Supplemental Information

S1 Table. List of all oxidative stress and antioxidant PCR primer targets on the PCR array. Red genes represent targets that are not expressed in CD8+ T cells.

Red gene	s represent taig	ets that	are not empress		or i cens.		
ALB	Albumin	GLRX 2	Glutaredoxin 2	MTL5	Metallothionein -like 5, testis- specific (tesmin)	PTGS1	Prostaglandin- endoperoxide synthase 1 (prostaglandin G/H synthase and cyclooxygenase)
ALOX12	Arachidonate 12- lipoxygenase	GPR 156	G protein- coupled receptor 156	NCF1	Neutrophil cytosolic factor 1	PTGS2	Prostaglandin- endoperoxide synthase 2 (prostaglandin G/H synthase and cyclooxygenase)
ANGPTL 7	Angiopoietin- like 7	GPX1	Glutathione peroxidase 1	NCF2	Neutrophil cytosolic factor 2	PXDN	Peroxidasin homolog (Drosophila)
AOX1	Aldehyde oxidase 1	GPX2	Glutathione peroxidase 2 (gastrointestinal)	NME5	Non-metastatic cells 5, protein expressed in (nucleoside- diphosphate kinase)	PXDNL	Peroxidasin homolog (Drosophila)-like
APOE	Apolipoprotein E	GPX3	Glutathione peroxidase 3 (plasma)	NOS2	Nitric oxide synthase 2, inducible	RNF7	Ring finger protein 7
ATOX1	ATX1 antioxidant protein 1 homolog (yeast)	GPX4	Glutathione peroxidase 4 (phospholipid hydroperoxidase)	NOX5	NADPH oxidase, EF- hand calcium binding domain 5	SCARA 3	Scavenger receptor class A, member 3
BNIP3	BCL2/adenoviru s E1B 19kDa interacting protein 3	GPX5	Glutathione peroxidase 5 (epididymal androgen-related protein)	NUDT1	Nudix (nucleoside diphosphate linked moiety X)-type motif 1	SELS	Selenoprotein S
CAT	Catalase	GPX6	Glutathione peroxidase 6 (olfactory)	OXR1	Oxidation resistance 1	SEPP1	Selenoprotein P, plasma, 1
CCL5	Chemokine (C-C motif) ligand 5	GPX7	Glutathione peroxidase 7	OXSR1	Oxidative-stress responsive 1	SFTPD	Surfactant protein D
CCS	Copper chaperone for superoxide dismutase	GSR	Glutathione reductase	PDLIM 1	PDZ and LIM domain 1	SGK2	Serum/glucocorticoid regulated kinase 2
CSDE1	Cold shock domain containing E1, RNA-binding	GSS	Glutathione synthetase	IPCEF 1	Interaction protein for cytohesin exchange factors 1	SIRT2	Sirtuin 2
СҮВА	Cytochrome b-245, alpha	GSTZ1	Glutathione transferase zeta 1	PNKP	Polynucleotide kinase 3'- phosphatase	SOD1	Superoxide dismutase 1, soluble

	polypeptide						
CYGB	Cytoglobin	GTF2I	General transcription factor IIi	PRDX1	Peroxiredoxin 1	SOD2	Superoxide dismutase 2, mitochondrial
DGKK	Diacylglycerol kinase, kappa	KRT1	Keratin 1	PRDX2	Peroxiredoxin 2	SOD3	Superoxide dismutase 3, extracellular
DHCR2 4	24- dehydrocholestero l reductase	LPO	Lactoperoxidase	PRDX3	Peroxiredoxin 3	SRXN1	Sulfiredoxin 1
DUOX1	Dual oxidase 1	MBL2	Mannose- binding lectin (protein C) 2, soluble	PRDX4	Peroxiredoxin 4	STK25	Serine/threonine kinase 25
DUOX2	Dual oxidase 2	MGST 3	Microsomal glutathione S- transferase 3	PRDX5	Peroxiredoxin 5	ТРО	Thyroid peroxidase
DUSP1	Dual specificity phosphatase 1	MPO	Myeloperoxidas e	PRDX6	Peroxiredoxin 6	TTN	Titin
EPHX2	Epoxide hydrolase 2, cytoplasmic	MPV17	MpV17 mitochondrial inner membrane protein	PREX1	Phosphatidylinositol -3,4,5-trisphosphate- dependent Rac exchange factor 1	TXNDC 2	Thioredoxin domain containing 2 (spermatozoa)
EPX	Eosinophil peroxidase	MSRA	Methionine sulfoxide reductase A	PRG3	Proteoglycan 3	TXNRD 1	Thioredoxin reductase 1
FOXM1	Forkhead box M1	MT3	Metallothionein	PRNP	Prion protein	TXNRD	Thioredoxin reductase 2

S2 Table. Exhaustive list of fold changes and their corresponding p-values in targets expressed in CD8+ T cells. A fold change below 1 corresponds to a downregulation $(2^{-\Delta CT})$.

Symbol	Protein Name	Fold Change	p-value
DDI IMI		Old/Young	0.01
PDLIMI		0.17	0.01
ANGPIL/	Angiopoleun-like /	2.95	0.01
MIL5	Metallothionein-like 5, testis-specific (tesmin)	0.45	0.01
NUDTI	motif 1	0.99	0.01
TTN	Titin	2.14	0.01
DUOX1	Dual oxidase 1	1.68	0.03
GPX3	Glutathione peroxidase 3 (plasma)	2.55	0.04
GLRX2	Glutaredoxin 2	0.47	0.06
PRNP	Prion protein	1.45	0.07
TXNRD1	Thioredoxin reductase 1	0.31	0.08
PRDX1	Peroxiredoxin 1	0.74	0.13
CCL5	Chemokine (C-C motif) ligand 5	3.81	0.15
TXNRD2	Thioredoxin reductase 2	1.15	0.18
GPX2	Glutathione peroxidase 2 (gastrointestinal)	2.15	0.20
SEPP1	Selenoprotein P, plasma, 1	0.34	0.21
ATOX1	ATX1 antioxidant protein 1 homolog (yeast)	0.50	0.22
BNIP3	BCL2/adenovirus E1B 19kDa interacting protein 3	1.12	0.23
CCS	Copper chaperone for superoxide dismutase	0.22	0.23
PX1	Glutathione peroxidase 1	0.18	0.23
PREX1	Phosphatidylinositol-3,4,5-trisphosphate-dependent Rac exchange factor 1	0.90	0.24
PRDX5	Peroxiredoxin 5	0.80	0.29
CSDE1	Cold shock domain containing E1, RNA-binding	0.49	0.30
SIRT2	Sirtuin 2	1.25	0.32
FOXM1	Forkhead box M1	1.78	0.33
GTF2I	General transcription factor IIi	0.52	0.34
GSS	Glutathione synthetase	0.26	0.37
NCF1	Neutrophil cytosolic factor 1	9.32	0.42
ALOX12	Arachidonate 12-lipoxygenase	0.60	0.46
STK25	Serine/threonine kinase 25	4.65	0.46
GPX4	Glutathione peroxidase 4 (phospholipid hydroperoxidase)	0.02	0.47
SELS	Selenoprotein S	0.10	0.48
MPV17	MpV17 mitochondrial inner membrane protein	0.57	0.49
MGST3	Microsomal glutathione S-transferase 3	0.49	0.50
GSR	Glutathione reductase	0.73	0.51
PRDX2	Peroxiredoxin 2	0.75	0.51
SOD1	Superoxide dismutase 1, soluble	6.52	0.51
SRXN1	Sulfiredoxin 1	0.12	0.52
CAT	Catalase	122.49	0.56
MSRA	Methionine sulfoxide reductase A	0.79	0.57
MT3	Metallothionein 3	0.28	0.59
NCF2	Neutrophil cytosolic factor 2	1.49	0.59

OXR1	Oxidation resistance 1	0.02	0.59
GPX7	Glutathione peroxidase 7	0.17	0.60
EPX	Eosinophil peroxidase	0.83	0.64
СҮВА	P22-phox	0.41	0.65
GSTZ1	Glutathione transferase zeta 1	1.89	0.65
PRDX6	Peroxiredoxin 6	3.93	0.65
IPCEF1	Interaction protein for cytohesin exchange factors 1	0.65	0.66
PRDX4	Peroxiredoxin 4	2.87	0.70
SOD2	Superoxide dismutase 2, mitochondrial	0.33	0.70
DUSP1	Dual specificity phosphatase 1	4.56	0.71
RNF7	Ring finger protein 7	6.55	0.71
SFTPD	Surfactant protein D	3.35	0.72
DHCR24	24-dehydrocholesterol reductase	0.57	0.82
PRDX3	Peroxiredoxin 3	1.60	0.84
OXSR1	Oxidation resistance 1	2.20	0.95
PNKP	Polynucleotide kinase 3'-phosphatase	1.59	0.98

Symbol	Protein Name	Mean ΔCt ± SEM	% CV
СҮВА	P22-phox	3.77 ± 0.4	10.64
CCL5	Chemokine (C-C motif) ligand 5	3.58 ± 0.68	19.06
SOD1	Superoxide dismutase 1, soluble	4.19 ± 0.23	5.56
GPX4	Glutathione peroxidase 4	4.38 ± 0.40	9.24
	(phospholipid hydroperoxidase)		- 10
PRDX6	Peroxiredoxin 6	4.60 ± 0.30	6.48
GTF2I	General transcription factor IIi	4.79 ± 0.28	5.93
PRDX3	Peroxiredoxin 3	4.61 ± 0.46	9.98
PRDX2	Peroxiredoxin 2	4.78 ± 0.35	7.24
GPX1	Glutathione peroxidase 1	5.17 ± 0.23	4.48
CSDE1	Cold shock domain containing E1, RNA-binding	5.19 ± 0.23	4.48
FOXM1	Forkhead box M1	5.39 ± 0.18	3.44
PRDX5	Peroxiredoxin 5	5.38 ± 0.28	5.25
ATOX1	ATX1 antioxidant protein 1 homolog (yeast)	5.60 ± 0.21	3.70
TXNRD1	Thioredoxin reductase 1	6.17 ± 0.08	1.33
DHCR24	24-dehydrocholesterol reductase	6.38 ± 0.18	2.83
RNF7	Ring finger protein 7	6.58 ± 0.21	3.13
SOD2	Superoxide dismutase 2, mitochondrial	6.59 ± 0.21	3.12
OXSR1	Oxidative-stress responsive 1	6.98 ± 0.10	1.47
CAT	Catalase	6.78 ± 0.4	5.92
PRDX4	Peroxiredoxin 4	6.99 ± 0.36	5.18
MGST3	Microsomal glutathione S-transferase 3	7.18 ± 0.23	3.22
MPV17	MpV17 mitochondrial inner membrane protein	7.18 ± 0.23	3.21
GLRX2	Glutaredoxin 2	7.39 ± 0.18	2.49
GSS	Glutathione synthetase	7.37 ± 0.28	3.81
BNIP3	BCL2/adenovirus E1B 19kDa interacting protein 3	7.38 ± 0.34	4.62
SELS	Selenoprotein S	7.78 ± 0.28	3.62
STK25	Serine/threonine kinase 25	7.98 ± 0.10	1.24
TXNRD2	Thioredoxin reductase 2	7.79 ± 0.34	4.43
NUDT1	Nudix (nucleoside diphosphate linked moiety X)-type motif 1	8.18 ± 0.09	1.04
PNKP	Polynucleotide kinase 3'-phosphatase	7.99 ± 0.31	3.89
GPX7	Glutathione peroxidase 7	8.00 ± 0.31	3.89
OXR1	Oxidation resistance 1	8.19 ± 0.23	2.85
PRDX1	Peroxiredoxin 1	8.19 ± 0.30	3.70
CCS	Copper chaperone for superoxide dismutase	8.80 ± 0.19	2.13
IPCEF1	Interaction protein for cytohesin exchange factors 1	8.57 ± 0.58	6.78
MTL5	Metallothionein-like 5, testis-specific (tesmin)	8.97 ± 0.31	3.46
SIRT2	Sirtuin 2	9.01 ± 0.38	4.20
PREX1	Phosphatidylinositol-3,4,5-trisphosphate-dependent	9.18 ± 0.23	2.49
	Rac exchange factor 1		
GSTZ1	Glutathione transferase zeta 1	9.19 ± 0.37	4.07
NCF2	Neutrophil cytosolic factor 2	9.38 ± 0.39	4.20
NCF1	Neutrophil cytosolic factor 1	9.00 ± 0.89	9.91

S3 Table S3. Normalized mRNA levels of individual genes expressed in young CD8+ T cells, ranked in descending order of expression (n=6).

SRXN1	Sulfiredoxin 1	10.18 ± 0.23	2.26
MSRA	Methionine sulfoxide reductase A	10.18 ± 0.3	2.93
DUSP1	Dual specificity phosphatase 1	10.19 ± 0.43	4.19
GPX3	Glutathione peroxidase 3 (plasma)	10.37 ± 0.49	4.76
SEPP1	Selenoprotein P, plasma, 1	10.57 ± 0.29	2.80
PRNP	Prion protein	11.01 ± 0.10	0.90
GSR	Glutathione reductase	10.78 ± 0.34	3.17
PDLIM1	PDZ and LIM domain 1	10.98 ± 0.3	2.79
TTN	Titin	11.19 ± 0.42	3.79
MT3	Metallothionein 3	11.4 ± 0.73	6.40
ANGPTL7	Angiopoietin-like 7	12.17 ± 0.23	1.87
ALOX12	Arachidonate 12-lipoxygenase	12.17 ± 0.42	3.48
EPX	Eosinophil peroxidase	12.58 ± 0.41	3.29
DUOX1	Dual oxidase 1	12.78 ± 0.34	2.68
SFTPD	Surfactant protein D	12.57 ± 0.76	6.07
PXDN	Peroxidasin homolog (Drosophila)	13.21 ± 0.62	4.70
GPX2	Glutathione peroxidase 2 (gastrointestinal)	13.99 ± 0.47	3.36

S1 Fig. Example flow cytometry median calcium traces from a donor obtained at Day 4 and Day 24. After cold-binding with anti-CD3 and anti-CD28 as described in Materials and Methods, cells were sampled for 3 minutes by flow cytometry before addition of anti-mouse IgG. The average of the fluorescence ratio was calculated at 4 second intervals and then normalized to the average baseline value to provide a fold-change value.



S2 Fig. Optimization of Jurkat T Cell Model using data obtained by no inhibitor, TMB-8, and EGTA conditions. Plots represent 17 different optimized parameter sets that were obtained

from comparing the model prediction to experimental data.





S3 Fig. Jurkat T Cell model behavior by parameter set used to fit Young CD8+ model

S4 Table. Optimized parameter set obtained from the Jurkat T Cell Model fitting used for the seeding the initial population of parameter values for the genetic algorithm optimization of the Young CD8⁺ T Cell Model to experimental data.

Parameter	Jurkat T cells
β_i^*	0.056
β_{er}^*	0.049
β_{mit}	0.033
ρ_{er}	0.015
ρ_{mit}	0.08
k _{PLCact}	0.0033 s ⁻¹
k _{PLCdeact}	0.042 s ⁻¹
k _{IP3prod}	$0.48 \ \mu M^{-1} s^{-1}$
k_{IP3deg}^{*}	0.010 s ⁻¹
V _{IP3}	4.0 s ⁻¹
K _{IP3}	0.57 µM
K _{act}	0.13 µM
Α	0.079
K _{inh}	1 μM
K _{IP3inh} *	0.82 µM
K _{ERleak} *	0.0043 s ⁻¹
V _{serca} *	$112.75 \ \mu M \ s^{-1}$
K _{serca}	0.43 µM
V _{mitin}	$388.6 \ \mu M \ s^{-1}$
K _{mitin}	0.81 µM
V _{mitout} *	$188.9 \ \mu M \ s^{-1}$
K _{mitout} *	4.03 µM
V _{crac}	$2.4 \ \mu M \ s^{-1}$
K _{soc} *	363.5 µM
K _{stim}	178.1 μM

K _{PMleak}	1.1e-6 s ⁻¹
V _{pmca} *	$2.14 \ \mu M \ s^{-1}$
K _{pmca}	0.11 µM

S5 Table. Optimized parameter set obtained from fitting the Young CD8⁺ T Cell Model to experimental data. This parameter set was used for all sensitivity analysis performed on the Young CD8⁺ T Cell Model.

Parameter	Primary CD8 ⁺ T cells
β_i^*	0.047
β_{er}^*	0.98
β_{mit}	0.033
$ ho_{er}$	0.015
$ ho_{mit}$	0.08
k _{PLCact}	0.0033 s ⁻¹
k _{PLCdeact}	0.042 s ⁻¹
k _{IP3prod}	$0.48 \ \mu M^{-1} \ s^{-1}$
k_{IP3deg}^*	$0.018 \ \mu s^{-1}$
V _{IP3}	4.0 µs ⁻¹
K _{IP3}	0.57 μΜ
K _{act}	0.13 μM
A	0.079
K _{inh}	1 μ M
K _{IP3inh} *	1.5 μM
K _{ERleak} *	0.048 s ⁻¹
V _{serca} *	103.88 μM s ⁻¹
K _{serca}	0.43 µM
V _{mitin}	$388.6 \ \mu M \ s^{-1}$
K _{mitin}	0.81 µM
V _{mitout} *	244.7 $\mu M s^{-1}$
K _{mitout} *	4.7 μΜ
V _{crac}	$2.4 \ \mu M \ s^{-1}$

K _{soc} *	358.8 µM
K _{stim}	178.1 μM
K _{PMleak}	1.1e-6 s ⁻¹
V _{pmca} *	$2.08 \ \mu M \ s^{-1}$
K _{pmca}	0.11 μM







S5 Fig. Best fit of Old CD8⁺ T Cell Model varying only two parameters, V_{crac} and V_{pmca} .

S6 Fig. Best fit of Old CD8⁺ Young T Cell Model varying the seven parameters as identified in the sensitivity analysis of the Young CD8⁺ Young T Cell Model.



S7 Fig. Varying K_{stim} from the Young CD8⁺ T Cell Model fit to investigate the effects on

calcium traces. K_{stim} was varied +/- 20% the fit value of 178.



S8 Fig. Varying V_{crac} from the Young CD8⁺ T Cell Model to investigate the effects on calcium traces. V_{crac} was varied +/- 20% the fit value of 2.37.



S9 Fig. Validation of RT-PCR results with Duox 1 expression. a) Representative Western Blot. b) Quantification of the Western Blots. Protein levels are normalized to the young cells protein expression level. * p<0.05 (paired 2-tail t-test).





S10 Fig. Expression of STIM1 in young and old primary human CD8+ T cells.