

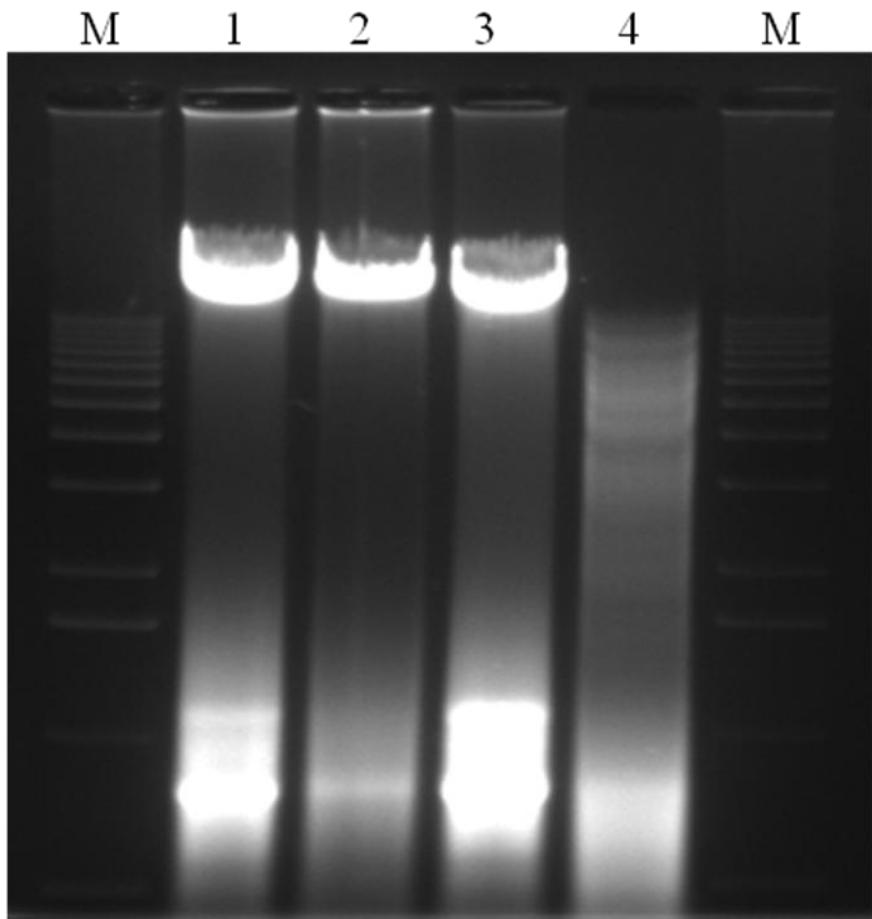
## Supplementary Figures and Tables

**Supplementary Table S1** Up-regulated and down-regulated genes in the hemH1 deletion mutant as compared to the parental strain of *Shewanella loihica* PV-4

Function	Gene	Log2R	Z-score	Annotation
Iron uptake	Shew_3097	7.29	9.42	TonB-dependent siderophore receptor
	Shew_0861	7.04	7.67	extracellular solute-binding protein
	Shew_3841	7.69	5.15	TonB-dependent siderophore receptor
	Shew_3166	6.02	7.3	extracellular solute-binding protein
	Shew_1181	5.87	7.26	putative iron regulated membrane protein
	Shew_2517	4.45	5.46	ferrous iron transport protein B
	Shew_2518	4.19	4.18	ferrous iron transport protein A
Cytochrome c	Shew_0552	4.47	3.11	Bacterioferritin associated ferrodoxin
	Shew_3112	-3.77	-5.09	flavocytochrome c
	Shew_0572	-3.51	-2.53	cytochrome c1
	Shew_2526	-3.19	-4.51	cytochrome c family protein
	Shew_2525	-2.98	-3.85	decaheme cytochrome c
PPIX and heme biosynthesis	Shew_0918	-6.75	-5.25	decaheme cytochrome c
	Shew_0678	-4.21	-7.71	protoporphyrinogen oxidase (hemG)
	Shew_0073	-4.38	-4.16	coproporphyrinogen III oxidase (hemN)
	Shew_2913	-3.54	-4.5	Glutamyl-tRNA reductase (hemA)
Flagella synthesis	Shew_3382	-3.4	-3.17	ALA dehydratase (hemB)
	Shew_1345	-7.20	-6.69	flagellar basal body rod protein FlgB
	Shew_1346	-6.76	-7.3	flagellar basal body rod protein FlgC
	Shew_1347	-6.73	-7.75	flagellar basal body rod modification protein
	Shew_1348	-6.12	-6.47	flagellar hook protein FlgE
	Shew_1350	-5.20	-7.19	flagellar basal body rod protein FlgG
	Shew_1352	-5.08	-7.69	flagellar basal body P-ring protein
	Shew_1354	-6.26	-8.17	flagellar hook-associated protein FlgK
	Shew_1355	-4.22	-5.58	flagellar hook-associated protein FlgL
	Shew_1356	-5.79	-3.96	flagellin domain-containing protein

**Supplemental Table S2** The presence of *hemH1*, *hemH2*, *rpoE2*, *chrR* and *pgpD* orthologs in sequenced *Shewanella* genomes

Strains	Genes				
	<i>chrR</i>	<i>rpoE2</i>	<i>pgpD</i>	<i>hemH2</i>	<i>hemH1</i>
<i>S. oneidensis</i>	SO_1985	SO_1986	SO_3348	SO_3349	SO_2019
<i>S. denitrificans</i>	Sden_3373	Sden_3374	Sden_2679	Sden_2678	
<i>S. frigidimarina</i>	Sfri_2319	Sfri_2318	Sfri_2853	Sfri_2852	
<i>S. amazonensis</i>	Sama_2037	Sama_2036	Sama_2474		Sama_1316
<i>S. baltica OS155</i>	Sbal_2598	Sbal_2597	Sbal_3022	Sbal_3021	Sbal_2545
<i>S. baltica OS185</i>	Shew185_2637	Shew185_2636	Shew185_3037	Shew185_3036	Shew185_2583
<i>S. baltica OS195</i>	Sbal195_2712	Sbal195_2711	Sbal195_3180	Sbal195_3179	Sbal195_2660
<i>S. baltica OS223</i>	Sbal223_1748	Sbal223_1749	Sbal223_1341	Sbal223_1342	Sbal223_1801
<i>S. baltica OS678</i>	Sbal678_2717	Sbal678_2716	Sbal678_3186	Sbal678_3185	Sbal678_2663
<i>S. baltica OS117</i>	Sbal117_2734	Sbal175_1733	Sbal175_1323	Sbal175_1324	Sbal117_2682
<i>S. baltica BA175</i>	Sbal175_1732	Sbal175_1733	Sbal175_1323	Sbal175_1324	Sbal175_1783
<i>S. loihica</i>	Shew_1477	Shew_1476		Shew_2741	Shew_2229
<i>S. putrefaciens CN-32</i>	Sputcn32_2323	Sputcn32_2322	Sputcn32_2684	Sputcn32_2683	Sputcn32_2296
<i>S. putrefaciens 200</i>	Sput200_2349	Sput200_2348	Sput200_2821	Sput200_2820	Sput200_2322
<i>S. sediminis</i>				Ssed_1235	Ssed_2846
<i>S. pealeana</i>				Spea_1130	Spea_1529
<i>S. sp. MR-4</i>	Shewmr4_2276	Shewmr4_2275	Shewmr4_1196	Shewmr4_1197	Shewmr4_2248
<i>S. sp. MR-7</i>	Shewmr7_2348	Shewmr7_2347	Shewmr7_1267	Shewmr7_1268	Shewmr7_2320
<i>S. sp. ANA-3</i>	Shewana3_2466	Shewana3_2465	Shewana3_1197	Shewana3_1198	Shewana3_2440
<i>S. sp. W3-18-1</i>	Sputw3181_1685	Sputw3181_1686	Sputw3181_1327	Sputw3181_1328	Sputw3181_1712
<i>S. halifaxensis</i>				Shal_1175	Shal_1599
<i>S. woodyi</i>	Swoo_2779	Swoo_2780		Swoo_1335	
<i>S. piezotolerans</i>				swp_3670	swp_1736
<i>S. violacea</i>					SVI_2775



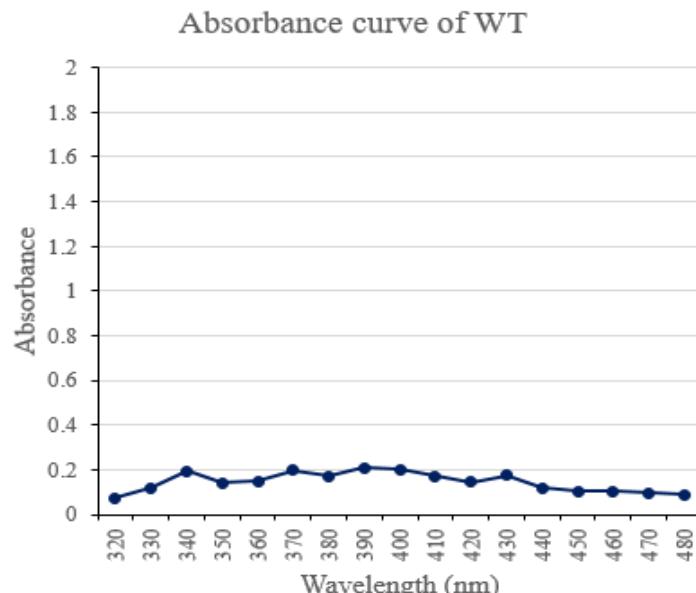
**Supplementary Figure S1** PstI restriction digestion of chromosomal DNAs of the wild-type PV-4 strain (lane 2), and PV-4 $\Delta$ *pstI* $\Delta$ *pstM* strain (lane 4). M: 1 kb DNA markers; lanes 1 and 3 are the undigested chromosomal DNAs used as control. The chromosomal DNA of the wild-type PV-4 strain could not be digested by a commercial PstI endonuclease probably due to methylation of recognition sites. To facilitate genetic manipulation, we deleted the putative PstI-like endonuclease (designated PstI, encoded by the Shew\_0993 locus) and DNA methylase (designated PstM, Shew\_0992) genes in the PV-4 strain, resulting in the successful digestion of chromosomal DNA of the resultant PV-4 $\Delta$ *pstI* $\Delta$ *pstM* (used as the parental strain of PV-4 hereafter) by the PstI endonuclease. These results confirmed that PstM mediated methylation of chromosomal DNA of PV-4.

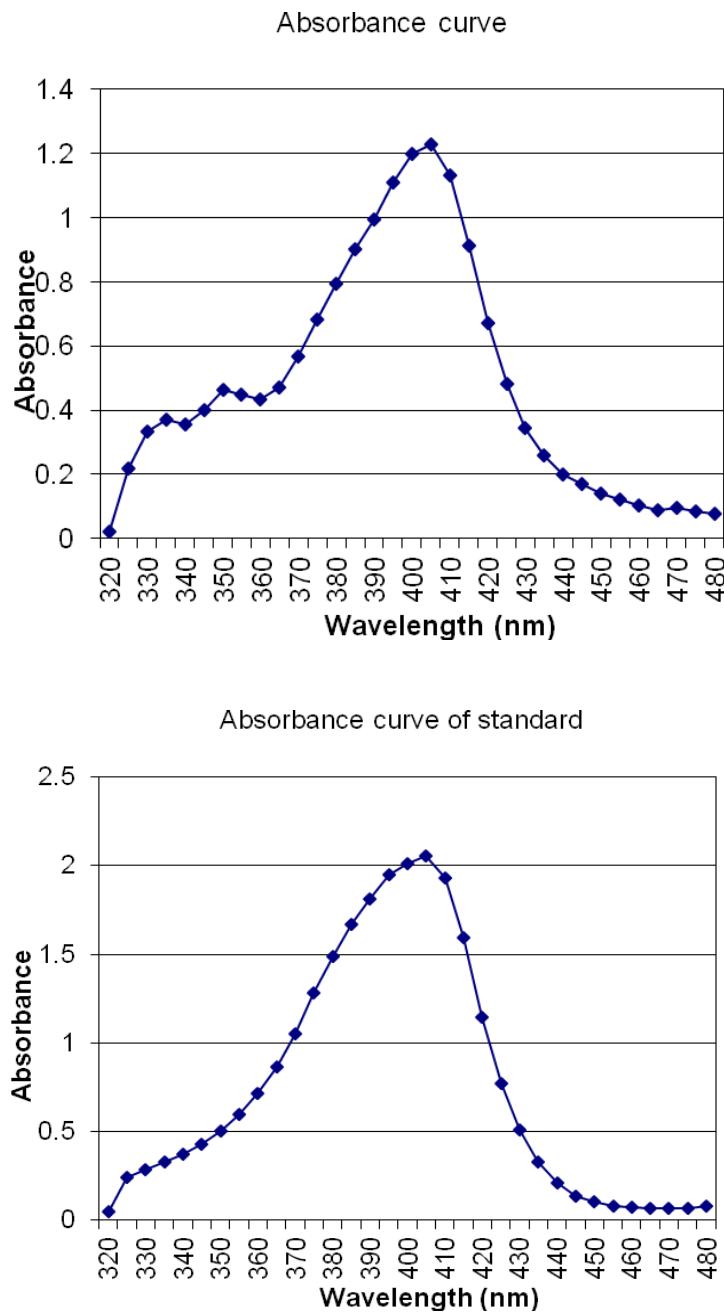
Start codon

TTGAATAAAGTCGACAAC **TAT**CTCCGCTAACGCTGGTCTGCTGGTAATCTCGG  
CACACCCGATACACCAACGCCTAAAGATGTAAGCAGTTCTAAAACAGTTCTCAGC  
GACCCGCGCTGGTCATCTAAACCTGGATCTGGAAACCCATACTCAATGGCATCAT  
ATTAAATACGCGCCCCAAGGCCGTAGCCAAGCTCTATGAGTCTATCTGGTGGCCCGAGG  
GCTCGCCTCTGATGGTAGTCAGTGAGGCCAACGTGAAGCGCTGAGTCGATTCTAAA  
AGCGCGTCATGGTAGCGATATTCCGGTGGAGTTGGGGATGAGTTATGGCAATCCCTCAC  
TGTCGTCGGGGATAGACAAGCTGGTGGCTCAGGGTGGTGAAGCGCCTGGTGGTGCCTGCC  
GCTATATCCTCAGTATTCCCTGCTCCACCGTGGCACCGTATTGATGCCATGCCAGTGA  
CTATAAGGGCAGACGCAACTATCCTGAGACCCGCTCAGTAAAGAGTATTCGAGCAC  
CCGGCCTATATCGCGCGCTGGCAGGTTCTGTGCGCGCTCATGGCAAGATAAGGGCC  
AGGGCGATTGCCTCTTGAATGTCCTTCATGGCGTCCCCCTGCGT **TA**TGTGACCGAGGGT  
GACCCCTATCAGCGTCAGTGTCAACGTACTGCCAGCTGCTGGCCGGCCTGGGCT  
TTACAGAGTCTCAGTGGCGCCTGTGTTCAATCGAAATTGGTAAGGAGGAGTGGCT  
GACGCCGGCAACGGATGCGCTGCTCGAGAGTTGCCGGTAAGGGGTTAACGCGCT  
GGATATTCTCTGTCCGGCCTTGCCGTGGATTGTTGGAGACGCTTAAGAGATCTCCA  
TTGGCGGCAAGGAGAGCTTATCGAGGCCGGCGGTAGGACTATCATTCAATCCCTGT  
TTGAACGAGGATGAAGCGCATATGCAGCTGCTCGCGGACTGGTCGATCAACAGGCGG  
CGGGCTGGCTTAAATCTTAA

Stop codon

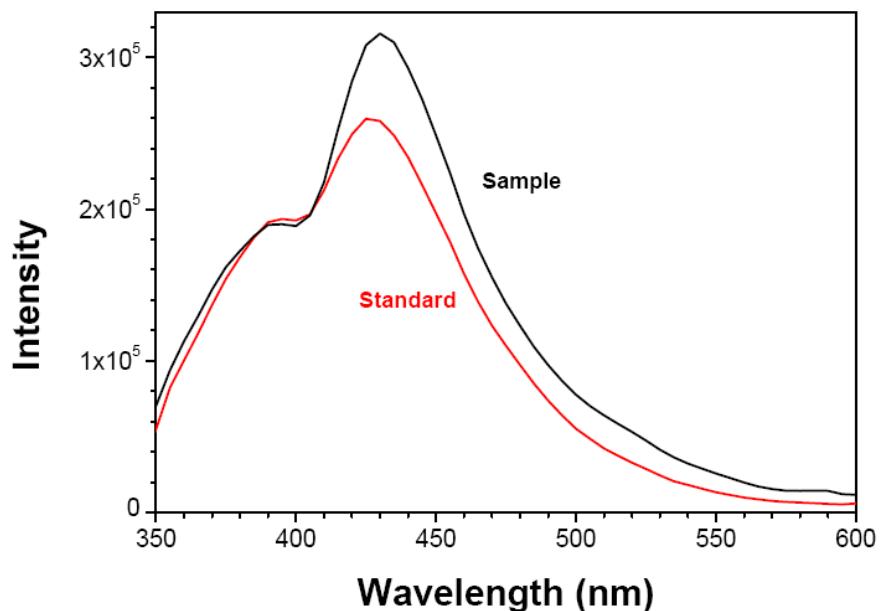
**Supplementary Figure S2** Mapping of insertional sites of transposon in the red pigment-overproducing transposon mutants of *Shewanella loihica* PV-4. The open reading frame of *hemH1* (locus Shew\_2229) is shown. The *mariner* transposon recognizes the dinucleotide TA for insertion and the insertional sites of two transposon insertional mutants are highlighted.



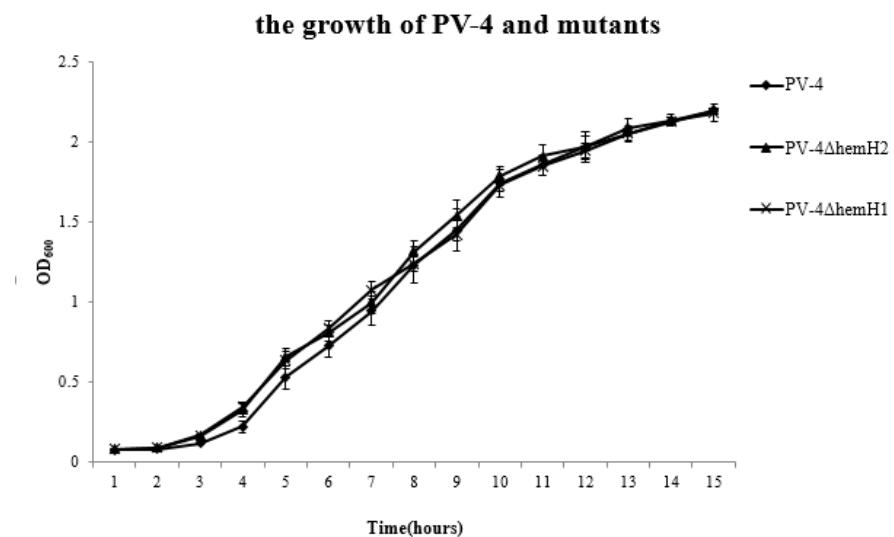


**Supplementary Figure S3** Ultraviolet-visible spectrograms of the bacterial extract (the wildtype and the *hemH1* mutant) and PPIX standard. Absorbance was measured at every 5 or 10 nanometers (nm).

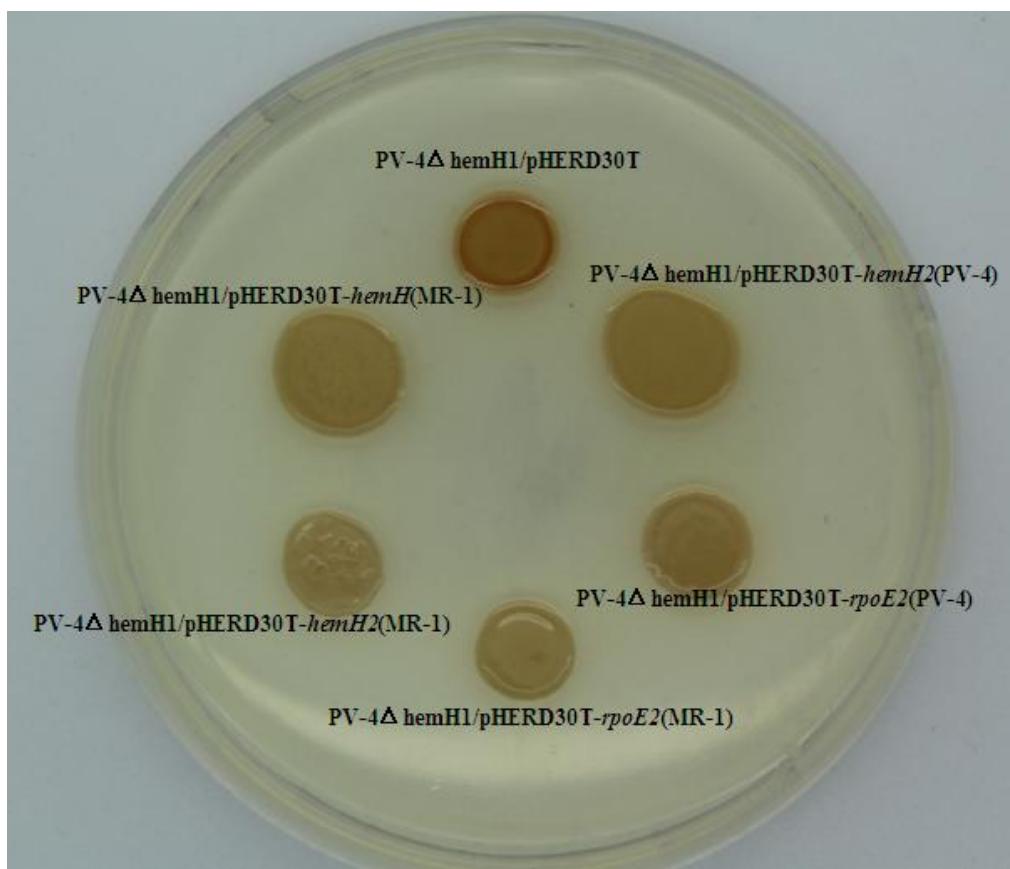
### Fluorescence Spectra of PPIX



**Supplementary Figure S4** The fluorescence spectra of the PPIX standard and the extracted sample showing their similarity.

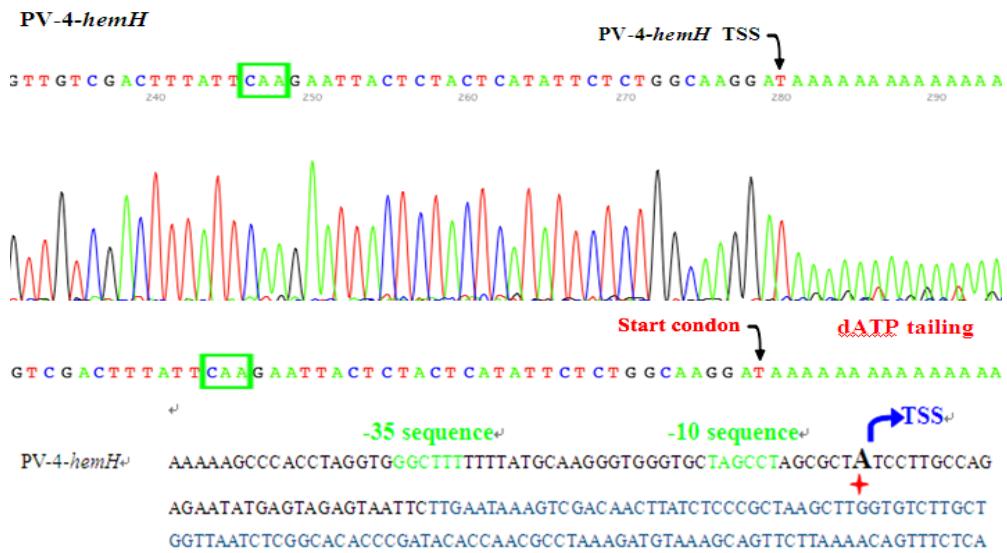


**Supplementary Figure S5** Bacterial growth (optical density at 600 nm) of PV-4 and the PV-4ΔhemH1 and PV-4ΔhemH2 mutants in the LB broth. Culture tubes were incubated at 28 °C, 220n/min.



**Supplementary Figure S6** Overexpression of plasmid borne *hemH1*, *hemH2* or *rpoE2* genes from PV-4 and MR-1 suppressed the PPIX-overproducing phenotype of the PV4 $\Delta$ *hemH1* strain.

(a)

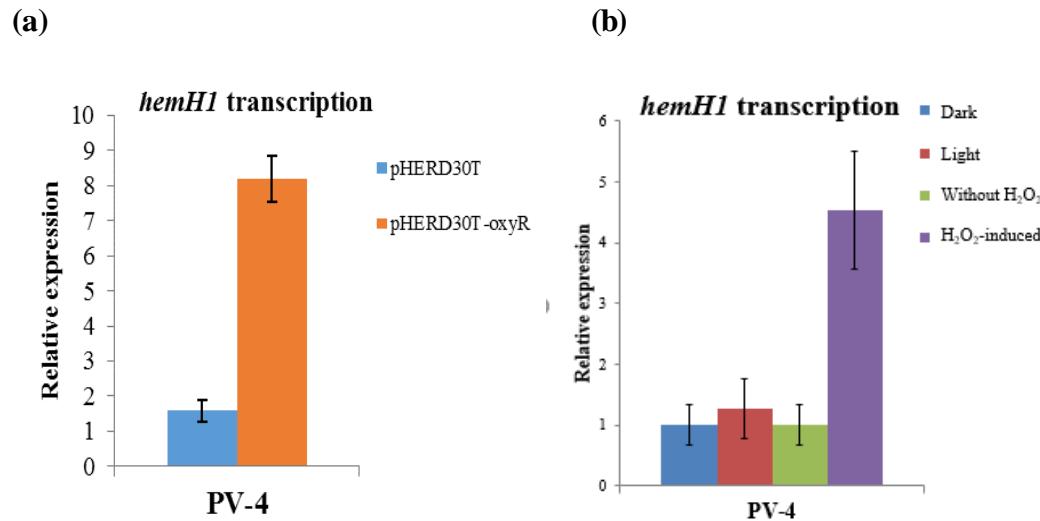


(b)

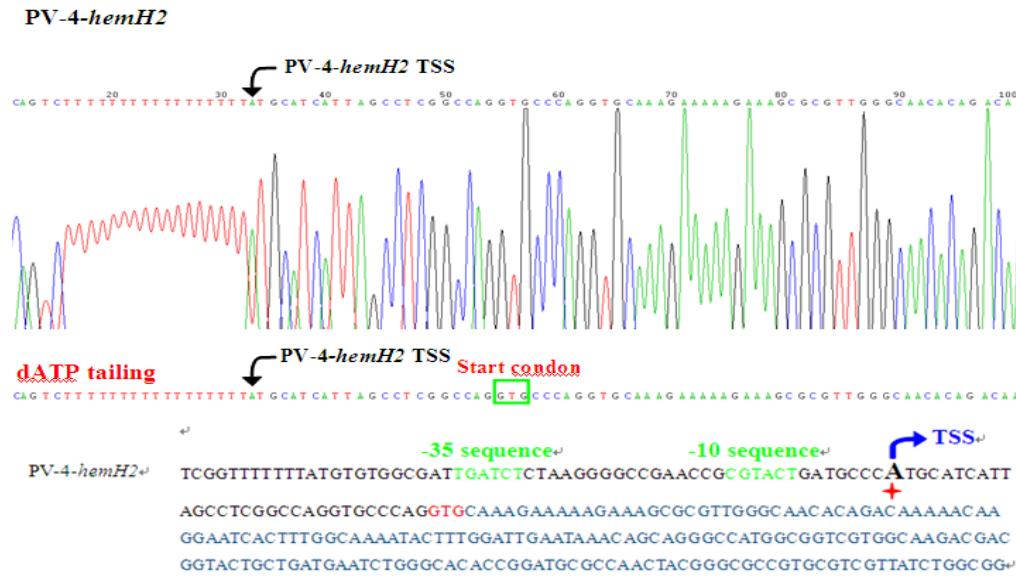
PV-4	GCTATATGAGCTGCTTCTCATATTG-AAAAAGCCACCTAGGTGGCTTTATGCA	81
Ssed	ACTAAATGAGCTAGTTCTCATATTTT-AAAAAGCTACTTAAGTGGCTTTATGGG	90
MR1	----CGAGAACAAAGTTCTCATATTAAGAAAAAGCTGCTTAAGCAGGCCCTTTATATG	72
W3181	----CGAGAACAAAGTTCTCATATTAAGAAAAAGCTGCCTAACAGCAGGCCCTTTATGTG	73
Svio	----TATGAGCCAGTTCTCATATTTT-AAGAAGCTCACTTAGGTGGCTTTATATG	77
OS155	----CGAGAACAGTTCTCATATTAGAAAAAGCTGCCGATGCAGGCCCTTTATGGG	74
Spea	TGTTTATGAGCTAGTTCTCATATTTT-AAAAGCCATGAAATGGCTTTATGCT	121
WP3	TGTTTATGAGCTAGTTCTCATATTTA-ATAAAGCCATGTAAGTGGCTTTATGCT	299
Shal	TGTTTATGAGCTAGTTCTCATATTTT-AAAAGCCATGAAATGGCTTTATGCT	126
	*** * ***** * *** * *****	
	OxyR (shew_1035)	-35
PV-4	AGGGTGGGTGCTAGCCTAG-----CGCTATCCTG-----	111
Ssed	GTTTTGGATGCTAGCCTAGA-----TTTTTCGCTGCTACTGGTCGTT-----	133
MR1	CGACTGAATGTTAGCCTG-----CCGGTATTG-----	103
W3181	TGACTGAATGTTAGCCTG-----CCGAGGTAAATTG-----	104
Svio	ATTTGAATGTTAGCCTAGGGCTTGTATCTAGAAATACTAGACTACAAATCTAGCC	137
OS155	TATCGCAATGTTAGCCT-----CCAAGTGAACCTCA-----	105
Spea	TTTGCGGTGATAGCCTGAC-----CTGAATATCTG-----	153
WP3	TTTCCAATGATAGCCTGACG-----CAGAATATCTG-----	332
Shal	TTTCGCAATGATAGCCTGAGG-----T-GAATATCTG-----	158
	** *****	*
	-10	Start site

**Supplementary Figure S7** Analysis of the promoter of the *hemH1* gene in *Shewanella* strains. (a) Primer extension analysis of transcriptional start site (TSS) of *hemH1* (Shew\_2229) in *S. loihica* PV-4. (b) Multiple alignment analysis on the nucleotide sequences upstream of the *hemH1* gene of *Shewanella* strains using

ClustalW2 to identify the promoter motifs and the putative OxyR (encoded by shewShew\_1035)-recognized motif. The shadowed motifs are supposedly predicted OxyR-recognized elements, and the underlined motifs are RpoD-recognized elements.

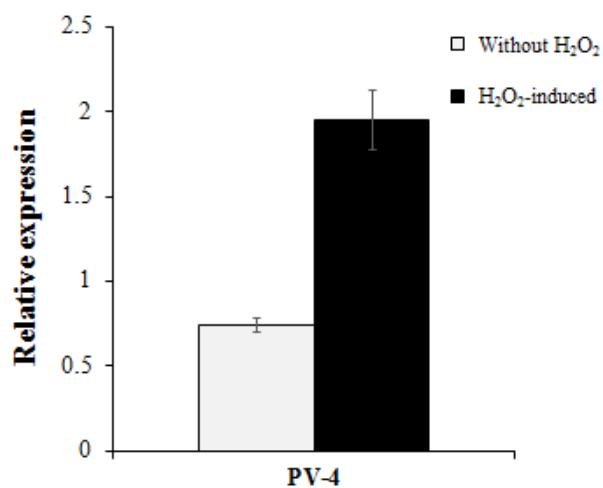


**Supplementary Figure S8** **(a)** Expression of the *oxyR* gene *in trans* activated expression of *hemH1* in the *S. loihica* PV-4 strain. **(b)** Effects of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and visible light on the transcription of the *hemH1* in the *S. loihica* PV-4.

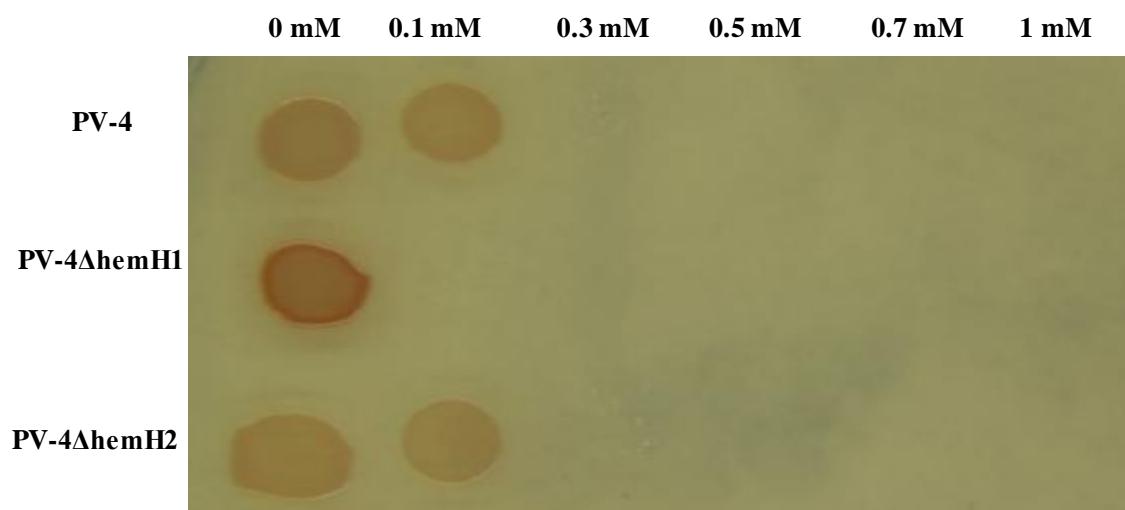


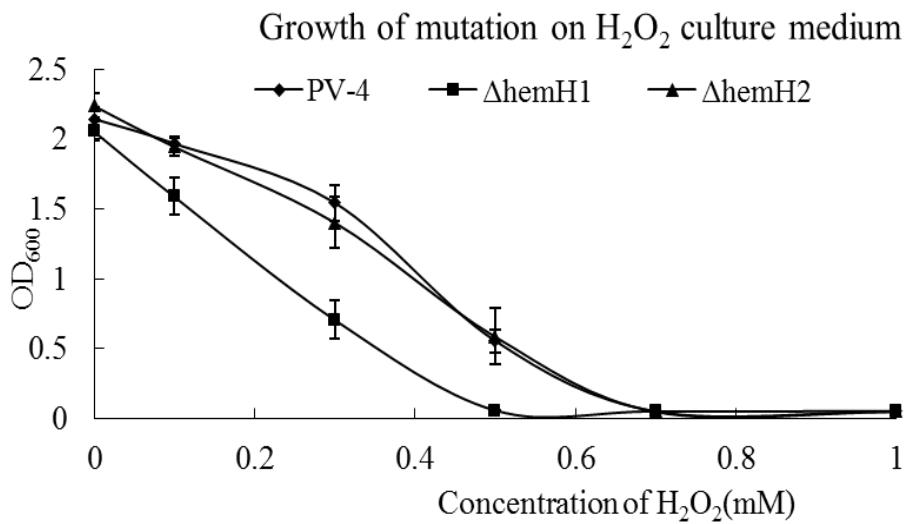
**Supplementary Figure S9** Primer extension analysis of transcriptional start site (TSS) of *hemH2* (Shew\_1140) in the *S. loihica* PV-4 strain.

### *rpoE2* transcription



**Supplementary Figure S10** Expression of *rpoE2* in PV-4 under different  $\text{H}_2\text{O}_2$ -induced oxidative stress condition.

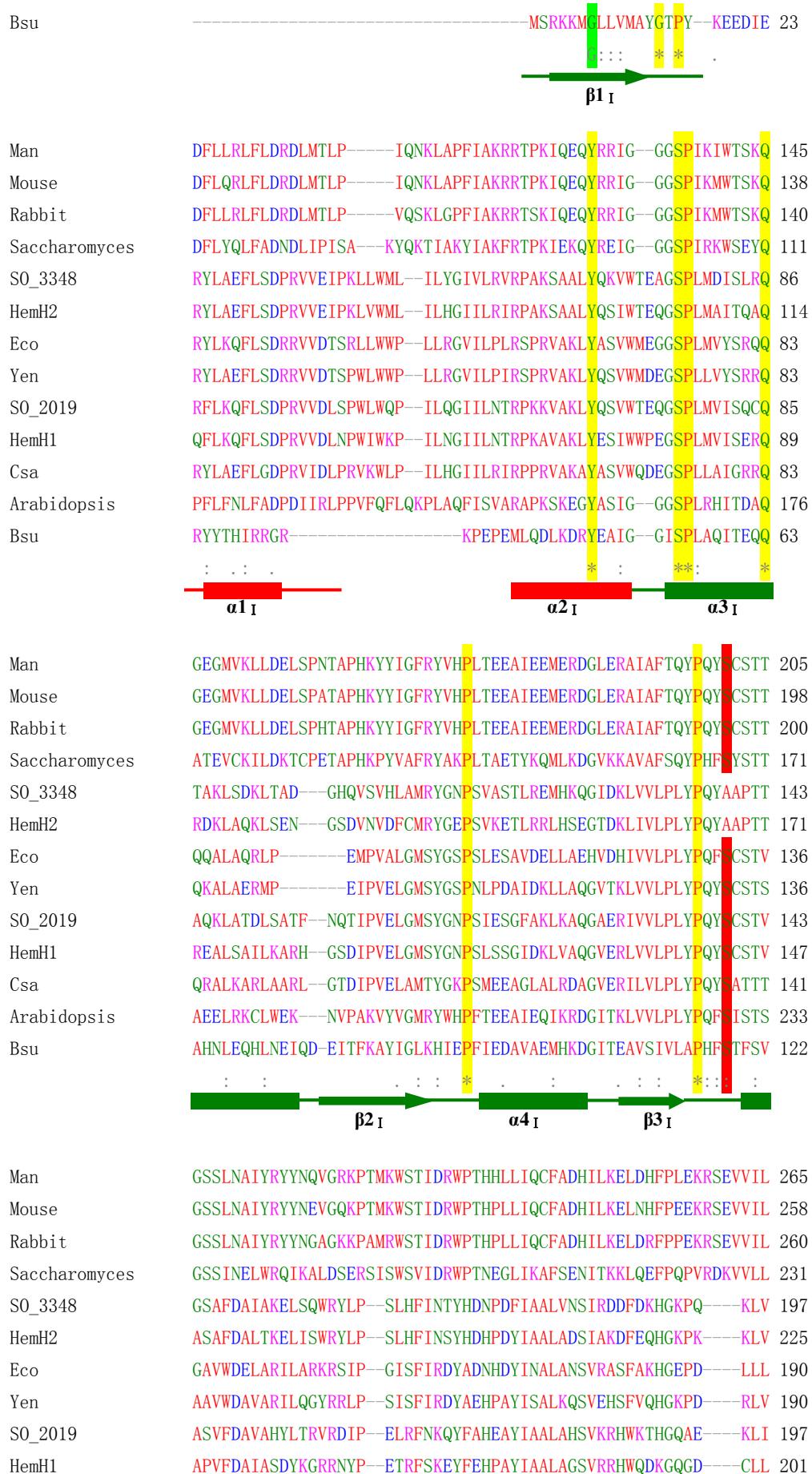


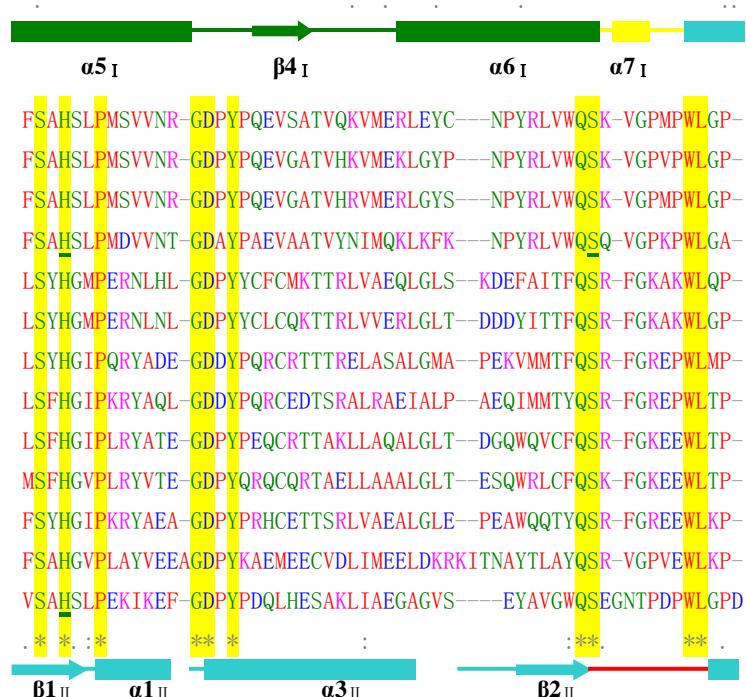
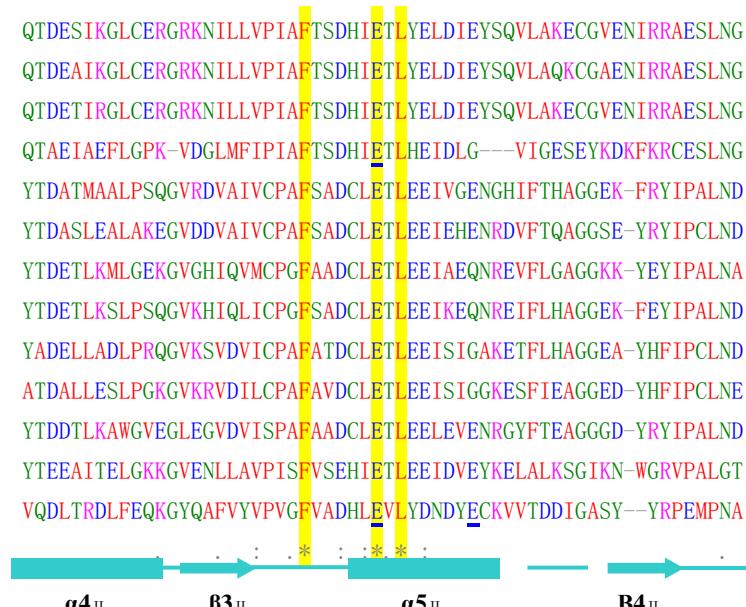
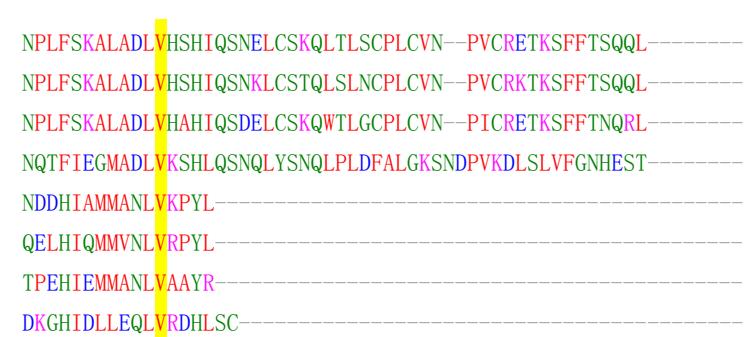


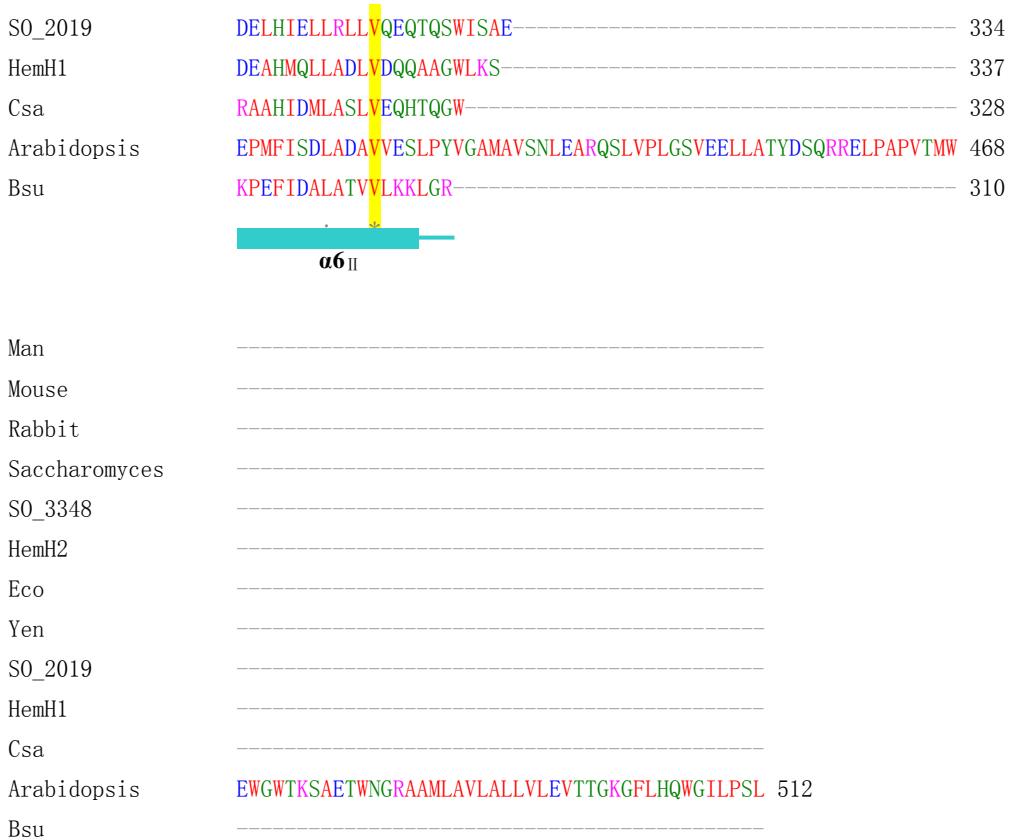
**Supplementary Figure S11** Effects of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) on the bacterial growth of the *Shewanella loihica* strains. Wild type PV-4 strain and the *hemH1* and *hemH2* mutants were grown in the LB broth containing 0, 0.1, 0.3, 0.5, 0.7 and 1 mM of hydrogen peroxide and incubated at 28 °C for 18 hrs.

Man	-----MRSLGANMAAALRAAGVLLRDPLASSSWRVCQPWR	35
Mouse	-----MLSASANMAAALRAAGALLREPLVHGSSRACQPWR	35
Rabbit	-----MLSAGTNMAAALRAAGALFRCPSVHGSSRAHQWPWR	35
Saccharomyces	-----MLSR-----TIRTQGSFLRRSQLTITR-----	22
S0_3348	-----	
HemH2	-----	
Eco	-----	
Yen	-----	
S0_2019	-----	
HemH1	-----	
Csa	-----	
Arabidopsis	MNC PAM TASPSSSSSSYSTFRPPPLLPQLSNDSQRSVVMHCTR LPFE AFAATSSNRL	60
Bsu	-----	

Man	WKSGA-AAAATTEAQHAQGAKPQVQPQKRYESNIRKPKT	ILMLNMGGPE	TLGDVH	92	
Mouse	CQSG-AAVAATTEKVHHAKTTKPQAQPER	RKPKT	ILMLNMGGPE	TLGEVQ	85
Rabbit	CQSGTAAAAAAAATTQRQSTKPQVQPEK	RKPKT	ILMLNMGGPE	TLGEVH	87
Saccharomyces	SFSVTFMNMQNA	KRSPT	IVLMNMGGPS	-KVEETY	56
S0_3348	-----MGHAARGKV	VLLLNLGTPDAPTA	SARV	28	
HemH2	-MQRKRKRV	QHRQKQGITLAKYFGLNKQQGHGGRGKTT	VLLMNLGTPDAPTTGAVR	56	
Eco	-----MRQTKT	VILLANLGTPDAPTP	PEAVK	25	
Yen	-----MKQSKL	VLMVNVLGTPDAPTP	QAVK	25	
S0_2019	-----MTSPSPA-F	VLLVNLGTPDEPTPKAVK	27		
HemH1	-----MNKVVDNLSPA	KL	VLLVNLGTPDTPTPKDVK	31	
Csa	-----MGSAPF	VMLVNVLGTP	EAFTTSAVR	25	
Arabidopsis	GKHSPLRAALVTSNPLNISSSVISDAI	SSSSVITDDAKI	VLLLNLLGGPE	TLDDVQ	118



Csa	GAVFDRLARALSPCPHLP--ELRFVRDYHDHPAYIEALAESIREHWETHGRQP----RLL	195
Arabidopsis	GSSLRLLERIFREDEYLVNMQHTVIPSWYQREGYIKAMANLIQSELGKFGSPNQ--VVIF	291
Bsu	QSYNKRAKEEAEKLGGLT---ITSVESWYDEPKFVTYWWDRVKETYASMPEDERENAMLI	179
		
Man	FSAHSILPMSVVNR-GDPYPQEVSATVQKVMERLEYC---NPYRLVWQSK-VGPMPWLGP-	319
Mouse	FSAHSILPMSVVNR-GDPYPQEVGATVHKVMEKLGYC---NPYRLVWQSK-VGPVPWLGP-	312
Rabbit	FSAHSILPMSVVNR-GDPYPQEVGATVHRVMERLGYS---NPYRLVWQSK-VGPMPWLGP-	314
Saccharomyces	FSAHSILPMDVVNT-GDAYPAEVAATVYNIMQKLKF---NPYRLVWQSQ-VGPKPWLGA-	285
S0_3348	LSYHGMPERNLHL-GDPYYCFMCKTRLVAEQLGLS--KDEFAITFQSR-FGKAKWLQP-	252
HemH2	LSYHGMPERNLNL-GDPYYCLCQKTRLVVERLGLT--DDYITTFQSR-FGKAKWLGP-	280
Eco	LSYHGIPQRYADE-GDDYPQRCRTTRELASALGMA--PEKVMMTFQSR-FGREPWLMR-	245
Yen	LSFHGIPKRYAQL-GDDYPQRCEDTSRALRAEIALP--AEQIMMTYQSR-FGREPWLT-	245
S0_2019	LSFHGIPRLRYATE-GDPYYPEQCRTTAKLLAQALGLT--DGQWQVCFQSR-FGKEEWLTP-	252
HemH1	MSFHGVPLRYVTE-GDPYQRQCQRTAELLAAALGLT--ESQWRLCFQSK-FGKEEWLTP-	256
Csa	FSYHGIPKRYAEA-GDPYPRHCETTSRLVAEALGLE--PEAWQQTQYSR-FGREEWLKP-	250
Arabidopsis	FSAHGVPLAYVEEAGDPYKAEMEECVDLIMEELDKRKITNAYTLAYQSR-VGPVEWLKP-	349
Bsu	VSAHSLPEKIKEF-GDPYPDQLHESAKLIAEGAGVS---EYAVGWQSEGNTPDPWLGP-	234
		
Man	QTDESIKGLCERGRKNILLVPIAFTS DHIELTYELDIEYSQVLAKECGVENIRRAESLNG	379
Mouse	QTDEAIKGLCERGRKNILLVPIAFTS DHIELTYELDIEYSQVLAKQCGAENIRRAESLNG	372
Rabbit	QTDETIRGLCERGRKNILLVPIAFTS DHIELTYELDIEYSQVLAKECGVENIRRAESLNG	374
Saccharomyces	QTAEIAEFLGPK-VDGLMFPIAFTS DHIELTLEIDLG---VIGESEYKDKFKRICESLNG	341
S0_3348	YTDA TMAALPSQGVDRV AIVCPAFA SADC LETLEEIVGENGHIFT HAGGEK-FRYIPALND	311
HemH2	YT DAS L EALAK EG VDD V AIVCPAFA SADC LETLEEIEHENRDVFTQAGGSE-YRYIPCLND	339
Eco	YTDE TLKMLGEKGVGHIQVMCPGFA ADC LETLEEIAEQN REVFLGAGGKK-YEYIPALNA	304
Yen	YTDE TLKSLPSQGVKH IQLICPGF SADC LETLEEIKEQN REIFLHAGGEK-FEYIPALND	304
S0_2019	YADELLADLPRQGVKSVDVICPAF ATDC LETLEEISIGAKETFLHAGGEA-YHFIPCLND	311
HemH1	ATD ALLES LPGKGVK RV DILCPA FAV DC LETLEEISIGG KESFIE AGGED-YHFIPCLNE	315
Csa	YT DDT LKA WGV E GLEG VDV I SPAFA A DC LETLEEISIGG KESFIE AGGED-YHFIPCLND	309
Arabidopsis	YTEE AIT ELG KKG VEN LL A VPISF VSE HI ET LEE ID VEY KEL ALK SG I KN-WGRVPALGT	408
Bsu	VQDLTRDLFEQKGYQAFVYVPVG FVADH L E LVYDND Y ECK VVT DDIG ASY-YRP E MPNA	292
		
Man	NPLFSKALADLVHSHIQSNELCSKQLTLSCPLCVN--PVC RETKS FFTS QQL-----	429
Mouse	NPLFSKALADLVHSHIQSNKLCSTQLSNCPLCVN--PVC RKT KS FFTS QQL-----	422
Rabbit	NPLFSKALADLVH AHIQ SDE LCSK QWT LG C PL CVN--P C RETKS FFTN QRL-----	424
Saccharomyces	NQT FIE GMAD LVK SHL QSN QLY SN QL P LD F AL G K S ND PV K DL S LV FG N HEST-----	393
S0_3348	NDDHIAMMANL VPK YL-----	327
HemH2	QELHIQMMVNL VRPYL-----	355
Eco	TPEHIE MMANL VAA YR-----	320
Yen	DKGHIDLLEQLV RDH LSC-----	322



**Supplementary Figure S12** Alignment of ferrochelatase amino acid sequences of various organism. Conserved regions of the sequence are highlighted in yellow, and minor differences are boxed in green (Thr and Gly) and red (Ser and Ala). Secondary structure elements are shown as determined from the *B. subtilis* enzyme (1). The alignment was generated by ClustalW2. The aligned sequences were from ferrochelatase amino acid sequences of a variety of organisms: Eco (*Escherichia coli*), Yen (*Yersinia enterocolitica*), Csa (*Cucumis sativus*), man, mouse, Rabbit (*Oryctolagus cuniculus*), Saccharomyces (*Saccharomyces cerevisiae*), Arabidopsis (*Arabidopsis thaliana*), Bsu (*B. subtilis*), the two HemH paralogues of the *S. loihica* PV-4 and the *S. oneidensis* MR-1.

## REFERENCES

1. Al-Karadaghi S, Hansson M, Nikonorov S, Jönsson B, Hederstedt L. 1997. Crystal structure of ferrochelatase: the terminal enzyme in heme biosynthesis. Structure 5: 1501-1510.