

Supplementary data:

Suppl. Figure 1: Spontaneous motor activity of DM at ages of 3 to 18 months.

Documentation of spontaneous open-field behaviour as the most sensitive and reliable measure of abnormal movements in PD mouse models revealed highly significant marked reductions of horizontal activity, vertical activity, and stereotype movements for the DM mice relative to their appropriate WT controls, already at the age of 3 months (columns at the left side). While this early onset low exploration activity of DM animals remained quite constant across their lifespan, the corresponding F1-hybrid controls showed later age-progressive reduction in spontaneous motor activity.

Suppl. Figure 2: pSer129-SNCA immunohistochemistry of DM mice at ages 15-17 months.

Bright field immunohistochemical stainings of pSer129-SNCA-immunoreactivity were performed in old DM mice with paralysis (A-C) or without paralysis (D/E). In paralytic DM animals, (A) thoracic / cervical levels of the spinal cord showed milder stages of inclusion formation (black arrows), (B) motor cortex showed discrete aggregation pathology (black arrows), while (C) striatum remained free of detectable pathology. In non-paralytic old DM mice, (D) the lumbar spinal cord showed mild inclusion pathology with only few immunopositive neurites (black arrow), and (E) the ventral tegmental area of the midbrain also showed only discrete aggregation (black arrow).

Suppl. Figure 3: Immunohistochemistry of *Pink1*^{-/-} mice at ages 15-17 months.

Bright field immunohistochemical stainings were performed in age-matched *Pink1*^{-/-} spinal cord as a control for Figure 4 spinal cord (upper panels) and for Figure 5 midbrain (lower panels). Protein aggregation was not detected by antibodies against pSer129-SNCA, p62 or ubiquitin.

Suppl. Figure 4: Double immunofluorescence in DM midbrain beyond age 1 year demonstrates aggregates in non-dopaminergic cells.

The colocalization with TH as a marker of midbrain dopaminergic (DAergic) neurons in the substantia nigra pars compacta (SNc) and the ventral tegmental area (VTA) was assessed. The 4B12 antibody detects human

alpha-synuclein selectively and demonstrates transgenic overexpression with nuclear staining throughout the midbrain DAergic neurons (A-C, white arrows). However, the pSer129-SNCA staining shows the protein aggregation to occur preferentially in proximity rather than in DAergic SN / VTA neurons (D-F). The p62 staining shows also protein aggregates again in neurons adjacent to DAergic cells (G-I). Similarly, ubiquitin staining in spite of high background immunoreactivity localizes the protein aggregates mostly outside the DAergic neurons (J-L). In low magnifications of all three stainings, the aggregates formed in a band of neurons and neurites just dorsal to the SN / VTA line of DAergic nerve cells.

Suppl. Figure 5: Double immunofluorescence in DM midbrain beyond age 1 year demonstrates aggregates to occur not in astrocytes, but in neurons.

The pSer129-SNCA staining in the VTA of aged DM mice was assessed by double immunofluorescence for colocalization with markers of glial and neuronal differentiation. GFAP (glial fibrillary acidic protein) as a marker of astrocytes clearly was not present in the cells affected by aggregates (A-C), while NeuN (neuronal nuclear antigen Rbfox3) as a general marker of neurons did colocalize with these cells (D-F, white arrows). Parvalbumin staining as a marker of interneurons produced immunoreactivity in multiple affected cells (G-I, white arrows), while GAD65 (glutamate decarboxylase encoded by *GAD2*) as a marker of GABAergic neurons were not present in the cells with aggregates (J-L), and VGLUT2 (vesicular glutamate transporter 2) as a marker of subcortical glutamatergic neurons was observed only in individual aggregate-containing cells (M-O, white arrows).

Suppl. Figure 6: Global transcriptome GSEA downregulation for DM cerebellum at age 6 weeks.

Details for the NOUSHMEHR_GBM_SILENCED_BY METHYLATION gene set.

Suppl. Figure 7: Global transcriptome GSEA upregulation for DM cerebellum at age 6 weeks. Details for the NAGASHIMA_EGF_SIGNALING_UP gene set.

Suppl. Figure 8: Global transcriptome GSEA upregulation for DM cerebellum at age 6 months.

Details for the NAGASHIMA_EGF_SIGNALING_UP gene set.

Suppl. Figure 9: Global transcriptome GSEA downregulation for DM cerebellum at age 6 months.

Details for the MITOCHONDRIAL_MATRIX gene set.

Suppl. Figure 10: Global transcriptome GSEA downregulation for DM midbrain at age 6 months.

Details for the QI_HYPOXIA_TARGETS_OF_HIF1A_AND_FOXA2 gene set.

Suppl. Figure 11: Global transcriptome GSEA upregulation for DM midbrain at age 6 months. Details for the PID_ATM_PATHWAY gene set.

Suppl. Movie 1: Progressive paralysis of DM mice beyond age 1 year.

The progressive phenotype of locomotor deficits was documented by video for several DM mice at ages beyond 1 year, before they had to be sacrificed to avoid animal suffering. Later their tissue was found to exhibit alpha-synuclein- / p62- / ubiquitin- positive aggregates in immunohistochemistry.

Suppl. Table 1: Global ubiquitination changes of DM brain hemispheres at age 18 months.

(A) Primary UbiScan® results are represented. The average fold change is provided in column “-/- TgA : Control”, while the fold change for each of the 3 DM:WT-F1-hybrid pairs is shown in the subsequent columns. Dysregulation of ubiquitination sites with increase above 2.0-fold were illustrated in green, while sites with decrease below -2.0-fold were highlighted in red (see a list in Excel Tab: Fold Change by Protein Type). Fold changes should be more accurate with higher maximal intensity values. Peak measurements reflect either area or height. An index number from the Details tab is used for easy lookup of peptides between tabs. Blue letters in Gene Names indicate siRNA products commercially available; in Protein Names they indicate site-specific antibodies commercially available. Within sequences in the peptide column, the ubiquitination site is indicated by *, while oxidized methionine is indicated by #. Count in Details refers to the number of MS/MS identifications in the Details tab for each protein/site. Average RT refers to average retention time for each peptide. In the Raw Intensity column, bold letters indicate intensities manually reviewed, while red letters indicate multiple identifications for 1 m/z measurement due

to ambiguous methylation site localization. The Normalized Log 2 Ratios were calculated by subtracting the median Log2 fold change for each comparison. The Details tab contains the MS/MS (blue box) and MS (red box) scan numbers, the Retention Time, the Xcorr values, the measured peptide mass accuracy, the DeltaCN value, the Rsp value, and the Peptide Prophet Probability (PP Probability) as a reference and measure of confidence to each peptide assignment. (B) Bioinformatics information is provided on the UbiScan® primary data.

Suppl. Table 2: Global transcriptome changes of DM brain hemispheres at ages 6 weeks and 6 months.

The columns from left show the microarray spot identifier, the transcript symbol, the transcript name, the average expression level, the adjusted p-value and then significant down-regulations (-1, green) and up-regulations (+1, red) for different organs and ages: This decision matrix assesses cerebellum (Cbll), midbrain+brainstem (MidB), striatum (Str), at the ages 6 weeks and 6 months, comparing in each column 4 DM versus 4 WT-F1-hybrid tissues, with a cutoff value of at least logarithmic fold changes of ± 0.6 .

Suppl. Table 3: Global transcriptome bioinformatics by Gene Set Enrichment Analysis.

The automatized output from the GSEA software for each of the three brain tissues (midbrain/brainstem = MB, striatum = ST, cerebellum = CB) at two ages (6 months = 6M, 6 weeks = 6W), separated for upregulation and for downregulation, is shown in twelve datasheets of this file, in parameters defined by the Guide To Interpret Results at the GSEA internet page.

Fig S1

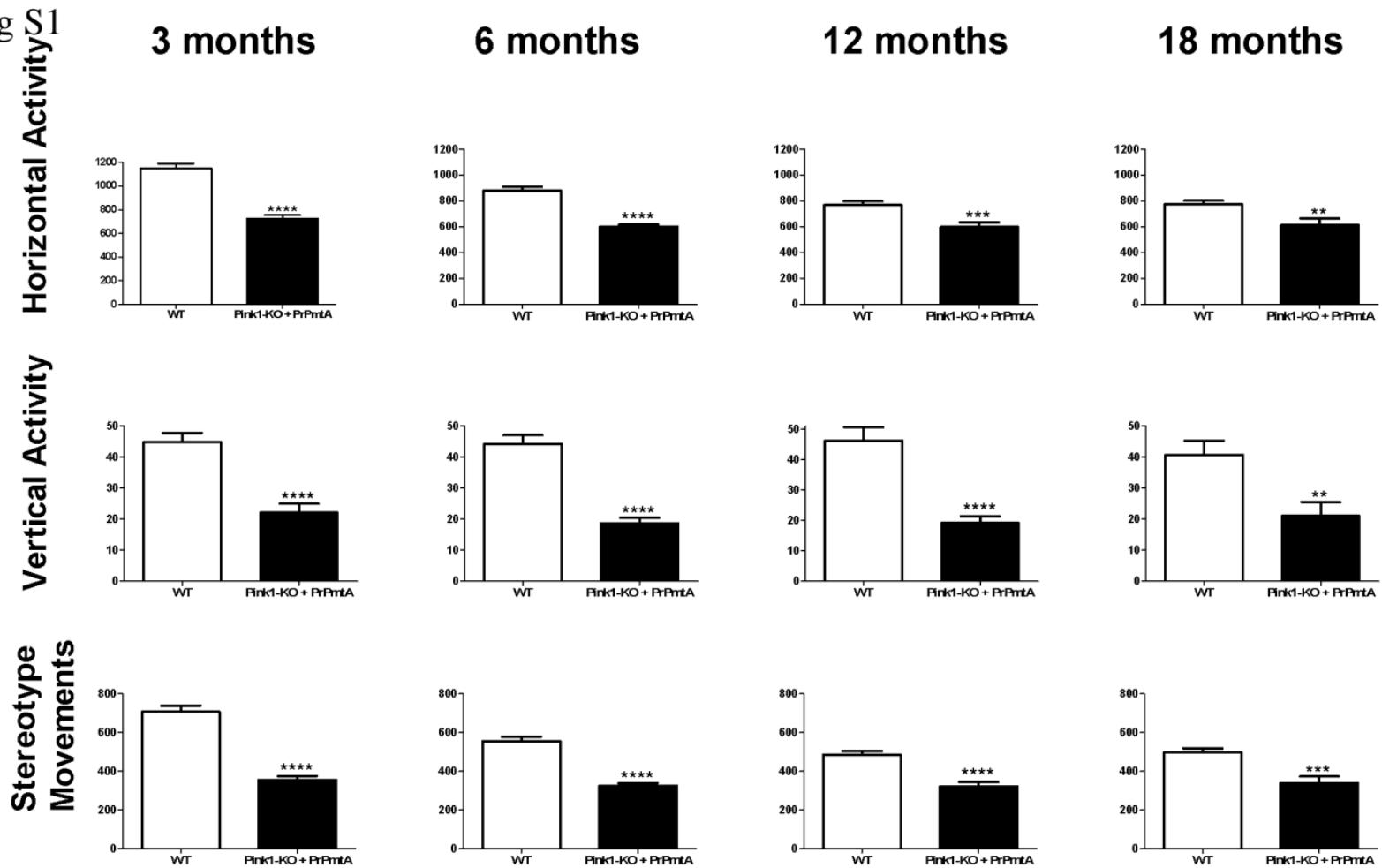


Fig S2

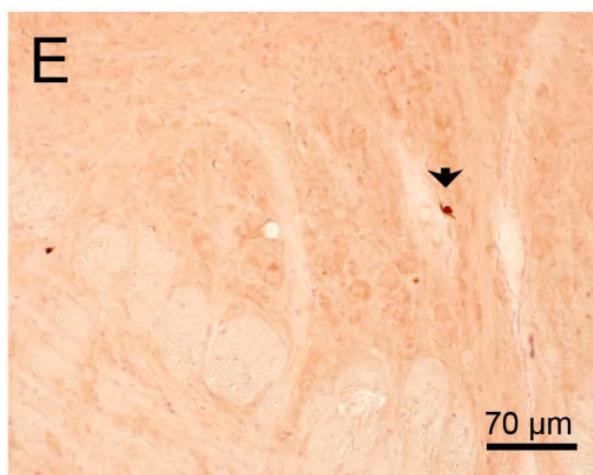
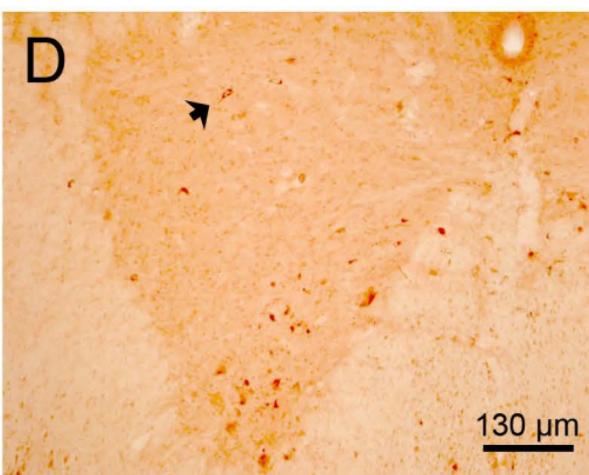
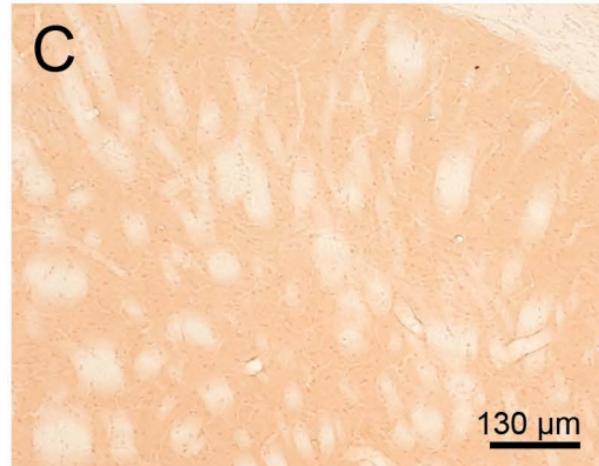
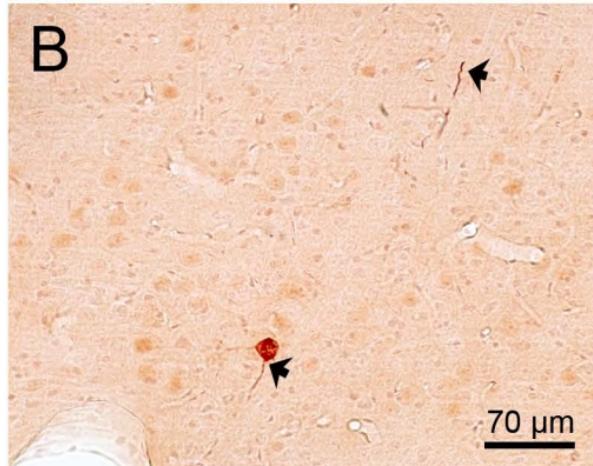
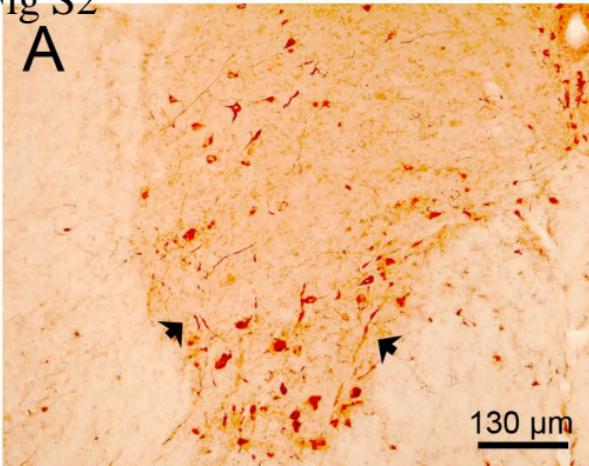
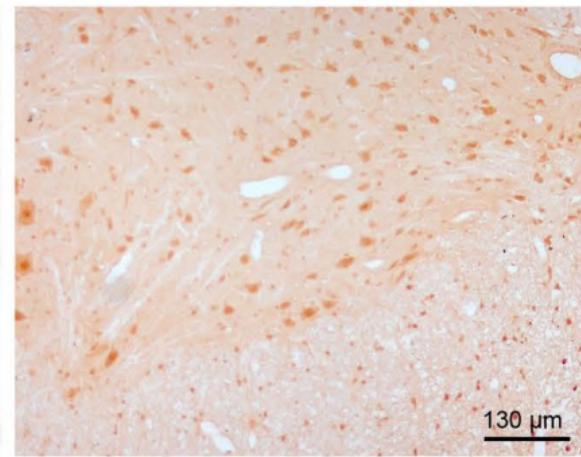
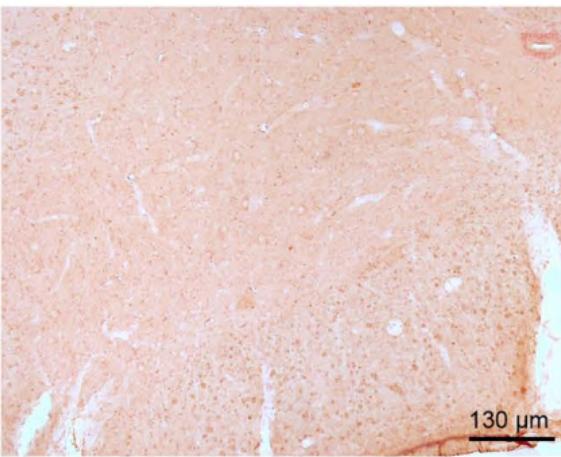
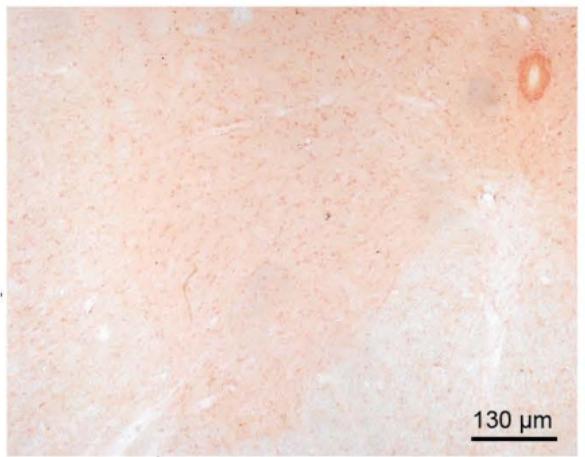


Fig S3 Phospho Ser129 SNCA

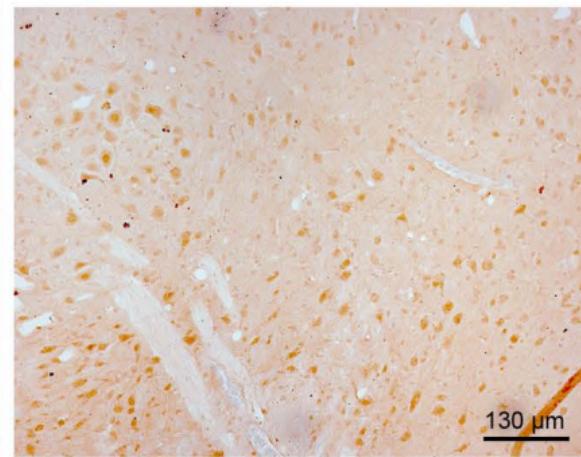
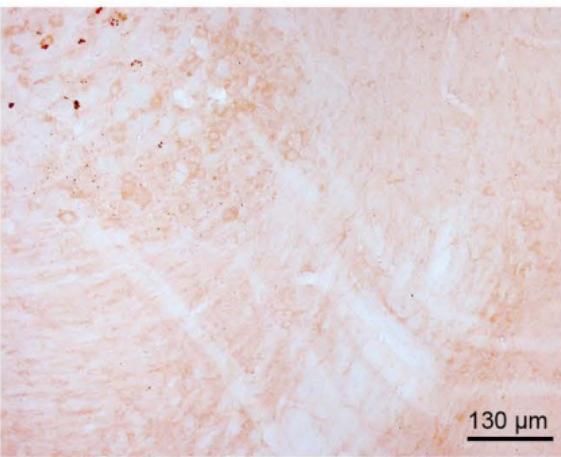
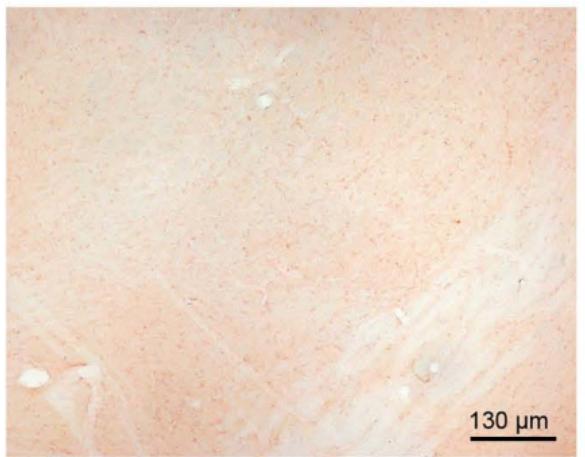
p62/SQSTM1

Ubiquitin

Spinal cord

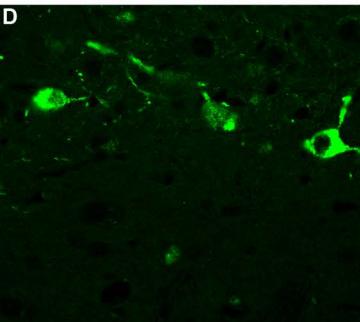


Midbrain



A Fig S4

4B12



B

TH



C

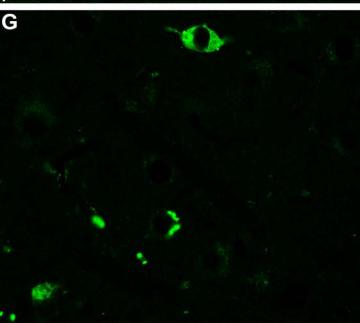
Merge



20 μ m

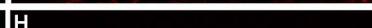
D

pSer129-SNCA



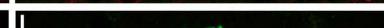
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TH



F

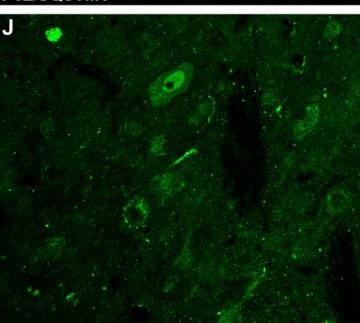
Merge



20 μ m

G

P62/SQSTM1



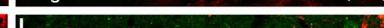
H

TH



I

Merge



20 μ m

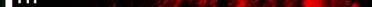
J

Ubiquitin



K

TH



L

Merge



20 μ m

A Fig S5

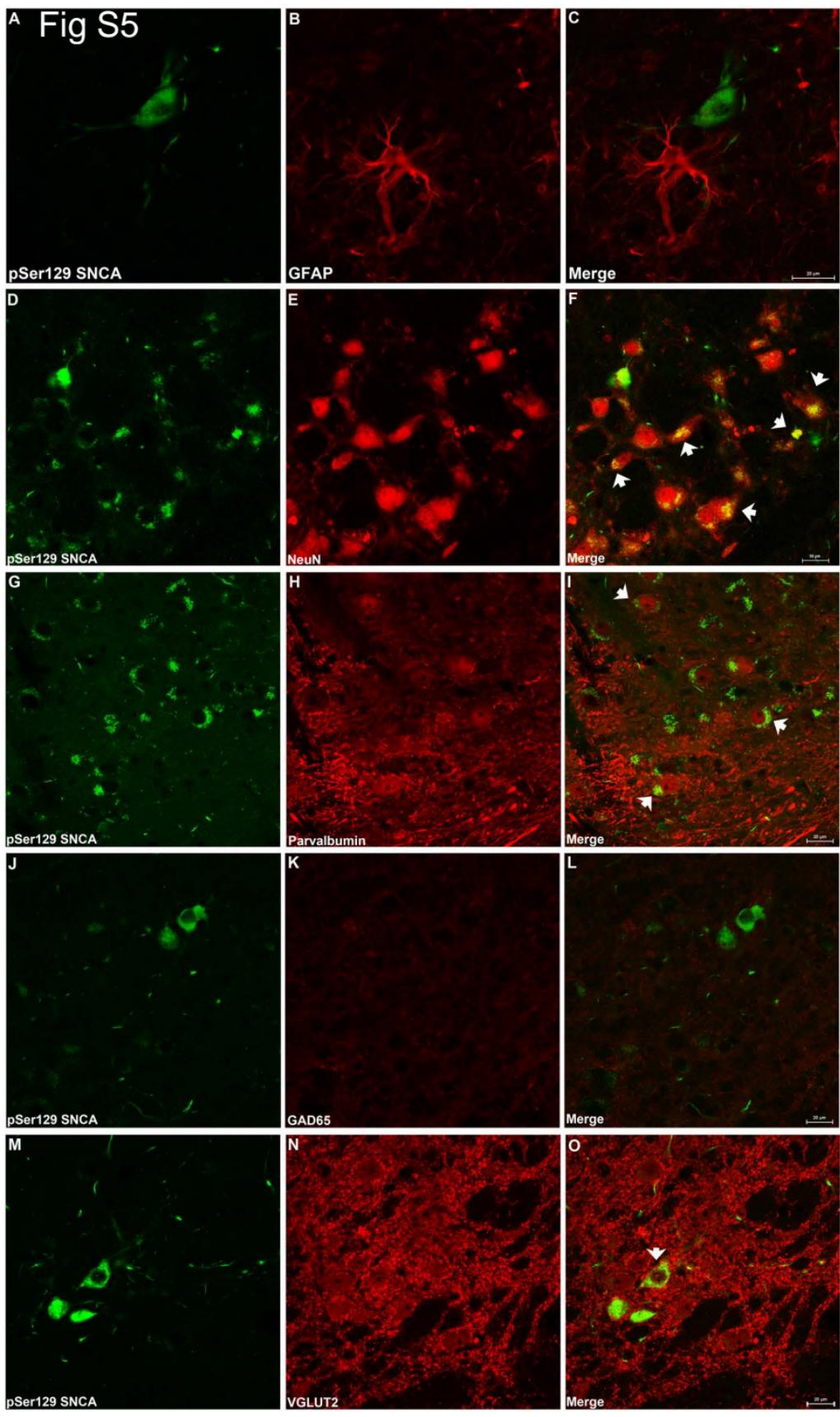


Fig S6

Table: GSEA Results Summary	
Dataset	CB_6W_collapsed_to_symbols.general.cls#mut_versus_WT
Phenotype	general.cls#mut_versus_WT
Upregulated in class	WT
GeneSet	NOUSHMEHR_GBM_SILENCED_BY METHYLATION
Enrichment Score (ES)	-0.6654536
Normalized Enrichment Score (NES)	-2.131186
Nominal p-value	0.0
FDR q-value	0.016506651
FWER p-Value	0.034

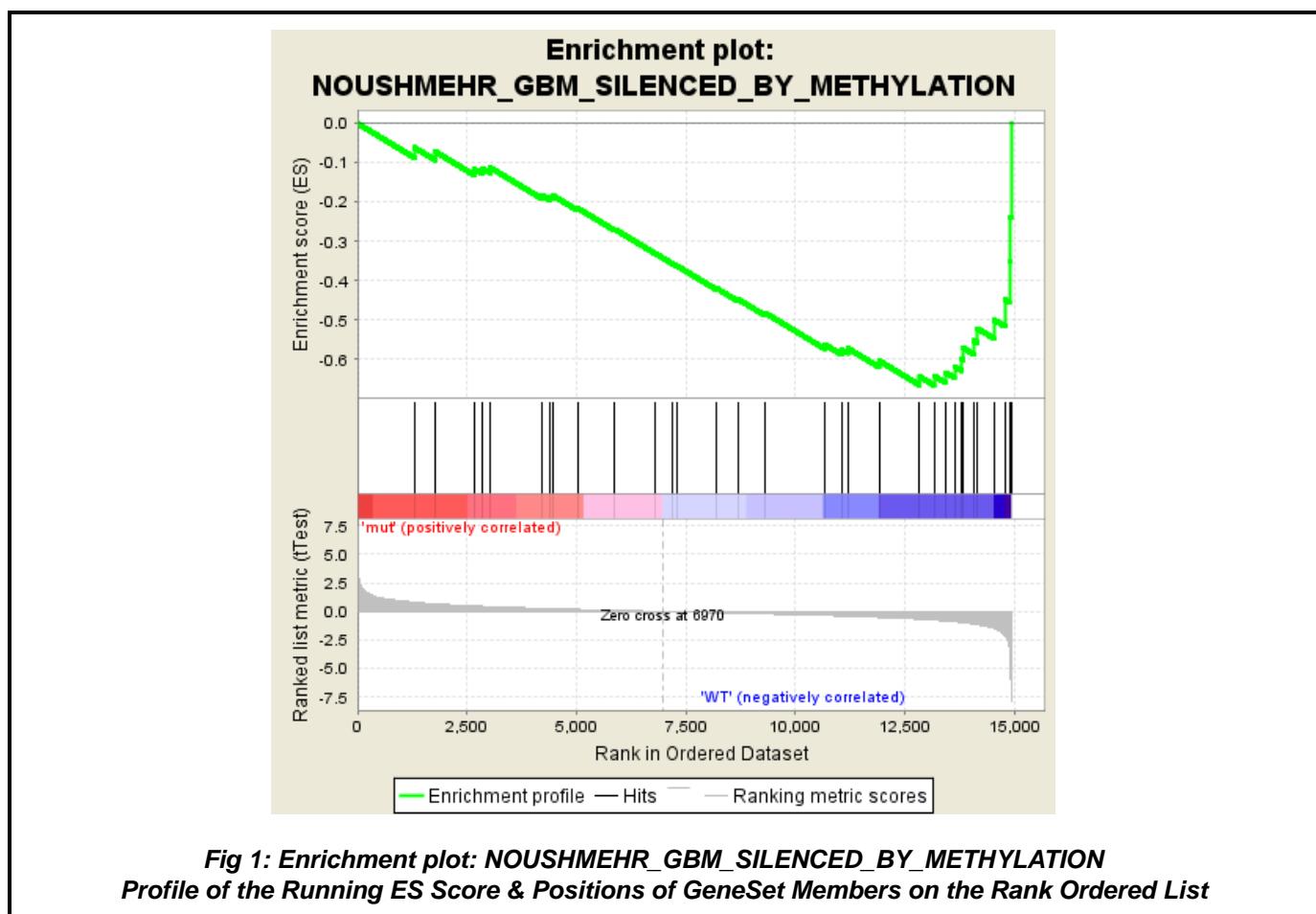


Table: GSEA details [plain text format]							
	PROBE	GENE SYMBOL	GENE_TITLE	RANK IN GENE LIST	RANK METRIC SCORE	RUNNING ES	CORE ENRICHMENT
1	FLNC	FLNC Entrez , Source	filamin C, gamma (actin binding protein 280)	1296	0.809	-0.0611	No
2	LDHA	LDHA Entrez , Source	lactate dehydrogenase A	1780	0.665	-0.0722	No
3	EPHX2	EPHX2 Entrez , Source	epoxide hydrolase 2, cytoplasmic	2676	0.482	-0.1168	No

4	TOM1L1 Entrez , Source	TOM1L1 target of myb1-like 1 (chicken)	2843	0.457	-0.1133	No
5	SEMA3E Entrez , Source	SEMA3E sema domain, immunoglobulin domain (Ig), short basic domain, secreted, (semaphorin) 3E	3037	0.427	-0.1126	No
6	FABP5 Entrez , Source	FABP5 fatty acid binding protein 5 (psoriasis-associated)	4221	0.271	-0.1833	No
7	CBR1 Entrez , Source	CBR1 carbonyl reductase 1	4389	0.253	-0.1864	No
8	CA3 Entrez , Source	CA3 carbonic anhydrase III, muscle specific	4473	0.245	-0.1841	No
9	OCIAD2 Entrez , Source	OCIAD2 OCIA domain containing 2	5023	0.187	-0.2150	No
10	TRIP4 Entrez , Source	TRIP4 thyroid hormone receptor interactor 4	5877	0.100	-0.2690	No
11	G0S2 Entrez , Source	G0S2 G0/G1switch 2	6786	0.017	-0.3294	No
12	NMNAT3 Entrez , Source	NMNAT3 nicotinamide nucleotide adenylyltransferase 3	7199	-0.020	-0.3564	No
13	MMP9 Entrez , Source	MMP9 matrix metalloproteinase 9 (gelatinase B, 92kDa gelatinase, 92kDa type IV collagenase)	7309	-0.032	-0.3627	No
14	LGALS8 Entrez , Source	LGALS8 lectin, galactoside-binding, soluble, 8 (galectin 8)	8196	-0.111	-0.4187	No
15	CHI3L1 Entrez , Source	CHI3L1 chitinase 3-like 1 (cartilage glycoprotein-39)	8699	-0.156	-0.4474	No
16	CHRDL2 Entrez , Source	CHRDL2 chordin-like 2	9310	-0.211	-0.4815	No
17	RARRES1 Entrez , Source	RARRES1 retinoic acid receptor responder (tazarotene induced) 1	10670	-0.348	-0.5616	No
18	SLC25A20 Entrez , Source	SLC25A20 solute carrier family 25 (carnitine/acylcarnitine translocase), member 20	11064	-0.391	-0.5755	No
19	B3GNT5 Entrez , Source	B3GNT5 UDP-GlcNAc:betaGal beta-1,3-N-acetylglucosaminyltransferase 5	11199	-0.407	-0.5714	No
20	FKBP5 Entrez , Source	FKBP5 FK506 binding protein 5	11914	-0.508	-0.6030	No
21	SLC39A12 Entrez , Source	SLC39A12 solute carrier family 39 (zinc transporter), member 12	12843	-0.677	-0.6436	Yes
22	RBP1 Entrez , Source	RBP1 retinol binding protein 1, cellular	13169	-0.747	-0.6415	Yes
23	PDPN Entrez , Source	PDPN podoplanin	13435	-0.822	-0.6330	Yes
24	FMOD Entrez , Source	FMOD fibromodulin	13637	-0.882	-0.6182	Yes

25	BIRC3	BIRC3 Entrez , Source	baculoviral IAP repeat-containing 3	13793	-0.938	-0.5986	Yes
26	RARRES2	RARRES2 Entrez , Source	retinoic acid receptor responder (tazarotene induced) 2	13836	-0.961	-0.5706	Yes
27	F3	F3 Entrez , Source	coagulation factor III (thromboplastin, tissue factor)	14082	-1.086	-0.5522	Yes
28	GJB2	GJB2 Entrez , Source	gap junction protein, beta 2, 26kDa (connexin 26)	14161	-1.129	-0.5213	Yes
29	RAB36	RAB36 Entrez , Source	RAB36, member RAS oncogene family	14537	-1.478	-0.4991	Yes
30	LGALS3	LGALS3 Entrez , Source	lectin, galactoside-binding, soluble, 3 (galectin 3)	14797	-2.145	-0.4478	Yes
31	LECT1	LECT1 Entrez , Source	leukocyte cell derived chemotaxin 1	14893	-3.196	-0.3518	Yes
32	FZD6	FZD6 Entrez , Source	frizzled homolog 6 (Drosophila)	14903	-3.540	-0.2390	Yes
33	CTHRC1	CTHRC1 Entrez , Source	collagen triple helix repeat containing 1	14930	-7.518	0.0001	Yes

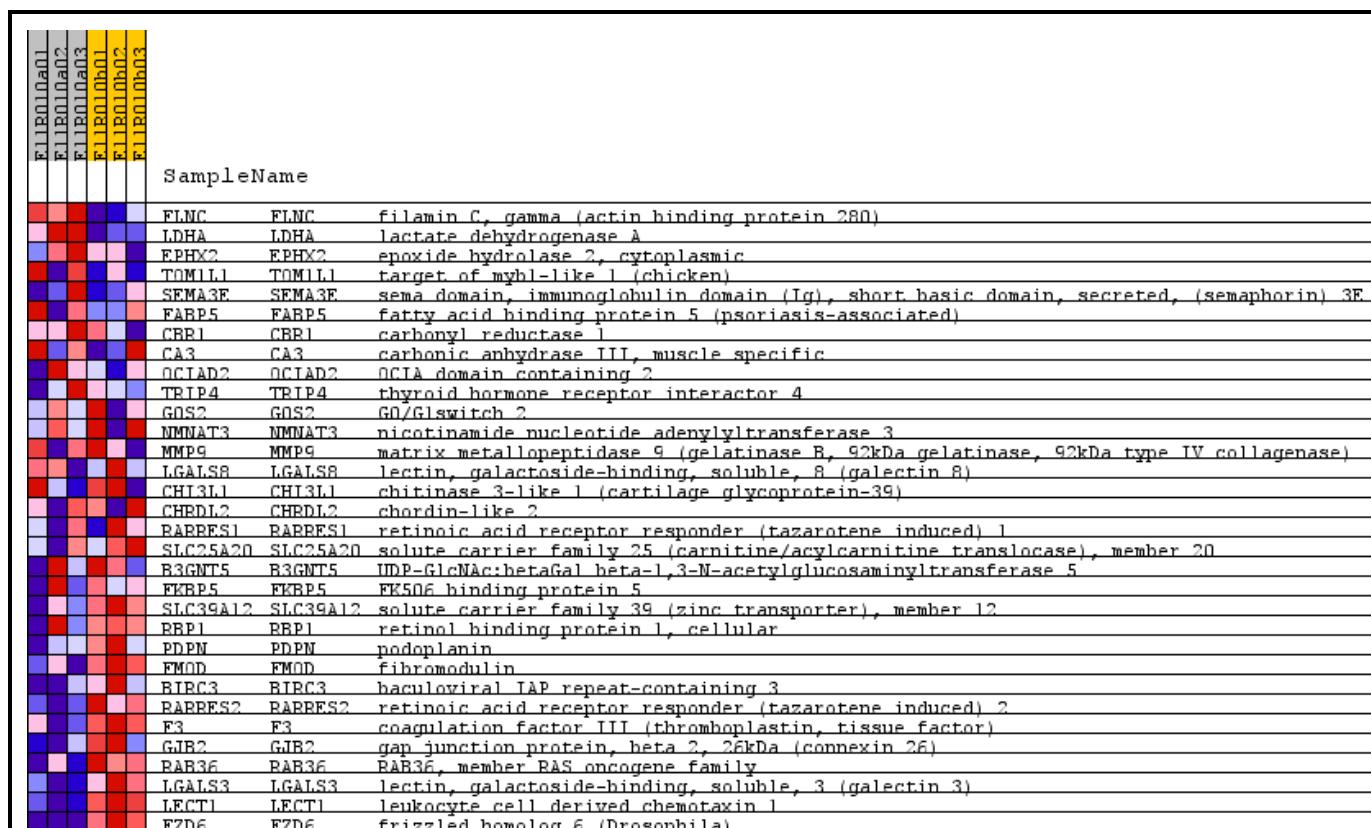


Fig 2: NOUSHMEHR_GBM_SILENCED_BY METHYLATION
Blue-Pink O' Gram in the Space of the Analyzed GeneSet

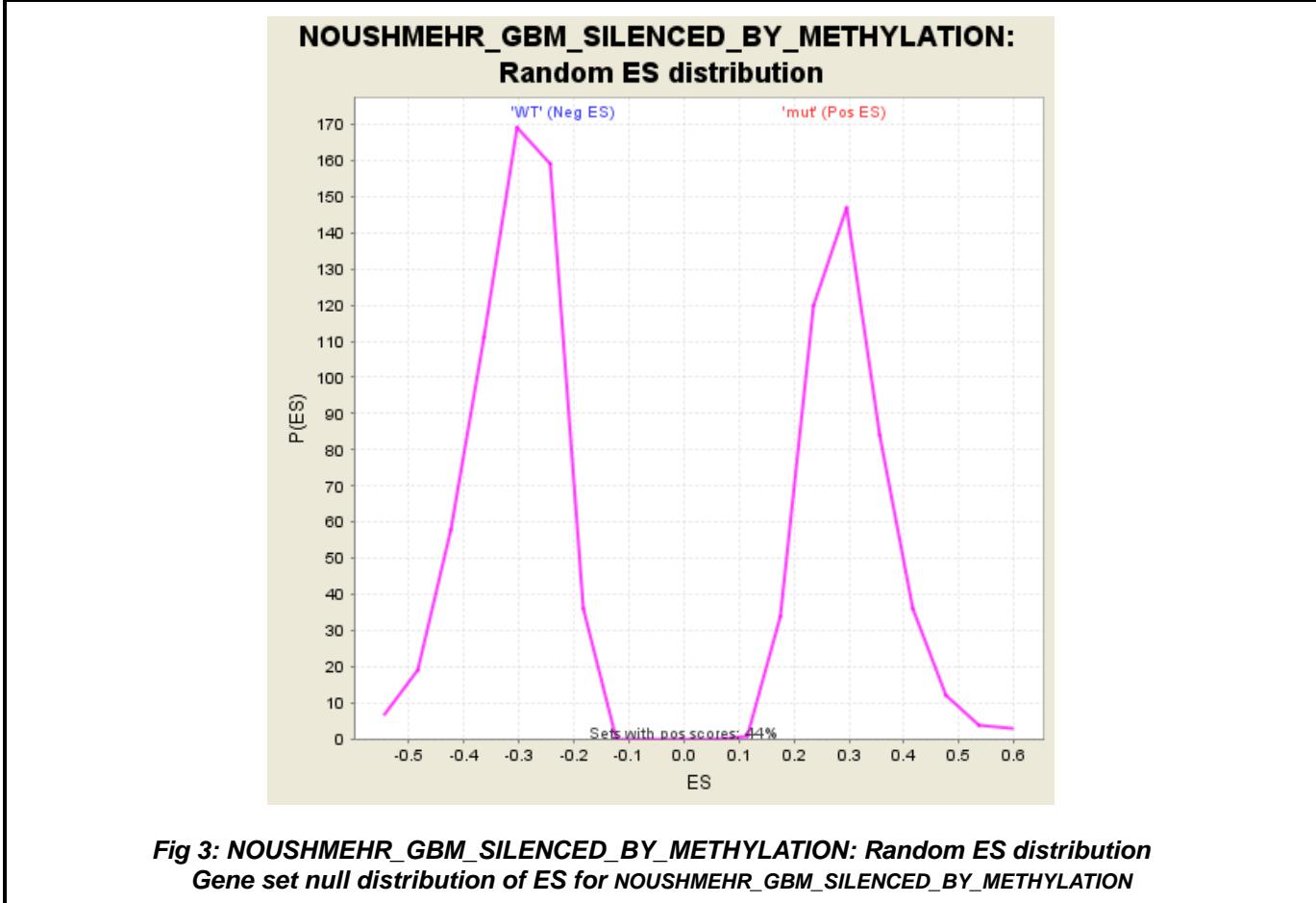


Fig S7

Table: GSEA Results Summary	
Dataset	CB_6W_collapsed_to_symbols.general.cls#mut_versus_WT
Phenotype	general.cls#mut_versus_WT
Upregulated in class	mut
GeneSet	NAGASHIMA_EGF_SIGNALING_UP
Enrichment Score (ES)	0.6003276
Normalized Enrichment Score (NES)	2.117522
Nominal p-value	0.0
FDR q-value	0.0585707
FWER p-Value	0.054

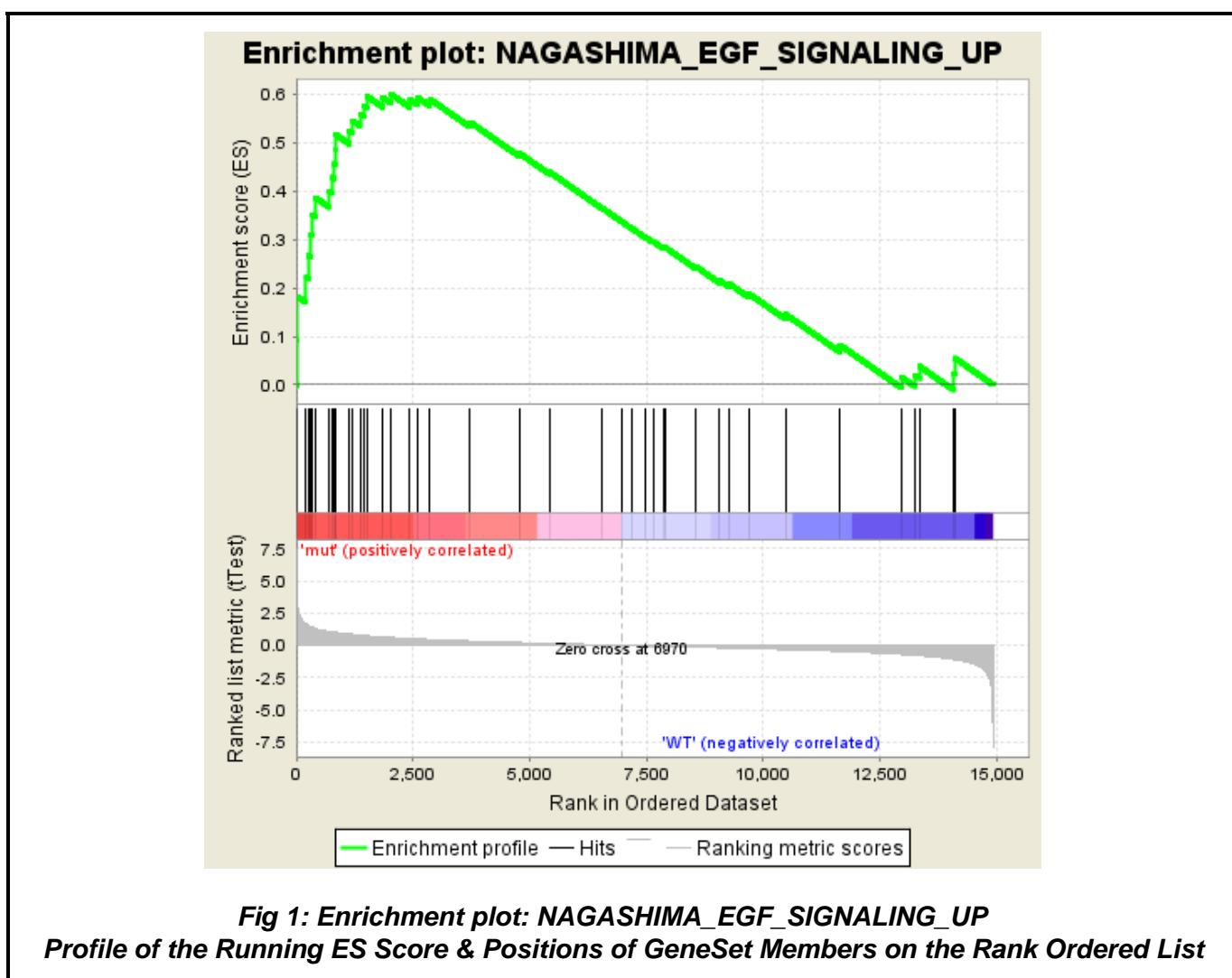


Table: GSEA details [plain text format]							
	PROBE	GENE SYMBOL	GENE_TITLE	RANK IN GENE LIST	RANK METRIC SCORE	RUNNING ES	CORE ENRICHMENT
1	NR4A1	NR4A1 Entrez , Source	nuclear receptor subfamily 4, group A, member 1	24	3.032	0.0923	Yes

2	JUNB	JUNB Entrez , Source	jun B proto-oncogene	28	2.903	0.1821	Yes
3	EGR4	EGR4 Entrez , Source	early growth response 4	211	1.682	0.2220	Yes
4	IER3	IER3 Entrez , Source	immediate early response 3	256	1.566	0.2676	Yes
5	FOS	FOS Entrez , Source	v-fos FBJ murine osteosarcoma viral oncogene homolog	300	1.459	0.3099	Yes
6	CTGF	CTGF Entrez , Source	connective tissue growth factor	328	1.415	0.3519	Yes
7	EGR1	EGR1 Entrez , Source	early growth response 1	412	1.296	0.3865	Yes
8	PHLDA1	PHLDA1 Entrez , Source	pleckstrin homology-like domain, family A, member 1	709	1.069	0.3998	Yes
9	HBEGF	HBEGF Entrez , Source	heparin-binding EGF-like growth factor	756	1.044	0.4291	Yes
10	NAB2	NAB2 Entrez , Source	NGFI-A binding protein 2 (EGR1 binding protein 2)	813	1.015	0.4568	Yes
11	ID3	ID3 Entrez , Source	inhibitor of DNA binding 3, dominant negative helix- loop-helix protein	835	1.005	0.4865	Yes
12	KLF10	KLF10 Entrez , Source	Kruppel-like factor 10	839	1.002	0.5173	Yes
13	DUSP5	DUSP5 Entrez , Source	dual specificity phosphatase 5	1136	0.875	0.5246	Yes
14	NR4A2	NR4A2 Entrez , Source	nuclear receptor subfamily 4, group A, member 2	1210	0.845	0.5459	Yes
15	DUSP1	DUSP1 Entrez , Source	dual specificity phosphatase 1	1362	0.784	0.5600	Yes
16	ARC	ARC Entrez , Source	activity-regulated cytoskeleton- associated protein	1455	0.755	0.5772	Yes
17	GEM	GEM Entrez , Source	GTP binding protein overexpressed in skeletal muscle	1510	0.739	0.5965	Yes
18	HES1	HES1 Entrez , Source	hairy and enhancer of split 1, (<i>Drosophila</i>)	1852	0.647	0.5936	Yes
19	DUSP2	DUSP2 Entrez , Source	dual specificity phosphatase 2	2033	0.606	0.6003	Yes

20	TRIB1	TRIB1 Entrez , Source	tribbles homolog 1 (Drosophila)	2435	0.525	0.5897	No
21	EDN1	EDN1 Entrez , Source	endothelin 1	2611	0.492	0.5932	No
22	FOSB	FOSB Entrez , Source	FBJ murine osteosarcoma viral oncogene homolog B	2867	0.454	0.5901	No
23	TNFRSF12A	TNFRSF12A Entrez , Source	tumor necrosis factor receptor superfamily, member 12A	3720	0.329	0.5431	No
24	EGR2	EGR2 Entrez , Source	early growth response 2 (Krox-20 homolog, Drosophila)	4783	0.212	0.4783	No
25	LIF	LIF Entrez , Source	leukemia inhibitory factor (cholinergic differentiation factor)	5421	0.146	0.4401	No
26	MYC	MYC Entrez , Source	v-myc myelocytomatosis viral oncogene homolog (avian)	6559	0.039	0.3649	No
27	TIPARP	TIPARP Entrez , Source	TCDD-inducible poly(ADP-ribose) polymerase	6968	0.000	0.3375	No
28	DLX2	DLX2 Entrez , Source	distal-less homeobox 2	7206	-0.021	0.3222	No
29	MCL1	MCL1 Entrez , Source	myeloid cell leukemia sequence 1 (BCL2-related)	7478	-0.046	0.3055	No
30	MAFF	MAFF Entrez , Source	v-maf musculoaponeurotic fibrosarcoma oncogene homolog F (avian)	7644	-0.062	0.2963	No
31	ID1	ID1 Entrez , Source	inhibitor of DNA binding 1, dominant negative helix-loop-helix protein	7860	-0.081	0.2844	No
32	KLF6	KLF6 Entrez , Source	Kruppel-like factor 6	7906	-0.085	0.2840	No
33	IER2	IER2 Entrez , Source	immediate early response 2	8546	-0.141	0.2455	No
34	EPHA2	EPHA2 Entrez , Source	EPH receptor A2	9067	-0.189	0.2164	No
35	RYBP	RYBP Entrez , Source	RING1 and YY1 binding protein	9274	-0.207	0.2090	No
36	EGR3	EGR3 Entrez , Source	early growth response 3	9709	-0.248	0.1875	No
37	DNAJB1	DNAJB1 Entrez , Source	DnaJ (Hsp40) homolog, subfamily B, member 1	10485	-0.330	0.1457	No

38	SPRED2	SPRED2 Entrez , Source	sprouty-related, EVH1 domain containing 2	11647	-0.469	0.0823	No
39	ETS2	ETS2 Entrez , Source	v-ets erythroblastosis virus E26 oncogene homolog 2 (avian)	12973	-0.704	0.0151	No
40	NEDD9	NEDD9 Entrez , Source	neural precursor cell expressed, developmentally down-regulated 9	13252	-0.769	0.0202	No
41	ZFP36	ZFP36 Entrez , Source	zinc finger protein 36, C3H type, homolog (mouse)	13351	-0.797	0.0383	No
42	CYR61	CYR61 Entrez , Source	cysteine-rich, angiogenic inducer, 61	14080	-1.086	0.0231	No
43	JUN	JUN Entrez , Source	jun oncogene	14109	-1.099	0.0553	No

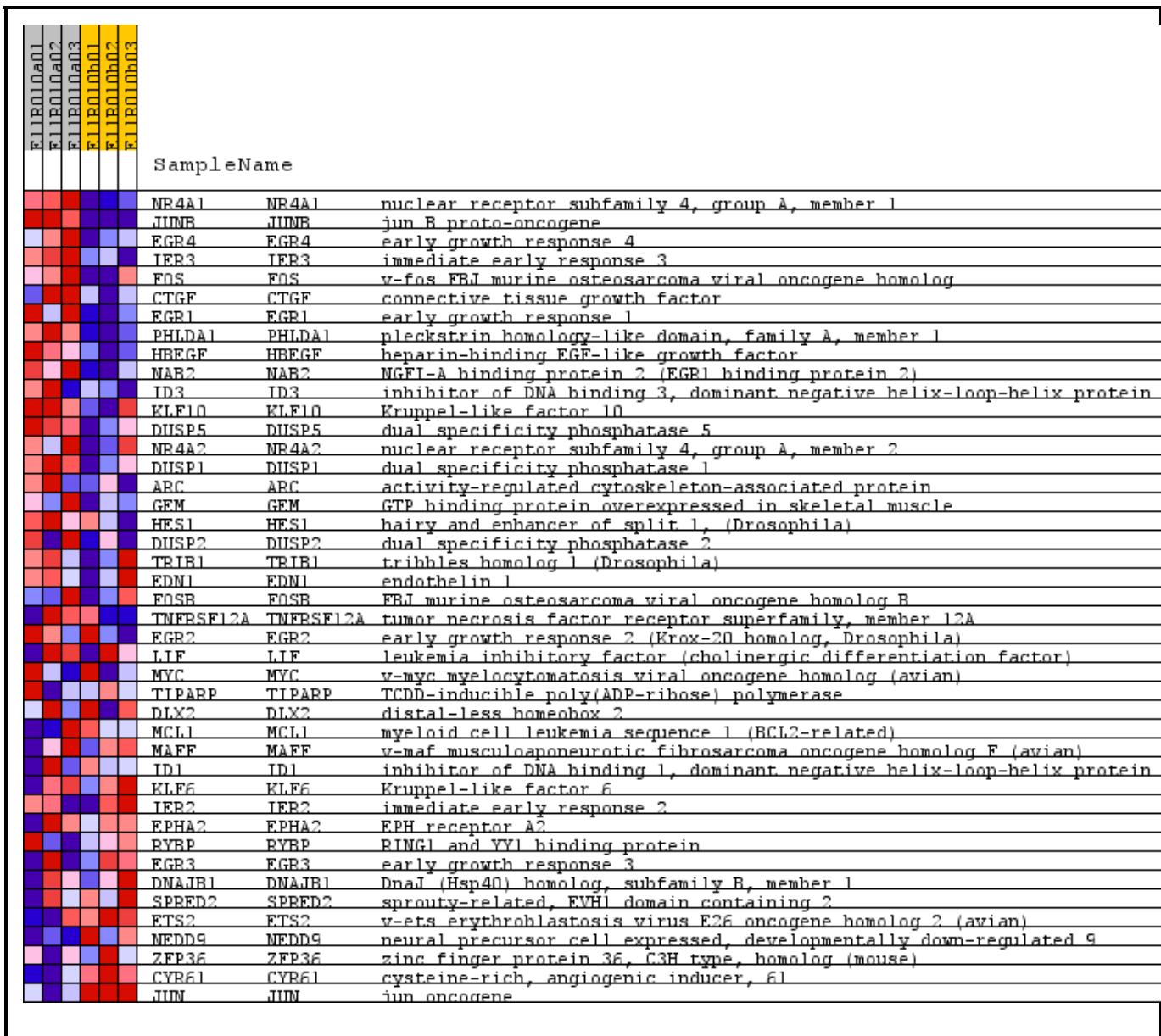


Fig 2: NAGASHIMA_EGF_SIGNALING_UP
Blue-Pink O' Gram in the Space of the Analyzed GeneSet

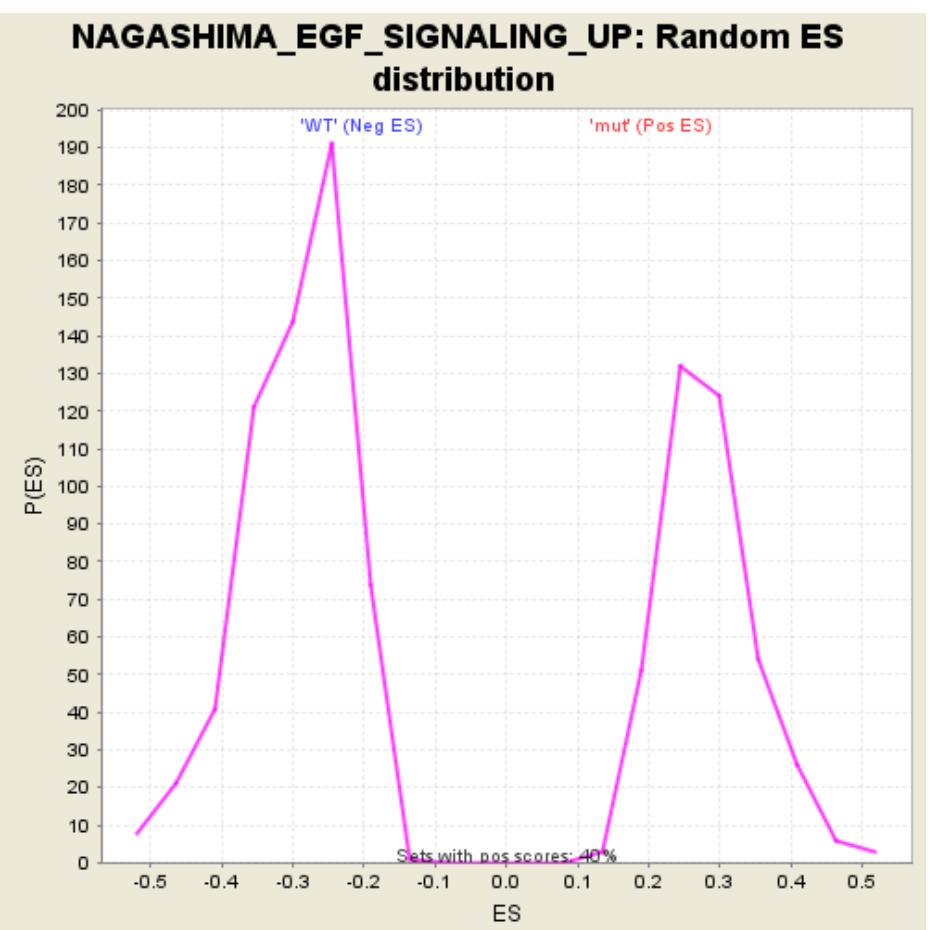


Fig 3: NAGASHIMA_EGF_SIGNALING_UP: Random ES distribution
Gene set null distribution of ES for NAGASHIMA_EGF_SIGNALING_UP

Fig S8

Table: GSEA Results Summary	
Dataset	CB_6M_collapsed_to_symbols.general.cls#mut_versus_WT
Phenotype	general.cls#mut_versus_WT
Upregulated in class	mut
GeneSet	NAGASHIMA_EGF_SIGNALING_UP
Enrichment Score (ES)	0.62230736
Normalized Enrichment Score (NES)	2.2834282
Nominal p-value	0.0
FDR q-value	0.0018626772
FWER p-Value	0.0030

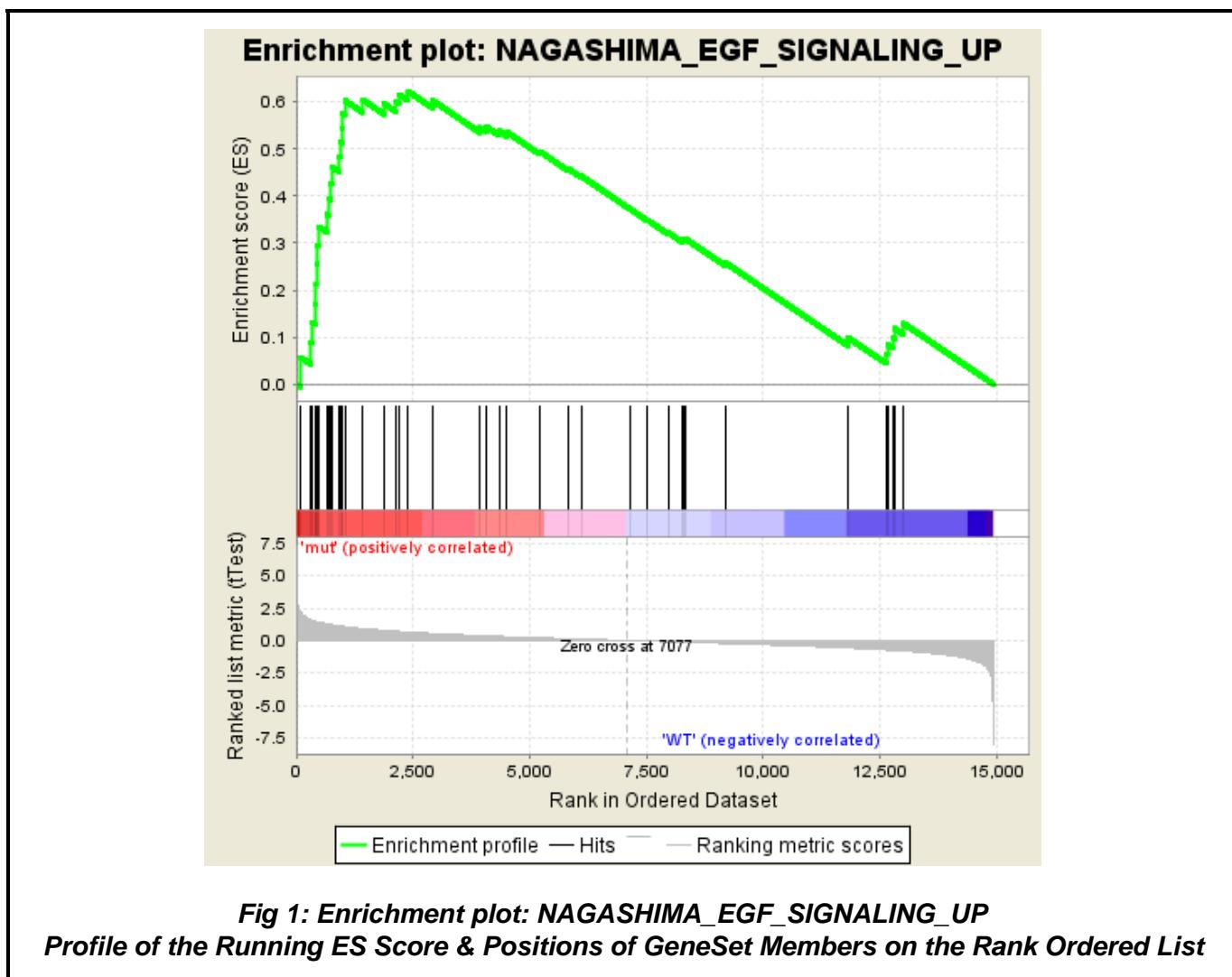


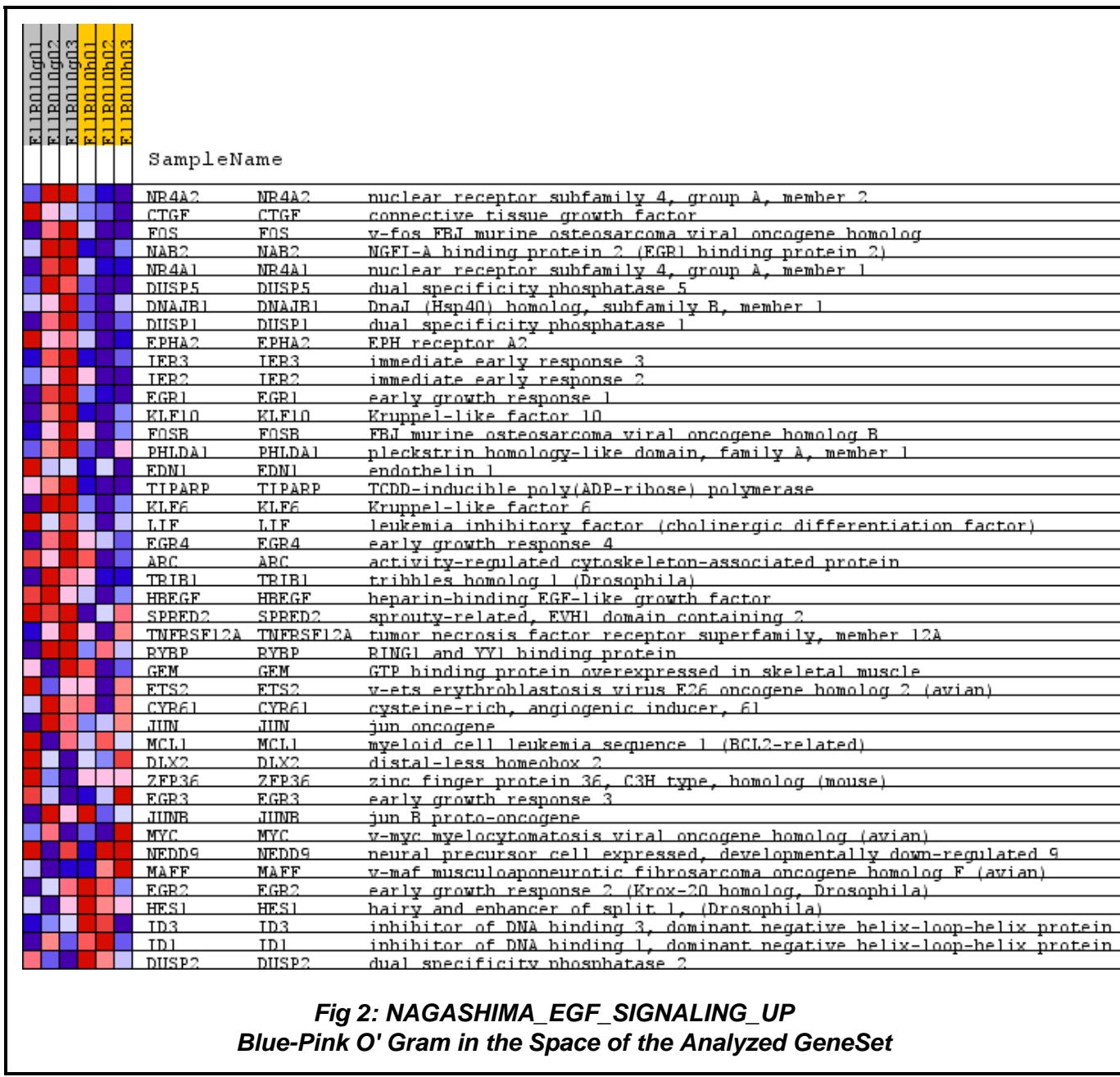
Table: GSEA details [plain text format]

	PROBE	GENE SYMBOL	GENE_TITLE	RANK IN GENE LIST	RANK METRIC SCORE	RUNNING ES	CORE ENRICHMENT
1	NR4A2	NR4A2 Entrez , Source	nuclear receptor subfamily 4, group A, member 2	99	2.157	0.0564	Yes

2	CTGF	CTGF Entrez , Source	connective tissue growth factor	304	1.598	0.0893	Yes
3	FOS	FOS Entrez , Source	v-fos FBJ murine osteosarcoma viral oncogene homolog	347	1.544	0.1316	Yes
4	NAB2	NAB2 Entrez , Source	NGFI-A binding protein 2 (EGR1 binding protein 2)	414	1.480	0.1704	Yes
5	NR4A1	NR4A1 Entrez , Source	nuclear receptor subfamily 4, group A, member 1	424	1.467	0.2126	Yes
6	DUSP5	DUSP5 Entrez , Source	dual specificity phosphatase 5	430	1.461	0.2550	Yes
7	DNAJB1	DNAJB1 Entrez , Source	DnaJ (Hsp40) homolog, subfamily B, member 1	448	1.446	0.2961	Yes
8	DUSP1	DUSP1 Entrez , Source	dual specificity phosphatase 1	489	1.408	0.3345	Yes
9	EPHA2	EPHA2 Entrez , Source	EPH receptor A2	654	1.289	0.3611	Yes
10	IER3	IER3 Entrez , Source	immediate early response 3	710	1.249	0.3939	Yes
11	IER2	IER2 Entrez , Source	immediate early response 2	742	1.230	0.4277	Yes
12	EGR1	EGR1 Entrez , Source	early growth response 1	770	1.213	0.4614	Yes
13	KLF10	KLF10 Entrez , Source	Kruppel-like factor 10	923	1.122	0.4839	Yes
14	FOSB	FOSB Entrez , Source	FBJ murine osteosarcoma viral oncogene homolog B	944	1.113	0.5151	Yes
15	PHLDA1	PHLDA1 Entrez , Source	pleckstrin homology-like domain, family A, member 1	977	1.093	0.5448	Yes
16	EDN1	EDN1 Entrez , Source	endothelin 1	990	1.090	0.5759	Yes
17	TIPARP	TIPARP Entrez , Source	TCDD-inducible poly(ADP-ribose) polymerase	1041	1.066	0.6037	Yes
18	KLF6	KLF6 Entrez , Source	Kruppel-like factor 6	1430	0.912	0.6042	Yes
19	LIF	LIF Entrez , Source	leukemia inhibitory factor (cholinergic differentiation factor)	1883	0.778	0.5966	Yes

20	EGR4	EGR4 Entrez , Source	early growth response 4	2142	0.708	0.6000	Yes
21	ARC	ARC Entrez , Source	activity-regulated cytoskeleton-associated protein	2197	0.695	0.6166	Yes
22	TRIB1	TRIB1 Entrez , Source	tribbles homolog 1 (Drosophila)	2398	0.654	0.6223	Yes
23	HBEGF	HBEGF Entrez , Source	heparin-binding EGF-like growth factor	2915	0.546	0.6036	No
24	SPRED2	SPRED2 Entrez , Source	sprouty-related, EVH1 domain containing 2	3941	0.376	0.5457	No
25	TNFRSF12A	TNFRSF12A Entrez , Source	tumor necrosis factor receptor superfamily, member 12A	4064	0.358	0.5480	No
26	RYBP	RYBP Entrez , Source	RING1 and YY1 binding protein	4342	0.319	0.5387	No
27	GEM	GEM Entrez , Source	GTP binding protein overexpressed in skeletal muscle	4516	0.296	0.5357	No
28	ETS2	ETS2 Entrez , Source	v-ets erythroblastosis virus E26 oncogene homolog 2 (avian)	5212	0.211	0.4952	No
29	CYR61	CYR61 Entrez , Source	cysteine-rich, angiogenic inducer, 61	5829	0.138	0.4579	No
30	JUN	JUN Entrez , Source	jun oncogene	6104	0.107	0.4426	No
31	MCL1	MCL1 Entrez , Source	myeloid cell leukemia sequence 1 (BCL2-related)	7147	-0.008	0.3729	No
32	DLX2	DLX2 Entrez , Source	distal-less homeobox 2	7529	-0.049	0.3487	No
33	ZFP36	ZFP36 Entrez , Source	zinc finger protein 36, C3H type, homolog (mouse)	7970	-0.099	0.3221	No
34	EGR3	EGR3 Entrez , Source	early growth response 3	8269	-0.127	0.3058	No
35	JUNB	JUNB Entrez , Source	jun B proto-oncogene	8290	-0.130	0.3082	No
36	MYC	MYC Entrez , Source	v-myc myelocytomatosis viral oncogene homolog (avian)	8331	-0.134	0.3094	No
37	NEDD9	NEDD9 Entrez , Source	neural precursor cell expressed, developmentally down-regulated 9	9193	-0.228	0.2583	No

38	MAFF	MAFF Entrez , Source	v-maf musculoaponeurotic fibrosarcoma oncogene homolog F (avian)	11825	-0.575	0.0984	No
39	EGR2	EGR2 Entrez , Source	early growth response 2 (Krox-20 homolog, Drosophila)	12645	-0.724	0.0645	No
40	HES1	HES1 Entrez , Source	hairy and enhancer of split 1, (Drosophila)	12668	-0.729	0.0843	No
41	ID3	ID3 Entrez , Source	inhibitor of DNA binding 3, dominant negative helix-loop-helix protein	12777	-0.750	0.0990	No
42	ID1	ID1 Entrez , Source	inhibitor of DNA binding 1, dominant negative helix-loop-helix protein	12818	-0.758	0.1185	No
43	DUSP2	DUSP2 Entrez , Source	dual specificity phosphatase 2	13012	-0.803	0.1289	No



NAGASHIMA_EGF_SIGNALING_UP: Random ES distribution

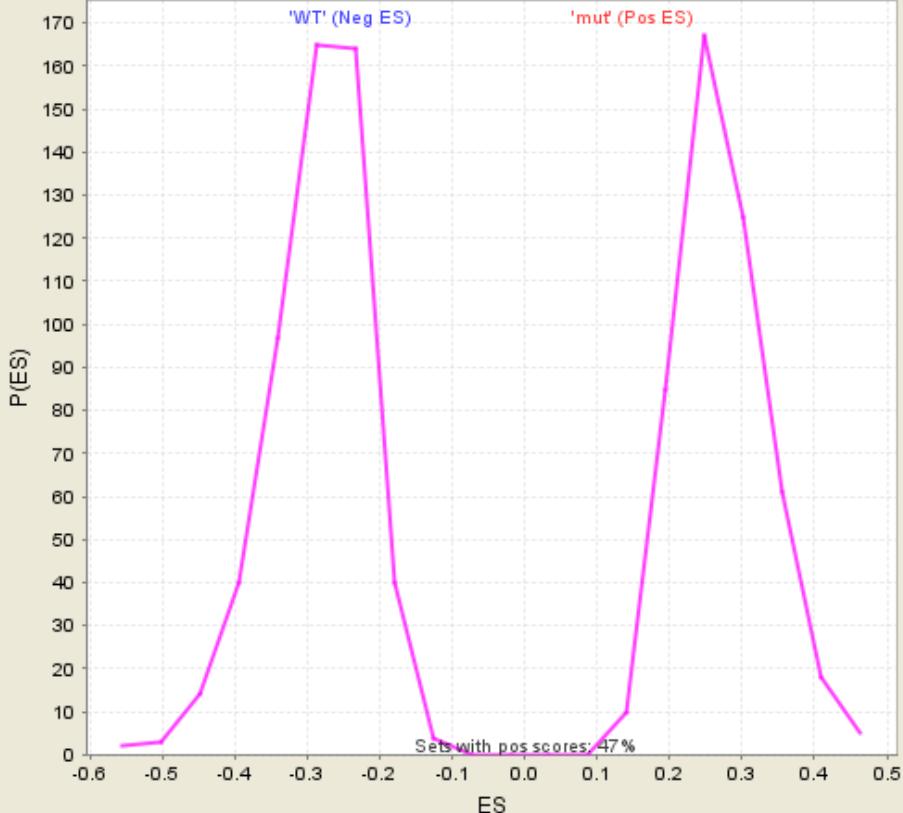
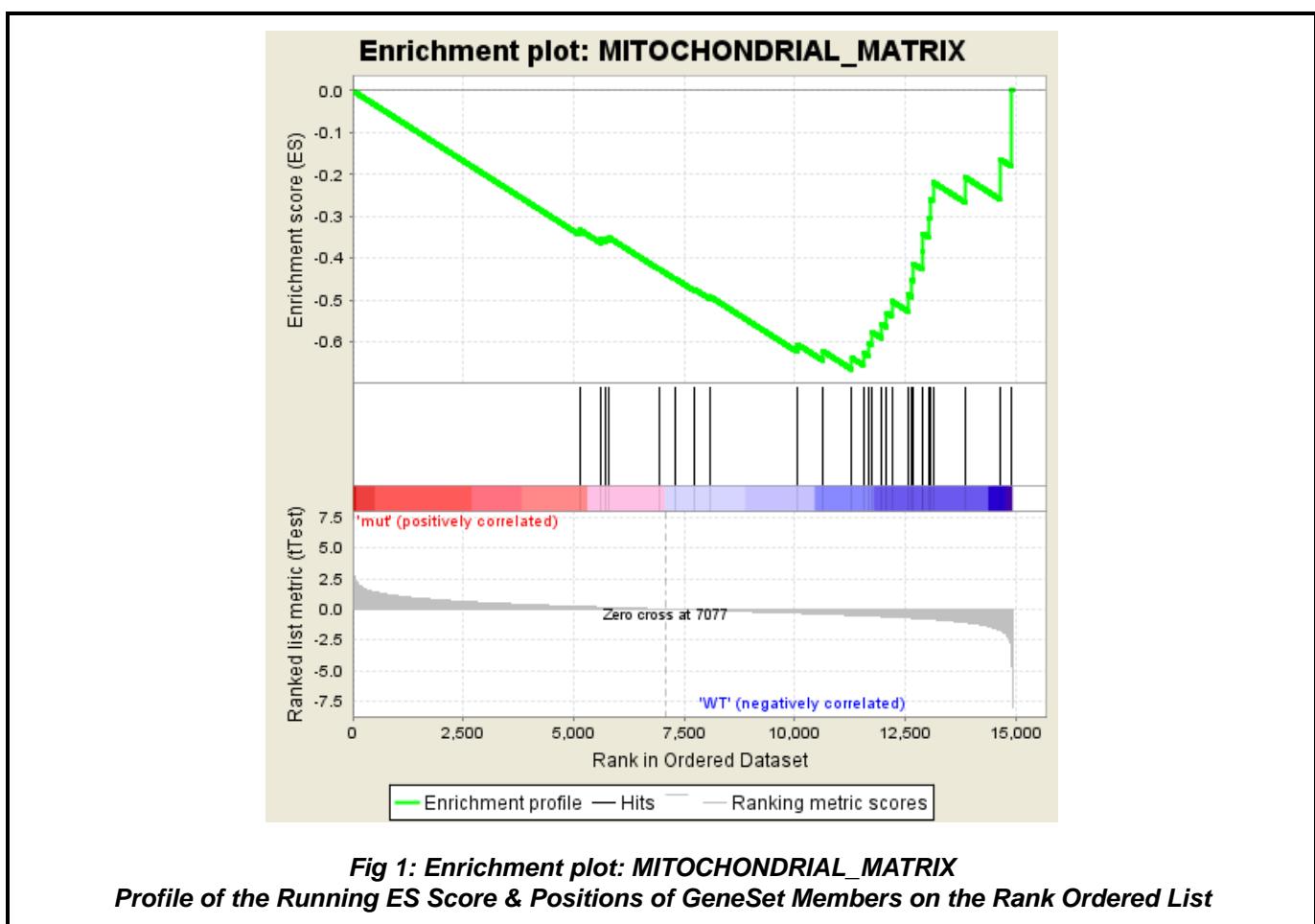


Fig 3: NAGASHIMA_EGF_SIGNALING_UP: Random ES distribution
Gene set null distribution of ES for NAGASHIMA_EGF_SIGNALING_UP

Fig S9

Table: GSEA Results Summary	
Dataset	CB_6M_collapsed_to_symbols.general.cls#mut_versus_WT
Phenotype	general.cls#mut_versus_WT
Upregulated in class	WT
GeneSet	MITOCHONDRIAL_MATRIX
Enrichment Score (ES)	-0.66644275
Normalized Enrichment Score (NES)	-2.174532
Nominal p-value	0.0
FDR q-value	0.014642635
FWER p-Value	0.018

**Table: GSEA details [plain text format]**

	PROBE	GENE SYMBOL	GENE_TITLE	RANK IN GENE LIST	RANK METRIC SCORE	RUNNING ES	CORE ENRICHMENT
1	NR3C1	NR3C1 Entrez , Source	nuclear receptor subfamily 3, group C, member 1 (glucocorticoid receptor)	5133	0.219	-0.3321	No
2	MRPS22	MRPS22 Entrez , Source	mitochondrial ribosomal protein S22	5613	0.164	-0.3550	No
3	CASQ1	CASQ1 Entrez , Source	calsequestrin 1 (fast-twitch, skeletal muscle)	5732	0.150	-0.3545	No

4	MRPS11	MRPS11 Entrez , Source	mitochondrial ribosomal protein S11	5807	0.141	-0.3516	No
5	CS	CS Entrez , Source	citrate synthase	6927	0.018	-0.4257	No
6	POLG2	POLG2 Entrez , Source	polymerase (DNA directed), gamma 2, accessory subunit	7290	-0.023	-0.4487	No
7	NFS1	NFS1 Entrez , Source	NFS1 nitrogen fixation 1 (<i>S. cerevisiae</i>)	7731	-0.071	-0.4742	No
8	PITRM1	PITRM1 Entrez , Source	pitrilysin metallopeptidase 1	8090	-0.110	-0.4921	No
9	MRPS15	MRPS15 Entrez , Source	mitochondrial ribosomal protein S15	10064	-0.327	-0.6061	No
10	TIMM44	TIMM44 Entrez , Source	translocase of inner mitochondrial membrane 44 homolog (yeast)	10635	-0.401	-0.6219	No
11	NDUFAB1	NDUFAB1 Entrez , Source	NADH dehydrogenase (ubiquinone) 1, alpha/beta subcomplex, 1, 8kDa	11300	-0.492	-0.6389	Yes
12	MRPL41	MRPL41 Entrez , Source	mitochondrial ribosomal protein L41	11553	-0.530	-0.6260	Yes
13	MRPS10	MRPS10 Entrez , Source	mitochondrial ribosomal protein S10	11689	-0.553	-0.6041	Yes
14	MRPS16	MRPS16 Entrez , Source	mitochondrial ribosomal protein S16	11735	-0.561	-0.5756	Yes
15	MRPL52	MRPL52 Entrez , Source	mitochondrial ribosomal protein L52	11972	-0.598	-0.5579	Yes
16	ATP5F1	ATP5F1 Entrez , Source	ATP synthase, H+ transporting, mitochondrial F0 complex, subunit B1	12087	-0.617	-0.5309	Yes
17	MRPL55	MRPL55 Entrez , Source	mitochondrial ribosomal protein L55	12199	-0.638	-0.5026	Yes
18	BCKDHB	BCKDHB Entrez , Source	branched chain keto acid dehydrogenase E1, beta polypeptide (maple syrup urine disease)	12567	-0.709	-0.4875	Yes
19	MRPS18A	MRPS18A Entrez , Source	mitochondrial ribosomal protein S18A	12653	-0.726	-0.4525	Yes
20	MRPL10	MRPL10 Entrez , Source	mitochondrial ribosomal protein L10	12687	-0.732	-0.4136	Yes
21	ETFA	ETFA Entrez , Source	electron-transfer-flavoprotein, alpha polypeptide (glutaric aciduria II)	12887	-0.773	-0.3836	Yes
22	MRPL40	MRPL40 Entrez , Source	mitochondrial ribosomal protein L40	12903	-0.778	-0.3410	Yes
23	MRPS21	MRPS21 Entrez , Source	mitochondrial ribosomal protein S21	13040	-0.810	-0.3047	Yes

24	BCKDHA	BCKDHA Entrez, Source	branched chain keto acid dehydrogenase E1, alpha polypeptide	13063	-0.815	-0.2604	Yes
25	MRPL51	MRPL51 Entrez, Source	mitochondrial ribosomal protein L51	13132	-0.837	-0.2181	Yes
26	DNAJA3	DNAJA3 Entrez, Source	DnaJ (Hsp40) homolog, subfamily A, member 3	13866	-1.077	-0.2068	Yes
27	DBT	DBT Entrez, Source	dihydrolipoamide branched chain transacylase E2	14644	-1.712	-0.1630	Yes
28	ALDH4A1	ALDH4A1 Entrez, Source	aldehyde dehydrogenase 4 family, member A1	14903	-3.250	0.0019	Yes

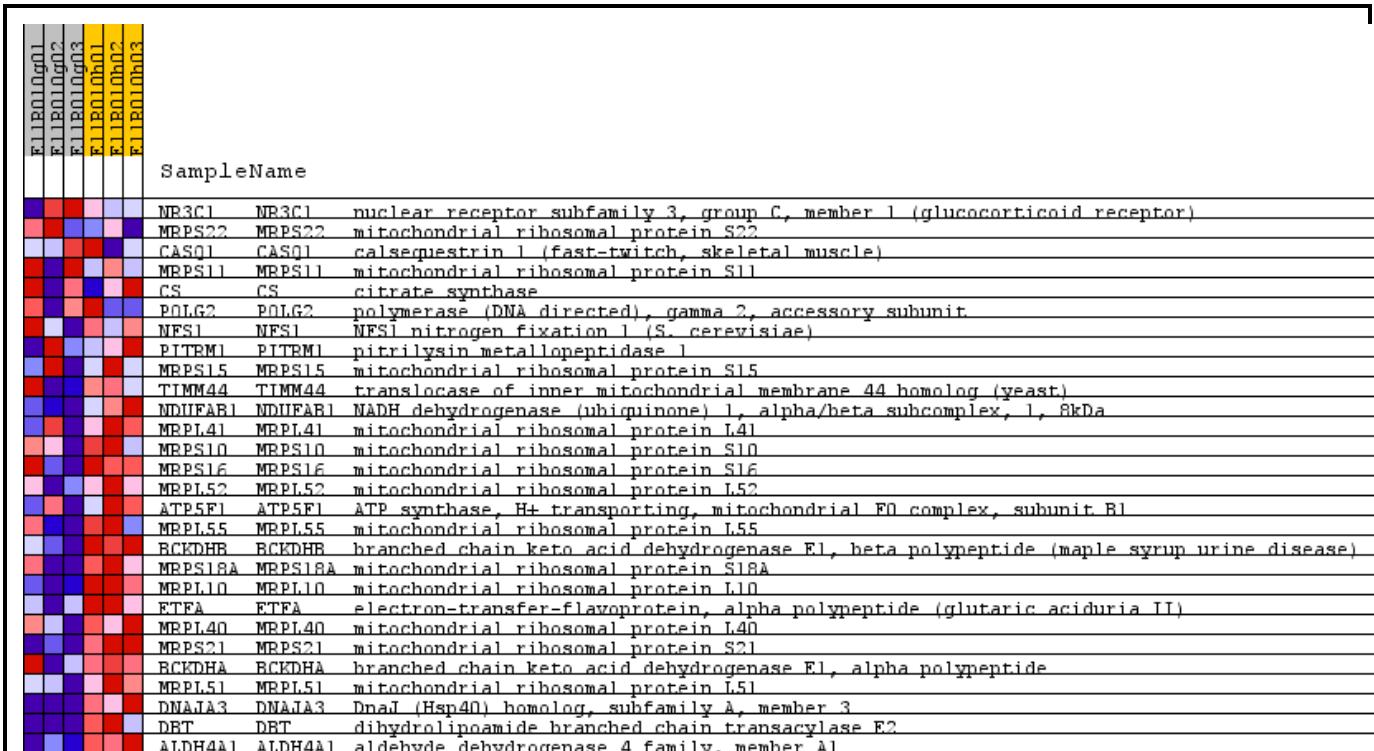


Fig 2: MITOCHONDRIAL MATRIX
Blue-Pink O' Gram in the Space of the Analyzed GeneSet

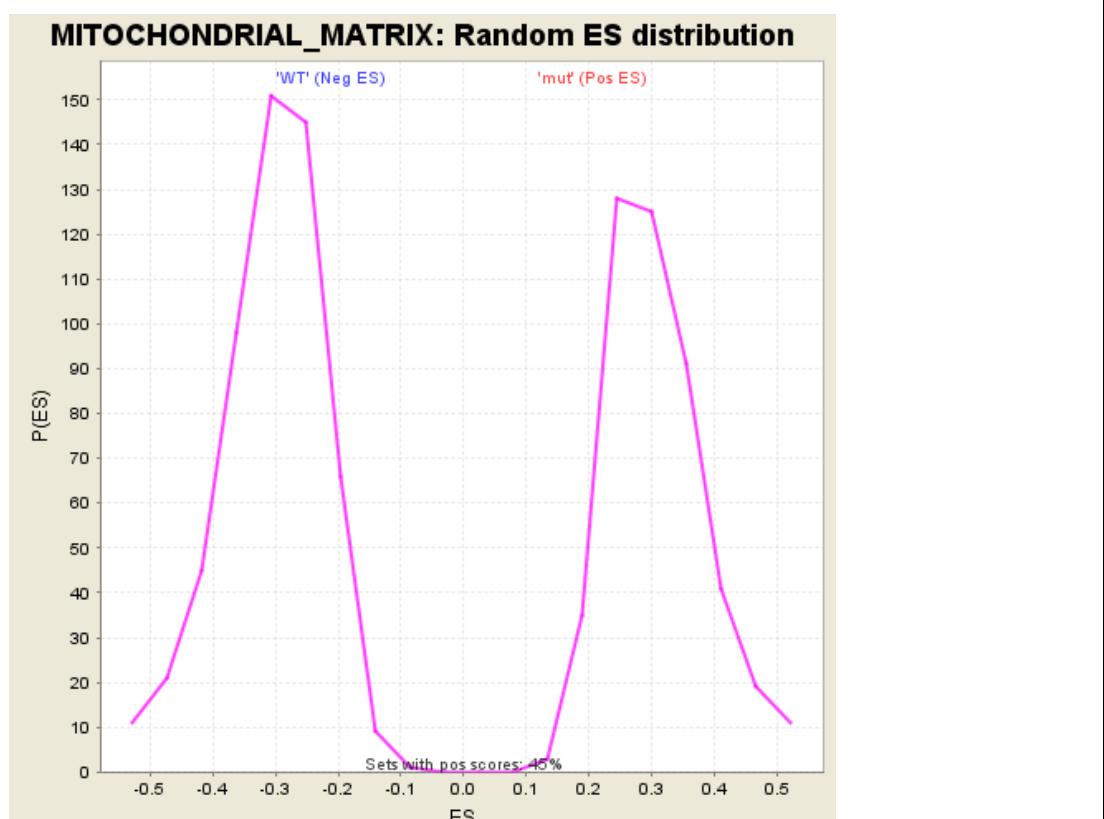


Fig 3: MITOCHONDRIAL_MATRIX: Random ES distribution
Gene set null distribution of ES for MITOCHONDRIAL_MATRIX

Fig S10

Table: GSEA Results Summary	
Dataset	MB_6M_collapsed_to_symbols.general.cls#mut_versus_WT
Phenotype	general.cls#mut_versus_WT
Upregulated in class	mut
GeneSet	QI_HYPOXIA_TARGETS_OF_HIF1A_AND_FOXA2
Enrichment Score (ES)	0.71504146
Normalized Enrichment Score (NES)	2.1899784
Nominal p-value	0.0
FDR q-value	0.014409064
FWER p-Value	0.015

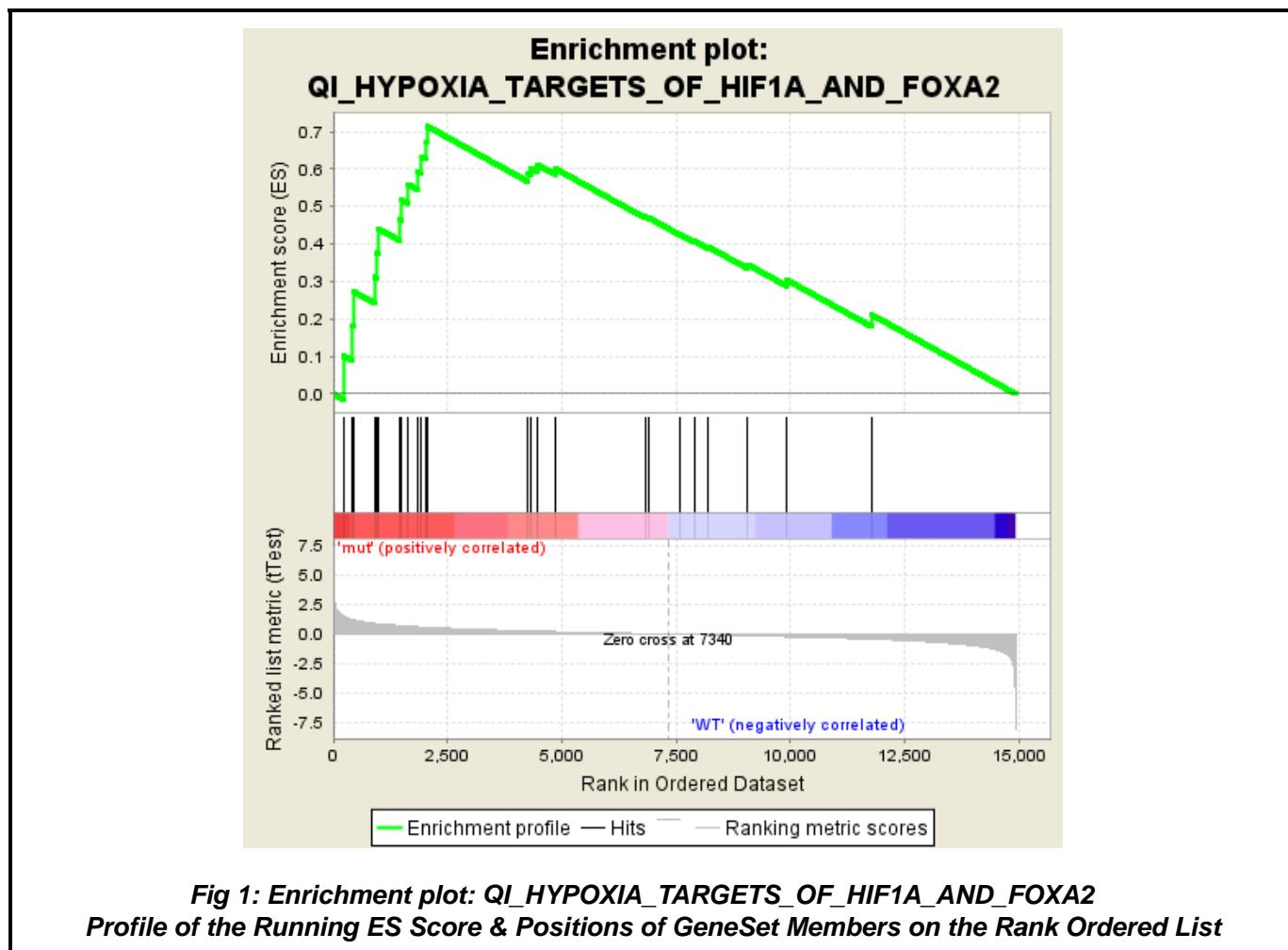
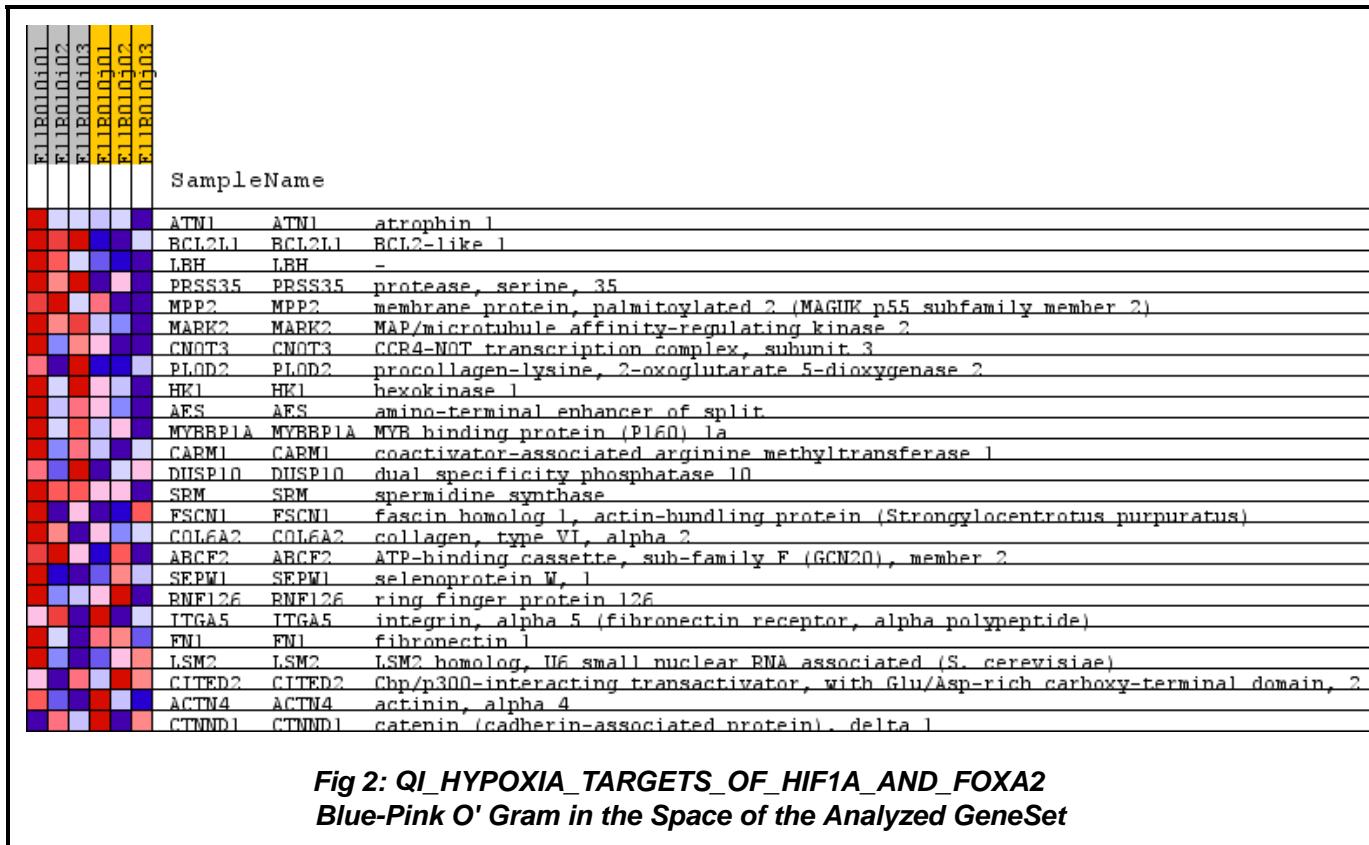


Table: GSEA details [plain text format]							
	PROBE	GENE SYMBOL	GENE_TITLE	RANK IN GENE LIST	RANK METRIC SCORE	RUNNING ES	CORE ENRICHMENT
1	ATN1	ATN1 Entrez , Source	atrophin 1	218	1.533	0.1012	Yes
2	BCL2L1	BCL2L1 Entrez , Source	BCL2-like 1	404	1.251	0.1832	Yes

3	LBH	LBH Entrez, Source	-	434	1.222	0.2735	Yes
4	PRSS35	PRSS35 Entrez, Source	protease, serine, 35	900	0.917	0.3115	Yes
5	MPP2	MPP2 Entrez, Source	membrane protein, palmitoylated 2 (MAGUK p55 subfamily member 2)	945	0.898	0.3764	Yes
6	MARK2	MARK2 Entrez, Source	MAP/microtubule affinity-regulating kinase 2	974	0.887	0.4415	Yes
7	CNOT3	CNOT3 Entrez, Source	CCR4-NOT transcription complex, subunit 3	1444	0.736	0.4656	Yes
8	PLOD2	PLOD2 Entrez, Source	procollagen-lysine, 2-oxoglutarate 5-dioxygenase 2	1485	0.725	0.5176	Yes
9	HK1	HK1 Entrez, Source	hexokinase 1	1636	0.685	0.5593	Yes
10	AES	AES Entrez, Source	amino-terminal enhancer of split	1835	0.633	0.5938	Yes
11	MYBBP1A	MYBBP1A Entrez, Source	MYB binding protein (P160) 1a	1934	0.611	0.6334	Yes
12	CARM1	CARM1 Entrez, Source	coactivator-associated arginine methyltransferase 1	2039	0.590	0.6709	Yes
13	DUSP10	DUSP10 Entrez, Source	dual specificity phosphatase 10	2045	0.589	0.7150	Yes
14	SRM	SRM Entrez, Source	spermidine synthase	4261	0.270	0.5868	No
15	FSCN1	FSCN1 Entrez, Source	fascin homolog 1, actin-bundling protein (Strongylocentrotus purpuratus)	4314	0.263	0.6032	No
16	COL6A2	COL6A2 Entrez, Source	collagen, type VI, alpha 2	4461	0.249	0.6122	No
17	ABCF2	ABCF2 Entrez, Source	ATP-binding cassette, sub-family F (GCN20), member 2	4850	0.209	0.6019	No
18	SEPW1	SEPW1 Entrez, Source	selenoprotein W, 1	6823	0.042	0.4728	No
19	RNF126	RNF126 Entrez, Source	ring finger protein 126	6904	0.035	0.4701	No
20	ITGA5	ITGA5 Entrez, Source	integrin, alpha 5 (fibronectin receptor, alpha polypeptide)	7597	-0.018	0.4250	No
21	FN1	FN1 Entrez, Source	fibronectin 1	7895	-0.042	0.4082	No

22	LSM2	LSM2 Entrez , Source	LSM2 homolog, U6 small nuclear RNA associated (<i>S. cerevisiae</i>)	8210	-0.067	0.3922	No
23	CITED2	CITED2 Entrez , Source	Cbp/p300-interacting transactivator, with Glu/Asp-rich carboxy-terminal domain, 2	9073	-0.139	0.3449	No
24	ACTN4	ACTN4 Entrez , Source	actinin, alpha 4	9929	-0.215	0.3038	No
25	CTNND1	CTNND1 Entrez , Source	catenin (cadherin-associated protein), delta 1	11781	-0.420	0.2114	No



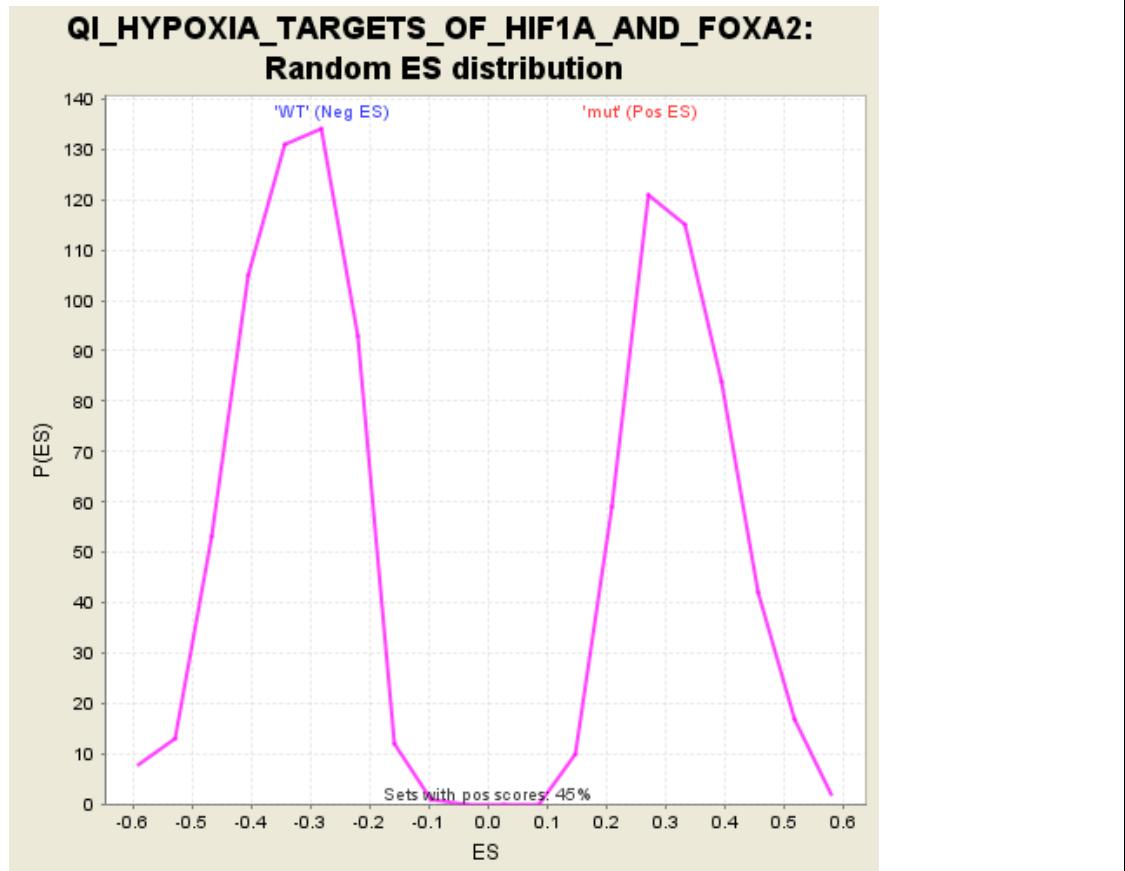


Fig 3: QI_HYPOXIA_TARGETS_OF_HIF1A_AND_FOXA2: Random ES distribution
Gene set null distribution of ES for QI_HYPOXIA_TARGETS_OF_HIF1A_AND_FOXA2

Fig S11

Table: GSEA Results Summary	
Dataset	MB_6M_collapsed_to_symbols.general.cls#mut_versus_WT
Phenotype	general.cls#mut_versus_WT
Upregulated in class	mut
GeneSet	PID_ATM_PATHWAY
Enrichment Score (ES)	0.5320049
Normalized Enrichment Score (NES)	1.6343192
Nominal p-value	0.006342495
FDR q-value	1.0
FWER p-Value	1.0

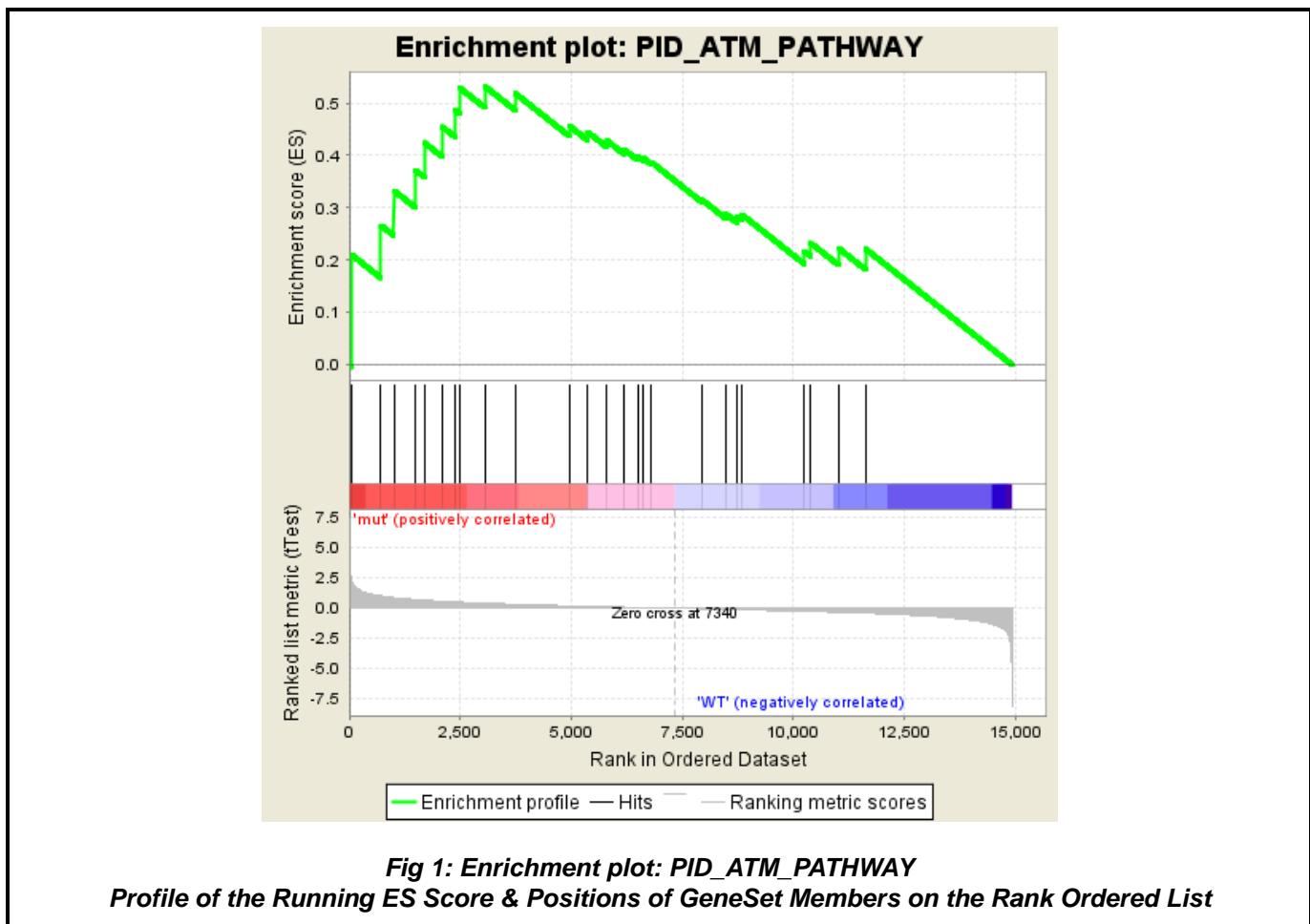


Table: GSEA details [plain text format]

	PROBE	GENE SYMBOL	GENE_TITLE	RANK IN GENE LIST	RANK METRIC SCORE	RUNNING ES	CORE ENRICHMENT
1	BRCA1	BRCA1 Entrez , Source	breast cancer 1, early onset	66	2.166	0.2090	Yes
2	BID	BID Entrez , Source	BH3 interacting domain death agonist	713	1.018	0.2660	Yes
3	RFWD2	RFWD2 Entrez , Source	ring finger and WD repeat domain 2	1002	0.874	0.3328	Yes

4	RAD50	RAD50 Entrez , Source	RAD50 homolog (S. cerevisiae)	1495	0.723	0.3711	Yes
5	CDC25A	CDC25A Entrez , Source	cell division cycle 25A	1692	0.673	0.4242	Yes
6	H2AFX	H2AFX Entrez , Source	H2A histone family, member X	2094	0.580	0.4545	Yes
7	ABL1	ABL1 Entrez , Source	v-abl Abelson murine leukemia viral oncogene homolog 1	2374	0.523	0.4873	Yes
8	RAD17	RAD17 Entrez , Source	RAD17 homolog (S. pombe)	2498	0.501	0.5284	Yes
9	XRCC4	XRCC4 Entrez , Source	X-ray repair complementing defective repair in Chinese hamster cells 4	3053	0.414	0.5320	Yes
10	YWHAB	YWHAB Entrez , Source	tyrosine 3-monooxygenase/tryptophan 5-monooxygenase activation protein, beta polypeptide	3739	0.329	0.5185	No
11	RBBP8	RBBP8 Entrez , Source	retinoblastoma binding protein 8	4954	0.199	0.4567	No
12	RNF8	RNF8 Entrez , Source	ring finger protein 8	5376	0.161	0.4443	No
13	ATM	ATM Entrez , Source	ataxia telangiectasia mutated (includes complementation groups A, C and D)	5809	0.126	0.4277	No
14	BLM	BLM Entrez , Source	Bloom syndrome	6187	0.093	0.4116	No
15	TRIM28	TRIM28 Entrez , Source	tripartite motif-containing 28	6499	0.068	0.3974	No
16	CTBP1	CTBP1 Entrez , Source	C-terminal binding protein 1	6630	0.058	0.3944	No
17	MDC1	MDC1 Entrez , Source	mediator of DNA damage checkpoint 1	6807	0.043	0.3869	No
18	RAD9A	RAD9A Entrez , Source	RAD9 homolog A (S. pombe)	7948	-0.046	0.3150	No
19	SMC1A	SMC1A Entrez , Source	structural maintenance of chromosomes 1A	8491	-0.091	0.2877	No
20	CHEK2	CHEK2 Entrez , Source	CHK2 checkpoint homolog (S. pombe)	8729	-0.112	0.2828	No
21	FANCD2	FANCD2 Entrez , Source	Fanconi anemia, complementation group D2	8839	-0.120	0.2873	No
22	TOP3A	TOP3A Entrez , Source	topoisomerase (DNA) III alpha	10252	-0.244	0.2166	No
23	SMC3	SMC3 Entrez , Source	structural maintenance of chromosomes 3	10393	-0.258	0.2326	No

24	UBE2N	UBE2N Entrez, Source	ubiquitin-conjugating enzyme E2N (UBC13 homolog, yeast)	11028	-0.327	0.2223	No
25	MRE11A	MRE11A Entrez, Source	MRE11 meiotic recombination 11 homolog A (<i>S. cerevisiae</i>)	11632	-0.401	0.2214	No

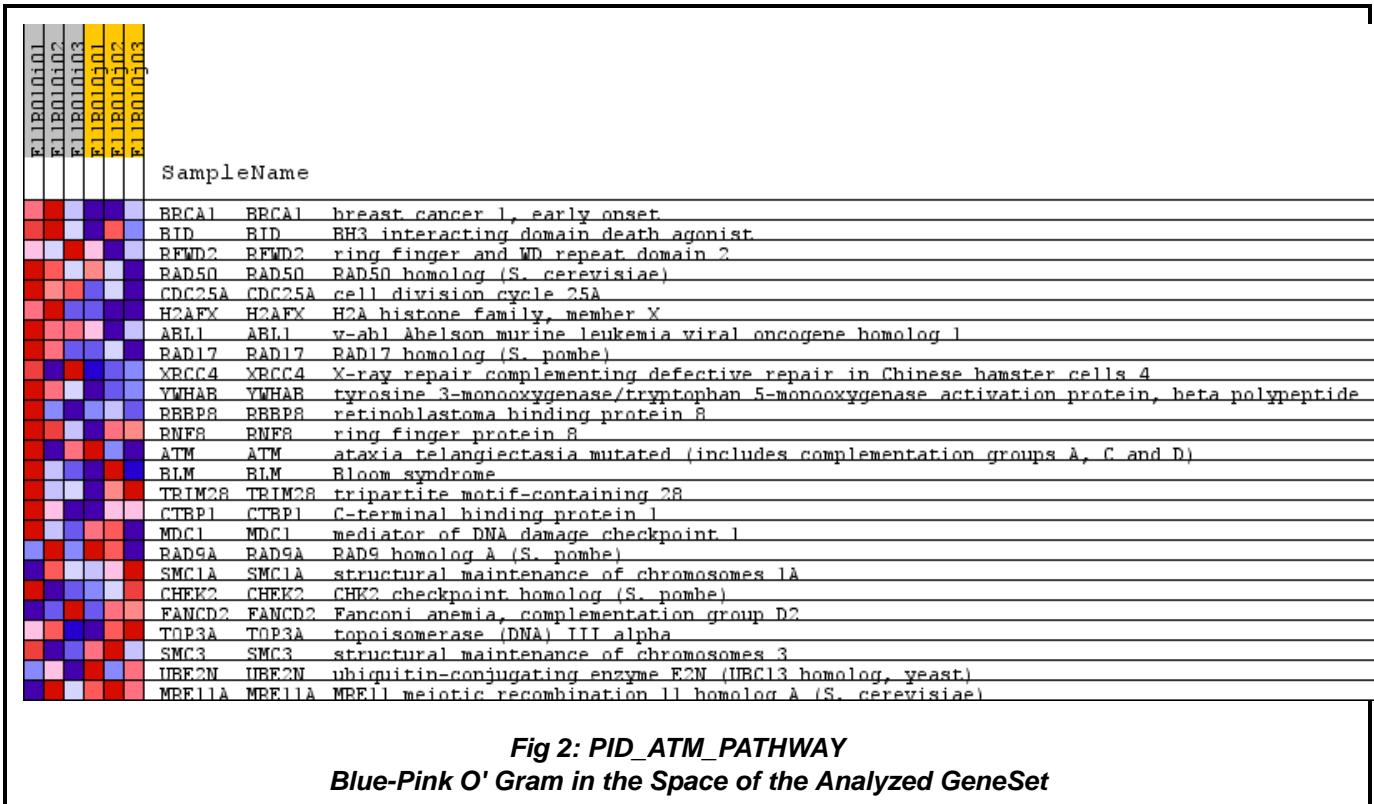


Fig 2: PID_ATM_PATHWAY
Blue-Pink O' Gram in the Space of the Analyzed GeneSet

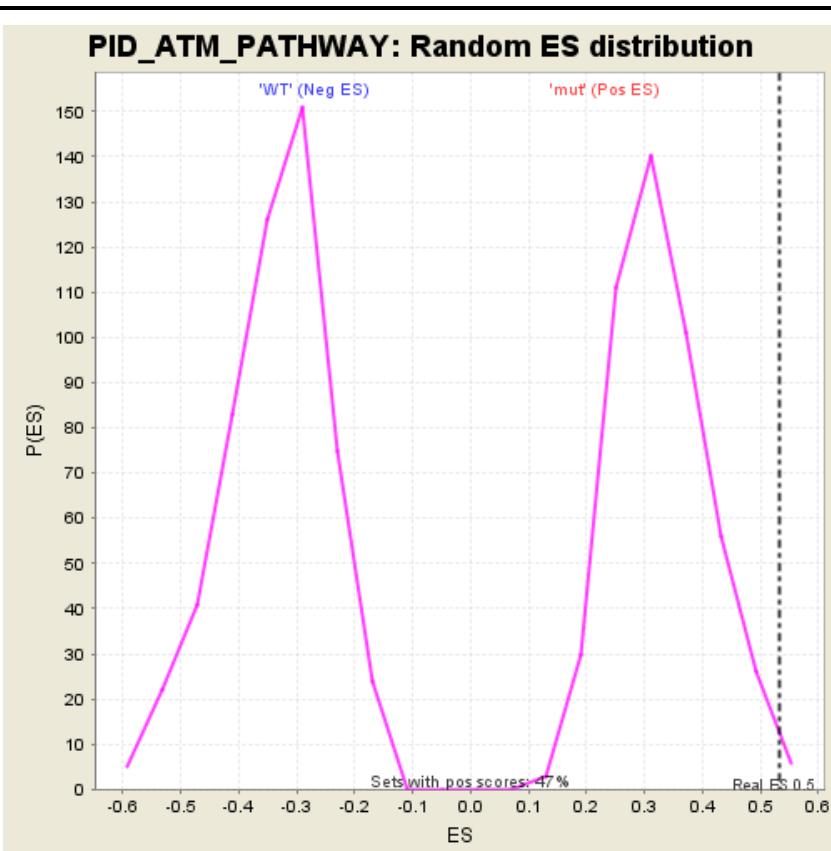


Fig 3: PID_ATM_PATHWAY: Random ES distribution
Gene set null distribution of ES for PID_ATM_PATHWAY