

Supporting information

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Cleavage of model substrates by *Arabidopsis* PRORP1 reveals new

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insights into its substrate requirements

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11 **Supporting Tables A-B and Figures A-C**

12 **Table A**
 13 Analysis of N₋₁:N₊₇₃ identities in mitochondrial and chloroplast tRNAs from eight
 14 different green algae and plants

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17 Composite (total of 423 tRNAs)

N ₋₁ \ N ₊₇₃	A	U	G	C
A	78	27	42	9
U	77	32	40	11
G	42	10	15	8
C	23	5	3	1

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20 *Arabidopsis thaliana*

N ₋₁ \ N ₊₇₃	A	U	G	C
A	7	3	5	4
U	5	2	5	1
G	9	2	1	2
C	2	1	1	0

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22 *Cyanophora paradoxa*

N ₋₁ \ N ₊₇₃	A	U	G	C
A	12	5	8	0
U	15	5	8	3
G	1	1	1	0
C	2	1	0	1

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24 *Ectocarpus siliculosus*

N ₋₁ \ N ₊₇₃	A	U	G	C
A	10	5	4	2
U	12	7	4	1
G	3	0	3	0
C	1	0	0	0

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26 *Oryza sativa japonica*

N ₋₁ \ N ₊₇₃	A	U	G	C
A	9	4	5	1
U	8	3	4	0
G	6	0	5	3
C	2	0	0	0

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28 *Ostreococcus tauri*

N ₋₁ \ N ₊₇₃	A	U	G	C
A	2	2	8	0
U	13	5	5	3
G	2	0	1	0
C	8	0	2	0

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30 *Phaeodactylum tricornutum*

N₋₁ \ N₊₇₃	A	U	G	C
A	14	2	4	1
U	10	5	3	1
G	2	1	1	0
C	5	1	0	0

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33 *Physcomitrella patens*

N₋₁ \ N₊₇₃	A	U	G	C
A	15	4	4	1
U	9	3	5	1
G	8	2	0	1
C	1	1	0	0

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36 *Solanum tuberosum*

N₋₁ \ N₊₇₃	A	U	G	C
A	9	2	4	0
U	5	2	6	1
G	11	4	3	2
C	2	1	0	0

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39 For the sequence analysis above (Tables S1 and S2), we first obtained tRNA
40 sequences from <http://plantrna.ibmp.cnrs.fr/>. We examined eight algal/plant species
41 that appear to have the entire suite of organellar tRNAs. We filtered duplicated genes,
42 classified as such by the presence of identical upstream and body sequences. All the
43 tRNA sequences were downloaded, collated using a Microsoft Excel spreadsheet, and
44 analyzed for identity distributions at desired locations.

45

46 **Table B**
 47 Analysis of N₊₁ identities of mitochondrial and chloroplast tRNAs from eight
 48 different green algae and plants
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	A	U	G	C
<i>Ath</i> (50)	10	4	34	2
<i>Cpa</i> (63)	9	5	44	5
<i>Esi</i> (52)	0	6	43	3
<i>Osa</i> (50)	7	4	38	1
<i>Ota</i> (51)	3	5	41	2
<i>Ptr</i> (50)	8	4	35	3
<i>Ppa</i> (55)	6	3	43	3
<i>Stu</i> (52)	7	5	38	2
Total (423)	50	36	316	21

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 51
 52 Abbreviations used: *Ath*, *Arabidopsis thaliana*; *Cpa*, *Cyanaphora paradoxa*; *Esi*,
 53 *Ectocarpus siliculosus*; *Osa*, *Oryza sativa*; *Ota*, *Ostreococcus tauri*; *Ptr*,
 54 *Phaeodactylum tricornutum*; *Ppa*, *Physcomitrella patens*; *Stu*, *Solanum tuberosum*.

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58 Supportin figure legends**59 Figure A**

60 Representative gels showing *At*PRORP1-mediated cleavage of pATSer variants, lacking
61 the 3' CCA-motif. Lanes 1 to 2 and 6 to 9 are negative controls (loaded in the same order
62 as with *At*PRORP1); M (size marker lanes 3 and 10) indicates cleavage of pATSerUG by
63 *Eco* RPR. Lane 4, pATSerCG_{Δ3'CCAC}; lane 5, pATSerCIno_{Δ3'CCAC}; lane 11,
64 pATSerUG_{Δ3'CCAC}; lane 12, pATSerUG_{Δ3'CAC}; lane 13, pATSerUG_{Δ3'AC}; and lane 14,
65 pATSerUG. The final concentration of *At*PRORP1 was 6.6 μM and the reactions were
66 performed in the presence of 10 mM Mg²⁺ for 60 min at 37°C (for details, see Materials
67 and Methods). The position of each 5' cleavage fragment (5' CL Frags) is indicated.

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69 Figure B

70 Representative data from fluorescence polarization binding assays performed with
71 *At*PRORP1 and 3'-fluorescein-labeled pSu1(-1U), pATSerUG and pATSerUG_{GAAA}.

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73 Figure C

74 Representative gels showing *At*PRORP1-mediated cleavage of pATSerUG and
75 pATSerCG with 2'OH and 2'H at N₋₁. Lanes 1 to 4 are negative controls (loaded in the
76 same order as with *At*PRORP1); and M (size marker, lane 5) indicates cleavage of
77 pATSerUG by *Eco* RPR. Lane 6, pATSerUG; lane 7, pATSerCG; lane 8,
78 pATSerU_{deoxy}G; and lane 9, pATSerC_{deoxy}G. The final concentration of *At*PRORP1 was
79 11 μM and the reactions were performed in the presence of 10 mM Mg²⁺ for 60 min at
80 37°C (for details, see *Materials and Methods*). The position of each 5' cleavage fragment
81 (5' CL Frags) is indicated.

Fig A

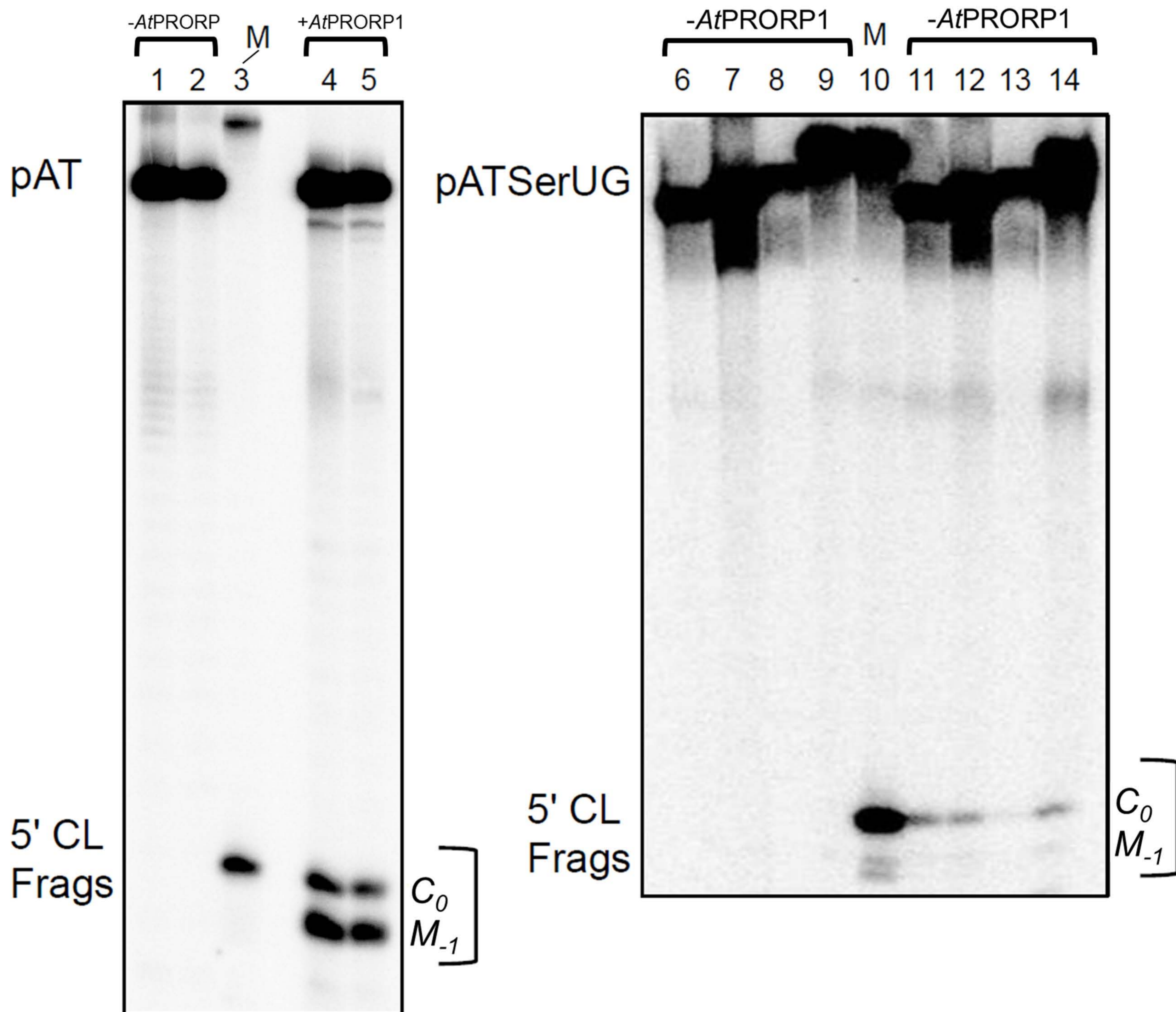


Fig B

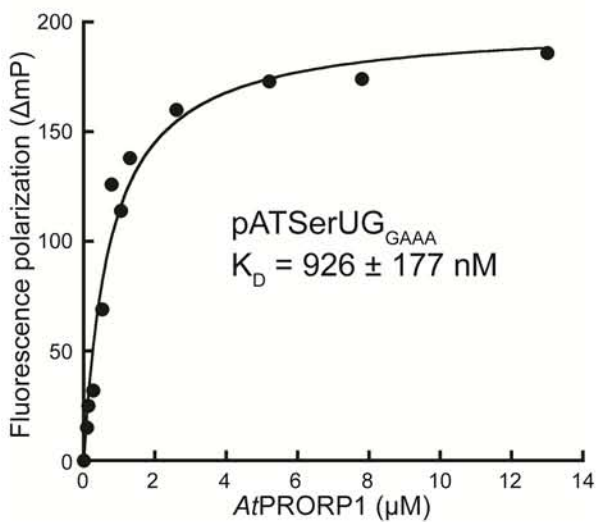
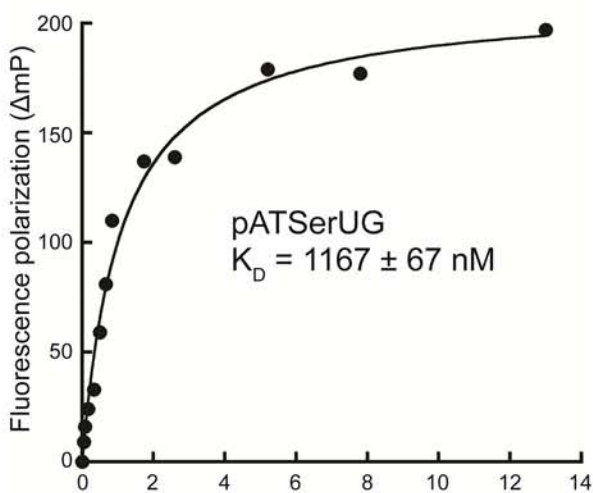
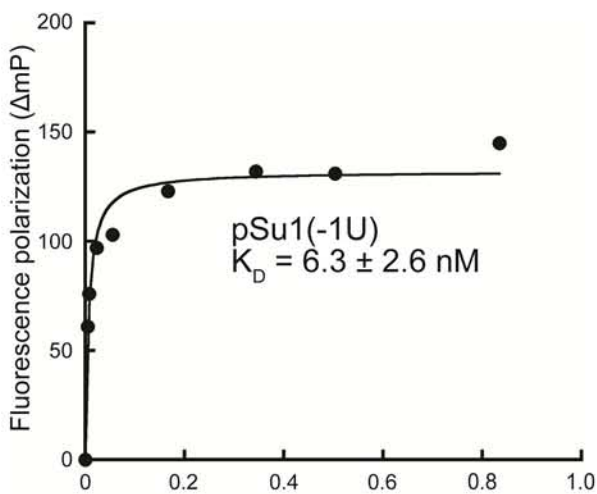


Fig C

