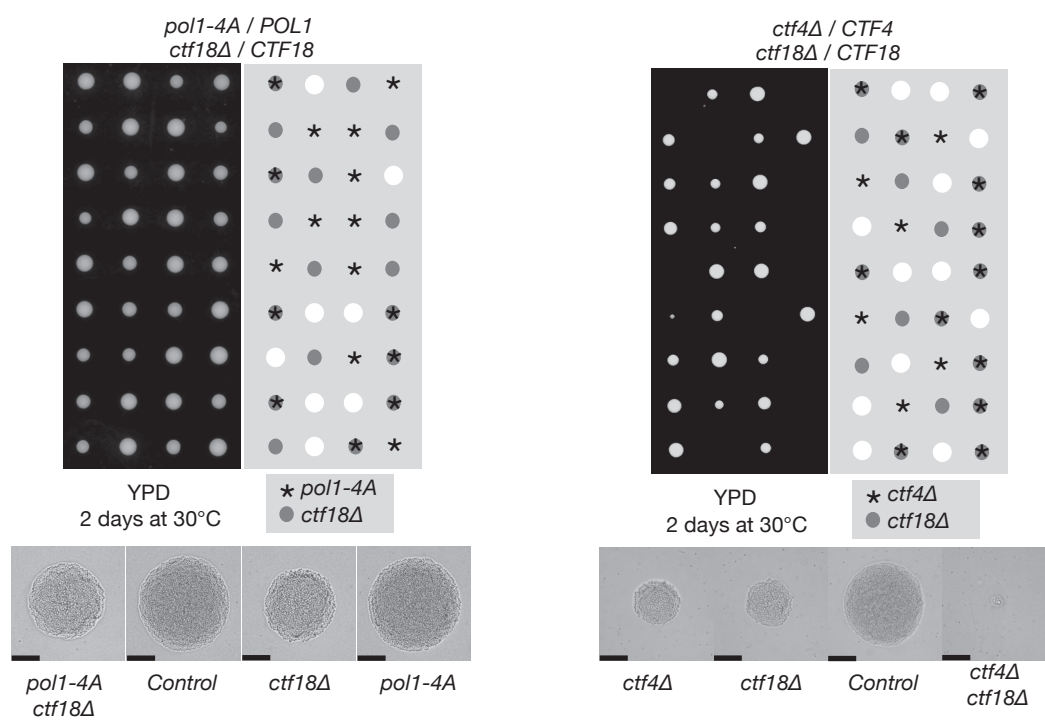


**Molecular Cell, Volume 63**

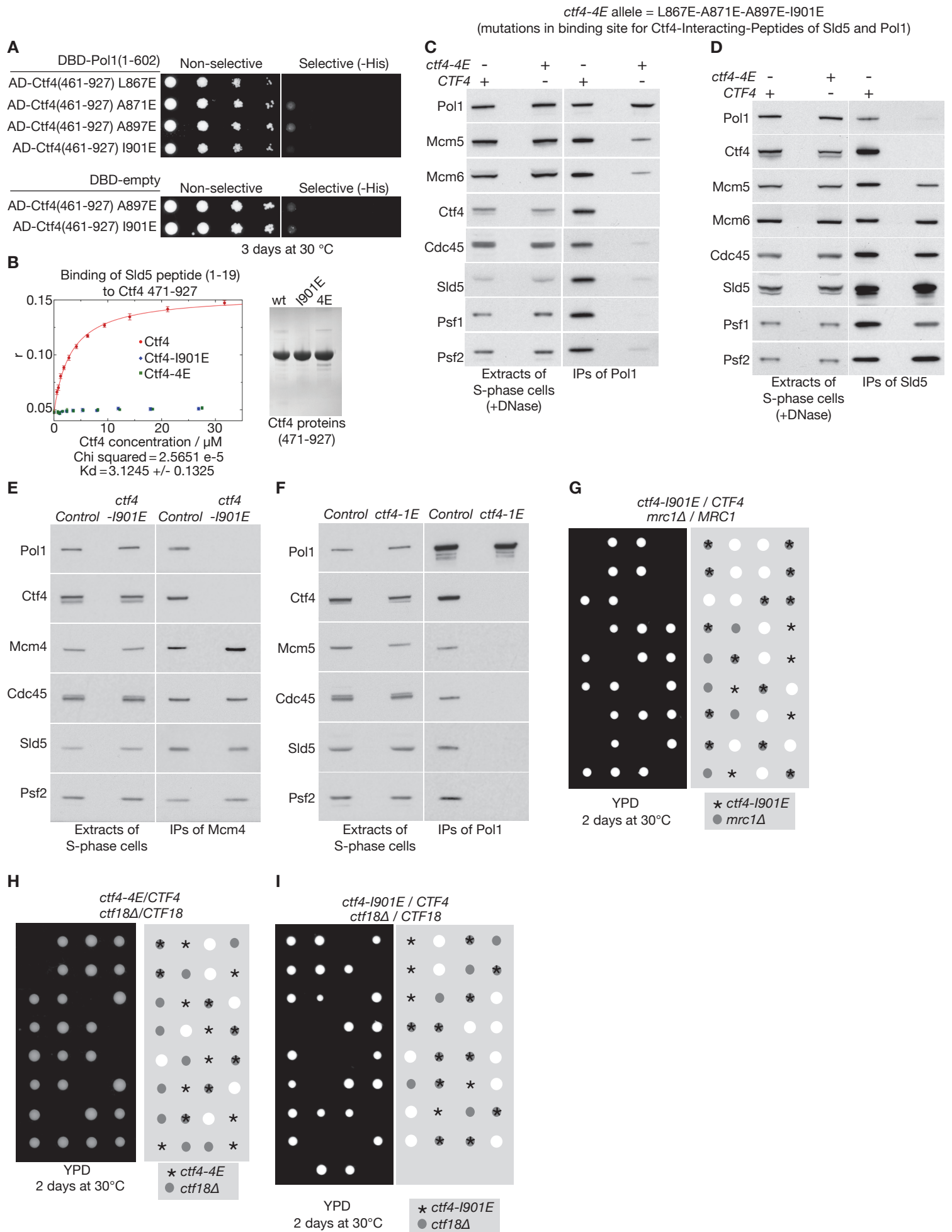
**Supplemental Information**

**Ctf4 Is a Hub in the Eukaryotic Replisome  
that Links Multiple CIP-Box Proteins  
to the CMG Helicase**

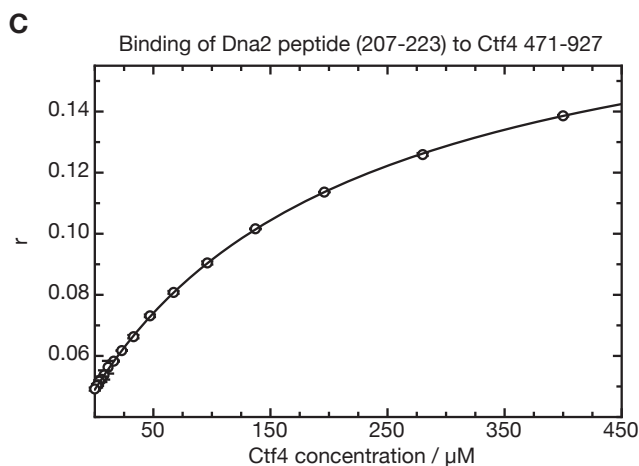
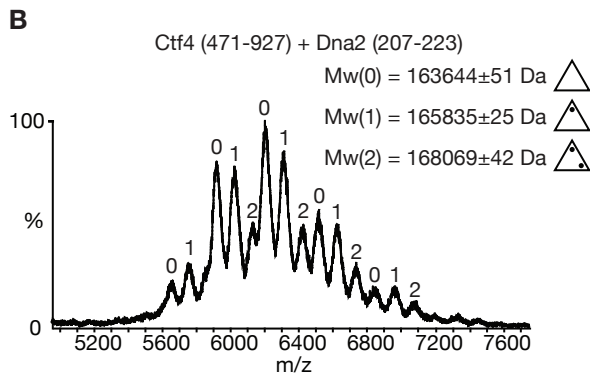
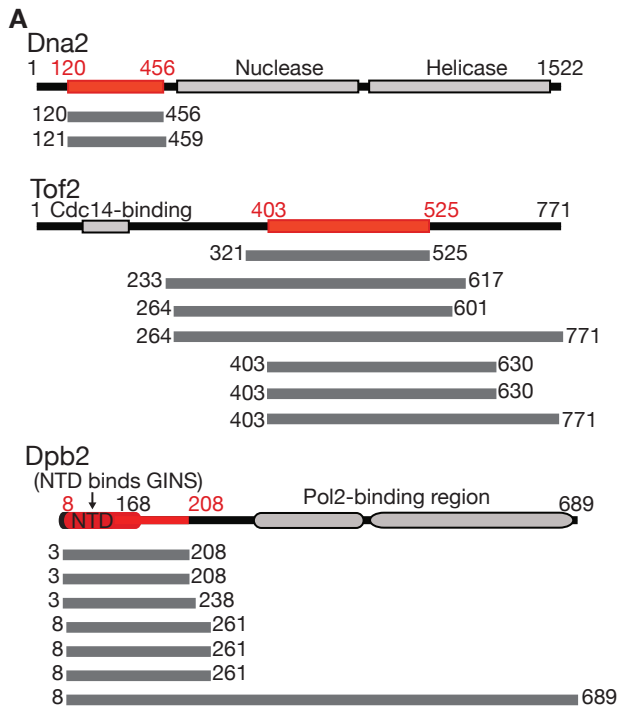
**Fabrizio Villa, Aline C. Simon, Maria Angeles Ortiz Bazan, Mairi L. Kilkenny, David Wirthensohn, Mel Wightman, Dijana Matak-Vinković, Luca Pellegrini, and Karim Labib**



Villa et al Supplementary Figure 2



Villa et al Supplementary Figure 3



Chi squared = 5.1970  
 Goodness of fit = 0.9903

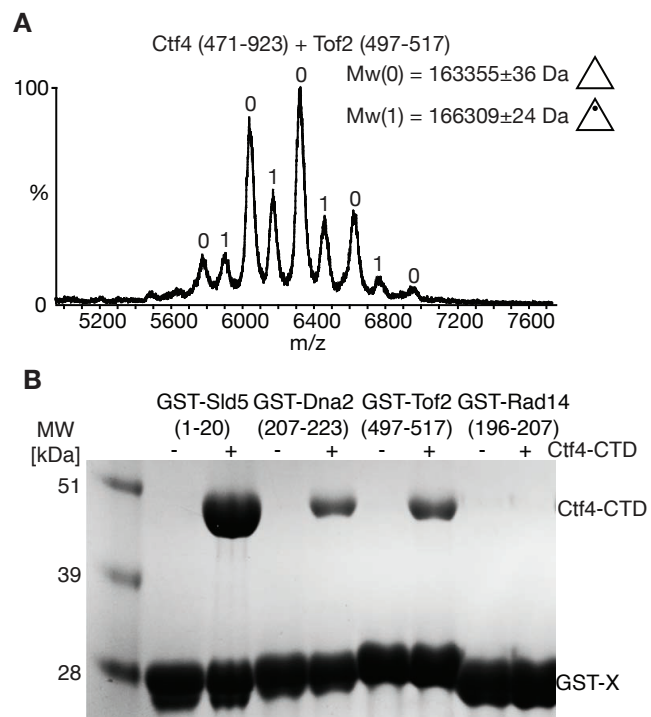
Parameters: Standard deviations:  
 Kd = 229.0088 Kd = 12.7404

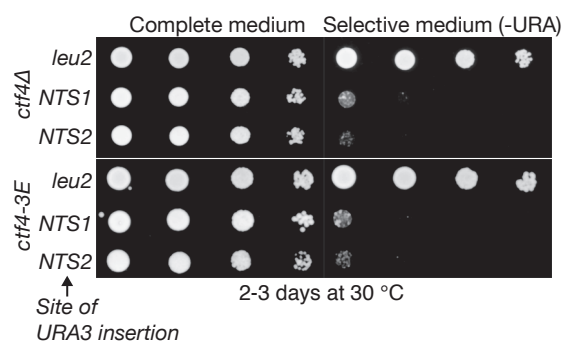
Kd 95% Confidence intervals 205.6937 ... 256.0578  
 (based on 500 converging iterations)

**D**

14 Budding yeast proteins with peptides matching the pattern:  
 [ND]-X-D-D-I-L-X-X-J,  
 where [ND] = N or D, J = hydrophobic residue and  
 X = any residue

- Slf5** 4-NIDDILAEL-12  
 (subunit of GINS component of CMG DNA helicase)
- Dna2** 210-NIDDILDDI-218  
 (Nuclease with role in DNA replication and recombination)
- Pol1** 139-NFDDILGEF-147  
 (Catalytic subunit of DNA polymerase alpha)
- Chl1** 701-DGDDILSGY-709  
 (DNA helicase important for establishment of sister chromatid cohesion)
- Lcd1/Ddc2** 13-DDDDILLEL-21  
 (essential partner of Mec1 protein kinase for S-phase checkpoint response)
- Isw2** 681-DIDDILKKG-689  
 (ATPase involved in chromatin remodeling)
- Irc20** 734-NIDDILISM-742  
 (E3 ubiquitin ligase and putative helicase involved in recombination)
- Rfx1** 681-NLDDILEMM-689  
 (Transcriptional repressor of DNA-damage induced genes)
- Pex19** 176-NIDDILSQL-184  
 (chaperone and import receptor for peroxisomal membrane proteins)
- Cog7** 9-DDDDILSMF-17  
 (Protein trafficking in the Golgi)
- Pdr8** 204-NQDKDDILSLL-212  
 (Transcription factor controlling membrane transporters)
- Vps45** 453-DKDDILTEL-461  
 (Vacuolar Protein Sorting)
- Tub4** 115-NQDDILNKI-123  
 (Gamma Tubulin)
- Sia1** 590-NLDDILDEI-598  
 (Unknown function)





## Legends to Supplemental Figures and Table

### Figure S1, related to main Figure 1

*pol1-4A* does not share the synthetic lethal phenotype of *ctf4Δ* with *ctf18Δ*.

The indicated diploids were sporulated and tetrads were dissected. The photos in the lower panels were taken after 20 hours growth at 30°C (scale bars = 50μm), and genotypes were determined by replica plating after two days of growth.

### Figure S2, related to main Figure 2

Additional phenotypes of mutations in the C-terminal peptide-binding domain of Ctf4. **(A)** Single mutations in the C-terminal peptide-binding domain of Ctf4 block interaction with Pol1 amino terminus in the yeast two-hybrid assay.

**(B)** Fluorescence anisotropy showing that the Ctf4-I901E and Ctf4-4E mutations blocked binding to a peptide containing the Sld5 CIP-box. **(C)**

Extracts of *ctf4-4E POL1-6HA* (YFV12) and *POL1-6HA* control cells (YMP262-1) were grown as in Figure 2D. Pol1-6HA was isolated from cell extracts by immunoprecipitation on anti-HA beads, and the associated proteins monitored by immunoblotting. **(D)** A similar experiment was

performed with *ctf4-4E TAP-SLD5* (YFV9) and *TAP-SLD5* control cells (YSS47). **(E)** Extracts of *ctf4-I901E MCM4-9MYC* (YFV33) and *MCM4-9MYC* control cells (YSS75) were grown as in Figure 2D. Mcm4-9MYC was isolated from cell extracts by immunoprecipitation on anti-MYC beads, and the associated proteins monitored by immunoblotting. **(F)** A similar experiment was performed with *ctf4-I901E POL1-6HA* (YFV32) and *POL1-6HA* control

cells (YMP262-1). **(G)** *ctf4-I901E* is synthetic lethal with *mrc1Δ*. Cells were processed as in Figure 1A. **(H)** *ctf4-4E* is synthetic lethal with *ctf18Δ*. **(I)** *ctf4-I901E* is synthetic lethal with *ctf18Δ*.

### **Figure S3, related to main Figure 3**

A screen for novel partners of Ctf4 461-927, and characterisation of the CIP-box of Dna2. **(A)** Summary of the novel hits in the two hybrid screen against Ctf4 461-927. Each grey line indicates a single hit in the screen, and the red segments indicate the minimal fragment of each protein that is predicted to interact with Ctf4. **(B)** Native mass spectrometry was used to show that Ctf4 471-927 binds directly to the Dna2 CIP *in vitro*. **(C)** Fluorescence anisotropy confirmed binding of Ctf4 471-927 with a  $K_d$  of  $229\mu\text{M}$ . **(D)** Results of Pattern Match search of the yeast proteome (<http://www.yeastgenome.org/cgi-bin/PATMATCH/nph-patmatch>), using the indicated sequence that is based on the CIP-boxes of Sld5-Pol1-Dna2.

### **Figure S4, related to main Figure 4**

The Tof2 CIP binds directly to Ctf4 471-927 *in vitro*. **(A)** Native mass spectrometry showed that Ctf4 471-927 binds directly to the Tof2 CIP-box. **(B)** GST-fusions of the indicated peptides were tested for binding to Ctf4 471-927. A peptide from the Rad14 protein provided a negative control.



**Figure S5, related to main Figure 6**

Transcriptional silencing in the rDNA repeats is not defective in *ctf4-3E* or *ctf4Δ* cells. Cells were processed as in Figure 6D.

**Table S1, related to main figure 1**

Strains used in this study - all based on the W303 yeast genetic background, except for the yeast two-hybrid strain PJ69-4A.

Strain	Genotype
W303-1	<i>MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100 / MAT<math>\alpha</math> ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100</i>
W303-1a	<i>MATa ade2-1 ura3-1 his3-1 trp1-1 leu2-3, 112 can1-100</i>
PJ69-4A	<i>MATa trp1-901 leu2-3,112 ura3-52 his3-200 gal4<math>\Delta</math> gal80<math>\Delta</math> LYS2::GAL1-HIS3 GAL2-ADE2 met2::GAL7-la</i>
YAG132	<i>MATa / MAT<math>\alpha</math> CTF4 / ctf4<math>\Delta</math>::kanMX MRC1 / mrc1<math>\Delta</math>::hphNT</i>
YCE449	<i>MATa pol1-4A (URA3)</i>
YDP14	<i>MATa / MAT<math>\alpha</math> CTF4 / ctf4<math>\Delta</math>::kanMX CTF18 / ctf18<math>\Delta</math>::hphNT</i>
YFV9	<i>MATa ctf4-4E TAP-SLD5 (kanMX) pep4<math>\Delta</math>::ADE2</i>
YFV11	<i>MATa ctf4-4E</i>
YFV12	<i>MATa ctf4-4E POL1-6HA (K.I. TRP1) pep4<math>\Delta</math>::ADE2</i>
YFV13	<i>MATa ctf4-4E MCM4-9MYC (kanMX) pep4<math>\Delta</math>::ADE2</i>
YFV17	<i>MATa dna2-4A-9MYC (K.I. TRP1)</i>
YFV21	<i>MATa GAL-DNA2-9MYC (kanMX &amp; K.I. TRP1) pep4<math>\Delta</math>::ADE2</i>
YFV22	<i>MATa GAL-DNA2-9MYC (kanMX &amp; K.I. TRP1) TAP-CTF4 (kanMX) pep4<math>\Delta</math>::ADE2</i>
YFV30	<i>MATa tof2<math>\Delta</math>::URA3</i>
YFV31	<i>MATa ctf4-3E</i>
YFV33	<i>MATa ctf4-I901E MCM4-9MYC (kanMX) pep4<math>\Delta</math>::ADE2</i>
YFV36	<i>MATa tof2-4A</i>
YFV47	<i>MATa ura3:: pRS306-GAL-ProteinA-CBP-TEV-TOF2 (URA3) pep4<math>\Delta</math>::ADE2</i>
YFV55	<i>MATa / MAT<math>\alpha</math> ctf4-4E MRC1 / mrc1<math>\Delta</math>::K.I. TRP1</i>
YFV56	<i>MATa / MAT<math>\alpha</math> ctf4-4E CTF18 / ctf18<math>\Delta</math>::K.I. TRP1</i>
YFV57	<i>MATa / MAT<math>\alpha</math> ctf4-I901E CTF18 / ctf18<math>\Delta</math>::K.I. TRP1</i>
YFV58	<i>MATa / MAT<math>\alpha</math> ctf4-I901E MRC1 / mrc1<math>\Delta</math>::K.I. TRP1</i>
YFV59	<i>MATa / MAT<math>\alpha</math> MRC1 / mrc1<math>\Delta</math>::K.I. TRP1 POL1 / pol1-4A (URA3)</i>
YFV60	<i>MATa / MAT<math>\alpha</math> CTF18 / ctf18<math>\Delta</math>::K.I. TRP1 POL1 / pol1-4A (URA3)</i>
YFV62	<i>MATa / MAT<math>\alpha</math> DNA2 / dna2-4A MEC1 / mec1<math>\Delta</math>::HIS sml1<math>\Delta</math>::ADE2/ sml1<math>\Delta</math>::ADE2</i>
YHM160	<i>MATa ctf4<math>\Delta</math>::kanMX</i>
YMP262-1	<i>MATa POL1-6HA (K.I. TRP1) pep4<math>\Delta</math>::ADE2</i>
YSS47	<i>MATa TAP-SLD5 (kanMX) pep4<math>\Delta</math>::ADE2</i>
YSS75	<i>MATa MCM4-9MYC (kanMX) pep4<math>\Delta</math>::ADE2</i>