

Differential expression of GS5 regulates grain size in rice

Chunjue Xu, Yu Liu, Yibo Li, Xiaodong Xu, Caiguo Xu, Xianghua Li, Jinghua Xiao and Qifa Zhang¹

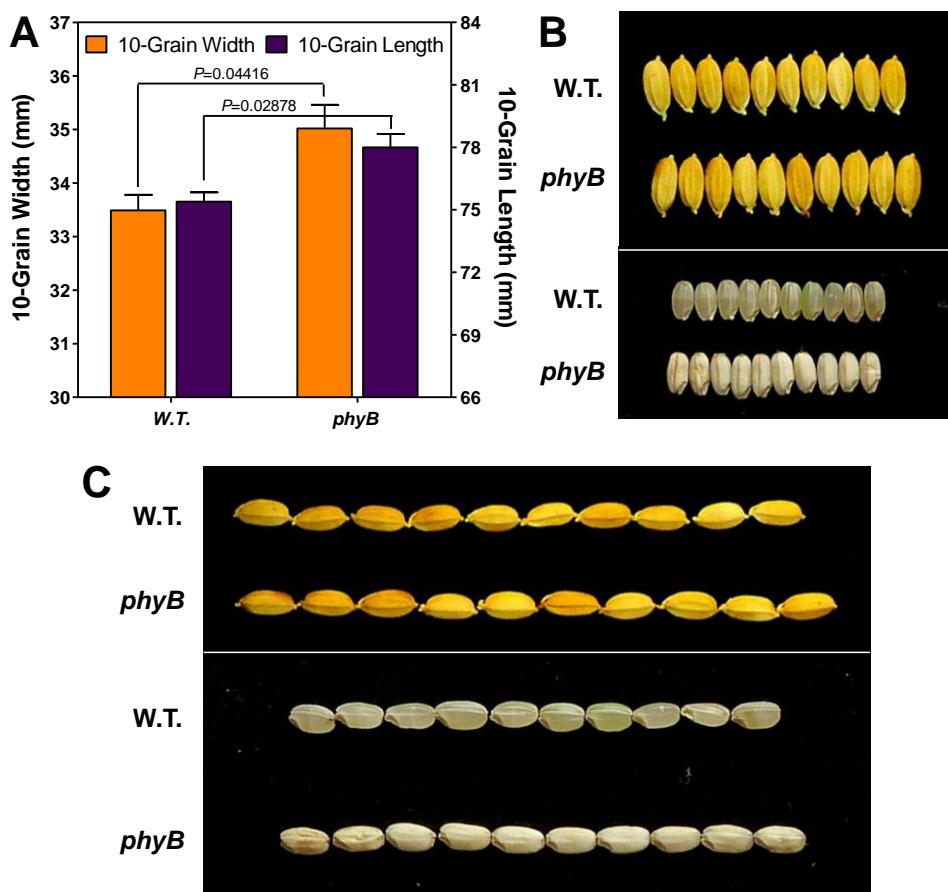


Figure S1. Grain size and chalkiness of *phyB* mutant. (A) Significant difference in grain size between *phyB* mutant and wild type (W.T.). All data are presented as mean \pm SE ($n \geq 3$). P values were calculated using Student's *t*-test. (B) Grain width and chalkiness of *phyB* mutant and wild type. (C) Grain length and chalkiness of *phyB* mutant and wild type.

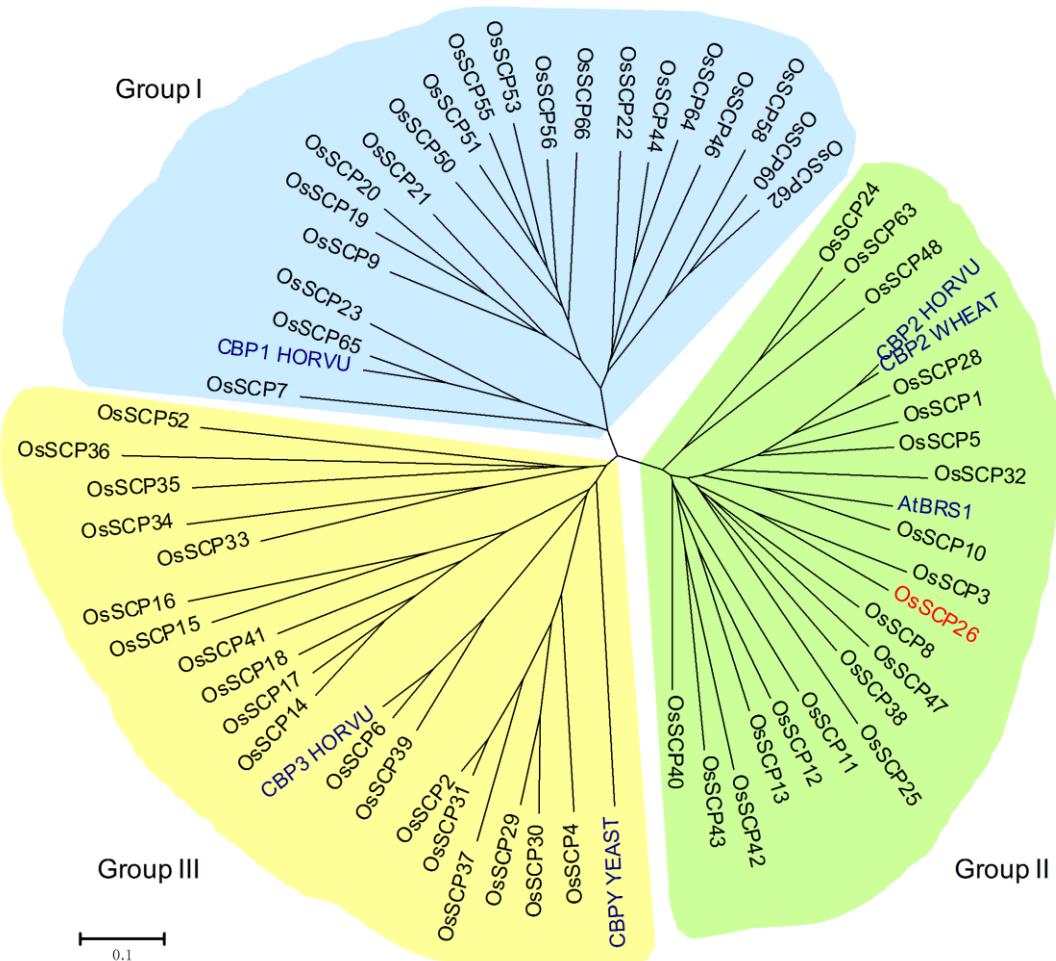


Figure S2. Phylogenetic tree of members of the serine carboxypeptidase-like protein family. Full-length protein sequences of OsSCPLs were downloaded from http://rice.plantbiology.msu.edu/ca/gene_fams/27_76.shtml and aligned with ClustalX and calculated with MEGA 3.1. *CBP1 HORVU* (NCBI accession NO. P07519), *CBP2 HORVU* (NCBI accession NO. P08818), *CBP3 HORVU* (NCBI accession NO. P21529) and *CBP2 WHEAT* (NCBI accession NO. P08819) were used to indicate the three groups of SCPL. *GS5* (*OsSCP26*) and *AtBRS1* (NCBI accession NO. NP_194790) belonging to SCPL group II; *CBPY YEAST* (NCBI accession NO. P00729) belongs to SCPL group III.

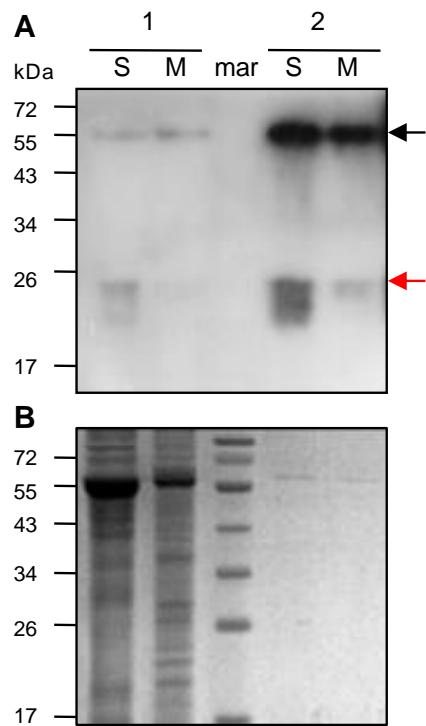


Figure S3. Assay of GS5-FLAG protein in *P_{Ubi}::GS5-FLAG* transgenic positive plants.

(A) Western blot assay of GS5-FLAG transgenic plants using anti-FLAG antibody showing the cleavage of GS5-FLAG in *P_{Ubi}::GS5-FLAG* transgenic positive plants into two chains (A and B). 1, Total protein extracted from transgenic positive plants; 2, Eluted proteins after anti-FLAG M2 agarose affinity purification; mar, Marker; S, Soluble protein sample; M, solubilized membrane protein sample; black arrow, full-length GS5-FLAG protein; red arrow, the C-terminal of GS5, B chain-FLAG peptide. (B) Coomassie Blue-stained protein samples of (A).

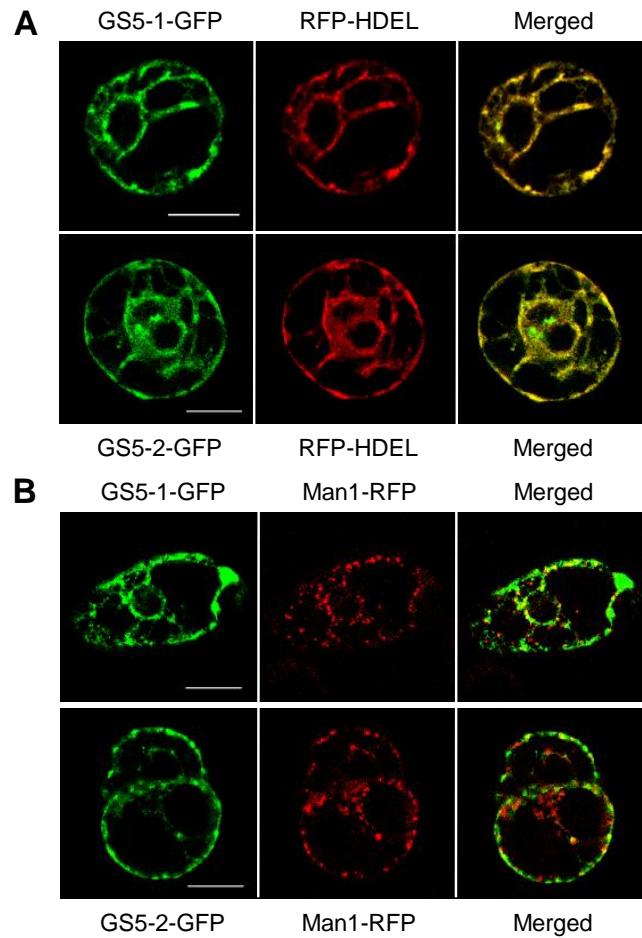
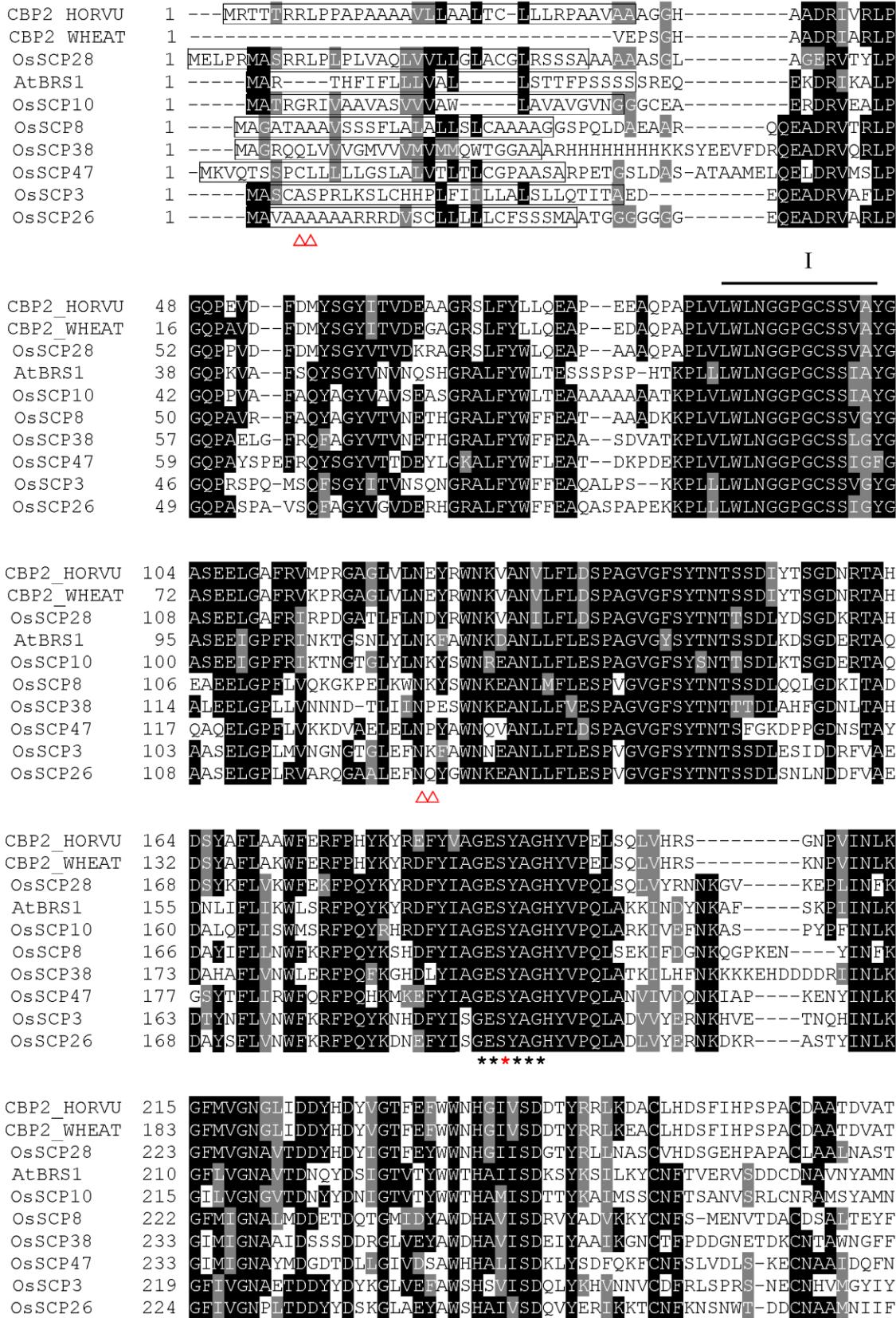
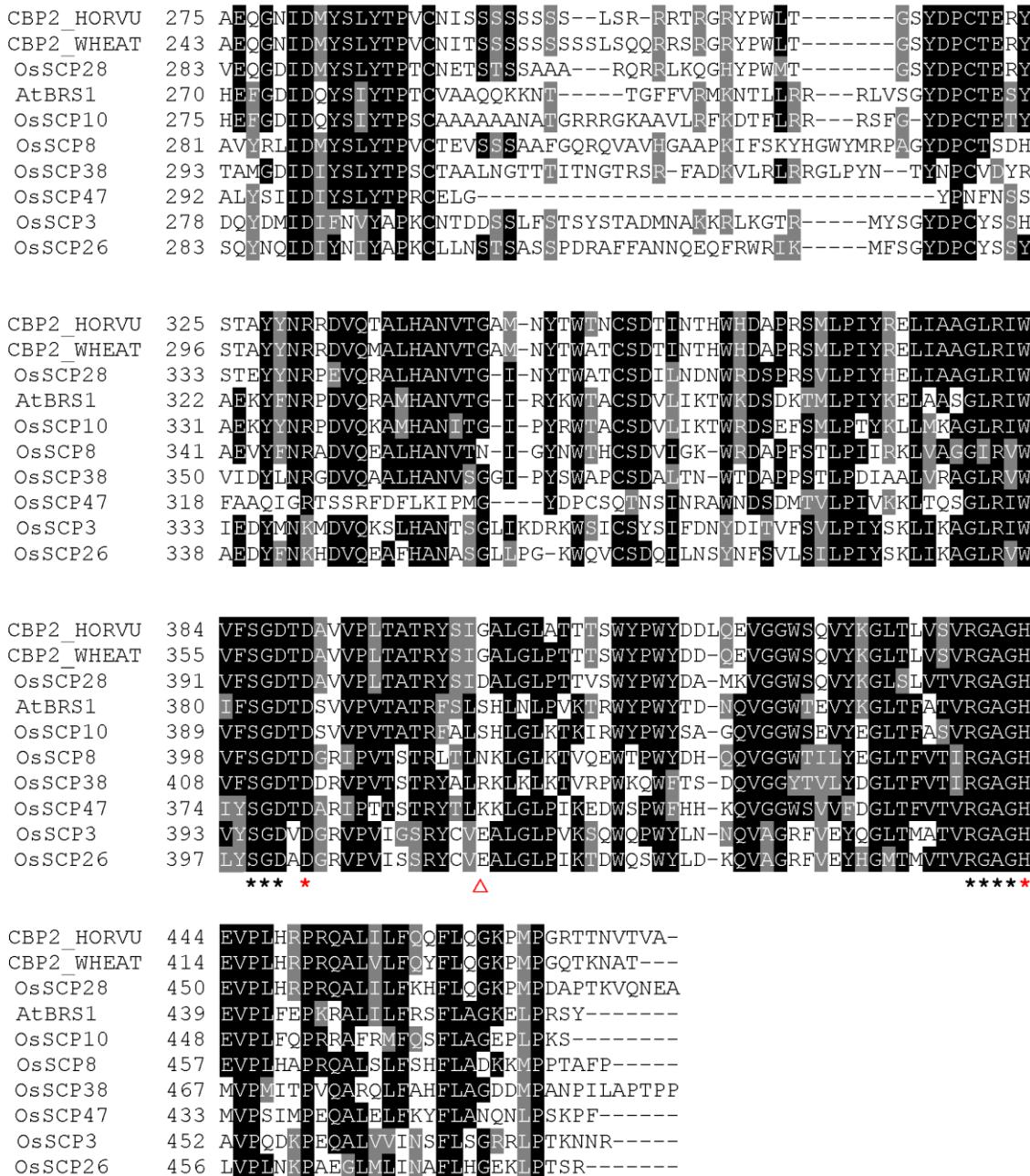


Figure S4. Co-localization assay of GS5 with endoplasmic reticulum marker inside the cell. (A) Co-expression of GS5-GFP and endoplasmic reticulum marker RFP-HDEL in tobacco BY-2 protoplasts showing co-localization of the two proteins. (B) Co-expression of GS5-GFP and *cis*-Golgi marker Man1-RFP in tobacco BY-2 protoplasts showing distinct localization of the two proteins. The experiments were repeated three times, yielding similar results, and representative images are shown. Scales bars = 20 μ m.



II

**Figure S5. Amino acid alignment of group II serine carboxypeptidase-like proteins.**

The boxed amino acids at the N terminus represent the putative signal peptide. I and II indicate the putative substrate binding region and linker peptide respectively. The A-chain is between signal peptide and linker peptide, and B-chain is downstream of the linker peptide. The red triangles indicate the different residues between GS5-1 and GS5-2 (OsSCP26). The red asterisks indicate the Ser-Asp-His catalytic triad of SCPL, and the black asterisks mark the residues essential for catalysis.

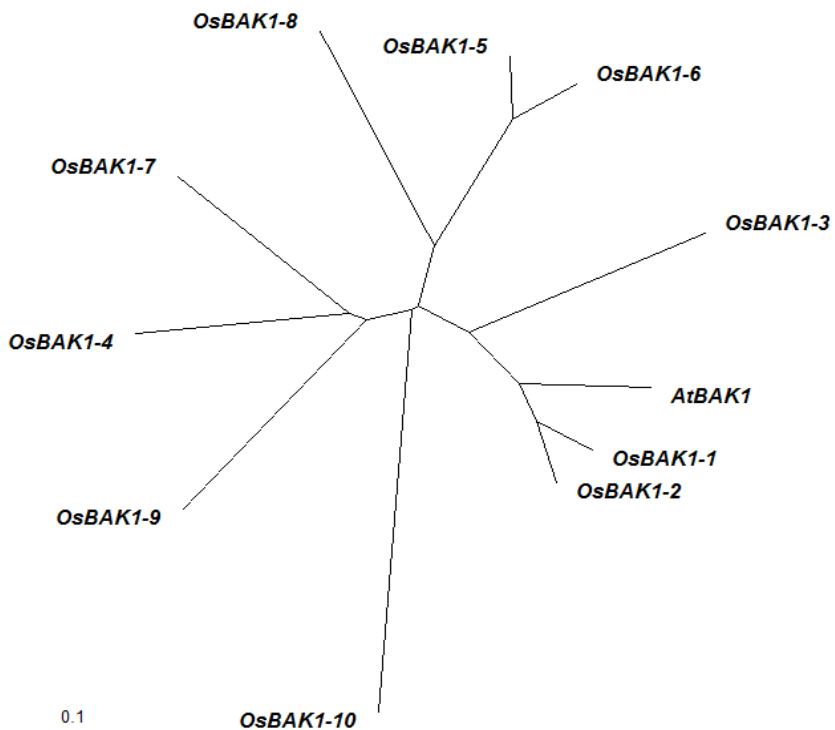


Figure S6. Phylogenetic tree of the rice BAK1 homologues. Full-length protein sequences of the rice BAK1 homologues were obtained by BLAST using the AtBAK1 (NCBI accession NO. NM_119497) sequence from <http://rice.plantbiology.msu.edu/index.shtml>, aligned with ClustalX and calculated with MEGA 3.1. OsBAK1-1, LOC_Os08g07760; OsBAK1-2, LOC_Os04g38480; OsBAK1-3, LOC_Os06g12120; OsBAK1-4, LOC_Os01g07630; OsBAK1-5, LOC_Os02g18320; OsBAK1-6, LOC_Os03g49620; OsBAK1-7, LOC_Os06g16330; OsBAK1-8, LOC_Os11g39370; OsBAK1-9, LOC_Os02g14120; and OsBAK1-10, LOC_Os03g16010.

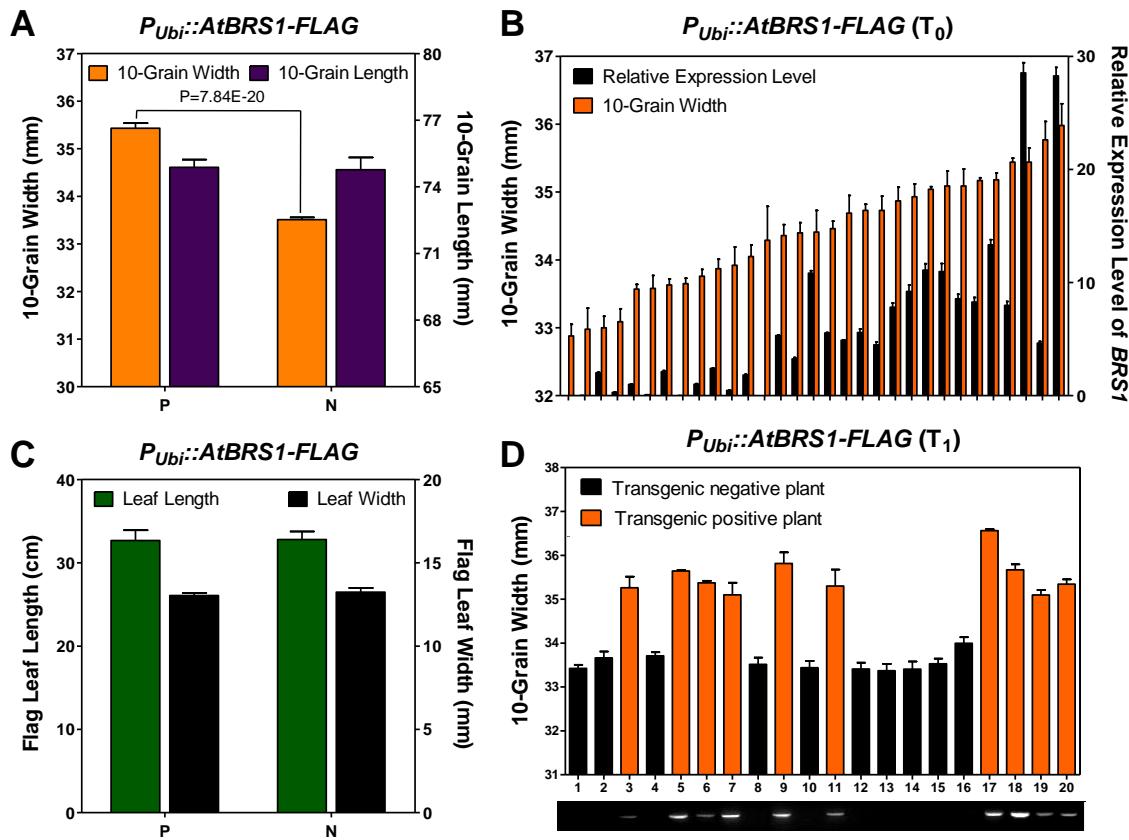


Figure S7. Grain size and leaf size of *P_{Ubi}::AtBRS1-FLAG* transgenic plants. (A) Grain width and grain length of T1 transgenic rice plants overexpressing *AtBRS1*. P, transgenic positive plants; N, transgenic negative plants. (B) Co-segregation of grain width with expression level of *AtBRS1* in T0 transgenic rice plants. (C) Flag leaf width and length of T1 transgenic rice plants overexpressing *AtBRS1*. P, transgenic positive plants; N, transgenic negative plants. (D) Co-segregation of grain width with *AtBRS1* genotype in T1 transgenic rice plants. All data are presented as mean \pm SE ($n \geq 3$). *P* values were calculated using Student's *t*-test.

Table S1. Polymorphisms and *cis* -acting elements in the 2-kb promoter of GS5 .

Position	SNPs/InDels		Sequence	<i>Cis</i> -acting elements	Organism	Function
	GS5-1	GS5-2				
-1961	/	/	CAANNNNATC	circadian	<i>Lycopersicon esculentum</i>	Circadian control
-1723	/	/	ATTAAT	Box4	<i>Petroselinum crispum</i>	Light responsiveness
-1656	/	/	AATCTAATCT	ATCT-motif	<i>Arabidopsis thaliana</i>	Light responsiveness
-1630	/	/	CAACTG	MBS E-box	<i>Arabidopsis thaliana</i>	MYB binding site, drought-inducibility Brassinosteroid responsiveness
-1620	/	/	GGAGATG	GAG-motif	<i>Hordeum vulgare</i>	part of a light responsive element
-1595	C	T	/	/	/	/
-1539	/	/	CATATG	E-box ABRE	<i>Arabidopsis thaliana</i>	Brassinosteroid responsiveness Abscisic acid responsiveness
-1518	/	/	CACGTG	G-box E-box	<i>Arabidopsis thaliana</i>	Light responsiveness Brassinosteroid responsiveness
-1514	/	/	TGACG	TGACG-motif	<i>Hordeum vulgare</i>	MeJA-responsiveness
-1489	/	/	CCGTCC	A-box CCGTCC-box	<i>Petroselinum crispum</i> <i>Arabidopsis thaliana</i>	<i>cis</i> -acting regulatory element Meristem specific activation
-1476	-	A	/	/	/	/
-1466	T	A	/	/	/	/
-1464	T	G	/	/	/	/
-1463	A	G	/	/	/	/
-1391	G	A	/	/	/	/
-1376	/	/	ATTAAT	Box4	<i>Petroselinum crispum</i>	Light responsiveness
-1345	/	/	ATTTCCTTCA	TC-rich repeats	<i>Nicotiana tabacum</i>	Defense and stress responsiveness
-1314	/	/	CGGTCA	MBS	<i>Zea mays</i>	MYB Binding Site
-1303	A	G	/	/	/	/
-1281	T	-	/	/	/	/
-1242	/	/	TGACG	TGACG-motif	<i>Hordeum vulgare</i>	MeJA-responsiveness
-1223	/	/	CACTTG	E-box	<i>Arabidopsis thaliana</i>	Brassinosteroid responsiveness
-1218	/	/	GTCAT	Skn-1-motif	<i>Oryza sativa</i>	Endosperm expression
-1204	/	/	CCG/ACCC	Spl	<i>Zea mays</i>	Light responsive element
-1164	/	/	CACTTG	E-box	<i>Arabidopsis thaliana</i>	Brassinosteroid responsiveness
-1159	/	/	GTCAT	Skn-1-motif	<i>Oryza sativa</i>	Endosperm expression
-1142	/	/	CATTTG	E-box	<i>Arabidopsis thaliana</i>	Brassinosteroid responsiveness
-1109	C	T	/	/	/	/
-1096	/	/	CAATTG	E-box	<i>Arabidopsis thaliana</i>	Brassinosteroid responsiveness
-1088	/	/	AAACAGA	GARE-motif	<i>Brassica oleracea</i>	Gibberellin-responsive element
-1076	A	G	/	/	/	/
-1032	A	T	/	/	/	/
-1015	G	T	/	/	/	/

Continuing Table S1

-1013	/	/	CAGATG	E-box	<i>Arabidopsis thaliana</i>	Brassinosteroid responsiveness
-895	G	GATAGGA	GATA-motif	<i>Arabidopsis thaliana</i>	part of a light responsive element	
	A	GATAAGATT	I-box	<i>Arabidopsis thaliana</i>	part of a light responsive element	
-879	/	/	TTTCAAA	BoxI	<i>Pisum sativum</i>	Light responsive element
-847	T	-	/	/	/	/
-825	T	TCTTAC	TCT-motif	<i>Arabidopsis thaliana</i>	part of a light responsive element	
	C	/	/	/	/	/
-788	C	CCG/ACCC	Spl	<i>Zea mays</i>	Light responsive element	
	A	/	/	/	/	/
-756	/	/	ATTAAT	Box4	<i>Petroselinum crispum</i>	Light responsiveness
-710	/	/	GTCAT	Skn-1-motif	<i>Oryza sativa</i>	Endosperm expression
-671	G	TGACG	TGACG-motif	<i>Hordeum vulgare</i>	MeJA-responsiveness	
	A	/	/	/	/	/
-409	/	/	AAACAGA	GARE-motif	<i>Brassica oleracea</i>	Gibberellin-responsive element
-375	/	/	TAACTG	MBS	<i>Arabidopsis thaliana</i>	MYB binding site, drought-inducibility
-371	/	/	TGAGTCA	GCN4-motif	<i>Oryza sativa</i>	Endosperm expression
-368	/	/	GTCAT	Skn-1-motif	<i>Oryza sativa</i>	Endosperm expression
-326	-	C	/	/	/	/
-325	-	A	/	/	/	/
-324	-	A	/	/	/	/
-323	-	A	/	/	/	/
-225	/	/	ATTTCTTCA	TC-rich repeats	<i>Nicotiana tabacum</i>	Defense and stress responsiveness
-100	/	/	TTTCTTCTCT	5'UTR Py-rich stretch	<i>Lycopersicon esculentum</i>	Conferring high transcription levels

-, single nucleotide deletion; /, not detected.

Table S2. Primers used in this work.

Name	Sequence
Expression analysis	
GS5qF	AGTGGACTGCTTCAGGGAAG
GS5qR	CACGCAGTACCGAGAACTGA
Act1F	TGCTATGTACGTGCCATCCAG
Act1R	AATGAGTAACCACGCTCCGTCA
Promoter constructs	
PAF	AAGCTTCGTATGGAATTACCTAGCTATGCA
PBF	AAGCTTTGACGAGAGGCAAGGACCCA
PCF	AAGCTTGATGGGAGAGATAGATAGACTC
PDF	AAGCTTGCAACCCGGATCTGGACAGA
PDR	GGATCCTGCTAACCGTGGTGCCGAGCTAGC
-1109mutF	ACACAGGATTTGACAGGAATAC
-1109mutR	GTATTCTGTCAAATCCTGTGT
-1032mutF	TGATTGGACCACTGAAAAAAC
-1032mutR	GTTTTTCAGTGGTCCAATCA
-825mutF	CTCCAAAATTCTTACATTTTCT
-825mutR	AGAAAAAAATGTAAGAATTTGGAG
B_GAmutF	ACAATTGTAACCAAGAGGATTGTA
B_GAmutR	TACAAATCCTCTTGGTTACAATTGT
C_LIGHTmutF	CTCCAAAATCCTTACATTTTTC
C_LIGHTmutR	GAAAAAAATGTAAGGATTGGAG
cDNA constructs	
GS5cDNAF	GGTACCTAGCTAGAGCTGGCACCA CGGGTAGCAA
GS5cDNAR	GGATCCTCTGTTGCGAACGTTCTACCATGAAG
OsBAK1-7cDNAF	GGTACCTACCTGAGCTCAGCAATGGA
OsBAK1-7cDNAR	GTAACTCTGGGCCGGAGAGCTC
OsMSBP1cDNAF	CTCGAGATGGCGGGCGGTGGCGGA
OsMSBP1cDNAR	GGATCCACTTCTTGGCGCCTCATCAGGTGCA
GS5AchainF	GAATTGGGATCCATATGGAGCAGGAGGCTGACAGGGT
GS5AchainR	CTGCAGCTCGAGGCCGCTTAGAAGGCAC TTTGGGCA
GS5BchainF	GAATTGGGATCCATATGTTTCAGGCTATGATCCATGC
GS5BchainR	CTGCAGCTCGAGGCCGCTTGCTTGTGGAAGCTT
OsBAK1-4LRRF	ACTAGTATGGAATTCAACTACGAAGTGGGGCGCTGA
OsBAK1-4LRRR	CTCGAGGATATCATCTCCAGACTGACACCACAGATC
OsBAK1-7LRRF	ACTAGTATGGAATTCAACAACGAAGTGAAGCTCTGATTG
OsBAK1-7LRRR	CGGGCCGGGATCCATCCTGCTCCCTGTTGGCGTC
OsBAK1-9LRRF	ACTAGTATGGAATTCAACTACGAAGTGGCGCGCTGATG
OsBAK1-9LRRR	CTCGAGGGATCCGAACCATGCATAATTGATGAATTGC
OsBAK1-4KDF	GGATCCCATATGAGGCGCAATCAGCAGATCTT
OsBAK1-4KDR	ACTAGTGAATTCCCTCGGTCCAGAGAGCTCCA
OsBAK1-7KDF	GGATCCCATATGCGCCGCAACCGGCAGATCCT
OsBAK1-7KDR	ACTAGTGAATTCTCTGGGCCGGAGAGCTCGA
OsBAK1-9KDF	CATATGGAATTCTGCAGATGGCGCTTGCTTT
OsBAK1-9KDR	ACTAGTCTCGAGTCTGGGACCAGATAGCTCAATTGG