Crystal structure of *Clostridium difficile* Toxin A

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Supplementary information Table 1: Data collection and refinement statistics.

	Native	HgCl ₂ (90 min)	HgCl ₂ (3 days)	S1329C HgCl ₂ (3 days)
Data collection				(
Wavelength (A) Cell dimensions	1.28149	1.00273	1.00275	1.00798
a, b, c (Å)	303.61,	300.34,	302.0, 123.99,	300.87,
$\alpha \beta \gamma (^{\circ})$	124.34, 73.98	123.80, 75.83	/5./0	124.34, 70.34
$\alpha, p, \gamma()$	50, 97.5, 90 50 0-3 25 (3 31	50, 50.0, 50 50, 0-3, 6 (3, 66-	50,0-3,8 (3,87-	50 0-4 15 (4 22
Resolution (Å)	3 25)*	3 6)	3 8)	4 15)
R	6 2 (63 4)	69(507)	8 0 (68 2)	87 (57 1)
$I/\sigma I$	21.0(2.5)	9.6 (1.8)	10.3(1.9)	10.4 (2.3)
Completeness (%)	100 (99.9)	99.3 (99.3)	99.8 (99.8)	99.7 (99.5)
Redundancy	6.0 (6.0)	2.7 (2.7)	3.8 (3.8)	5.1 (5.1)
	K ₂ PtCl ₂	HAuCl ₄		
Wavelength (Å) Cell dimensions	1.06578	1.03501		
a h c (Å)	315.88,	297.66,		
<i>u</i> , <i>b</i> , <i>c</i> (<i>H</i>)	123.41, 75.43	123.92, 75.52		
α, β, γ (°)	90, 97, 90	90, 98.2, 90		
Resolution (Å)	50.0-4.1 (4.17- 4.1)	50.0-3.5 (3.56- 3.5)		
$R_{\rm merge}$	5.4 (51.9)	6.1 (44.1)		
I/oI	15.9 (2.2)	16.0 (2.5)		
Completeness (%)	99.9 (100.0)	99.4 (95.8)		
Redundancy	4.0 (4.0)	5.2 (4.5)		
Refinement				
Resolution (Å)	50-3.26			
No. reflections	43,813			
$R_{\text{work}}/R_{\text{free}}^{\dagger}$ (%)	18.21/23.74			
No. atoms	14412			
Protein	14411			
Zinc	1			
Water	0			
B-factors ($Å^2$)	10515			
Wilson B	106.15			
Protein Zine	123.95			
LINC P m s deviations	146.46			
Rond lengths (Å)	0.018			
Bond angles (°)	1 975			
Bond lengths (Å) Bond angles (°)	0.018 1.975			

*Highest resolution shell is shown in parenthesis.

[†] Rfree calculated with 4.57% of the reflections (2002) that were omitted from the refinement process.

Toxin	Ligands	PDB ID	Rmsd Å ²	No of Cα atoms	Reference
TcdA	Аро	4DMV	0.65	428	<u>46</u>
TcdA	Аро	3SS1	0.78	484	24
TcdA	UDP, Mn	4DMW	0.7	430	<u>46</u>
TcdA	UDP-glucose, Mn	3SRZ	0.92	455	<u>24</u>
TcdB	UDP, glucose, Mn	2BVL	1.22	469	<u>47</u>
TcsL	UDP-glucose, Mn	2VKD	0.96	461	48
Tcna	Аро	2VK9	1.99	420	48

Supplementary information Table 2: Alignment of enzymatic domains to isolated domain structures.

Supplementary information Table 3: Zinc to protein ratios determined by ICP-MS

Sample*	Zinc/Protein		Sample	Zinc/Protein			
TcdA ₁₇₉₅	0.8	0.83	0.72	TcdB	0.74	0.68	0.6
TcdA ₁₇₉₅ C700A	0.08			TcdB C698A	0.09		
TcdA ₁₇₉₅ H655A	0.06			TcdB H653A	0.04		
TcdA ₁₇₉₅ H759A	0.3			TcdB H757A	0.12	0.31	0.5
			_			_	
TcdA ₁₇₉₅ + InsP6	0.57	0.51		TcdB + InsP6	0.45	0.55	
TcdA ₁₇₉₅ + TPEN	0.04			TcdB + TPEN	0.05		
TcdA ₁₇₉₅ (apo) + ZnCl ₂	0.48			TcdB (apo) + ZnCl ₂	0.38		

* Each entry represents a measurement from a unique protein preparation.

Supplementary information Table 4: Primers used to generate the plasmids for this study.

Protein	Plasmid	Primers
TcdA ₁₈₃₂	redA ₁₈₃₂ pBL578 pBL578	5' -CTTACTCTTTATTATTATGTTCATATCCAATATCAACG-3'
S1329C		5'-CGTTGATATTGGATATGAACATAATAATAAAGAGTAAG-3'
TcdA ₁₇₉₅ pBL6:	nDI 656	5'-GATATTAAAAAACTATCATTAGGATATATAATGAGTGGAGGGCATGCCGGC-3'
	рыгозо	5'-GCCGGCATGCCCTCCACTCATTATATATCCTAATGATAGTTTTTTAATATC-3'
TcdA ₁₇₉₅	nDI 712	5'-GTAGAAGTAAACTTACTTGGAGCTAATATGTTTAGTTATGATTTTAATG-3'
C700A pBL/12	5'-CATTAAAATCATAACTAAACATATTAGCTCCAAGTAAGTTTACTTCTAC-3'	
TcdA ₁₇₉₅ H655A pBL713	nDI 712	5'-GTAAAAGTAACCTTTATTGGAGCGGGTAAAGATGAATTCAAC -3'
	pBL/13	5'-GTTGAATTCATCTTTACCCGCTCCAATAAAGGTTACTTTTAC-3'
TcdA ₁₇₉₅	nDI 714	5'-GAAAAGAACTTCTGGCTGCGTCAGGTAAATGGATAAATAA
Н759А РВL7	рыс/14	5'-CTTTATTTATCCATTTACCTGACGCAGCCAGAAGTTCTTTTC-3'
TedB	TedB H757A pBL689	5'-GGAAGAAGAGAATTATTGGATGCGTCTGGTGAATGGATAAATAA
H757A ^{pf}		5'-CTTTATTTATCCATTCACCAGACGCATCCAATAATTCTCTTCTCC-3'
TcdASAS V1109S, N1110A, N1111S	pBL675	5'- GCAGGAATACCTTCATTATCTGCTTCTGAATTAATATTGCATGATAAGGC-3' '5'-GCCTTATCATGCAATATTAATTCAGAAGCAGATAATGAAGGTATTCCTGC-3'



Supplementary information Figure 1: Wall-eyed stereo view of representative electron density. Weighted 2Fo-Fc map contoured at 1 sigma.



Supplementary information Figure 2: A truncated TcdA construct (TcdA₁₇₉₅) undergoes autoprocessing more efficiently than TcdA and TcdA₁₈₃₂. The three proteins were incubated with or without 10 mM InsP6 for 2 hours in a buffer containing 20 mM HEPES pH 6.9, 50 mM NaCl and then subjected to SDS-PAGE. Autoprocessing was most efficient in the TcdA₁₇₉₅ sample. Gel is representative of three independent experiments.



Supplementary information Figure 3: Zinc is present in the APD active sites of TcdA. **a.** Analysis of the anomalous zinc signal, reveals a single peak, shown here in purple at 7 sigma, located in the APD active site. The zinc is coordinated by H655 (2.7 Å), C700 (2.7 Å), and H759 (2.2 Å) and may have indirect contacts with E544 or D545. The blue mesh depicts a weighted 2Fo-Fc map contoured around the 5 active site amino acids at 1 sigma. **b.** X-ray absorption near-edge spectrum of TcdA in solution.



Supplementary information Figure 4: Topology of the TcdA delivery domain. Cylinders indicate α-

helices, arrows represent β -strands.



Supplementary information Figure 5: Full Western blots associated with Figures 3c and 3d. a. Proteins were separated by SDS-PAGE and transferred to PVDF membrane. The membrane was cut between the 37 kDa and 25 kDa molecular weight markers so that the top blot could be probed with an antibody against the TcdA CROPS domain and the middle blot could be probed for unglucosylated Rac1. After exposure to film, the middle blot was stripped and reprobed with an antibody against total Rac1. b. The membrane was initially probed with an antibody that recognizes unglucosylated Rac1. After exposure to film, the blot was stripped and reprobed with an antibody against total Rac1.

References

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