

SUPPLEMENTARY FIGURES

Negative Feedback Regulation of HIV-1 by Gene Editing Strategy

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Table S1

primer	sequence
1. Cloning pX260-LTR-Cas9 constructs	
Kpn1-LTR(-454)-F	5'- GGTACCT GGAAGGGCTAATTTGG-3'
Kpn1-LTR(-120)-F	5'- GGTACCT CGAGCTTTCTACAAGG-3'
Xba1-LTR(-80)-F	5'- TCTAGAGGAGGTGTGGCCTGGGC-3'
LTR(+66)-Nco1-R	5'- CCATGGT AAGCAGTGGGTTCC-3'
2. Cloning lentiLTR(-80/+66)-Cas9-Blast construct	
Nhe1-LTR(-80)-F	5'- GCTAGCGGAGGTGTGGCCTGGGC-3'
LTR(+66)-Xba1-R	5'- TCTAGATAAGCAGTGGGTTCC-3'
3. PCRs	
LTR -417/F	5'-GATCTGTGGATCTACCACACACA-3'
LTR -19/R	5'-GCTGCTTATATGTAGCATCTGAG-3'
LTR -374/F	5'-TTAGCAGAACTACACACCAGGGCC-3'
LTR +43/R	5'-CCGAGAGCTCCCAGGCTCAGATCT-3'
HIV-1 5'UTR +97/F	5'-AAGTAGTGTGTGCCCGTCTG-3'
HIV-1 5'UTR +235/R	5'-TCGAGAGATCTCCTCTGGCT-3'
HIV-1 Env +5828/F	5'- TCCTTGGGATGTTGATGATCT-3'
HIV-1 Env +5977/R	5'- TGGCCCAAACATTATGTACC-3'
b- actin/F	5'-CTACAATGAGCTGCGTGTGGC-3'
b-actin/R	5'-CAGGTCCAGACGCAGGATGGC-3'
4. Taqman qPCRs	
HIV-1 5'UTR F	5'-AAGTAGTGTGTGCCCGTCTG-3'
HIV-1 5'UTR R	5'-TCGAGAGATCTCCTCTGGCT-3'
HIV-1 5'UTR probe	5'-FAM-CTGTTCTGGGCGCCACTGCTA-ZEN-IowaBlackFQ-3'
HIV-1 env F	5'- TCCTTGGGATGTTGATGATCT-3'
HIV-1 env R	5'- TGGCCCAAACATTATGTACC-3'
HIV-1 env probe	5'-FAM-TGGTGGTGGTCTTCTTTCCACACA-ZEN-IowaBlackFQ-3'
Hs b-globin F	5'-CCCTTGGACCCAGAGGTTCT-3'
Hs b-globin R	5'-CGAGCACTTTCTTGCCATGA-3'
Hs b-globin probe:	5'-FAM-GCGAGCATCTGTCCACTCCTGATGCTGTTATGGGCGCTCGC-ZEN-IowaBlackFQ-3'
Hs b-actin F	5'-TGGACTTCGAGCAAGAGATG-3'
Hs b-actin R	5'-GAAGGAAGGCTGGAAGAGTG-3'
Hs b-actin probe:	5'-FAM-CGGCTGCTTCCAGCTCCTCC-ZEN-IowaBlackFQ-3'

SUPPLEMENTAL FIGURE LEGENDS

Figure S1. Position and nucleotide sequences of gRNA A/B targets within the LTR (highlighted in green, PAM in red) and LTR specific primers used in PCR on TZM-bl and in vitro infected Jurkat cells genomic DNA (highlighted in blue) in the reference HIV-1 NL4-3 genome. Sequences and sizes of LTR specific PCR products (full-length and truncated) and predicted edited fragment.

Figure S2. A representative agarose gel analyzing LTR specific PCR reactions used for quantification of Cas9/gRNA mediated LTR excision efficiency in experiments using the Jurkat 2D10 reporter cell line from Figures 3 and 4.

Figure S3. Position and nucleotide composition of LTR gRNA A/B targets (highlighted in green, PAM in red) and LTR specific primers used to analyze excision by PCR in Jurkat 2D10 cells (highlighted in blue) in the reference HIV-1 NL4-3 genome. Nucleotide sequences and sizes of amplicons (full-length and truncated LTR DNA) and predicted excised DNA fragment are shown.

Figure S4. (a) A representative fluorescence microscopy images of transduced/infected Jurkat cells at day 5 of infection. Expression of BFP is indicative of the presence a vector expressing gRNAs. HIV-1 infection was monitored by the level of GFP. **(b)** Quantitative comparison of cell numbers at various time points between the control and experimental samples treated with LTR-Cas9.

Figure S5. Primary human fetal astrocytes and microglia were transduced with lentiviral cocktails containing: lenti-LTR-80/+66-Cas9 (MOI 10), lenti-KLV-BFP-LTR A, B (MOI 3.3 of each). At day 3 post-transduction cells were infected with HIV-1_{NL4-3-GFP-P2A-Nef/VSV-G} at MOI 1. One week after HIV-1 infection cells were harvested and viral expression levels were quantified by GFP

expression in flow cytometry **(a)** viral DNA levels **(b)** and viral RNA **(c)** by Taqman qPCR and qRT-PCRs using primer set and probe specific for Gag gene.

Figure S1

TZMbl

F [-413/-391] (T361)
TGGAAGGGCTAATTTGGTCCCAAAAAAGACAAGAGATCCTT **GATCTGTGGATCTACCACACACA** AGGCTA
LTR A [-347/-328] PAM
CTTCCCTGATTGGCAGA ACTACACACCAGGGCCAGGG **ATCAGATATCCACTGACCTT** TGGATGGTGCTTC
AAGTTAGTACCAGTTGAACCAGAGCAAGTAGAAGAGGCCAATGAAGGAGAGAACAACAGCTTGTACACC
CTATGAGCCAGCATGGGATGGAGGACCCGGAGGGAGAAGTATTAGTGTGGAAGTTTGACAGCCTCCTAGC
PAM LTR B [-143/-124]
ATTTTCGTCACATGGCCCGAGAGCTGCAT **CCG GAGTACTACAAAGACTGCTG** ACATCGAGCTTTCTACAAG
R [-41/-19] (T363)
GGACTTCCGCTGGGGACTTTCCAGGGAGGTGTGGCCTGGGCGGGACTGGGGAGTGGCGAGCC **CTCAGAT**
Transcription start[+1]
GCTACATATAAGCAGC TGCTTTTTGCCTGTACTG **G**GTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGC
TCTC **T**GGCTAACTAGGGAACCCACTGCTTAAGCCTCAATAAAGCTTGCCTTGAGTGCTCAAAGTAGTGTG

PCR product full length LTR -413/-19: 395bp

GATCTGTGGATCTACCACACACA AGGCTACTTCCCTGATTGGCAGA ACTACACACCAGGGCCAGGG **ATCA**
GATATCCACTGACCTT TGGATGGTGCTTCAAGTTAGTACCAGTTGAACCAGAGCAAGTAGAAGAGGCCAA
TGAAGGAGAGAACAACAGCTTGTACACCCTATGAGCCAGCATGGGATGGAGGACCCGGAGGGAGAAGTA
TTAGTGTGGAAGTTTGACAGCCTCCTAGCATTTTCGTCACATGGCCCGAGAGCTGCAT **CCGGAGTACTACA**
AAGACTGCTG ACATCGAGCTTTCTACAAGGGACTTTCCGCTGGGGACTTTCCAGGGAGGTGTGGCCTGGG
CGGGACTGGGGAGTGGCGAGCC **CTCAGATGCTACATATAAGCAGC**

+SpCas9/gRNA LTR A+LTR B:

GATCTGTGGATCTACCACACACA AGGCTACTTCCCTGATTGGCAGA ACTACACACCAGGGCCAGGG **ATCA**
GATATCCACTGAC

Edited fragment: 190bp

↓ **CTT** **TGG** ATGGTGCTTCAAGTTAGTACCAGTTGAACCAGAGCAAGTAGAAGAGGCCAATGAAGGAGAGAA
CAACAGCTTGTACACCCTATGAGCCAGCATGGGATGGAGGACCCGGAGGGAGAAGTATTAGTGTGGAAG
TTTGACAGCCTCCTAGCATTTTCGTCACATGGCCCGAGAGCTGCAT **CCG** **GAG** ↓
TACTACAAAGACTGCTG ACATCGAGCTTTCTACAAGGGACTTTCCGCTGGGGACTTTCCAGGGAGGTGTG
GCCTGGGCGGGACTGGGGAGTGGCGAGCC **CTCAGATGCTACATATAAGCAGC**

PCR product truncated LTR -413/-19: 205bp

GATCTGTGGATCTACCACACACA AGGCTACTTCCCTGATTGGCAGA ACTACACACCAGGGCCAGGG **ATCA**
GATATCCACTGAC **T** **TACTACAAAGACTGCTG** ACATCGAGCTTTCTACAAGGGACTTTCCGCTGGGGACTTT
CCAGGGAGGTGTGGCCTGGGCGGGACTGGGGAGTGGCGAGCC **CTCAGATGCTACATATAAGCAGC**

Figure S2

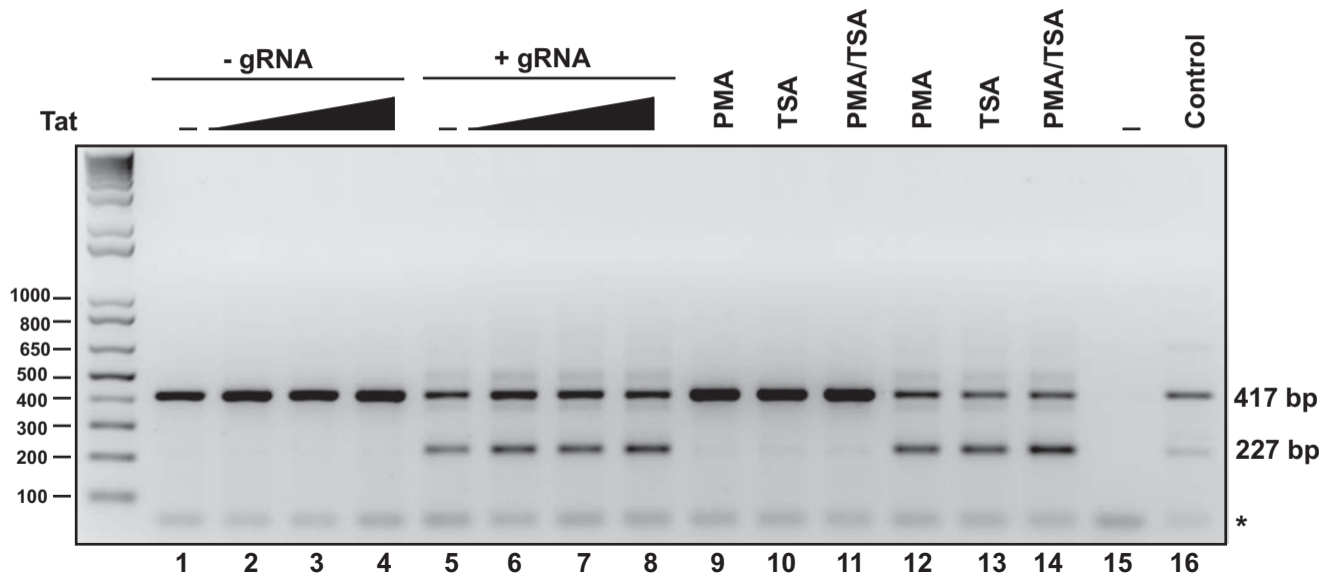


Figure S3

Jurkat 2D10

Sequence:

TGGAAGGGCTAATTTGGTCCCAAAAAAGACAAGAGATCCTTGATCTGTGGATCTACCACACACAAGGCTA
F [-375/-352] LTR A [-347/-328] PAM
CTTCCCTGATTGGCAGAACTACACACCAGGGCCAGGGATCAGATATCCACTGACCTTTGGATGGTGCTTC
AAGTTAGTACCAGTTGAACCAGAGCAAGTAGAAGAGGCCAATGAAGGAGAGAACAACAGCTTGTACACC
CTATGAGCCAGCATGGGATGGAGGACCCGGAGGGAGAAGTATTAGTGTGGAAGTTGACAGCCTCCTAGC
PAM LTR B [-143/-124]
ATTTTCGTCACATGGCCCGAGAGCTGCATCCGGAGTACTACAAAGACTGCTGACATCGAGCTTTCTACAAG
GGACTTTCCGCTGGGGACTTTCCAGGGAGGTGTGGCCTGGGCGGGACTGGGGAGTGCCGAGCCCTCAGAT
└─Transcription start[+1] R [+19/+43]
GCTACATATAAGCAGCTGCTTTTTGCCTGTACTGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGC
TCTCTGGCTAACTAGGGAACCCACTGCTTAAGCCTCAATAAAGCTTGCCTTGAGTGCTCAAAGTAGTGTG

PCR product full length LTR -375/+43: 417bp

TTGGCAGAACTACACACCAGGGCCAGGGATCAGATATCCACTGACCTTTGGATGGTGCTTCAAGTTAGTA
CCAGTTGAACCAGAGCAAGTAGAAGAGGCCAATGAAGGAGAGAACAACAGCTTGTACACCCTATGAGCC
AGCATGGGATGGAGGACCCGGAGGGAGAAGTATTAGTGTGGAAGTTGACAGCCTCCTAGCATTTTCGTCA
CATGGCCCGAGAGCTGCATCCGGAGTACTACAAAGACTGCTGACATCGAGCTTTCTACAAGGGACTTTCC
GCTGGGGACTTTCCAGGGAGGTGTGGCCTGGGCGGGACTGGGGAGTGCCGAGCCCTCAGATGCTACATAT
AAGCAGCTGCTTTTTGCCTGTACTGGTCTCTCTGGTTAGACCAGATCTGAGCCTGGGAGCTCTCTGG

+SpCas9/gRNA LTR A+LTR B:

TTGGCAGAACTACACACCAGGGCCAGGGATCAGATATCCACTGAC

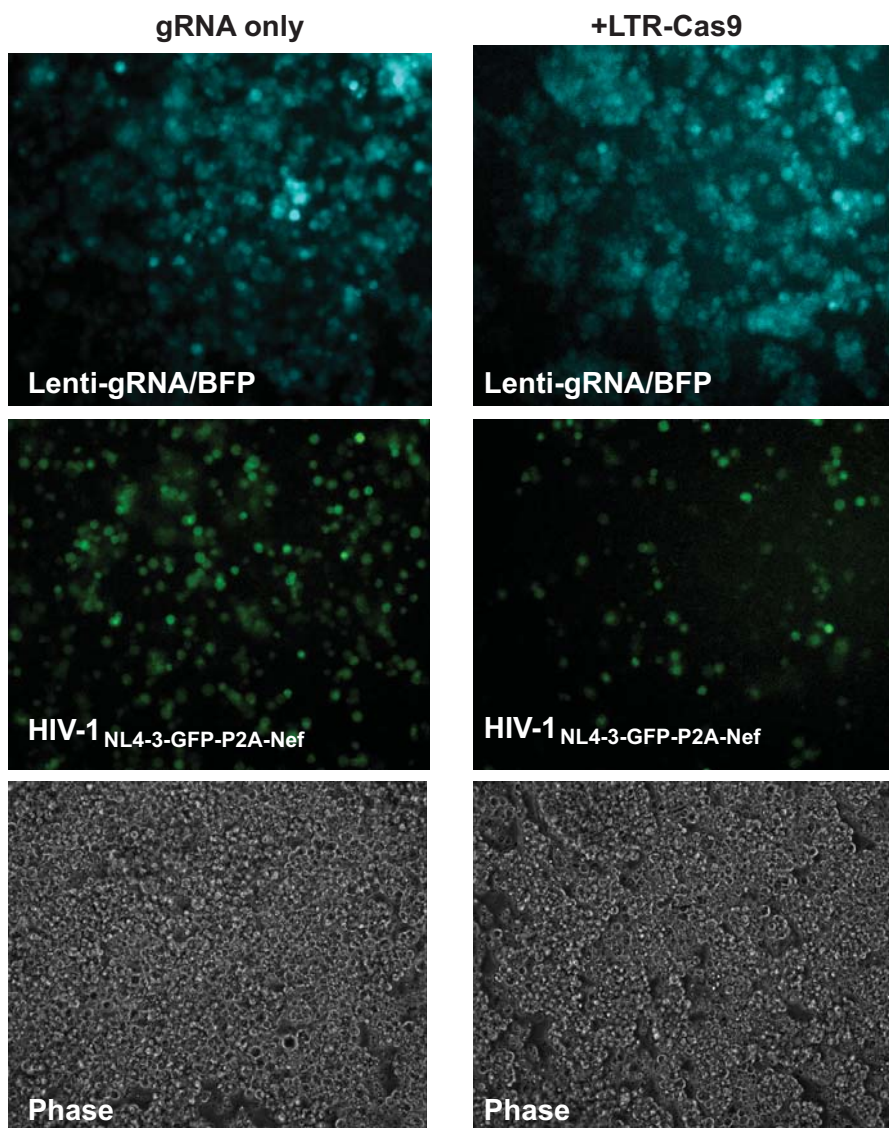
Edited fragment: 190bp

↓ CTTGGATGGTGCTTCAAGTTAGTACCAGTTGAACCAGAGCAAGTAGAAGAGGCCAATGAAGGAGAGAA
CAACAGCTTGTACACCCTATGAGCCAGCATGGGATGGAGGACCCGGAGGGAGAAGTATTAGTGTGGAAG
TTTACAGCCTCCTAGCATTTTCGTACATGGCCCGAGAGCTGCATCCGGAG↓

PCR product truncated LTR -375/-43: 227bp

TTGGCAGAACTACACACCAGGGCCAGGGATCAGATATCCACTGACTACTACAAAGACTGCTGACATCGAG
CTTTCTACAAGGGACTTTCCGCTGGGGACTTTCCAGGGAGGTGTGGCCTGGGCGGGACTGGGGAGTGCCG
AGCCCTCAGATGCTACATATAAGCAGCTGCTTTTTGCCTGTACTGGTCTCTCTGGTTAGACCAGATCTG
AGCCTGGGAGCTCTCTGG

a



b

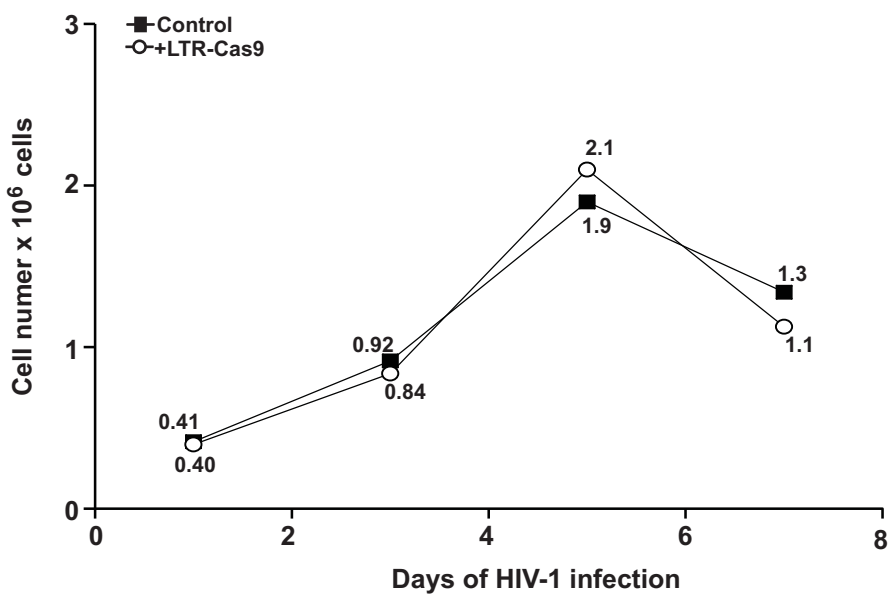
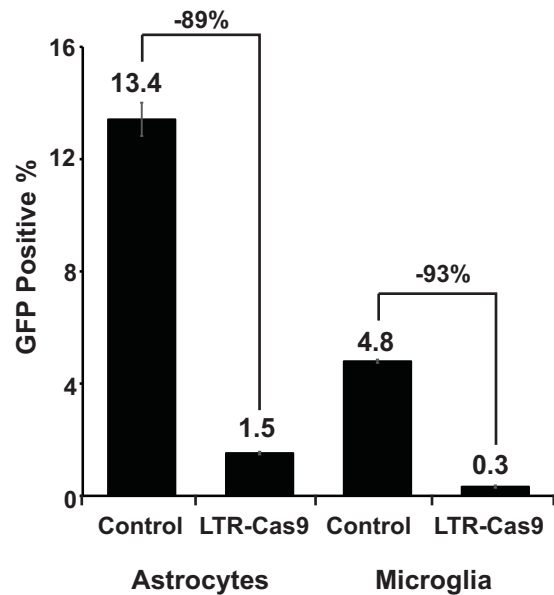
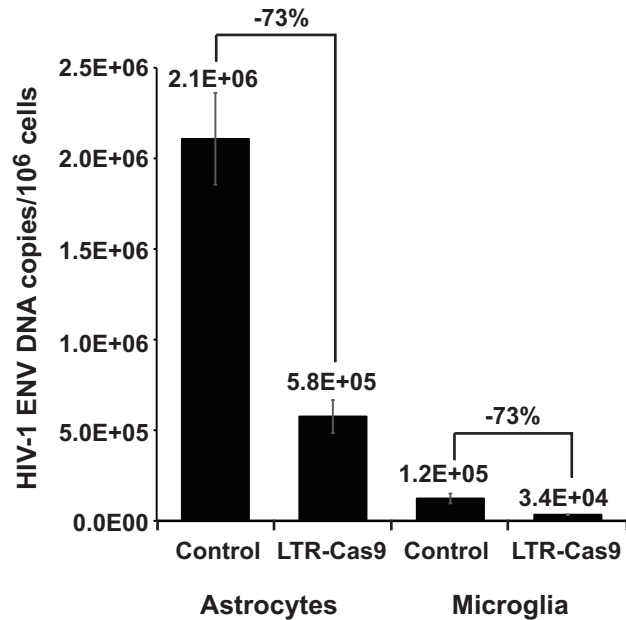


Figure S5

a**b****c**