

## SUPPLEMENTARY DATA

**TITLE:**

**Sucrose is an early modulator of the key hormonal mechanisms controlling bud outgrowth in *Rosa hybrida***

**Running title:** Sucrose regulates bud outgrowth hormonal network

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## Supplementary Table S1

**A**

Hormone	Retention time (min)	Ions mode	MSMS transition	Collision Energy (V)	Sampling cone voltage (V)
Z7G <sup>2</sup> H <sub>5</sub> -t-Z7G	2.41	-	382>220 387>225	35	20
ZOG <sup>2</sup> H <sub>5</sub> -t-ZOG	2.54	-	382>220 387>225	35	20
Z <sup>15</sup> N-t-Z	2.67	-	221>137 220>136	45	15
Z9G <sup>2</sup> H <sub>5</sub> -t-Z9G	2.88	-	382>220 387>225	35	20
ZROG <sup>2</sup> H <sub>5</sub> -t-ZROG	3.43	-	514>220 519>225	40	23
ZR <sup>2</sup> H <sub>5</sub> -t-ZR	3.71	-	352>220 357>225	40	17
ZRMP <sup>2</sup> H <sub>5</sub> -t-ZRMP	3.82	-	432>220 437>225	40	20
iPRMP <sup>2</sup> H <sub>5</sub> -iPRMP	4.25	-	416>204 422>210	40	20
iP <sup>2</sup> H <sub>5</sub> -iP	4.76	-	204>136 210>137	35	12
iPR <sup>15</sup> N-iPR	5.88	-	336>204 334>205	40	15

**B**

Hormone	LOQ ng/g DW	LOD ng/g DW
Z7G	ND	ND
ZOG *	8.66	2.6
Z	1.43	0.4
Z9G	ND	ND
ZROG *	2.85	0.86
ZR	1.11	0.33
ZRMP	11.71	3.51
iPRMP	2.99	0.9
iP	0.42	0.12
iPR	0.83	0.25

**Supplementary Table S1.** (A) Parameters used for LC-ESI-MS/MS analysis of cytokinin contents. (B) Summary of LOD and LOQ in samples (\* in CKs g<sup>-1</sup> DW).

## Supplementary Table S2

<b>Protein identified in <i>Rosa hybrida</i></b>	<b>Peptidic homology with <i>Arabidopsis thaliana</i></b>		<b>Conserved domains characteristic of the putative protein functions</b>
<b>RhIPT3</b> (KJ933389)	AtIPT3 (AT3G63110)	Identities : 59 %	- Adenylate isopentenyltransferase domains
<b>RhIPT5</b> (KJ933390)	AtIPT5 (AT5G19040)	Identities : 68 %	- Adenylate isopentenyltransferase domains
<b>RhCKX1</b> (RC001490*)	AtCKX1 (AT2G41510)	Identities : 65 %	- FAD and cytokinin binding domain - cytokinin dehydrogenase domain
<b>RhTAR1</b> (KJ933382)	AtTAR2 (AT4G24670)	Identities : 61 %	- Aminotransferase domains
<b>RhYUC1</b> (KJ933383)	AtYUC4 (AT5G11320)	Identities : 70 %	- NAD(P)-binding Rossmann-like domain - FAD binding domain - Flavin-binding monooxygenase-like - Flavin-containing monooxygenase FMO GS-OX
<b>RhPIN1</b> (KJ933386)	AtPIN1 (AT1G73590)	Identities : 73 %	- Auxin Efflux Carrier (AEC) Family - Membrane transport protein
<b>RhPID</b> (KJ933387)	AtPID (AT2G34650)	Identities : 81 %	- Serine/threonine protein kinase
<b>RhBRC1</b> (KF530803)	AtBRC1 (AT3G18550)	Identities : 71 %	- Transcription factor TCP24 (TEOSINTE BRANCHED1, CYCLOIDEA, AND PCF FAMILY 24)
<b>RhSUSI1</b> (KJ933388)	AtWRKY20 (AT4G26640)	Identities : 55 %	- 2 WRKY DNA-binding domains - 1 Nuclear Localization Signal (NLS)

**Supplementary Table S2.** Homologies between the protein sequences of the genes cloned in *Rosa hybrida* and those of *Arabidopsis thaliana*, and conserved domains detected on the protein sequences of *Rosa hybrida* that could be implied in their putative function. Multiple sequence alignment was first performed with the BLAST tool of The Arabidopsis Information Resource site (TAIR, arabidopsis.org) and conserved domains were determined with the CDD tool of the NCBI or with NLS mapper. Accession code is given for each gene. \* cluster number from *Rosa chinensis* EST data base (<https://iant.toulouse.inra.fr/plants/rosa/FATAL>).

### Supplementary Table S3

<b>Gene</b>	<b>Primers</b>
<b><i>RhIPT3</i></b>	Fwd : CCACCATTGCTTGGAGTT Rev : CTCCAACGATGA TTGGAAGC
<b><i>RhIPT5</i></b>	Fwd : GGCACAGTGGATCCTGAGTC Rev : CGTACTTCTCCGAAACTCG
<b><i>RhCKX1</i></b>	Fwd : CCAGTCAACAAAACCAAGTGGAA Rev : CCTGTGGAAGATGGCACCAC
<b><i>RhBRC1</i></b>	Fwd : TGCATTGTTTAACCCTCTGCA Rev : GTTCTTCTCTTGTCTCGCTCTT
<b><i>RwMAX2</i></b>	Fwd : GCTGCCTATCCCGTTTTCTC Rev : AATCCACAGTATCACCACAATCC
<b><i>RwMAX3</i></b>	Fwd : CAGCAGCTTCTTCGGTCT Rev : CTTCAACCACATGTCAAATGGA
<b><i>RwMAX4</i></b>	Fwd : CCCGTGTTGGCGTTATCG Rev : AGTTATGCTACCGTGAGTTCTC
<b><i>RhTAR1</i></b>	Fwd : TGTTTAGTCTGCCTGACTTTGCA Rev : GGCTGCGTCTCCTGTTGAA
<b><i>RhYUC1</i></b>	Fwd : CGGTTGGCTTCGGAAGAA Rev : TTGCTATCATAACGGCGTCGTA
<b><i>RhPIN1</i></b>	Fwd : ACCAAAGATCATAGCATGTG Rev : ACAAATGGGACAATTCCTTG
<b><i>RhPID</i></b>	Fwd : GGGCGTTCGATTACTTTTGA Rev : CTGGCAGGACTGTCTGAACA
<b><i>RhPP2A</i></b>	Fwd : TGTCACATGCATCAAAGGACAG Rev : GACGAATTGTCTTCTCCACCA
<b><i>RhSUS11</i></b>	Fwd : GCCTTGATCTTGGTGTGGAA Rev : TCGCAGGTGATTGTTGGATCT

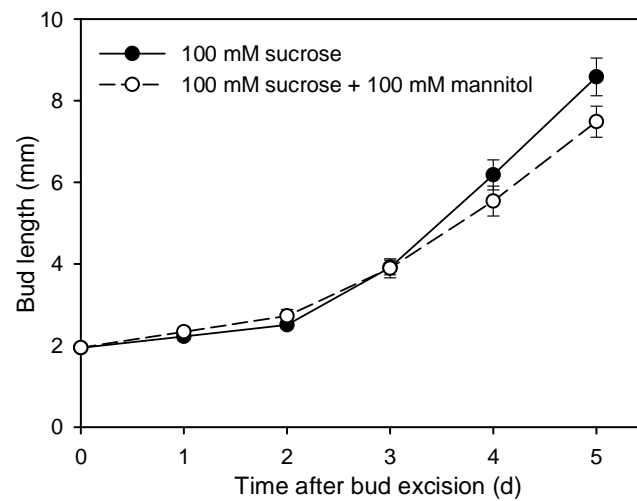
**Supplementary Table S3.** Primers used for quantitative real time PCR (Q-PCR).

## Supplementary Table S4

Sugar	Absorption	Absorption reduction (%)
Control	28,86	
10 mM sucrose	16,38	43,24
30 mM sucrose	9,57	66,84
10 mM turanose	27,83	3,57
30 mM turanose	25,36	12,13
10 mM lactulose	29,55	-2,39
30 mM lactulose	26,32	8,80
10 mM melibiose	20,99	13,34
30 mM melibiose	26,84	7,00

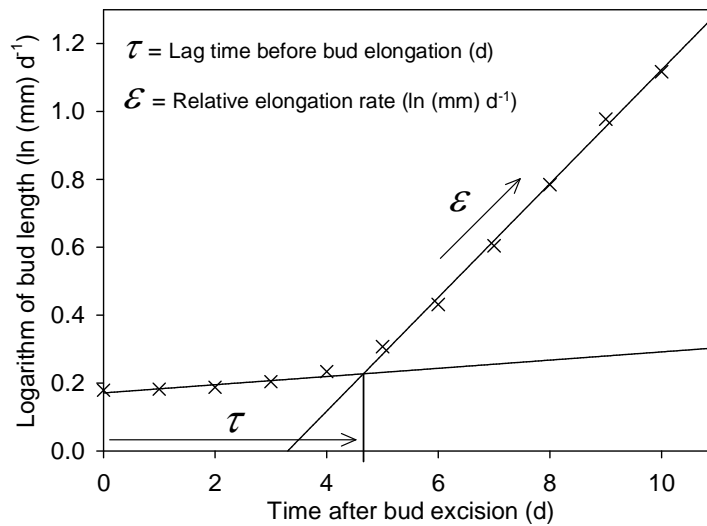
**Supplementary Table S4.** Sucrose uptake (3 mM <sup>14</sup>C-sucrose) was measured in yeast transformed with either pDR/RhSUC2 or pDR alone (background) in the presence of different sugars. Results were initially expressed as RhSUC2-dependent uptake (uptake by RhSUC2-expressing yeast minus uptake by pPDR-expressing yeast, in nmol min<sup>-1</sup> mg<sup>-1</sup> protein), and then converted into percentages of the control uptake in the absence of sugar (100% or 28,86 nmol min<sup>-1</sup> mg<sup>-1</sup> protein). Results are the means of 3 independent experiments with 4 replicates *per* experiment.

## Supplementary Figure S1



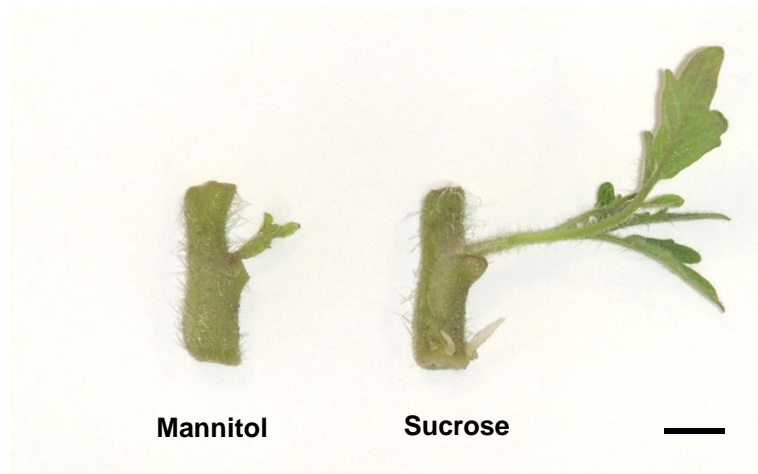
**Supplementary Figure S1.** Elongation time course of buds grown with 100 mM sucrose (black circles) or 100 mM sucrose combined with 100 mM mannitol (white circles). Data are the means  $\pm$  s.e. of n=10 replicates.

## Supplementary Figure S2



**Supplementary Figure S2.** Observed (crosses) and fitted (lines) elongation time courses for buds of nodal position 6 cultivated *in vitro* on classical MS medium after excision from *Rosa hybrida* primary axis at the FBV stage (time zero). Elongation is expressed in logarithmic scale.  $\tau$  is the lag time before rapid bud elongation and  $\epsilon$  is the relative elongation rate.  $\tau$  and  $\epsilon$  are used to characterize bud elongation.

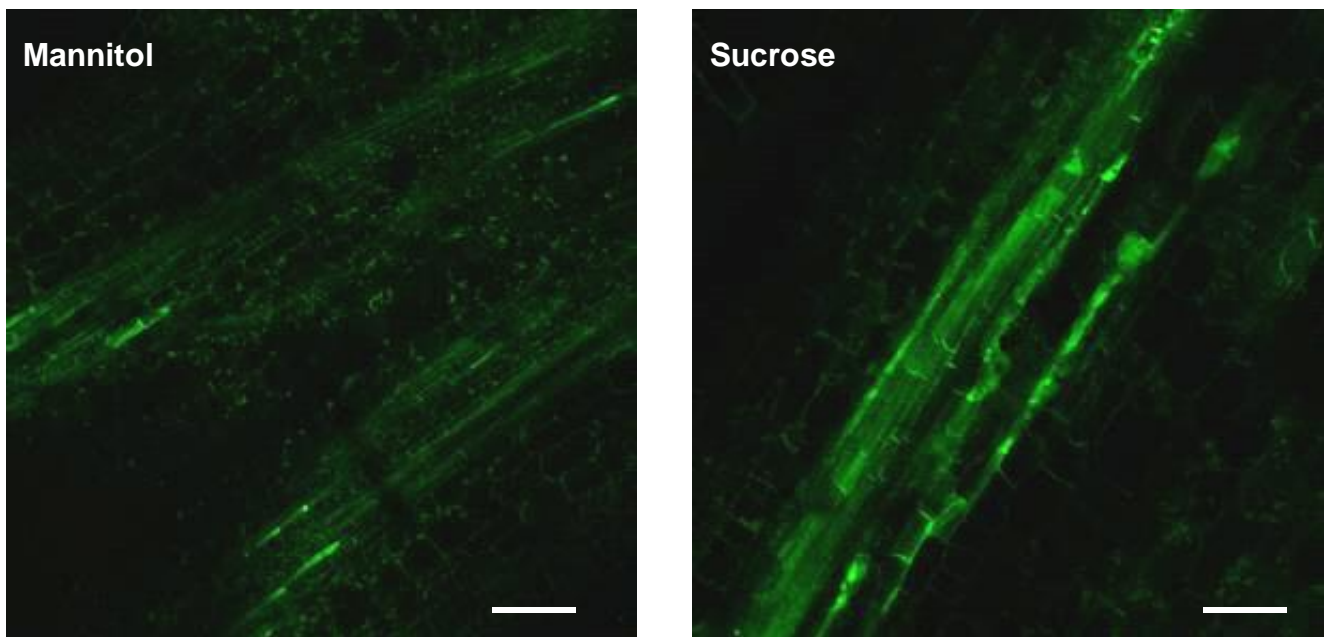
## Supplementary Figure S3



**Supplementary Figure S3.** Impact of 100 mM mannitol or sucrose after 8 days of treatment on bud elongation of *Solanum lycopersicum* Money Makerøcultivated *in vitro*. The picture is representative of 5 to 6 replicates. Scale bar: 1 cm.

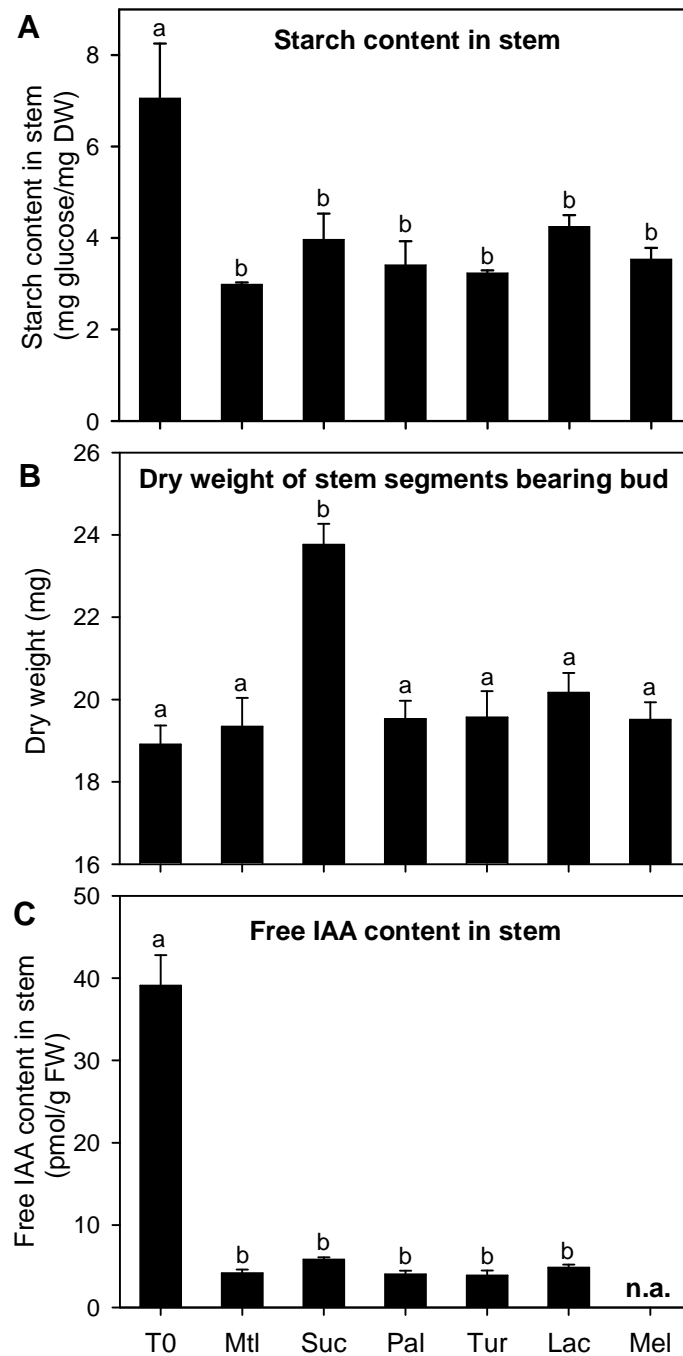


## Supplementary Figure S4



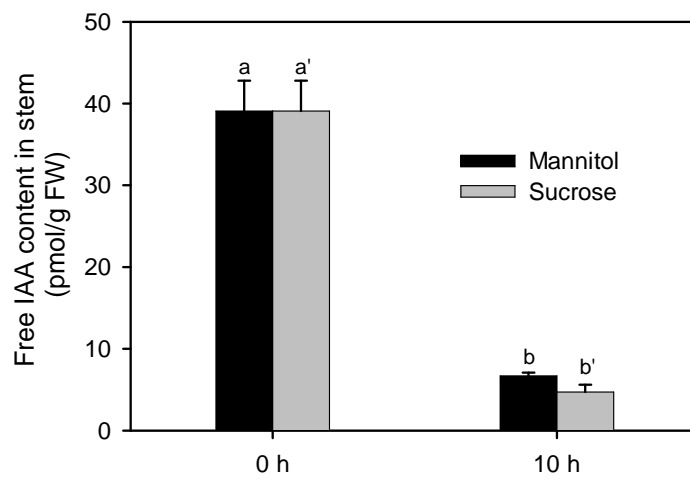
**Supplementary Figure S4.** PIN1 localization in bud stems of the *pPIN1::PIN1-GFP*-expressing tomato line grown 96 h with 100 mM mannitol (left) or sucrose (right). Micrographs are representative of 3 to 4 replicates. White bars represent 50 μm.

## Supplementary Figure S5



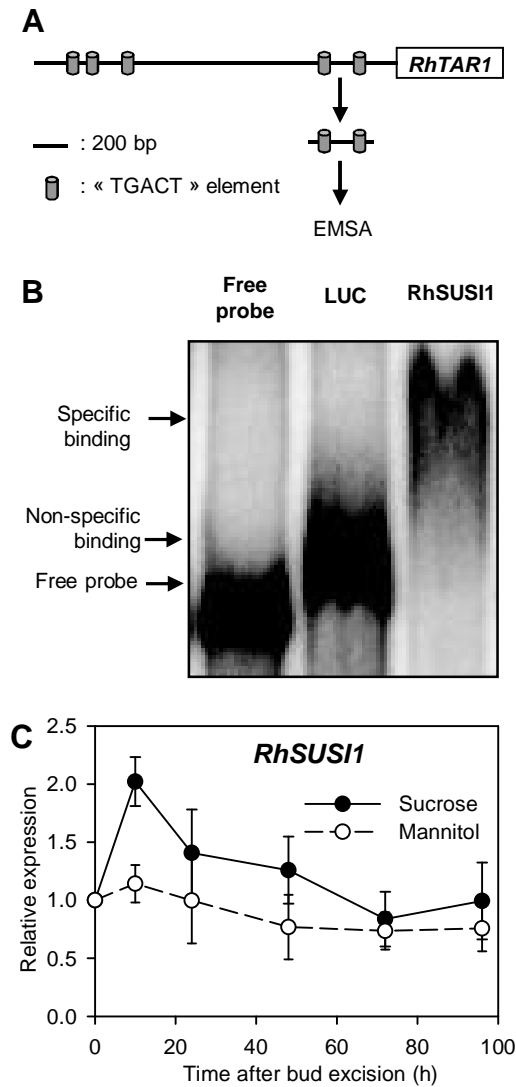
**Supplementary Figure S5.** (A) Stem starch content, (B) dry weight and (C) stem auxin content of bud-bearing stem segments grown for (A) 72 h or (B) and (C) 48 h with 80 mM mannitol (Mtl), sucrose (Suc), palatinose (Pal), turanose (Tur), lactulose (Lac) or melibiose (Mel). Data are the means  $\pm$  s.e. of  $n=9$  replicates for (A) and  $n=3$  replicates for (B) and (C). Letters indicate significant differences ( $\alpha = 0.05$ ). The statistical analysis in (B) and (C) separate the values at 48 h from those at T0.

## Supplementary Figure S6



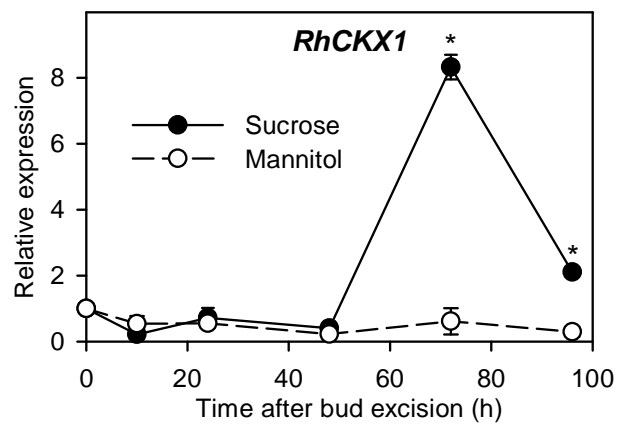
**Supplementary Figure S6.** Free IAA content in stem segments of intact plants before treatment (T0) or grown 10 h with 100 mM mannitol (black bar) or sucrose (gray bar). Data are the means  $\pm$  s.e. of  $n=3$  replicates. Letters indicate significant differences ( $\alpha = 0.05$ ).

## Supplementary Figure S7



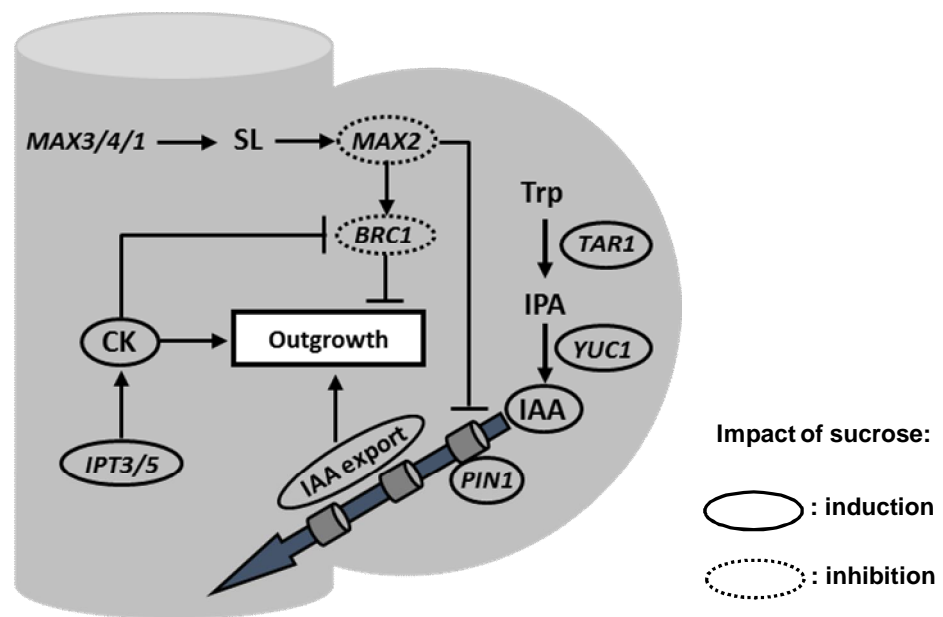
**Supplementary Figure S7.** (A) Localization of the  $\delta$ TGACT $\delta$  W-box elements on the promoter of *RhTAR1* and fragment used for EMSA. (B) EMSA performed on a native 387-bp fragment of *pRhTAR1*. Probe mobility was tested in the presence of LUCIFERASE (LUC) or RhSUS11 and compared to a free probe. (C) Relative expression of *RhSUS11* in buds grown with 100 mM mannitol or sucrose. Data are means  $\pm$  s.e. of  $n=3$  measurements on a pool of 60 buds.

## Supplementary Figure S8



**Supplementary Figure S8.** Relative expression of *RhCKX1* in nodal stems grown for 96 h with 100 mM mannitol or sucrose. Data are means  $\pm$  s.e. of  $n=3$  measurements on a pool of 60 buds. Asterisks indicate significant differences between sucrose and mannitol for each time-point.

## Supplementary Figure S9



**Supplementary Figure S9.** Schematic representation of the impact of sucrose on the hormonal regulation during bud release. Sucrose impacts the second messenger-based model by (i) up-regulating cytokinins (CK) synthesis through the induction of *IPT3/5* and (ii) down-regulating strigolactones (SL) signaling through the inhibition of *MAX2* and *BRC1*. Sucrose impacts the auxin canalization-based model by sucrose induces (i) auxin synthesis from tryptophan (Trp) and indole-3-pyruvate (IPA) by up-regulating *TAR1* and *YUC1/2* and (ii) auxin export from bud by up-regulating *PINI* polarization at the plasma membrane, finally leading to the establishment of an auxin transport canal between bud and stem. Ellipses in full line represent the actors up-regulated by sucrose and ellipses in dotted line represent those inhibited by sucrose.