

1 **Supporting Figure Legends**

2 **Figure S1. Competition assay of *E. faecalis* harboring plasmid vector.** The overnight
3 cultures of *E. faecalis* OG1S and *E. faecalis* OG1S harboring pAM401 (open bar) or
4 pHT1100 (closed bar) were mixed equally with a 100-dilution in fresh THB broth,
5 followed by co-cultivation at 37°C. At the indicated time point, the percentage of
6 plasmid-harboring cells in the entire population was determined by the ratio of cfu on
7 THB with chloramphenicol (plasmid-harboring cells) against that of THB without
8 antibiotics (sample of the entire population including cells with and without plasmid).

9

10 **Figure S2. Diversity of BacL₁ and BacL₂ homologues.** Phylogenetic tree of BacL₁ (A)
11 and BacL₂ (B) homologues of *E. faecalis* strains was constructed using JTT model by
12 MEGA7 based on the amino acid sequences alignment generated by ClustalW ver.2.0.
13 The tree with the highest log likelihood is shown. The percentages of trees in which the
14 associated taxa clustered together are shown next to the branches. Initial tree(s) for the
15 heuristic search were obtained automatically by applying Neighbor-Join and BioNJ
16 algorithms to a matrix of pairwise distances estimated using a JTT model, and then
17 selecting the topology with superior log likelihood value. The tree is drawn to scale,
18 with branch lengths measured in the number of substitutions per site. The analysis
19 involved 148 or 153 amino acid sequences for BacL₁ or BacL₂, respectively. All
20 positions containing gaps and missing data were eliminated. Names of the source strain
21 and plasmid were represented and colored by Bac41-type (clade) as shown in Fig. 2.
22 Type (clade) I, orange; type (clade) IIa, yellow; type (clade) IIb, pink; type (clade) III,
23 blue.

24

25 **Figure S3. Amino acid sequence alignment of BacA homologues.** Amino acid
26 sequences of various clades BacA were aligned. The regions predicted as peptidoglycan

27 binding (PGB) domain and glycosyl hydrolase 25 (GH25) domain are highlighted by
28 grey boxes. Immunity determinant region suggested in this study is highlighted by red
29 box. The region that shows low similarity between clades IIa- and IIb-BacA is
30 highlighted by orange box. Predicted transmembrane (TM) domain was highlighted by
31 yellow. Names of the source strain and plasmid are represented.

32

33 **Figure S4. Diversity of BacI1 and BacI2 homologues.** Phylogenetic tree of BacI1 (A)
34 and BacI2 (B) homologues of *E. faecalis* strains was constructed using JTT model by
35 MEGA6 based on the amino acid sequences alignment generated by ClustalW ver.2.0.
36 The tree with the highest log likelihood is shown. The percentages of trees in which the
37 associated taxa clustered together are shown next to the branches. Initial tree(s) for the
38 heuristic search were obtained automatically by applying Neighbor-Join and BioNJ
39 algorithms to a matrix of pairwise distances estimated using a JTT model, and then
40 selecting the topology with superior log likelihood value. The tree is drawn to scale,
41 with branch lengths measured in the number of substitutions per site. The analysis
42 involved 125 or 159 amino acid sequences for BacI1 or BacI2, respectively. All
43 positions containing gaps and missing data were eliminated. Names of the source strain
44 and plasmid were represented and colored by Bac41-type (clade) as shown in Fig. 2A.
45 Type (clade) I, orange; type (clade) IIa, yellow; type (clade) IIb, pink; type (clade) IIc,
46 cyan.

47

48 **Figure S5. Amino acid sequence alignment of BacI1 and BacI2 homologues.** Amino
49 acid sequences of various clades BacI1 (A) or BacI2 (B) were aligned. Predicted
50 transmembrane (TM) domains are highlighted by yellow. Inside or outside regions
51 predicted by TMHMM server are also indicated. Names of the source strain and plasmid
52 were represented.

