**Endosome-mediated endocytic mechanism replenishes the majority of synaptic vesicles at mature CNS synapses in an activity-dependent manner** 

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#### **SUPPLEMENTAL FIGURE LEGENDS**

**Supplemental Figure 1. A,** Hippocampal neurons at DIV19 were undergone two consecutive FM 1-43 dye loading/unloading cycles. Neurons were loaded with FM 1–43 by field stimulation (300 APs at 10 Hz) and kept for an additional 30 s after the stimulation to label all endocytosed vesicles. After a 10 min-resting period, a 1,200 APs at 10 Hz stimulus was given to unload FM 1–43. After the first round of the cycles, neurons were treated with DMSO for 30 min. **B,** Representative traces showing the average unloading responses of the control group in each first and second cycle. The extent of SV turnover was estimated from the total amount of unloaded dye in each first (ΔF1) and second cycle (ΔF2). FM1-43 signals were normalized to the intensity recorded in the first cycle (*n* = 29 neurons from 3 independent coverslips). Error bars indicate SEM. **C**, The amount of unloaded dye from each first (ΔF1) and second cycle  $(\Delta F2)$  over the first cycle. Data are presented as means  $\pm$  SEM, n.s. = not significant.

**Supplemental Figure 2. A,** Schematic figure showing synaptic vesicle recycling. At rest, SypHy fluorescence is quenched by the intraluminal acidic pH of the vesicle (1). Upon stimulation, vesicles fuse with the plasma membrane and expose their lumen to the neutral pH of the extracellular medium (pH  $\sim$  7.4), causing an increase in SypHy fluorescence (2). The fluorescence is then quenched once again after endocytosis (3) and reacidification (4, 5). **B**, Representative fluorescent time-lapse images of SypHy transfected neurons at DIV19 that were stimulated with 300 APs at 10 Hz. Individual regions were selected by hand and rectangular regions of interest were drawn around the synaptic boutons, then average intensities were calculated. Large puncta, which are typically interpreted as clusters of smaller synapses, were excluded from the selection procedure. Net fluorescence changes were obtained by subtracting the average intensity of the first four frames  $(F_0)$  from the intensity of each frame  $(F_t)$  for individual boutons. Representative images were selected for display from the entire time series; pre-stimulation, peak of response and post-stimulation. Right: Heat-map for pseudo-colored fluorescence intensity. Each of the numbers at the bottom corresponds to each stage in **A.** Scale bar: 5 μm. **C,** Changes in fluorescence intensity in response to electrical stimulation (300 APs at 10Hz, thick black bar) at individual synapses enclosed by rectangles from the image in **B**. Average of all displayed synapses is shown in red. **D,** Fluorescence intensity profile of each bouton was normalized to its peak value and plotted against time. Average of all displayed synapses is shown in red. Numbers at the bottom correspond to each stages in **A**.

**Supplemental Figure 3. A, C,** Schematic figures for two scenarios. Scenario-1, BFA does not block the budding of bulk endosomes from the plasma membrane but inhibits their subsequent re-acidification. Scenario-2, besides its effect on SV budding from the endosome, BFA actually blocks the budding of bulk endosomes from the plasma membrane (that is, the inside of the bulk endosomes should be accessible from outside the cell). **B,** Schematic figure for reacidification experiments If scenario-1 is true, then ΔF1 (before stimulation) and ΔF2 (after stimulation) should be the same in the BFA-treated neurons ( $\Delta F1 = \Delta F2$ ). If scenario-2 is true, a second application of pH 5.5 after stimulation should decrease SypHy fluorescence all the way to zero (ΔF1 < ΔF2). **D,** Schematic figures for TEV cleavage and QSY35 quenching experiments. If scenario-1 is true, TEV protease or QSY35 should not have access to the cytosol of the bulk endosomes; thus, the resulting amount of fluorescence loss in the BFA-treated neurons should be the same as that in the control neurons ( $\Delta F_{\text{Con}} = \Delta F_{\text{BFA}}$ ). If scenario-2 is true, TEV protease or QSY35 could cleave or quench the ecliptic pHluorin not only in the plasma membrane but also inside the bulk endosomes, and  $\Delta F_{BFA}$  should be larger than  $\Delta F_{Con}$ .

**Supplemental Figure 4. The roscovitine-sensitive SV retrieval becomes the major pathway with neuronal maturation. A-C,** Average SypHy fluorescence intensity profiles of the boutons from DIV9 (**A**), DIV14 (**B**) and DIV19 (**C**) neurons before (black) or after (red) 30 min treatment of 100  $\mu$ M roscovitine (Ros), a CDK5 inhibitor, followed by a 300 APs at 10 Hz stimulation. Note that regardless of maturation, the SVs retrieval in Ros-treated neurons is impaired, although the degrees of sensitivity differ in a similar manner with BFA-treated neurons ( $n = 69$  neurons from 4 independent coverslips for DIV9,  $n= 186$  neurons from 4 independent coverslips for DIV14,  $n = 95$  neurons from 4 independent coverslips for DIV19). **D,** The ratio of Ros-insensitive SV (RIS) / Ros-sensitive SV (RS):  $1.74 \pm 0.18$  for DIV9, 1.48  $\pm$  0.07 for DIV14, and 0.67  $\pm$  0.04 for DIV19 neurons. Data are presented as means  $\pm$  s.e. \* *p* < 0.05 (ANOVA and Tukey's HSD post hoc test)

**Supplemental Figure 5. shRNA-targeting AP-1 or AP-3 efficiently depletes AP-1 or AP-3 in hippocampal neurons, respectively. A,** Primary hippocampal neurons transfected with or without shRNA-tageting AP-1 or AP-3 were stained with anti-AP-1 or AP-3 antibody followed by Alexa 488-conjugated secondary antibody. Arrowheads indicate the cell bodies of shRNAtransfected cells. Expression levels of AP-1 and AP-3 were measured at cell bodies to avoid possible spatial overlap with other cells. Scale bar: 20 µm. **B,** In cells transfected with shRNAs, AP-1 or AP-3 was severely depleted compared with nontransfected cells: Expression levels over the nontransfected; AP-1 KD:  $0.20 \pm 0.06$  ( $n = 25$ ) whereas AP-3 KD:  $0.13 \pm 0.06$  ( $n =$ 20). \*\*\*p<0.001(Student's *t*-test).









 $\mathbf{A}$ 



B









# Statistical Parameters used in the Figures

#### **Fig. 1D**

**Kolmogorov\_Smirnov**

















## **Fig. 1F**

#### **Kolmogorov\_Smirnov**







**P valuee** 0.001240931

#### **Fig. 1G**

**Kolmog orov\_Smirnov**













**P value**



## **Fig. 2C**

#### **Kolmogorov\_Smirnov**







**P valuee** 0.03541777

## **Fig. 2E**







**P value**0.049608725

## **Fig. 2G**

#### **normality test (mean) DF Statistic p-value Decision at level(5%)** B 4 0.35462 0.59918 Can't reject normality **Controller 1 Can and 2 1 Can't reject normality** C 3 0.26939 1 Can't reject normality **Kolmogorov\_Smirnov**





**P value**0.00383386

#### **Fig. 3G**

**Kolmogorov\_Smirnov**













#### **Fig. 4C**

**Kolmog orov\_Smirnov**













## **Fig. 5G**

#### **Kolmogorov\_Smirnov**







**P value**9.013E-07

#### **Fig. 5H**

**Kolmog orov\_Smirnov**













5 0.32328 0.58233 C an't reject norm ality **Cont rol-second**

**Kolmogorov\_Smirnov**





**Control- first** Control- second

**P value**

0.093552554

**Kolmogorov\_Smirnov**



**mean +/- SEM**



 $Dne-way ANOVA$ 











**P value**