untagged TssB1C1	This study
pACYC- <i>AI-B1C1</i>	pACYC-Duet-1 expressing His <sub>6</sub> -tagged TssA1 and TssB1C1 without tag from two separate T7 promoters	This study
pKT25	BTH vector for fusion of target proteins to <i>B. pertussis cya</i> gene T25 fragment; P <sub>lac</sub> :: <i>cya</i> <sup>1-675</sup> p15ori, Km <sup>R</sup>	Karimova et al., 1998
pUT18C	BTH vector for fusion of target proteins to <i>B. pertussis cya</i> gene T18 fragment; P <sub>lac</sub> :: <i>cya</i> <sup>675-1197</sup> pUCori, Ap <sup>R</sup>	Karimova et al., 1998
pKT25- <i>AI</i>	Fusion of <i>tssAI</i> to <i>cya</i> gene T25 fragment in pKT25, Km <sup>R</sup>	Lossi et al., 2012
pUT18C- <i>AI</i>	Fusion of <i>tssAI</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	This study
pUT18C- <i>Hcp1</i>	Fusion of <i>hcp1</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	This study
pUT18C- <i>EI</i>	Fusion of <i>tssEI</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	Lossi et al., 2012
pUT18C- <i>F1</i>	Fusion of <i>tssF1</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	Lossi et al., 2012
pUT18C- <i>G1</i>	Fusion of <i>tssG1</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	Lossi et al., 2012
pUT18C- <i>K1</i>	Fusion of <i>tssK1</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	This study
pUT18C- <i>VgrG1a</i>	Fusion of <i>vgrG1a</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	This study
pUT18C- <i>ClpV1</i>	Fusion of <i>ClpV1</i> to <i>cya</i> gene T18 fragment in pUT18C, Ap <sup>R</sup>	Förster et al., 2014

Ap<sup>R</sup>, ampicillin-resistant; Cm<sup>R</sup>, chloramphenicol-resistant; Km<sup>R</sup>, kanamycin-resistant; Str<sup>R</sup>, streptomycin-resistant; Tc<sup>R</sup>, tetracyclin-resistant.



**Appendix Figure S1. The N-terminally His<sub>6</sub>-tagged TssA1 is functional. A.** Proteins from whole cell extracts and culture supernatants of PAK $\Delta$ *retS*, the derivative *tssA1* mutant ( $\Delta$ *A1*) (both carrying the pBBR plasmid) and the complemented  $\Delta$ *A1* mutant carrying pBBR-*tssA1* ( $\Delta$ *A1*+*A1*) or pBBR-*His<sub>6</sub>-tssA1*( $\Delta$ *A1*+His<sub>6</sub>*A1*) were analysed by Western blotting. Polyclonal antibodies directed against TssA1 (A1), Hcp1 and VgrG1a were used. The anti-VgrG1a antibody detects VgrG1a, VgrG1b and VgrG1c, as indicated on the left. RNA polymerase (RNAP) was monitored in both whole-cell lysates and culture supernatants using monoclonal antibody directed against its  $\beta$ -subunit. **B.** Western blot analysis showing the production of TssB1 in PAK $\Delta$ *retS*,  $\Delta$ *A1* (both carrying the pBBR plasmid) and the complemented strains  $\Delta$ *A1*+*A1* and  $\Delta$ *A1*+His<sub>6</sub>*A1*. Polyclonal antibody directed against TssB1 was used.

**A****B**

**Appendix Figure S2. The gp6/gp8 baseplate subcomplex of the T4 phage. (A), Cartoon and (B), surface representations of the dodecameric gp6 structure (PDB code 3H3W) and the structure of gp8 (PDB code 1PDM) are shown. In both representations, chain A of the gp6 dimers is shown in magenta and chain B in green and the TssA1/gp6 conserved regions are shown in orange in the gp6 ring. The TssA1/gp8 conserved blocks are shown in red in the 3D structure of gp8. The conservation is most significant in regions involved in gp8 dimerization and interaction with gp6.**