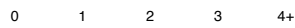
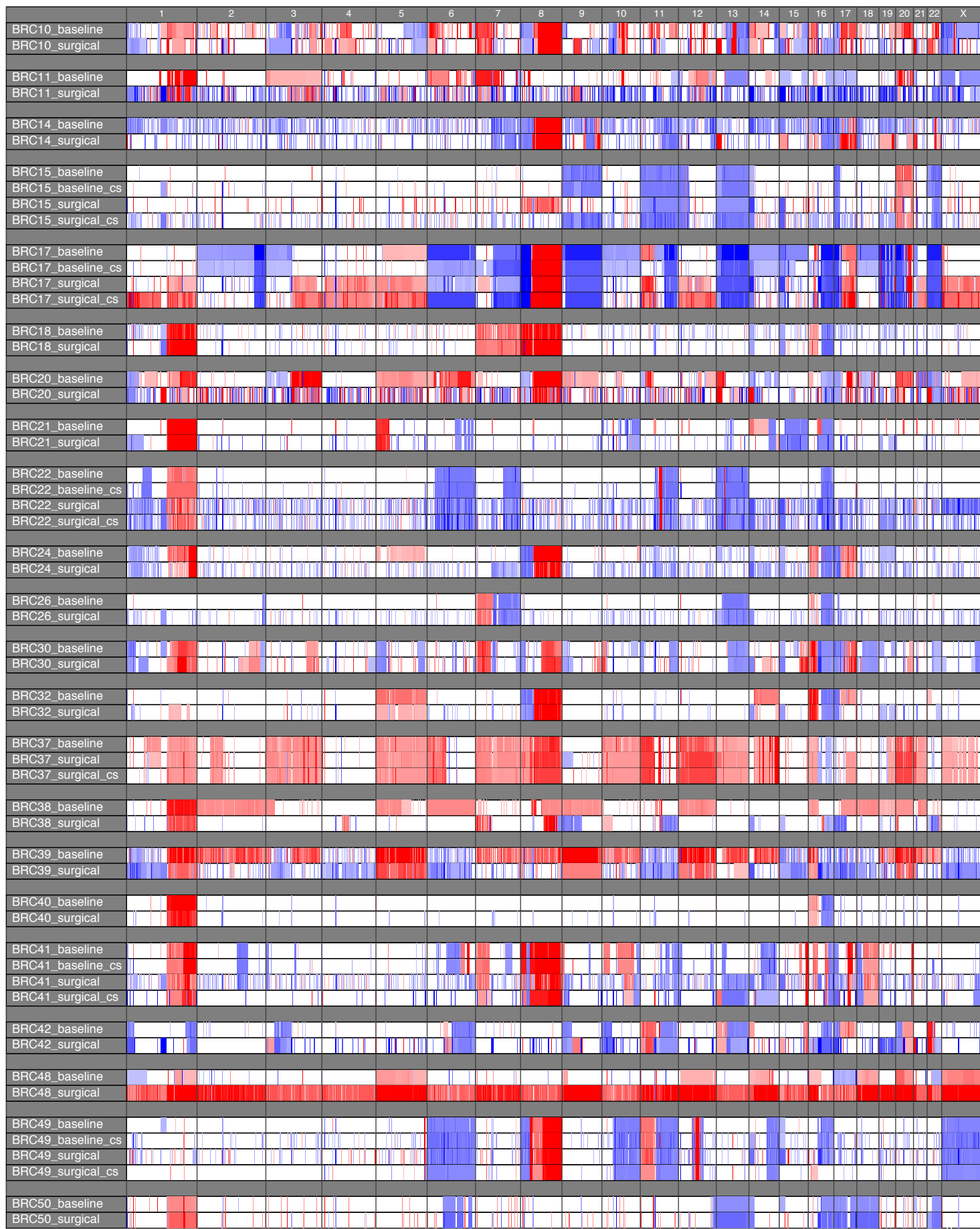
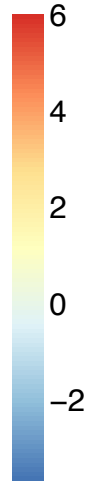
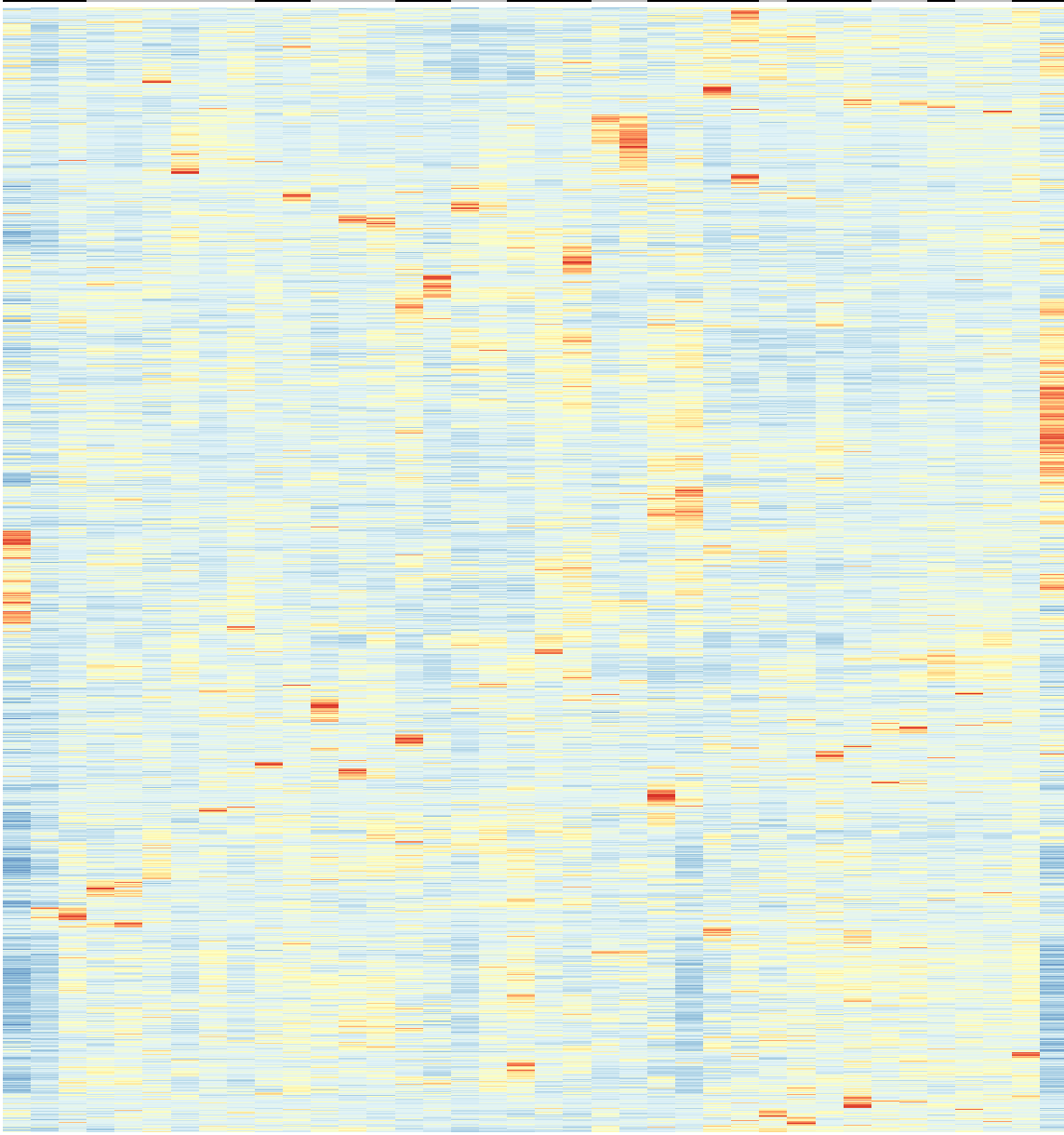
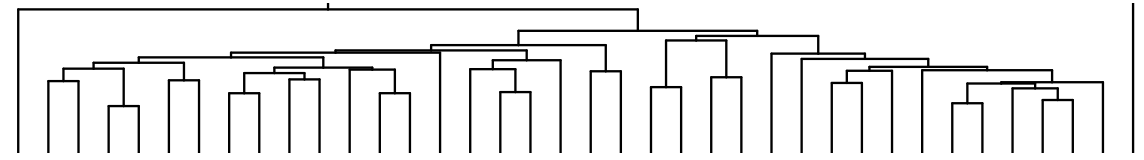


Supplementary Figures

Supplementary Figure 1: Somatic copy number alterations detected by CopyCat and displayed after purity correction.



Supplementary Figure 2: Expression Clustering of all samples. Heatmap display of hierarchical clustering of the top 5000 most variably-expressed genes in all samples

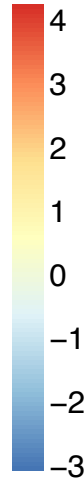
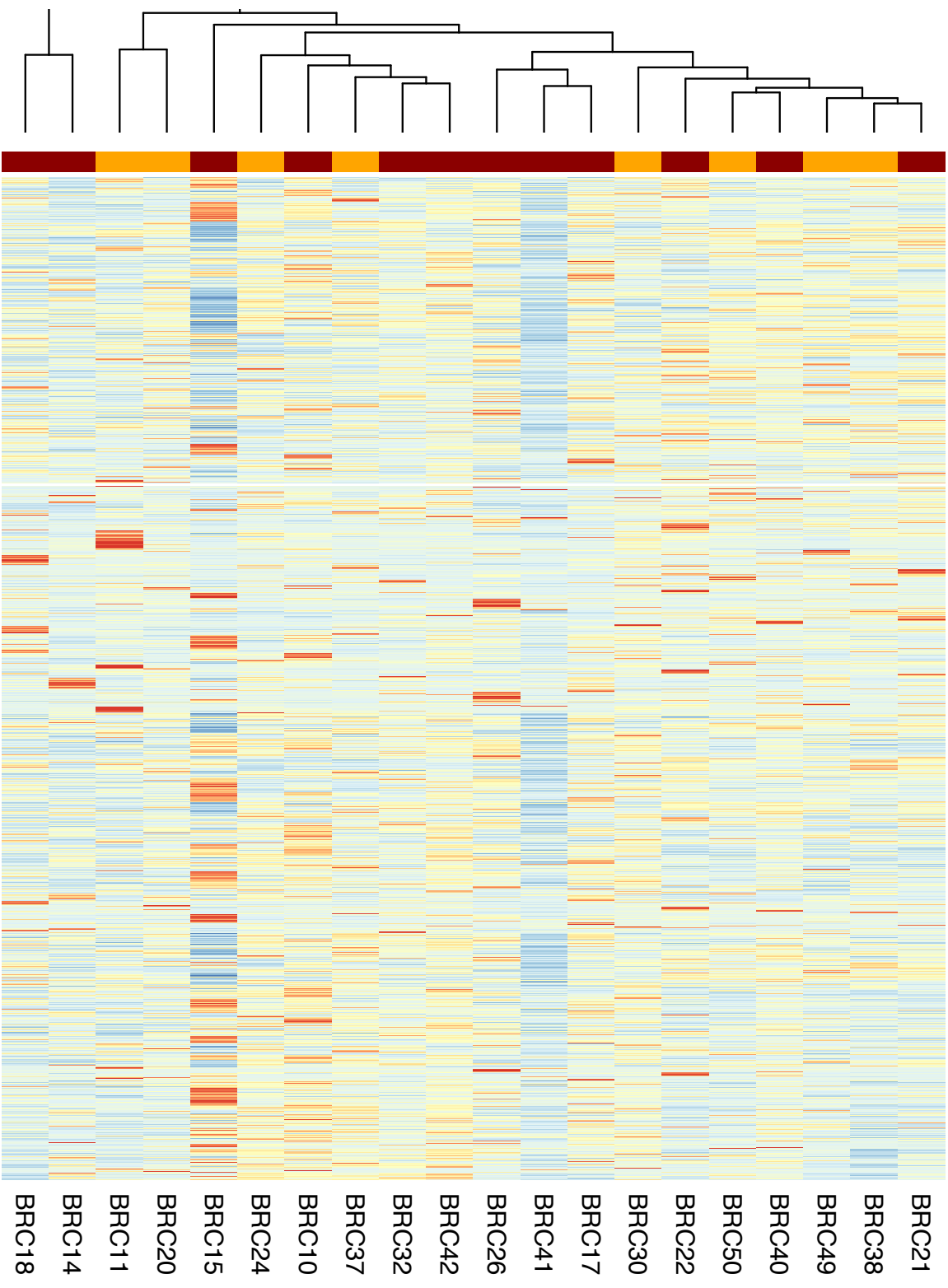


Sample
Surgical
Baseline

Response
AI-resistant
AI-sensitive

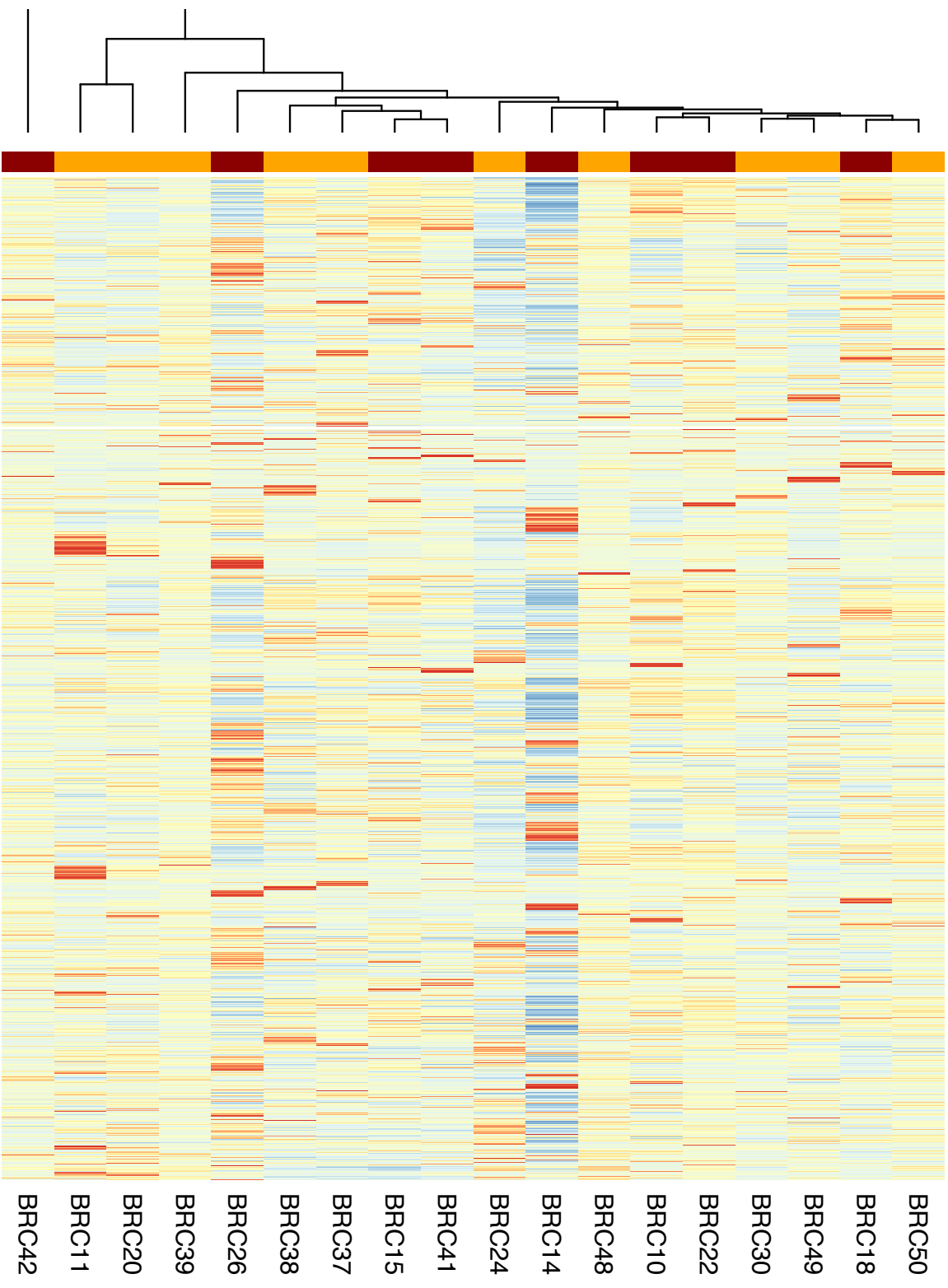
BRC15
BRC22
BRC48
BRC39
BRC42
BRC50
BRC50
BRC18
BRC15
BRC21
BRC38
BRC22
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BRC26
BRC26
BRC11
BRC11
BRC10
BRC42
BRC10
BRC24
BRC24
BRC14
BRC14
BRC17
BRC37
BRC37
BRC38
BRC40
BRC32
BRC30
BRC30
BRC20
BRC20
BRC49
BRC49
BRC41
BRC41
BRC14

Supplementary Figure 3: Expression Clustering of baseline samples. Heatmap display of hierarchical clustering of the top 5000 most variably-expressed genes in 20 baseline samples



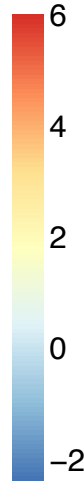
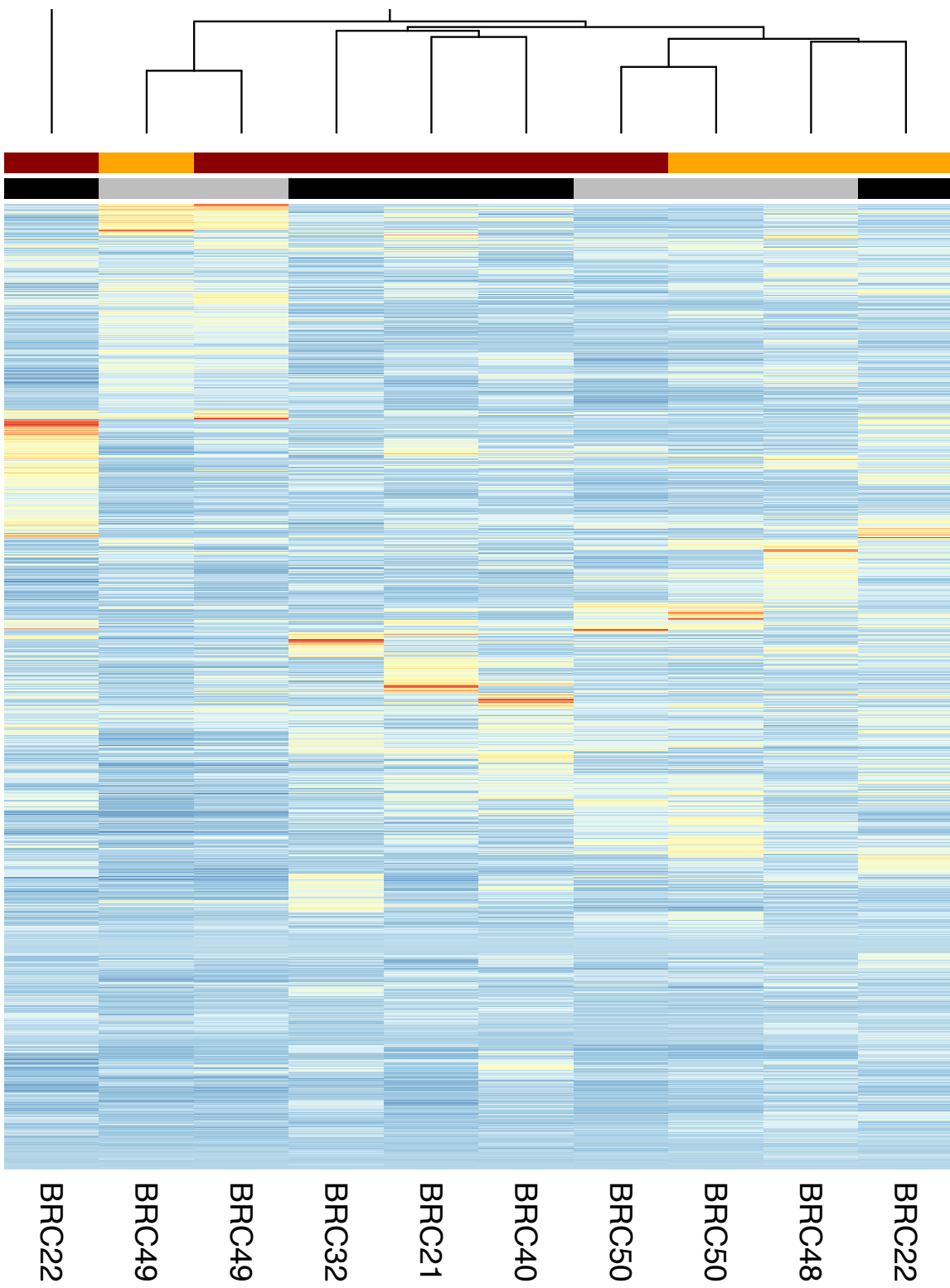
Therapy
AI-resistant
AI-sensitive

Supplementary Figure 4: Expression Clustering of surgical samples. Heatmap display of hierarchical clustering of the top 5000 most variably-expressed genes in 18 surgical samples



Therapy
■ AI-resistant
■ AI-sensitive

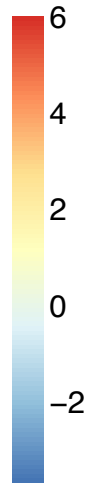
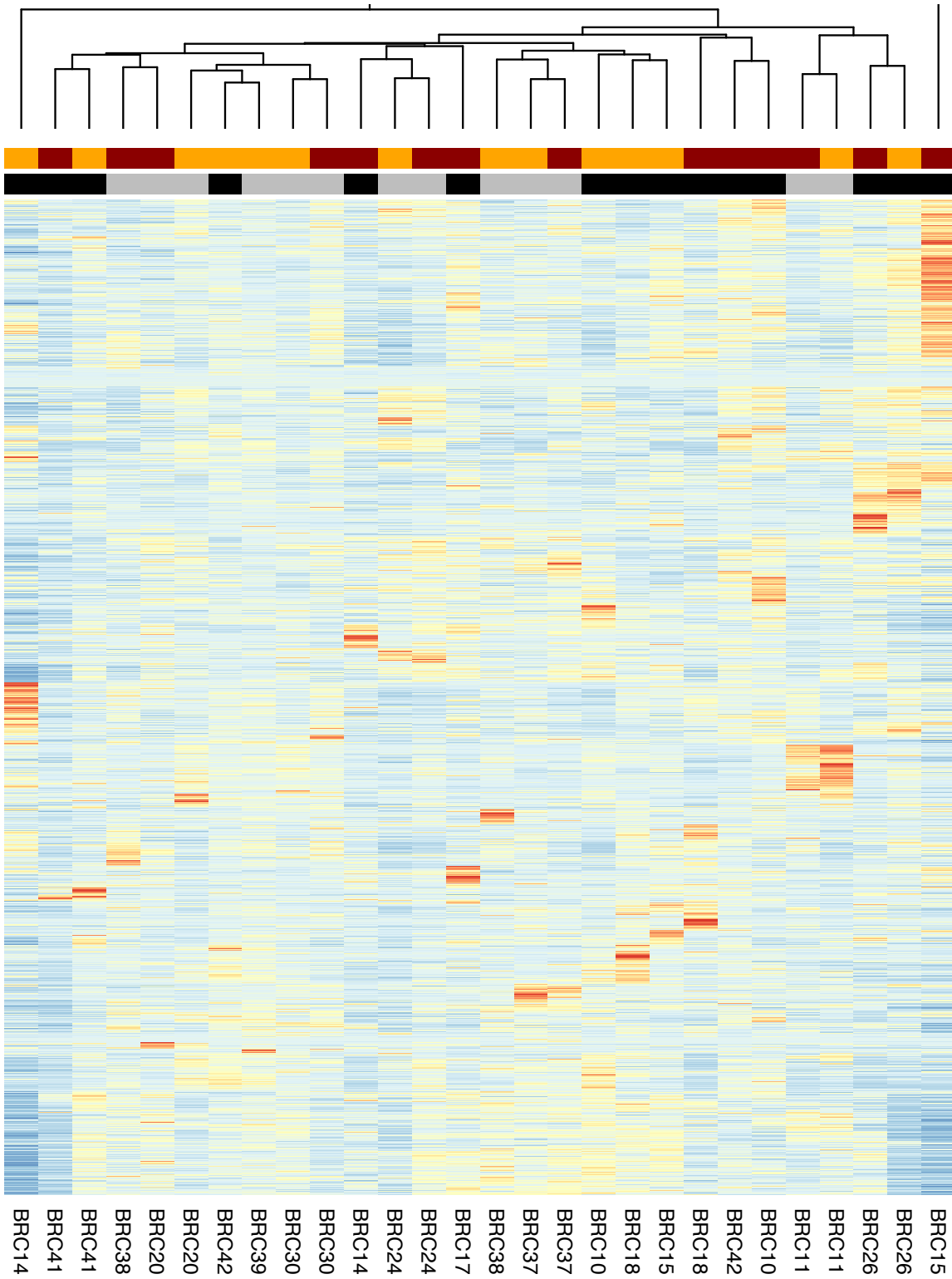
Supplementary Figure 5: Expression Clustering of baseline Luminal A samples. Heatmap display of hierarchical clustering of the top 5000 most variably-expressed genes in all baseline LumA samples



Sample
Surgical
Baseline

Response
AI-resistant
AI-sensitive

Supplementary Figure 6: Expression Clustering of baseline Luminal B samples. Heatmap display of hierarchical clustering of the top 5000 most variably-expressed genes in all baseline LumB samples

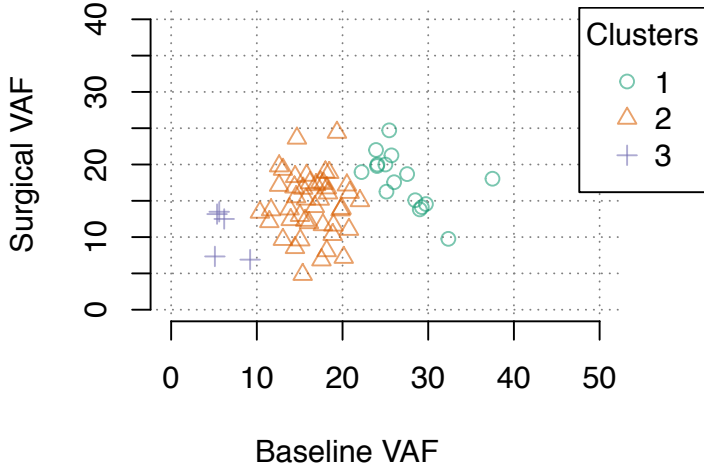


Sample
Surgical
Baseline

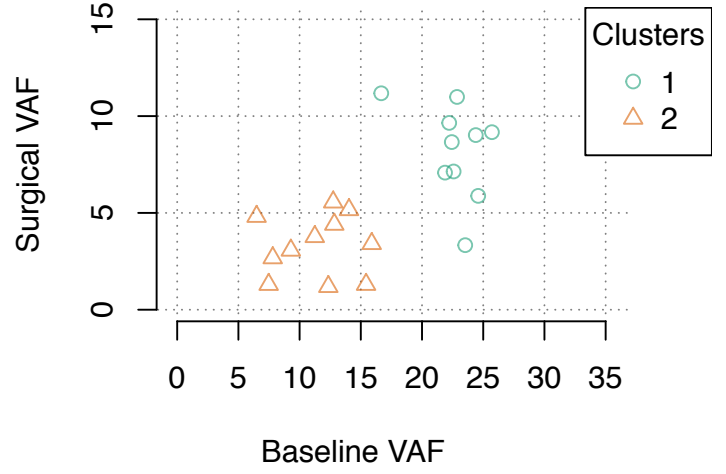
Response
AI-resistant
AI-sensitive

Supplementary Figure 7: Clonality inference for all samples

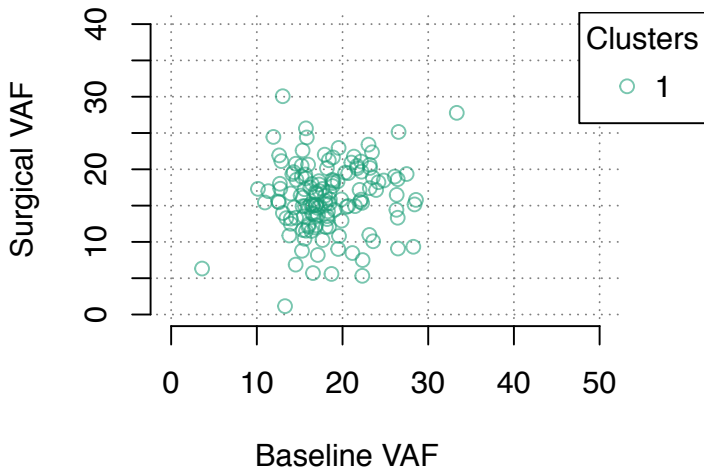
BRC10



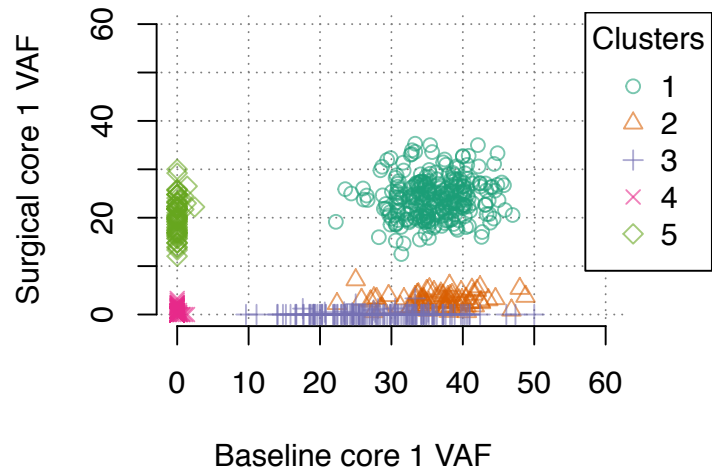
BRC11



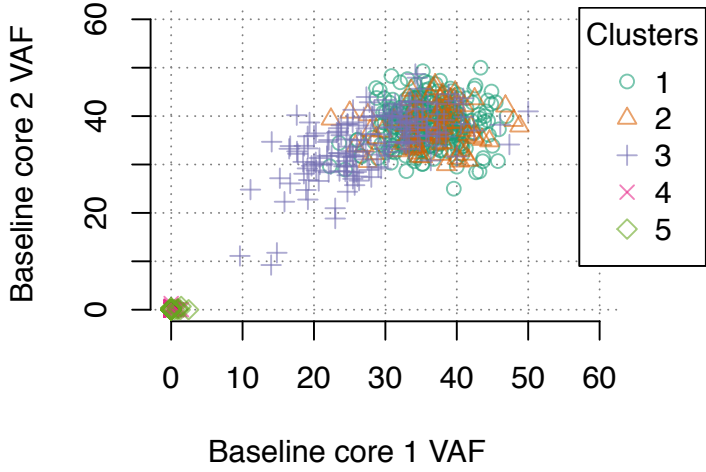
BRC14



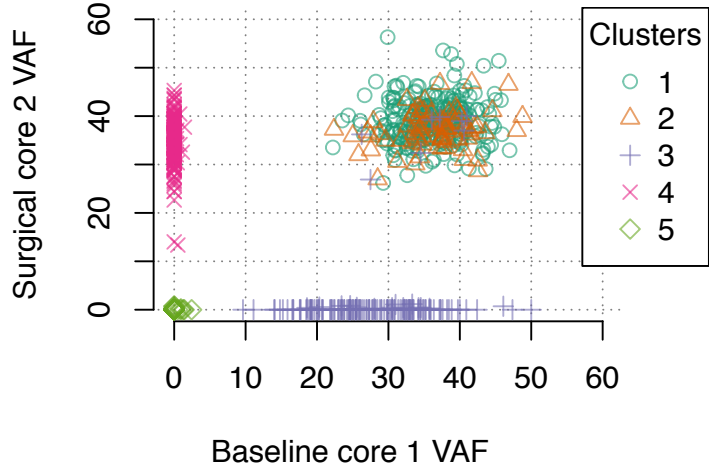
BRC15



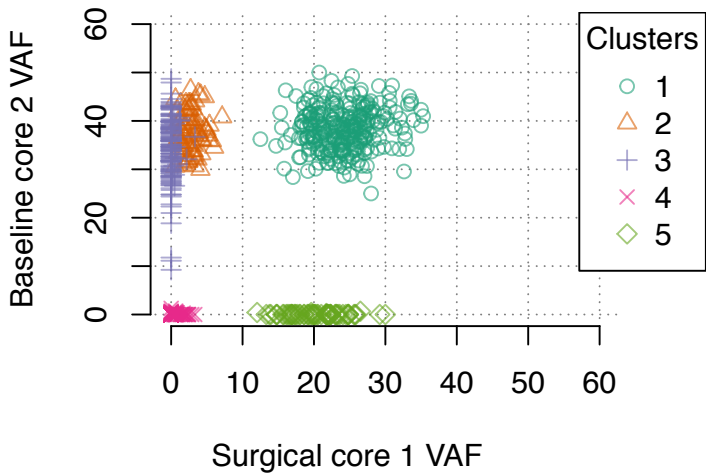
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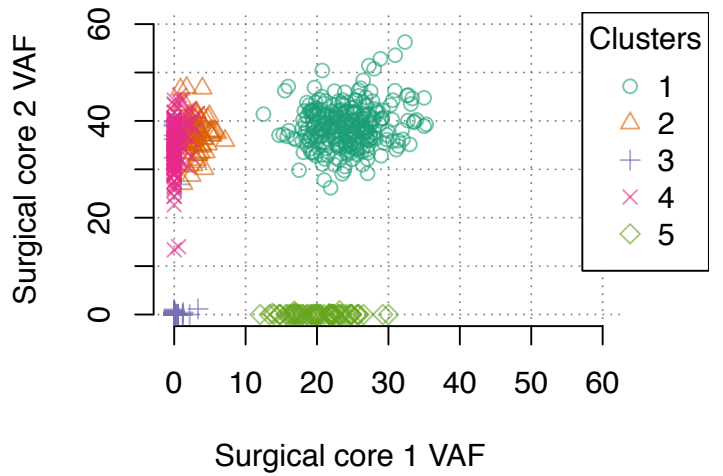
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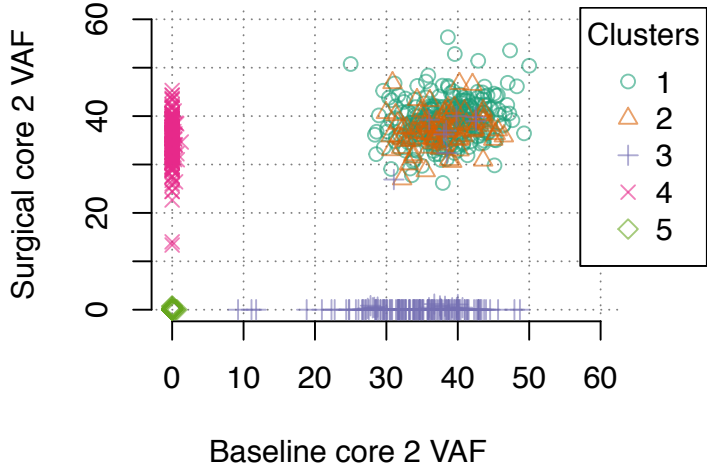
BRC15



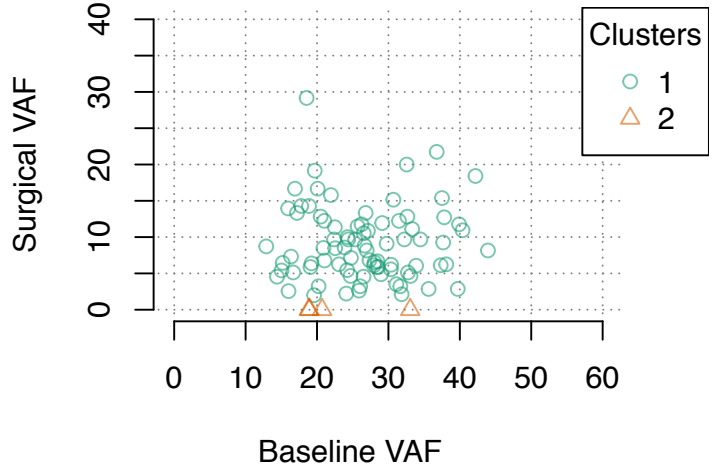
BRC15



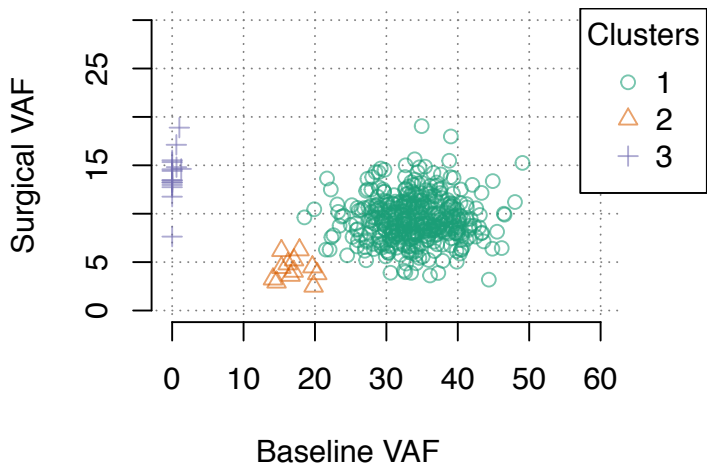
BRC15



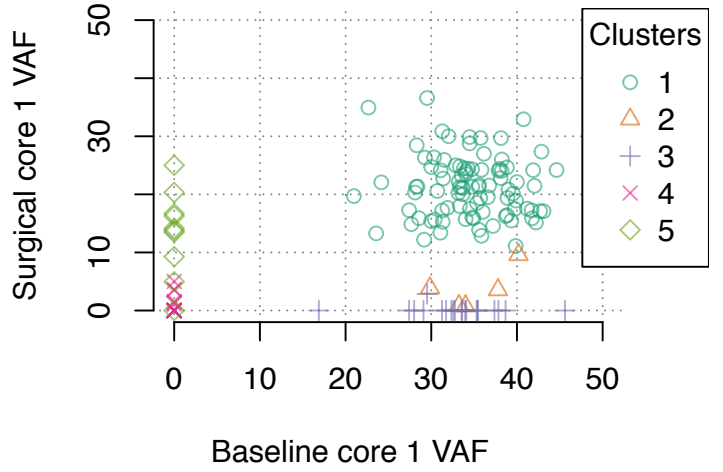
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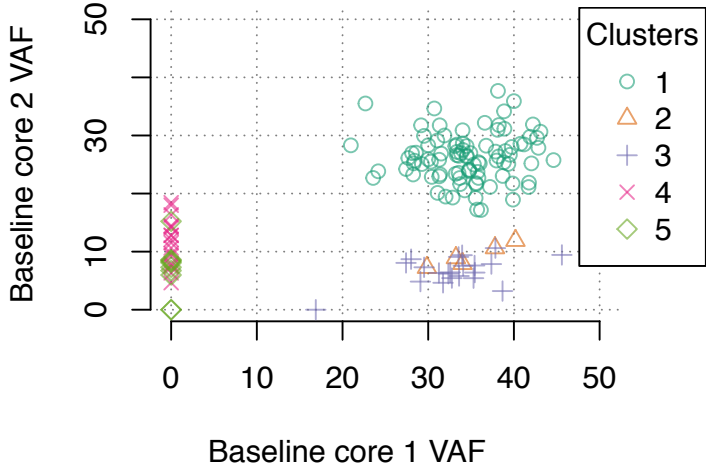
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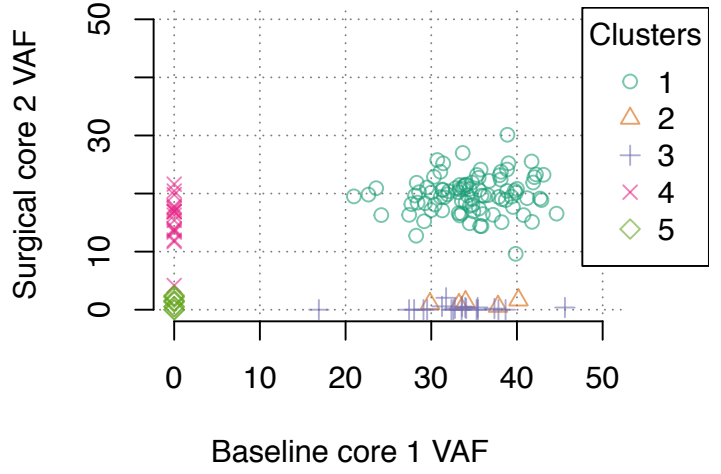
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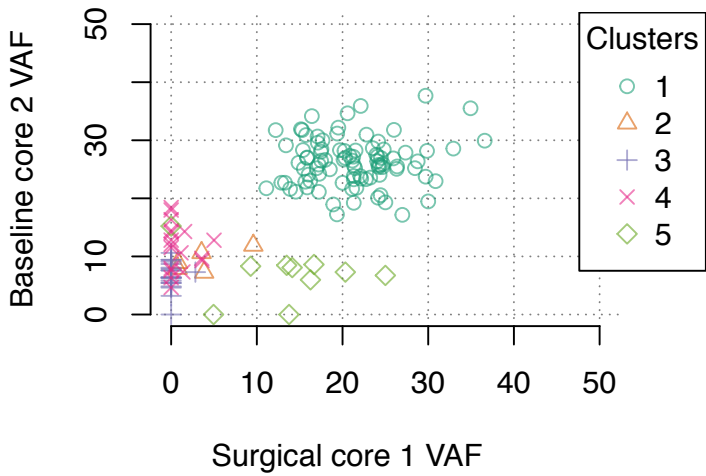
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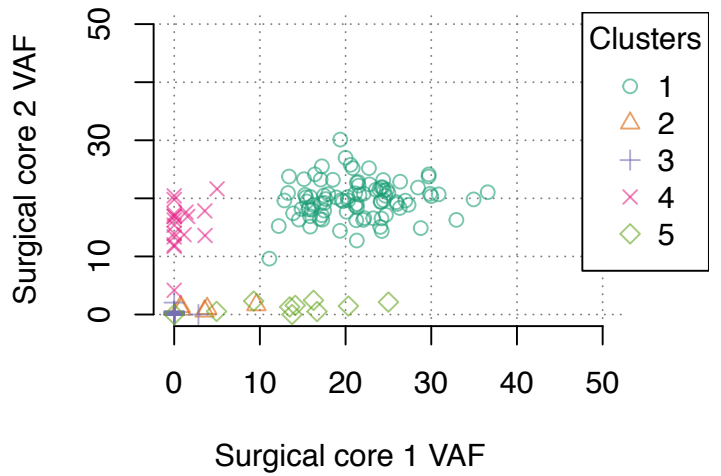
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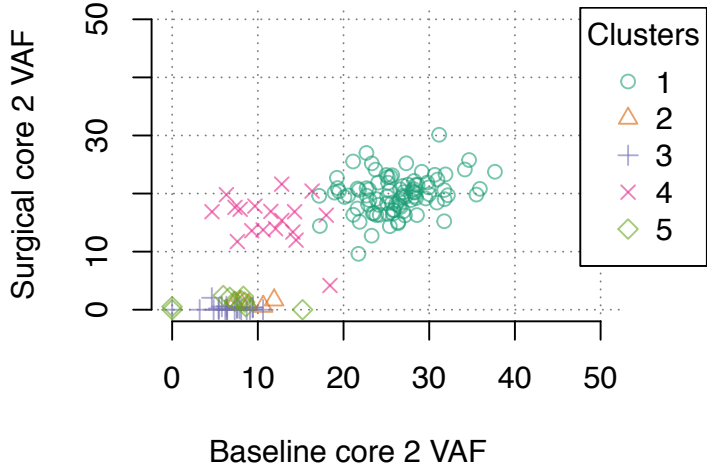
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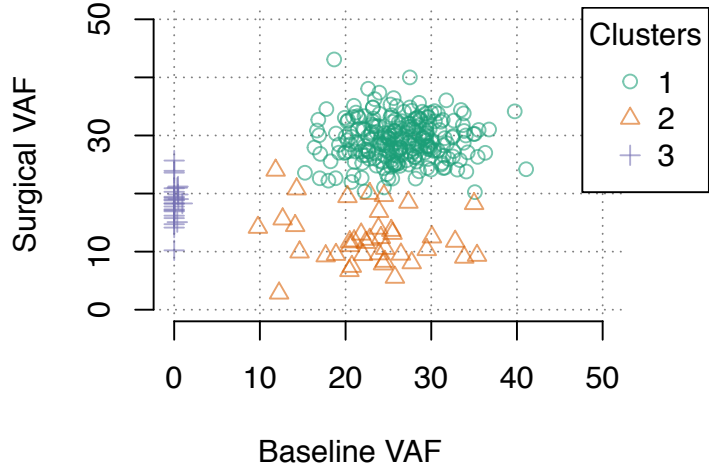
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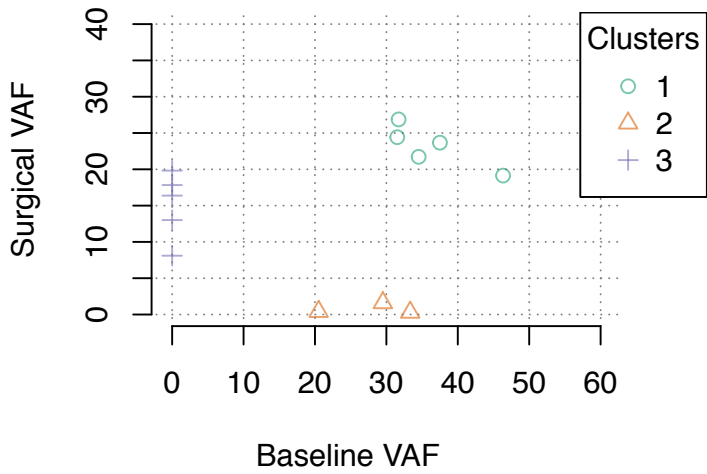
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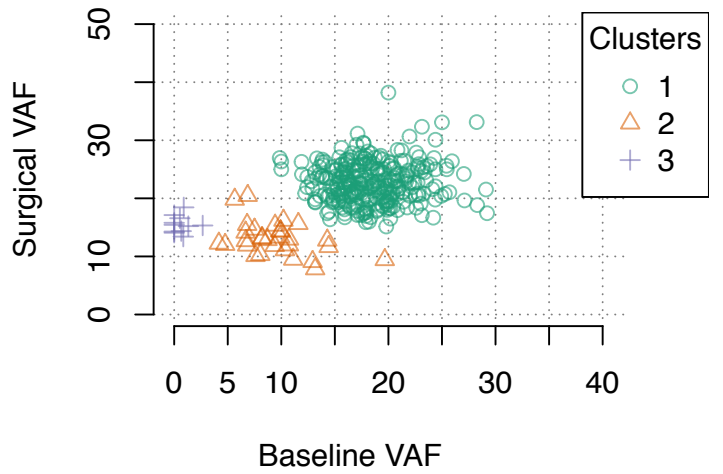
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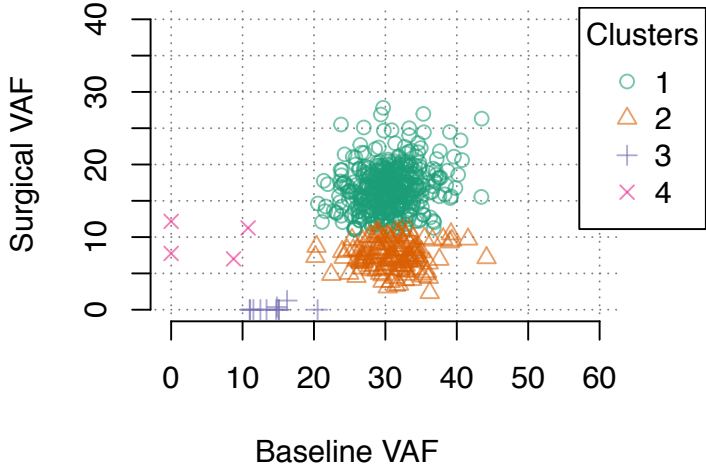
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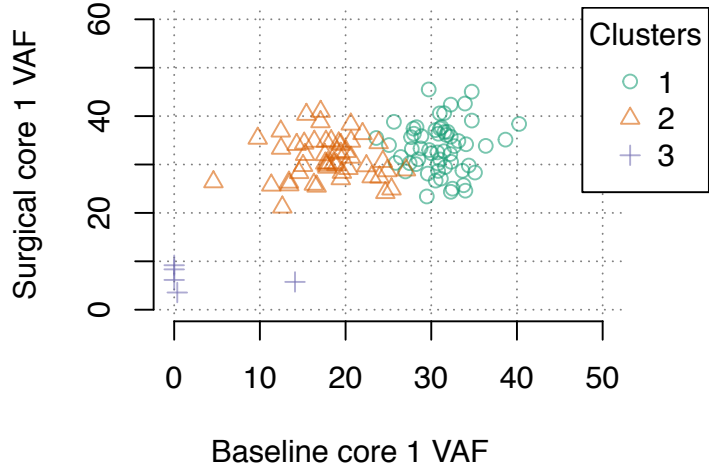
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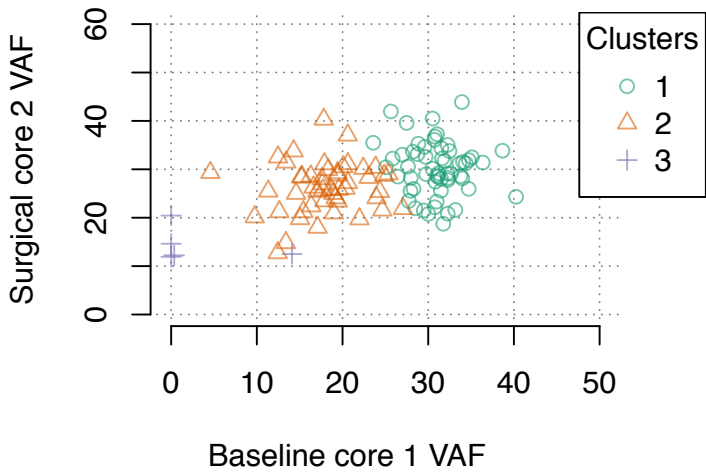
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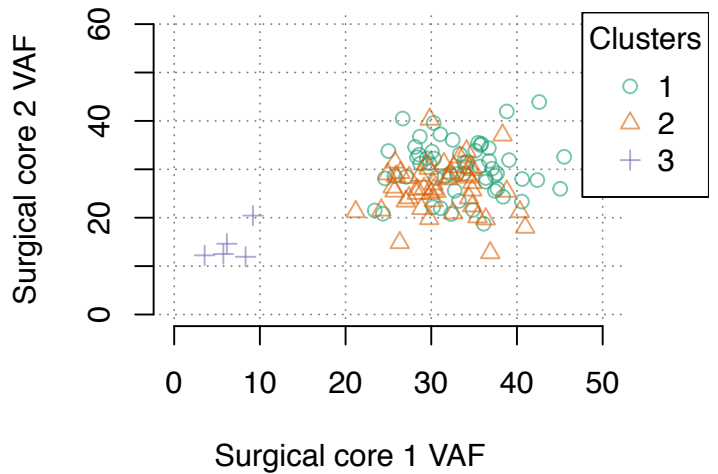
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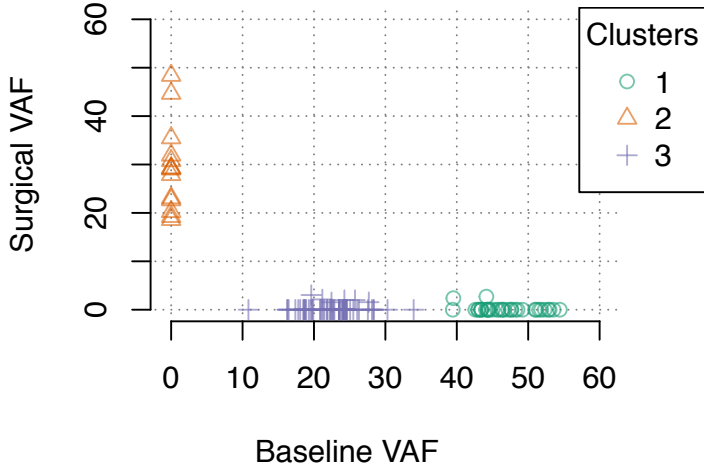
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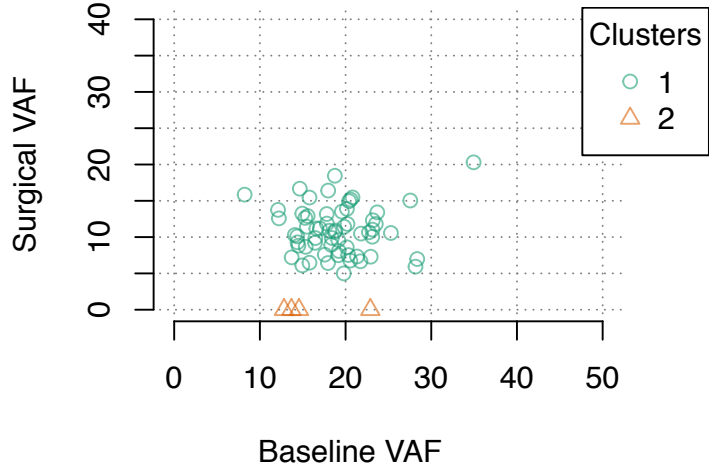
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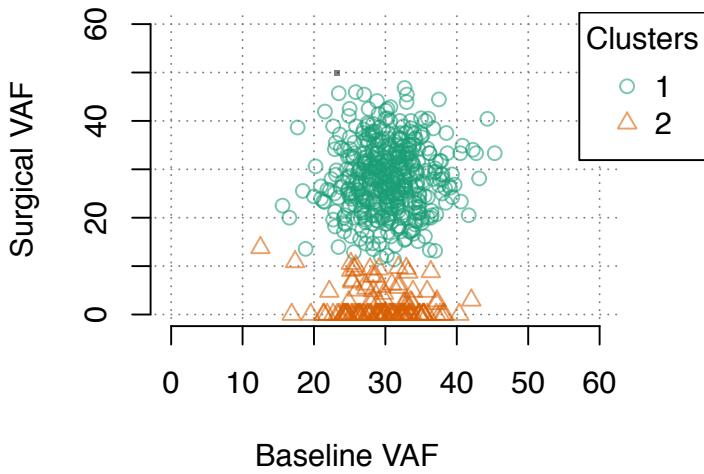
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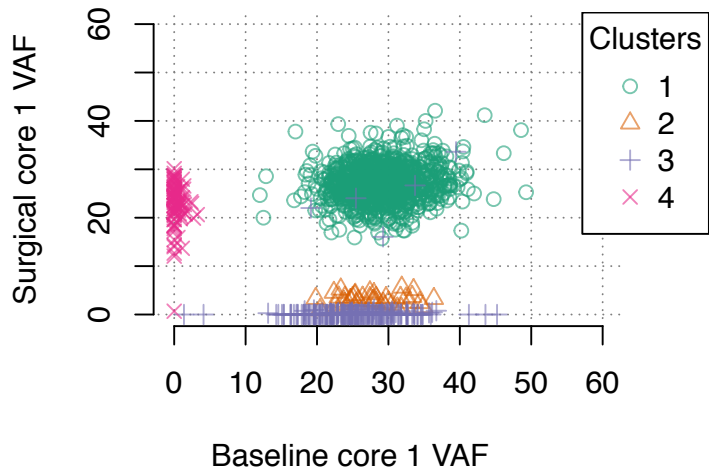
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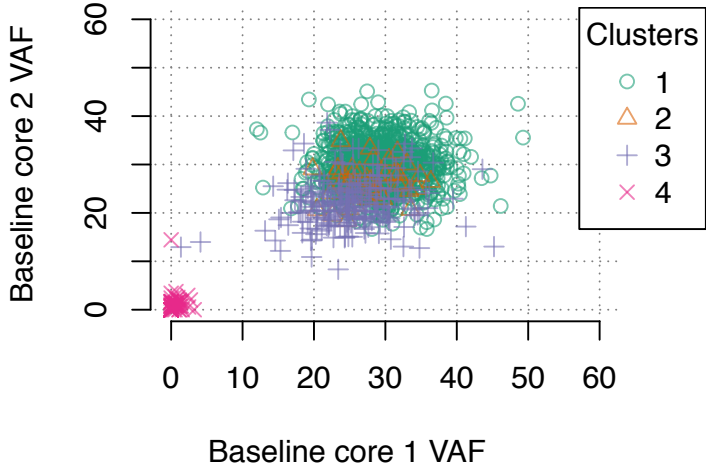
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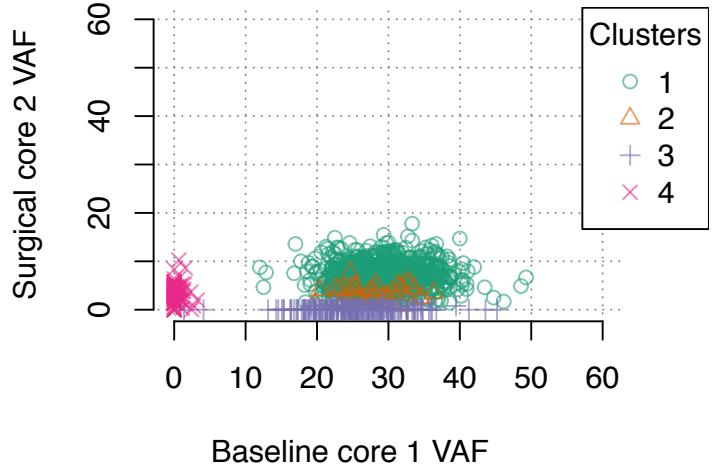
BRC41



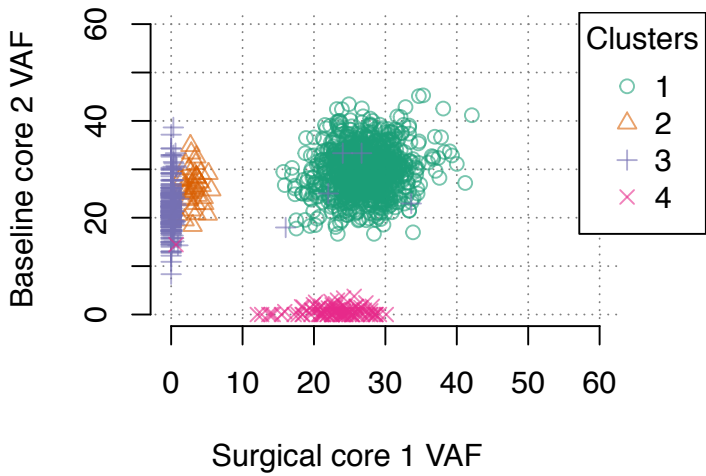
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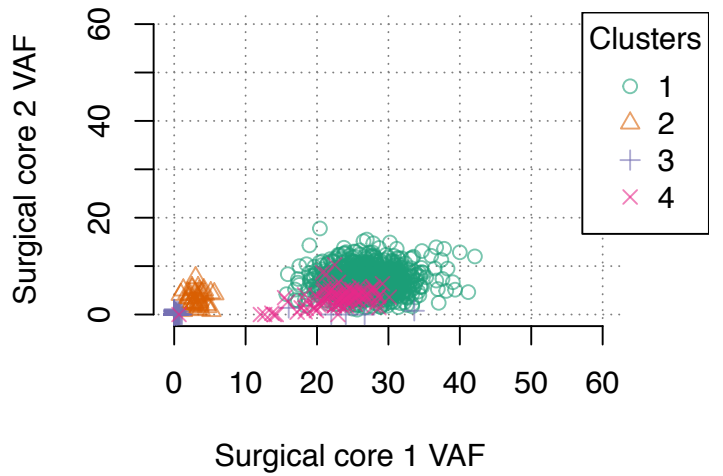
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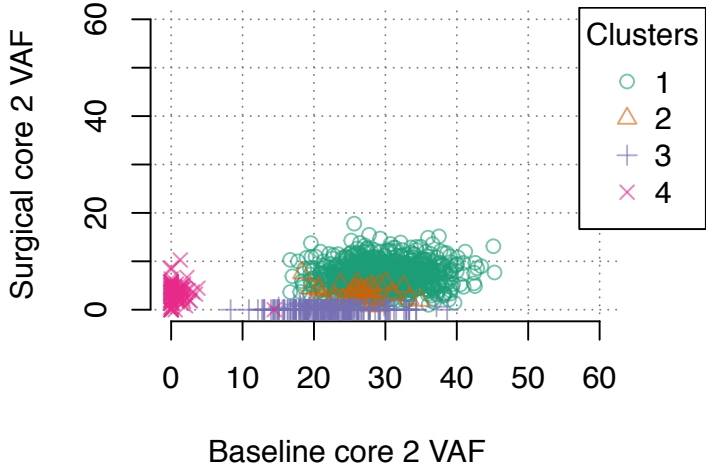
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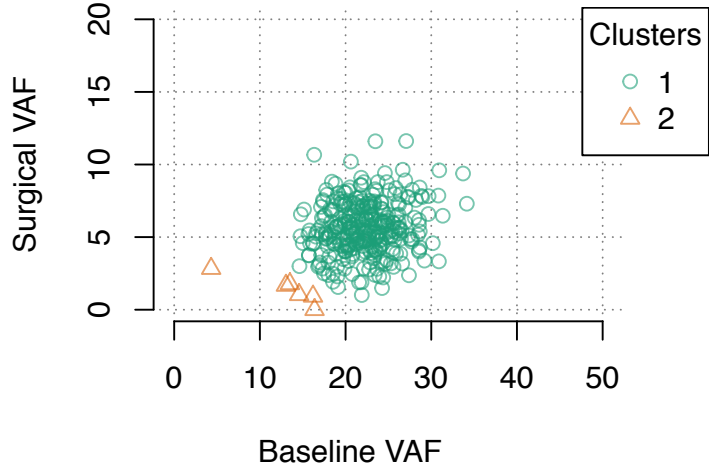
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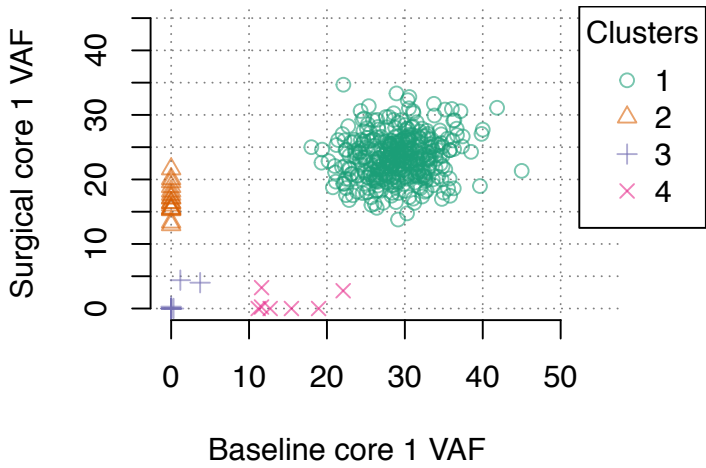
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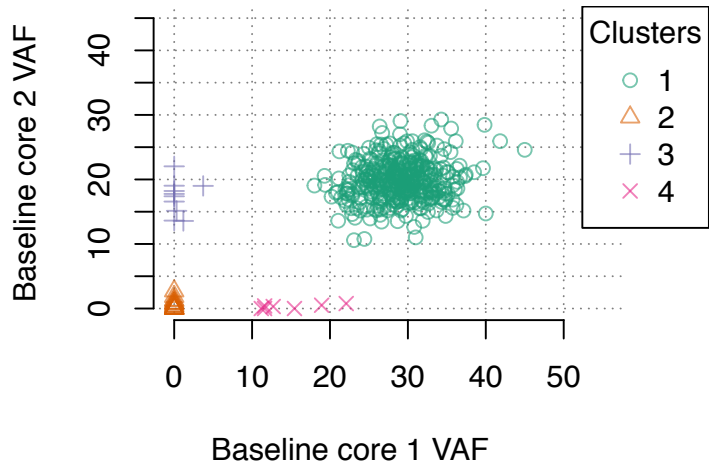
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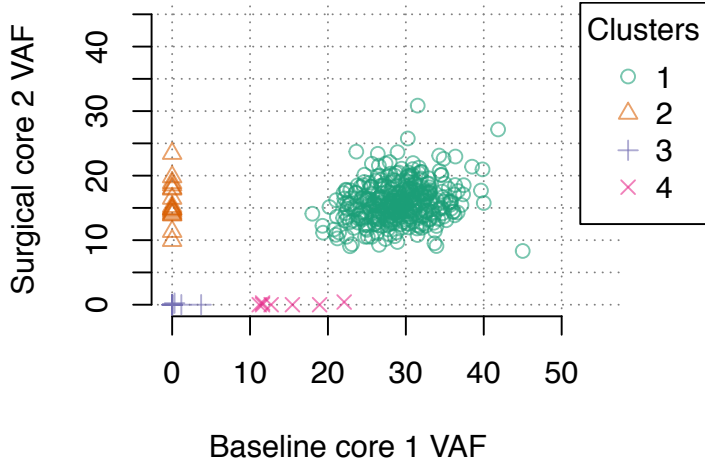
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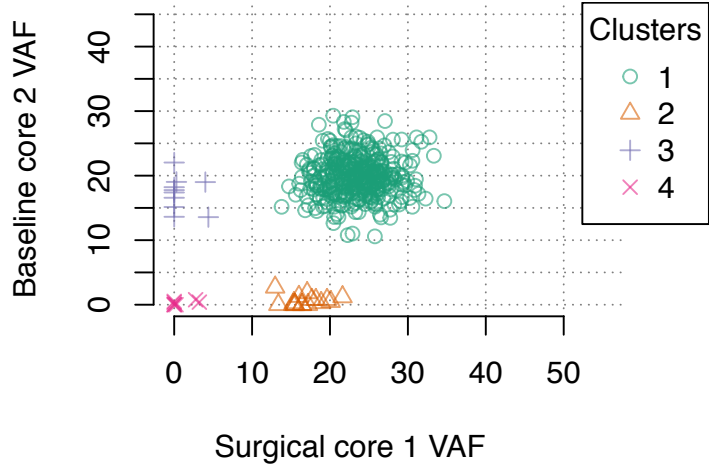
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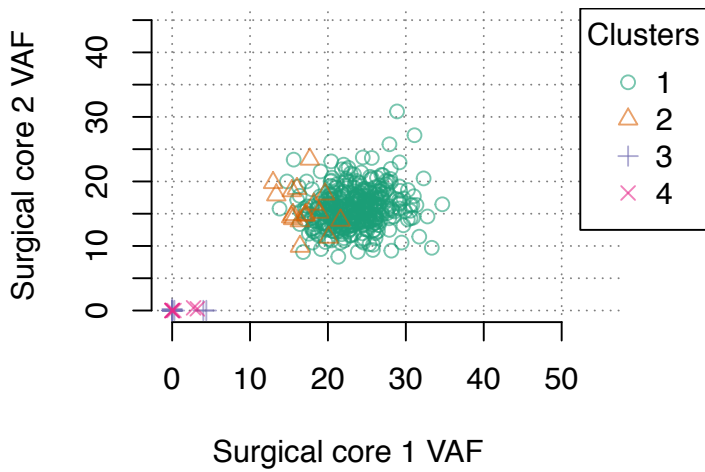
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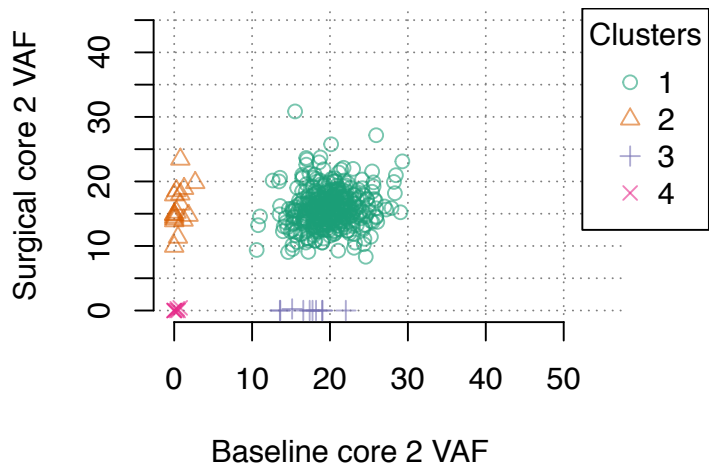
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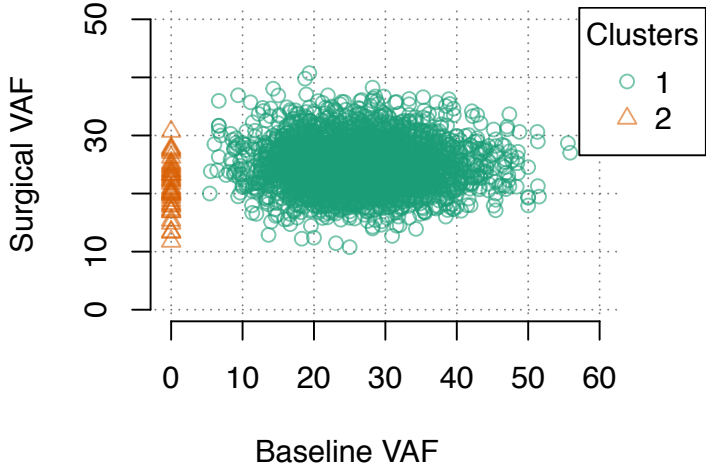
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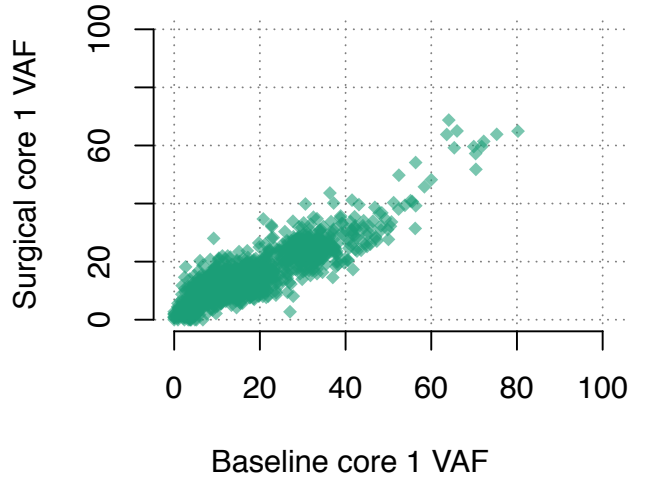
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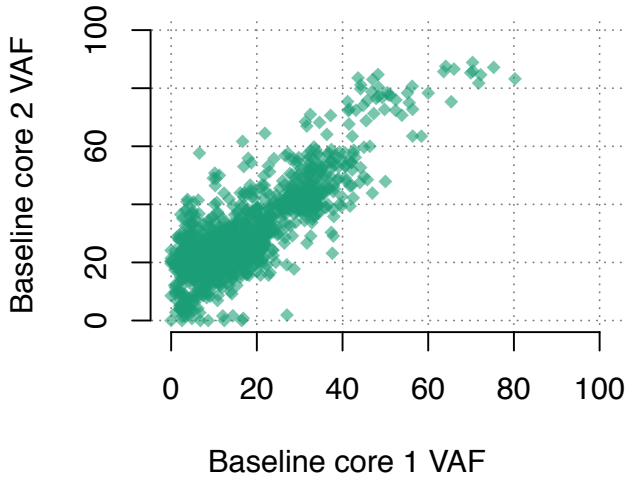
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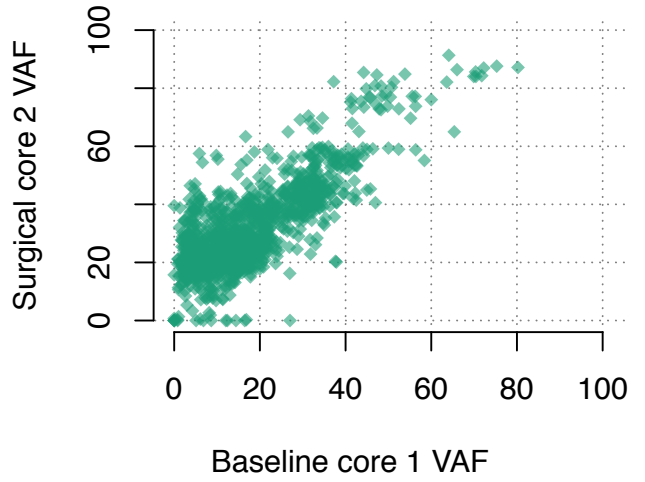
BRC17



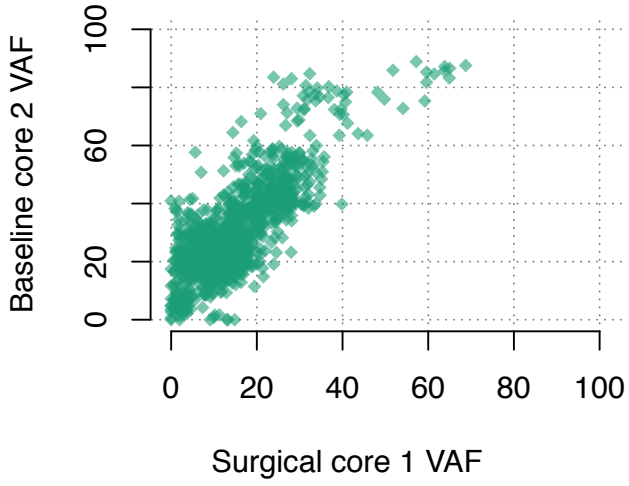
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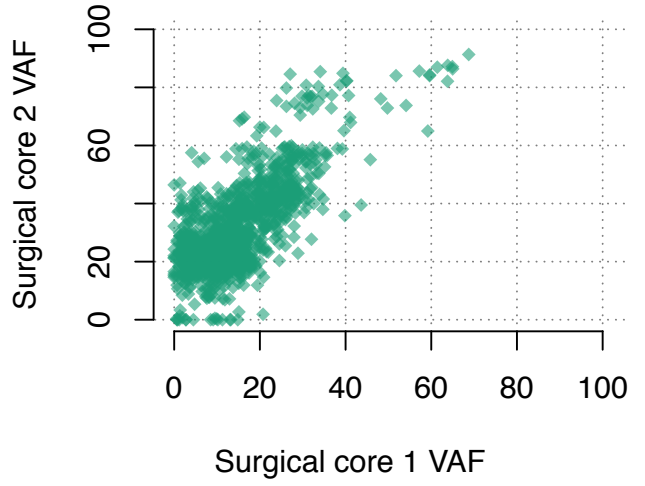
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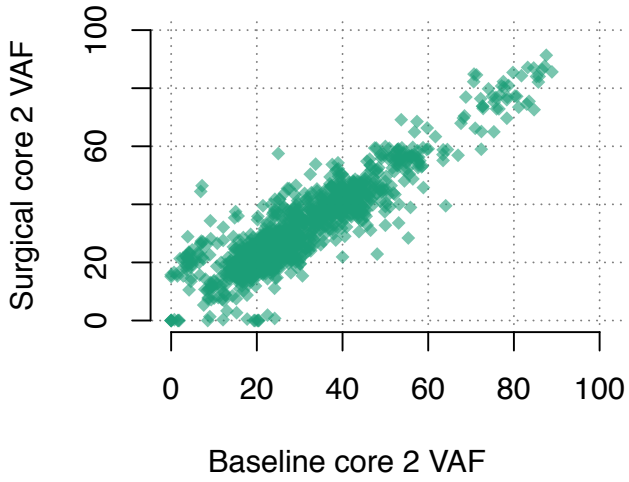
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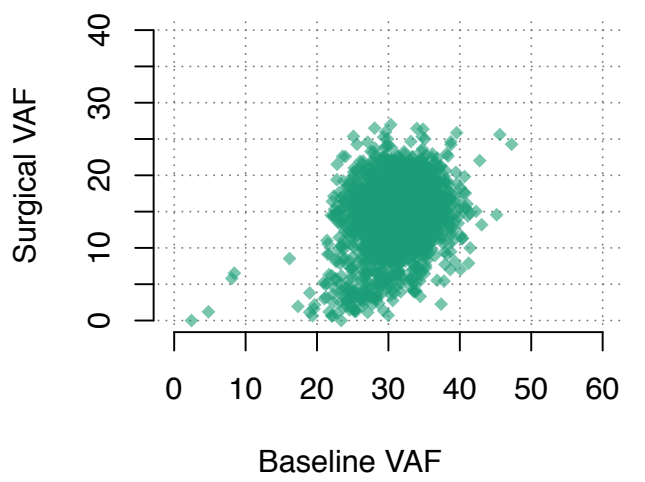
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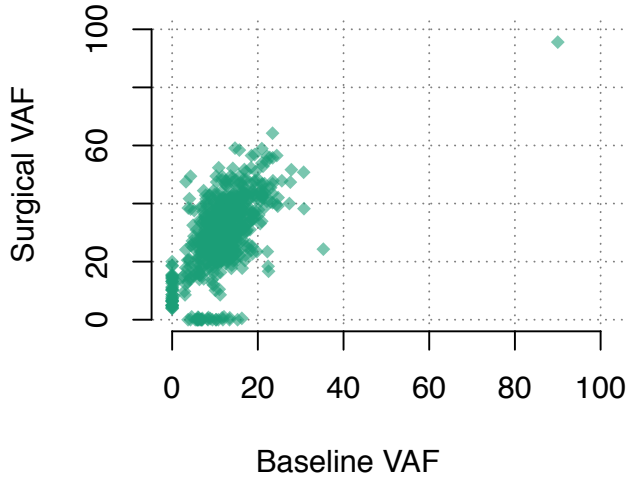
BRC17



BRC18

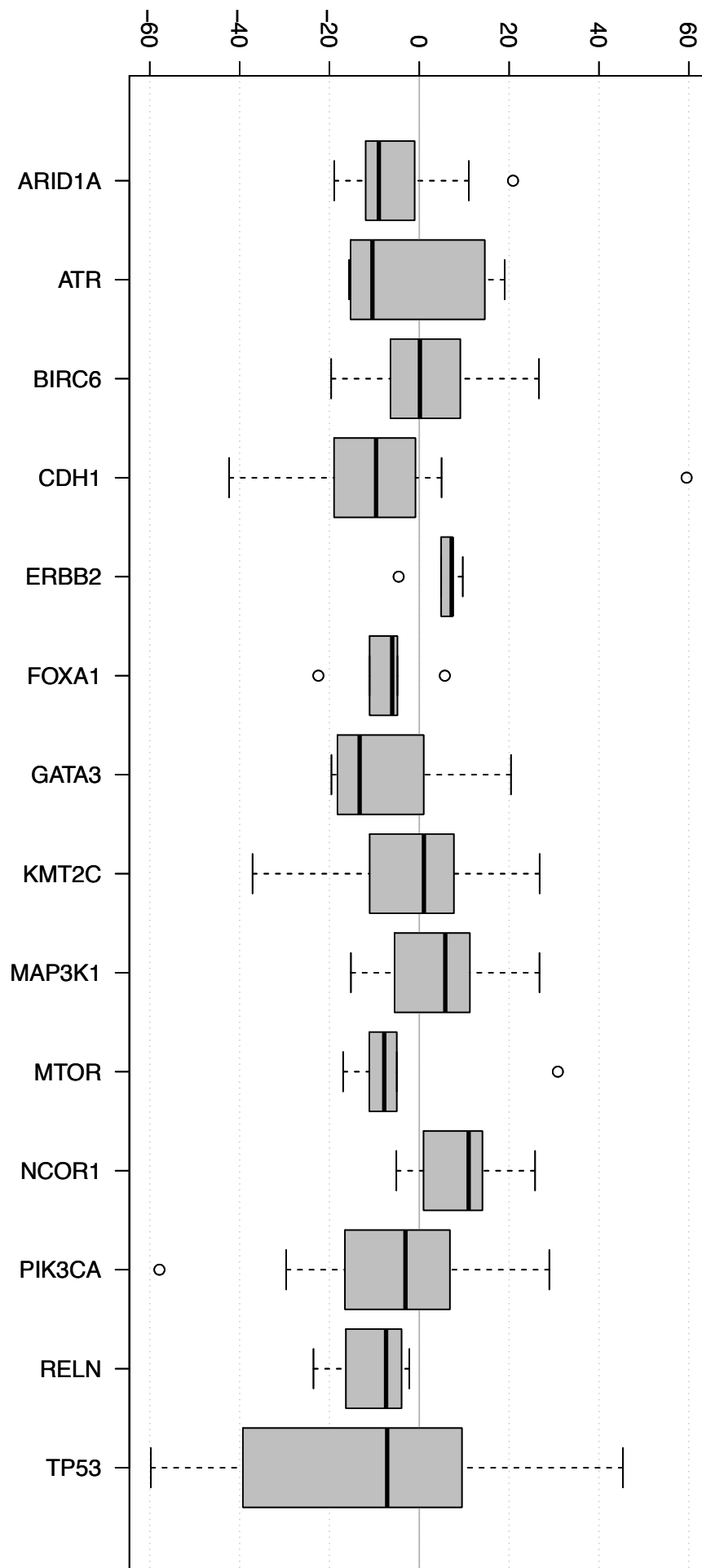


BRC48



Supplementary Figure 8: VAF changes of mutations in key genes in the extension cohort.
No gene had a shift in a positive or negative direction that reached significance in this cohort.

VAF change (Post-Pre)



Supplementary Notes

Supplementary Note 1: Integrated Analysis of DNA and RNA

30.6% of all coding mutations were expressed at detectable levels in the RNA data. In an attempt to identify specific drivers of response and resistance, baseline- and surgical-specific variants were identified and further subset to retain only expressed variants. Key cancer-related mutations found specifically in surgical samples include BCAS3 (p.K533N) and MLLT4 (p.M855L). Mutations observed in the baseline samples but completely absent in the surgical tumors include MSH6 (p.K1140*), BRAF (p.K601E), FOXA1 (p.R265H), as well as a splice-site mutation in CDH1. In BRC21, we also observe a large reduction in the VAF of a GATA3 splice-site mutation, declining from nearly every cell in the baseline-detected tumor to only about 19% of them. No changes of this type were detected in ESR1 or in related pathway genes. Sixty-two expressed coding mutations were specific to the surgical samples

Supplementary Note 2: Identification of Gene Fusions

With the INTEGRATE algorithm, we utilized both DNA and RNA sequence data to identify putative expressed gene fusion transcripts. Out of 38 samples with RNA sequence data, we detected a total of 612 fusion candidates with supporting reads/evidence in both RNA and DNA sequence data (Supplementary Data 6). Out of these candidates, 147 are “tier1”, having higher confidence due to identification of both encompassing and spanning reads. Cancer-related genes involved in fusions include TP53 (BRC30), JAK1 (BRC41, BRC15, and BRC20), and FLT3 (BRC26 post treatment).

Supplementary Note 3: Transcriptome analysis

RNA sequencing was performed on both pre and post-treatment tumors, allowing us to do detailed differential expression analysis. We identified 44 genes that were differentially expressed between the AI-sensitive and AI-resistant tumors (Supplementary Data 8): 24 genes with higher expression in AI-resistant samples and 20 genes with higher expression in AI-sensitive tumors.

Unsupervised hierarchical clustering of FPKM values grouped 16 of the 32 baseline and surgical samples from the same patient together (Supplementary Figure 4), indicating that in those cases, inter-tumor expression variability was greater than expression changes induced by therapy. Such clustering failed to find a clear signature that differentiates between AI-sensitive and AI-resistant tumors, whether given inputs of baseline, surgical, or all tumors (Supplementary Figures 5-6), and whether we evaluated all genes or a subset of the most variable genes. Clustering of Luminal A and B tumors independently also did not distinguish between response classes (Supplementary Figures 7-8).

While transcription levels of estrogen-receptor pathway genes (as defined by GO terms or KEGG pathways) were often variable between timepoints, differences in their transcription levels between baseline and surgical samples did not have a clear relationship to the clonal instability of the tumor or the response to therapy.

Supplementary Note 4: Spatial organization of the tumors

All cores were taken according to the study protocol, but the locations would have been essentially random due to two factors: The first is that when dealing with fresh tissue biopsies, the center and periphery of the tumor are only grossly identifiable by visual inspection. The second is that this was a multi-center trial and there were no specific guidelines about biopsy location communicated to the many surgeons through the study protocol. Thus, this study cannot answer questions about the spatial organization of the tumors (nor was it designed to).

Supplementary Note 5: DCIS Samples from BRC15

Three additional samples from the periphery of BRC15 tumor were obtained at surgery, all classified as Ductal Carcinoma in situ (DCIS) by pathology. Each was sequenced with a targeted panel of 83 breast-cancer related genes (Supplementary Data 11). Two of the DCIS samples contained *BIRC6* and *PIK3CA* mutations identical to those observed in the tumor's founder clone, suggesting that they share a genetic origin with the same pre-neoplastic cells that gave rise to the tumor. The third DCIS sample contained the same pair of founder clone mutations, as well as the observed *BRAF* mutation at a VAF of 13.11% (representing about 35% of the tumor cells in that sample). This indicates a shared ancestry between this lesion and the cells that comprised cluster three in the invasive cancer.