

Additional file 2: Supplementary figures for
Comparative genomics and transcriptomics of *Pichia pastoris*

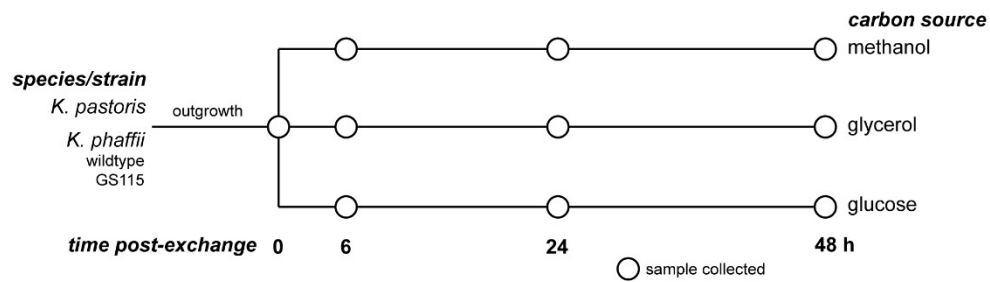


Figure S1: Experimental timeline for RNA sequencing of *Komagatella* strains.

Schematic timeline for collection of RNA samples during batch cultivation of strains in shake flasks on 3 different carbon sources. Three independent cultivations were sampled for each time point.

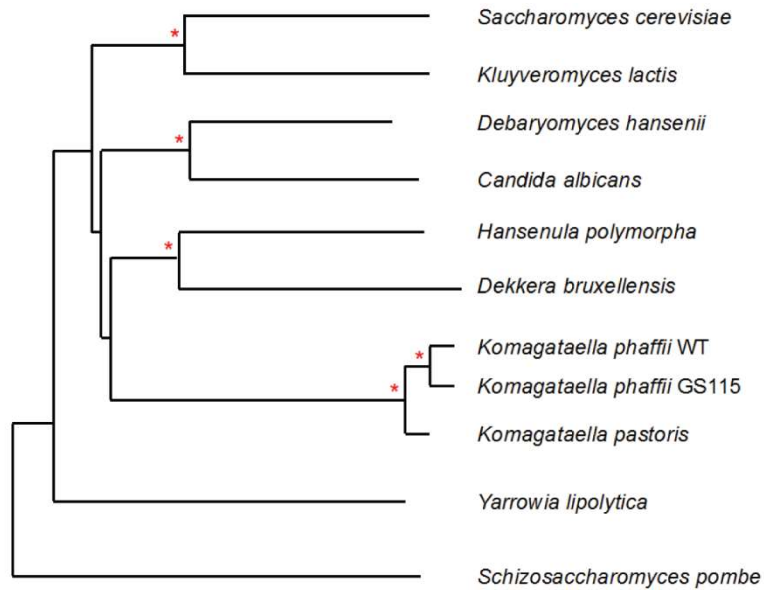
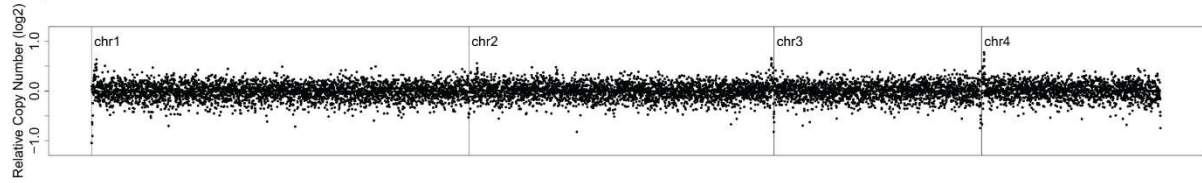
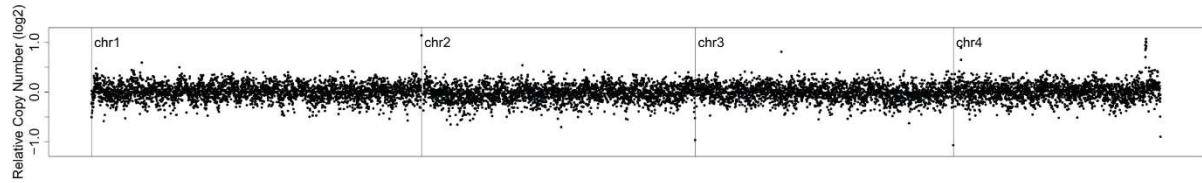


Figure S2: Phylogenetic comparison of *K. pastoris* and *K. phaffii* to other related yeasts. Phylogeny was generated using a concatenated, gap-free alignment of ten orthologous proteins. Phylogenetic tree was calculated using neighbor-joining, distance-based, maximum likelihood and maximum parsimony methods; reliability was assessed using bootstrapping. Clades marked with an asterisk are supported by 100% of bootstrap replicates in all four methods.

a) *K. pastoris*



b) *K. phaffii*



c) *K. phaffii* (GS115)

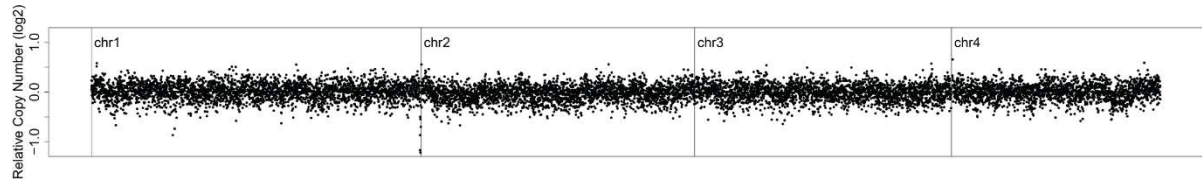


Figure S4: Copy number determination for major chromosomes in a) *K. pastoris*, b) *K. phaffii* wild-type and c) *K. phaffii* GS115 strains.

a) *K. pastoris*

		Second Base				Third Base			
		U	C	A	G				
U	UUU	1.14	UCU	1.56	UAU	1.03	UGU	1.24	U
	UUC	0.86	UCC	1.04	UAC	0.97	UGC	0.76	C
	UUA	1.07	UCA	1.25	UAA	Set 0.00	UGA	Set 0.00	A
	UUG	1.71	UCG	0.57	UAG	Set 0.00	UGG	1.00	G
C	CUU	1.02	CCU	1.35	CAU	1.23	CGU	0.84	U
	CUC	0.51	CCC	0.73	CAC	0.77	CGC	0.32	C
	CUA	0.76	CCA	1.51	CAA	1.20	CGA	0.72	A
	CUG	0.93	CCG	0.41	CAG	0.80	CGG	0.32	G
A	AUU	1.38	ACU	1.46	AAU	1.07	AGU	0.95	U
	AUC	0.91	ACC	0.96	AAC	0.93	AGC	0.63	C
	AUA	0.71	ACA	1.09	AAA	1.04	AGA	2.72	A
	AUG	Met 1.00	ACG	0.49	AAG	0.96	AGG	1.09	G
G	GUU	1.55	GCU	1.62	GAU	1.23	GGU	1.43	U
	GUC	0.87	GCC	0.97	GAC	0.77	GGC	0.63	C
	GUA	0.72	GCA	1.09	GAA	1.18	GGA	1.45	A
	GUG	0.87	GCG	0.32	GAG	0.82	GGG	0.49	G

b) *K. phaffii*

		Second Base				Third Base			
		U	C	A	G				
U	UUU	1.16	UCU	1.56	UAU	1.03	UGU	1.24	U
	UUC	0.84	UCC	1.04	UAC	0.97	UGC	0.76	C
	UUA	1.07	UCA	1.25	UAA	Set 0.00	UGA	Set 0.00	A
	UUG	1.74	UCG	0.57	UAG	Set 0.00	UGG	1.00	G
C	CUU	1.03	CCU	1.36	CAU	1.23	CGU	0.84	U
	CUC	0.51	CCC	0.72	CAC	0.77	CGC	0.31	C
	CUA	0.74	CCA	1.52	CAA	1.21	CGA	0.72	A
	CUG	0.91	CCG	0.41	CAG	0.79	CGG	0.32	G
A	AUU	1.40	ACU	1.46	AAU	1.07	AGU	0.95	U
	AUC	0.89	ACC	0.95	AAC	0.93	AGC	0.63	C
	AUA	0.71	ACA	1.10	AAA	1.04	AGA	2.72	A
	AUG	Met 1.00	ACG	0.49	AAG	0.96	AGG	1.09	G
G	GUU	1.56	GCU	1.60	GAU	1.23	GGU	1.43	U
	GUC	0.86	GCC	0.97	GAC	0.77	GGC	0.63	C
	GUA	0.72	GCA	1.10	GAA	1.19	GGA	1.46	A
	GUG	0.87	GCG	0.32	GAG	0.81	GGG	0.49	G

Figure S5: Codon usage for a) *K. pastoris* and b) *K. phaffii* as determined from all coding sequences identified in genome annotation. The relative abundance observed for each codon is represented as a percentage of total codon usage for the corresponding amino acid.

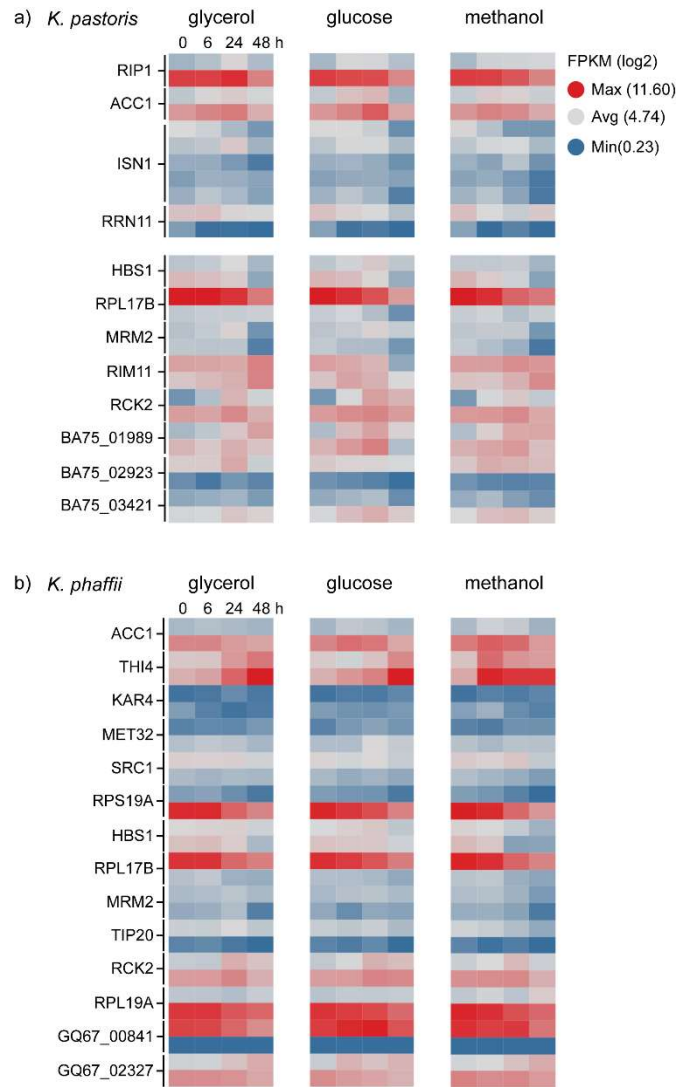


Figure S6: Isoform expression in *K. pastoris* and *K. phaffii* as a function of cultivation conditions. Heat maps of gene expression (log2 fpkm) for isoforms of alternatively spliced genes that alter coding sequences in a) *K. pastoris* and b) *K. phaffii* detected in initial genome annotation. Alternatively spliced genes with sufficient homology to *S. cerevisiae* are named, otherwise gene identifiers from genome annotation are used. Isoform expression is shown as a function of batch growth in glycerol, glucose or methanol during a 48 h cultivation period.

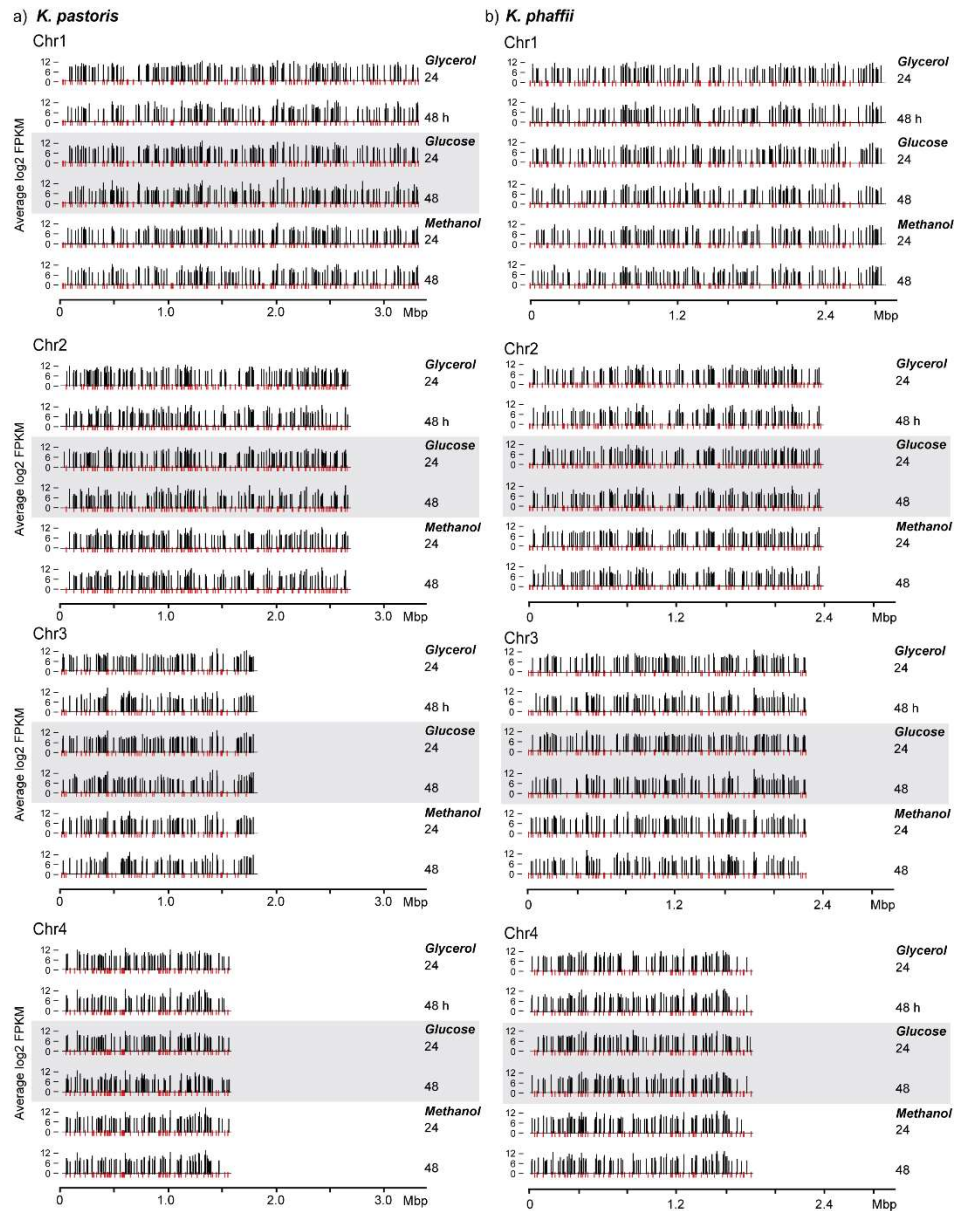


Figure S7: Chromosomal locations of highly expressed genes. Map of chromosomal location (base pair identity) for the most highly expressed genes (top 10% expression) in a) *K. pastoris* and b) *K. phaffii*. Black lines indicate gene expression level at 24 h time points during batch cultivation in either glycerol, glucose or methanol. Red lines indicate locations of GC-rich autonomously replicating sequence (GC-ARS) motifs identified by BLAST.

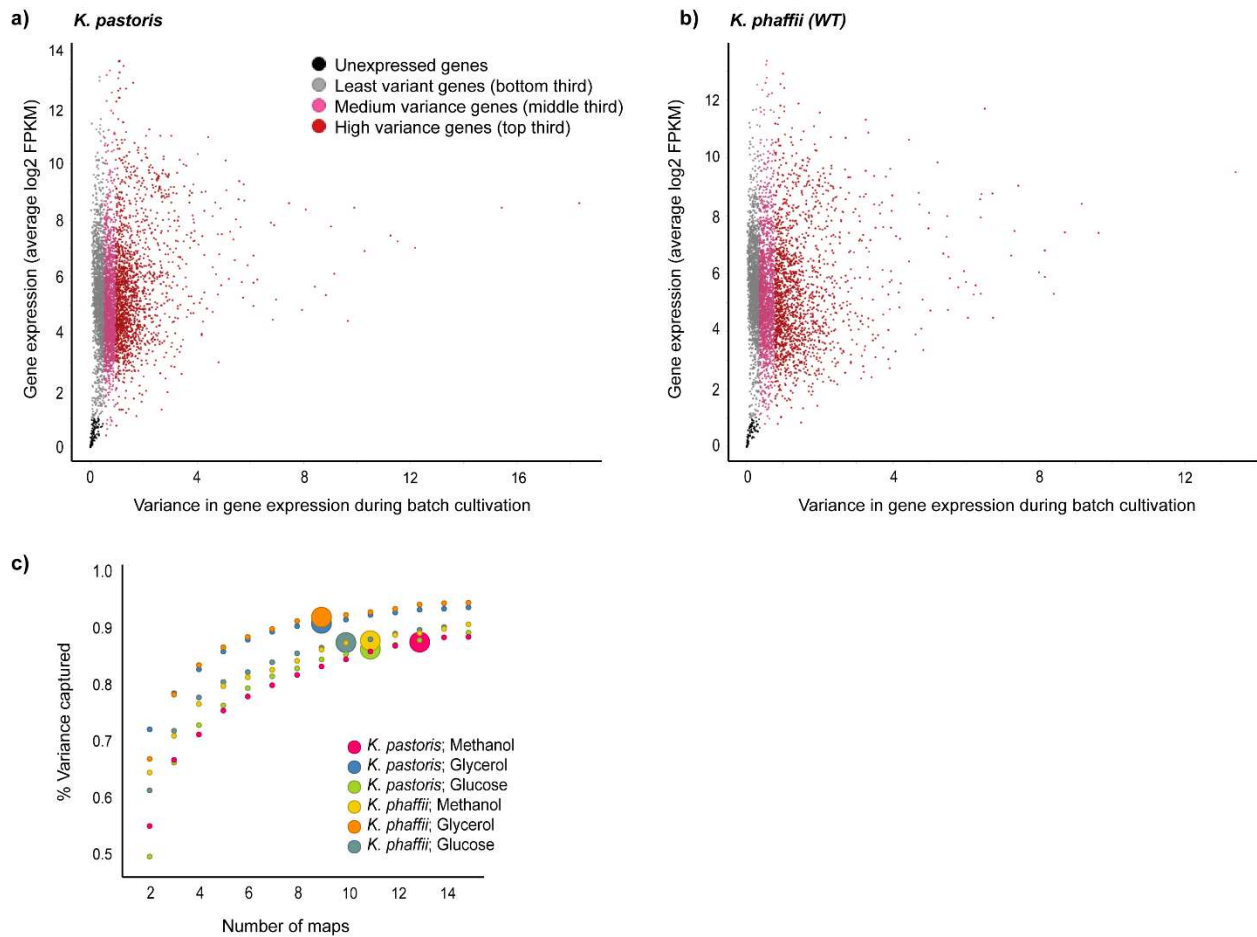


Figure S8: Variance in gene expression during batch cultivation of *K. pastoris* and *K. phaffii*. Scatter plots of the average variance versus expression observed for all annotated genes across the 10 conditional averages generated from either a) *K. pastoris* or b) *K. phaffii* expression data. Genes with average log2 fpkm <1 and variance < 0.05 were excluded from further analyses. c) Elbow analysis of input cluster number to identify optimal expression data clustering by self-organizing maps (SOMs). Large circles denote the number of clusters for each expression data set where the additional variance captured by further clustering was < 1%.

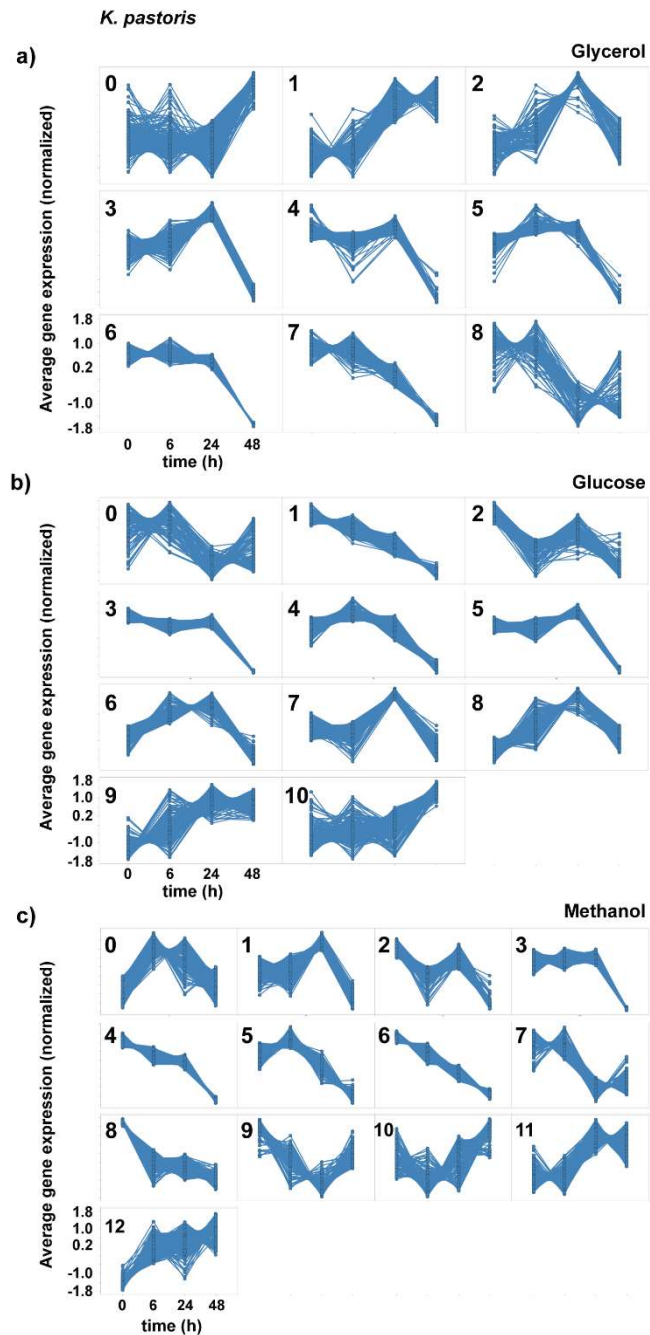


Figure S9: Gene expression phenotypes in *K. pastoris* as a function of cultivation conditions. Self-organizing maps (SOMs) of genes changing expression similarly in *K. pastoris* during a 48 h batch cultivation in a) glycerol, b) glucose or c) methanol.

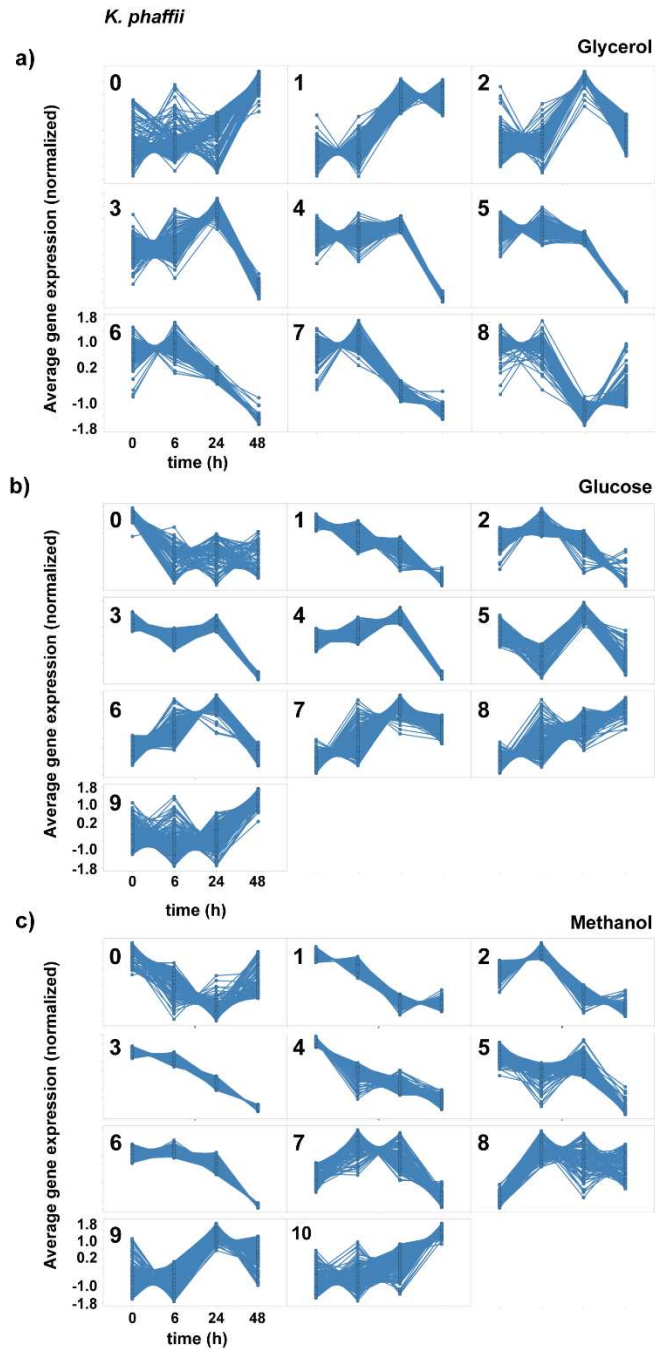


Figure S10: Gene expression phenotypes in *K. phaffii* as a function of cultivation conditions. Self-organizing maps (SOMs) of genes changing expression similarly in *K. phaffii* during a 48 h batch cultivation in a) glycerol, b) glucose or c) methanol.

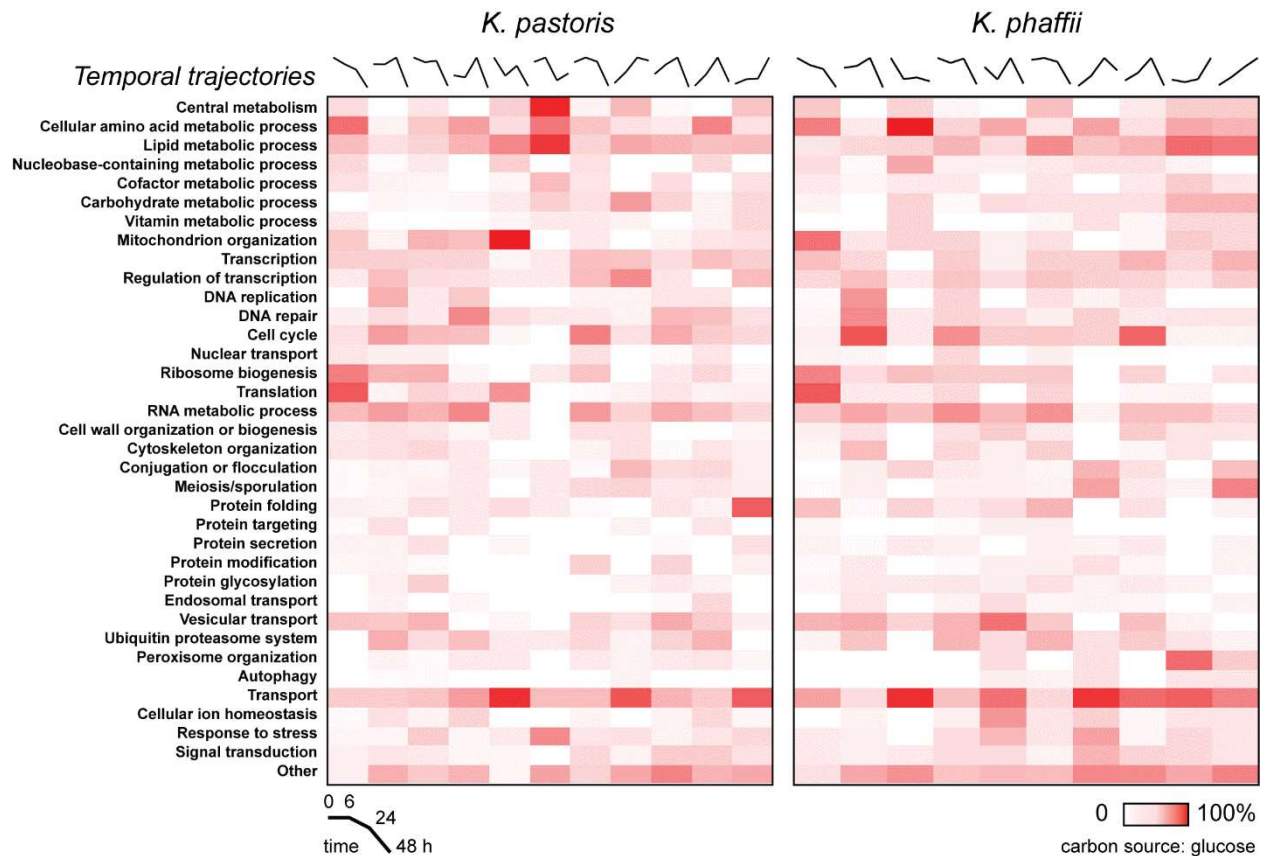


Figure S11: Biological process enrichment as a function of cultivation in glucose. Heat map representation of the enrichment of GO biological process terms for expression phenotypes observed in *K. pastoris* and *K. phaffii* during a 48 h batch cultivation in glucose system as characterized by self-organizing maps (SOMs). Representative temporal trajectories of gene expression were generated for each SOM by averaging expression data at each time point for genes present within a given map. Color density relates to the number of genes assigned to a particular process as a percentage of the total number genes present in a particular expression phenotype or map.

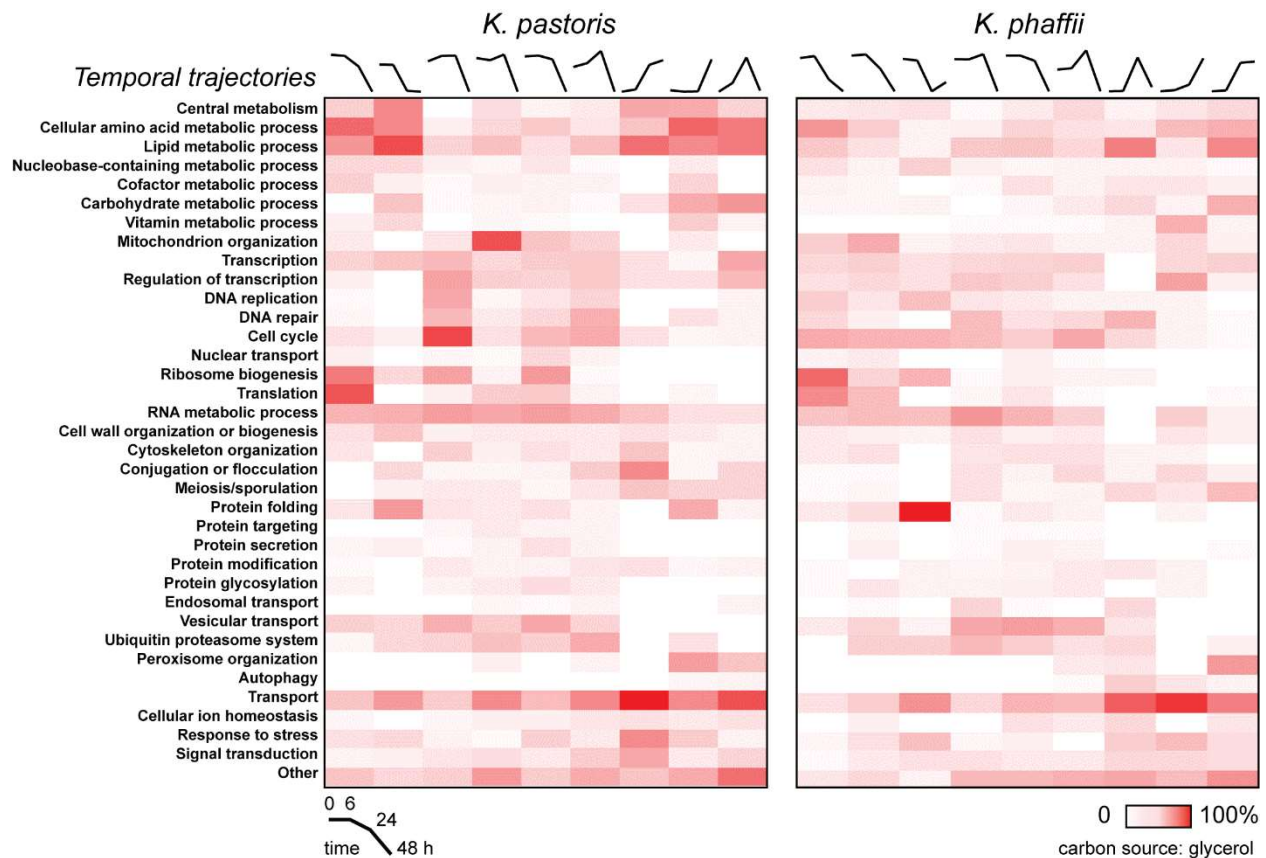


Figure S12: Biological process enrichment as a function of cultivation in glycerol. Heat map representation of the enrichment of GO biological process terms for expression phenotypes observed in *K. pastoris* and *K. phaffii* during a 48 h batch cultivation in glycerol as characterized by self-organizing maps (SOMs). Representative temporal trajectories of gene expression were generated for each SOM by averaging expression data at each time point for genes present within a given map. Color density relates to the number of genes assigned to a particular process as a percentage of the total number genes present in a particular expression phenotype or map.

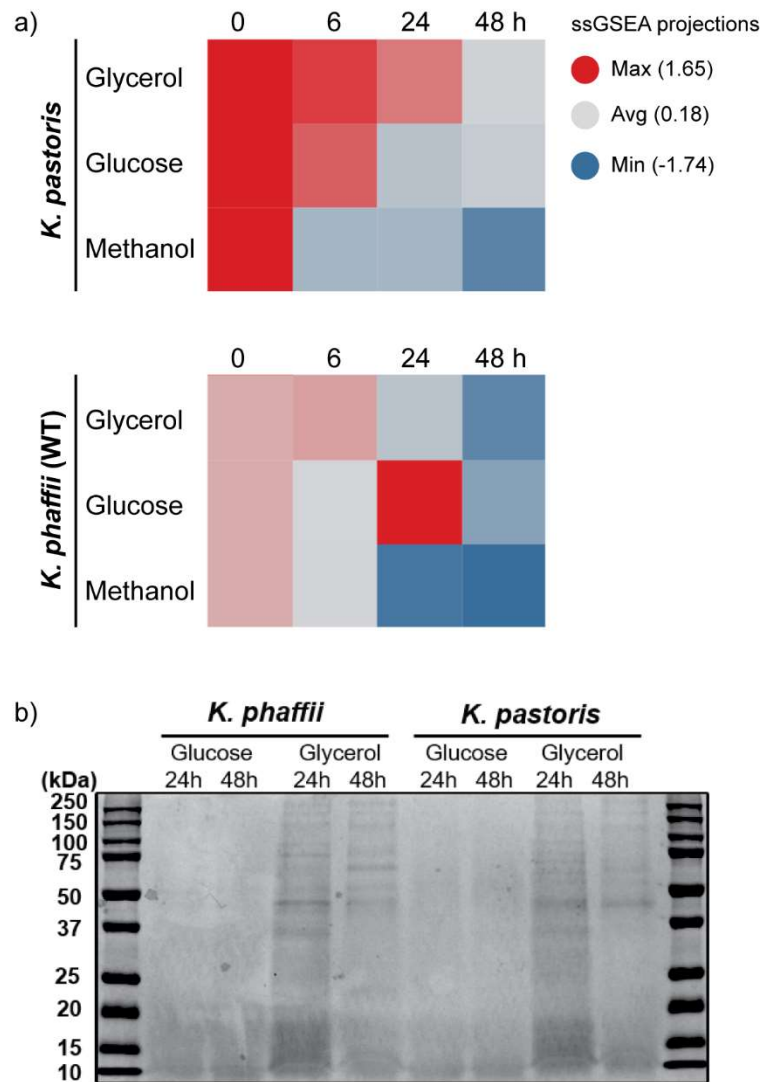


Figure S13: Secretory pathway protein expression in *K. pastoris* and *K. phaffii*.

a) Row normalized single set Gene Set Enrichment Analysis (ssGSEA) projections for 170 proteins bearing a signal peptide as identified by Signalp. b) SDS-PAGE analysis of host-cell protein expression in supernatants during batch cultivation of *K. pastoris* and *K. phaffii* for 48h in glucose or glycerol-containing media.

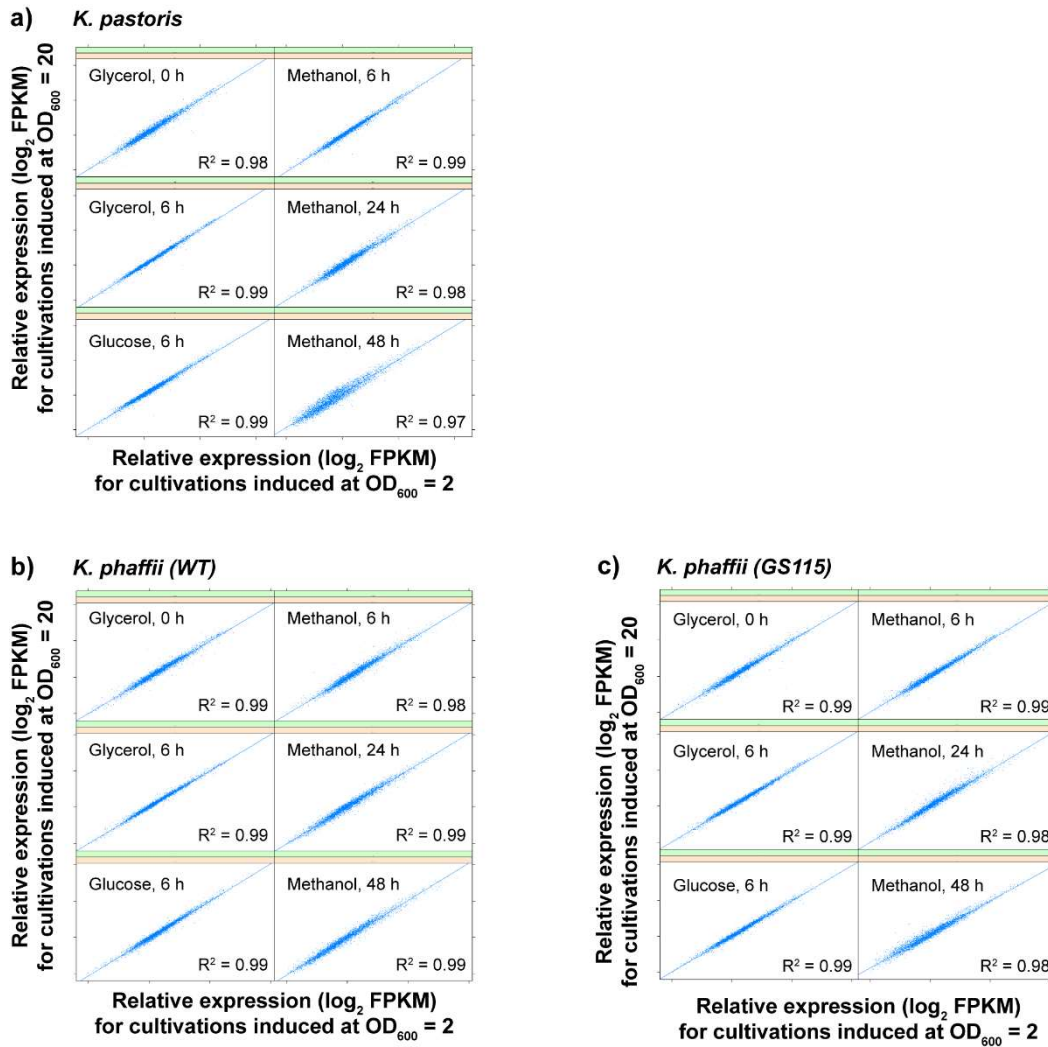


Figure S14: Correlation of gene expression at two different cultivation densities.

Scatter plots of gene expression between similar cultivation conditions for a) *K. pastoris*,

b) wildtype *K. phaffii*, and c) *K. phaffii* GS115 grown at two different cell densities.

Density A corresponds to cultures outgrown to $OD_{600} = 2.0$ prior to sampling and

Density B corresponds to cultures outgrown to $OD_{600} = 20$ prior to sampling. Pearson

correlation coefficients were calculated from expression vectors that were averages of

three biological replicates for each cultivation condition and density.