

**The modulation of MiR-155 and MiR-23a manipulates
Klebsiellapneumoniae Adhesion on Human pulmonary Epithelial
cells via Integrin $\alpha 5\beta 1$ Signaling**

Yan Teng^{1a}, Junming Miao^{1a}, Xiaofei Shen¹, Xiaolong Yang¹, Xinyuan Wang¹,
Laibing Ren¹, Xiaoying Wang¹, Junli Chen¹, Jingyu Li¹, Shanze Chen¹, Yi Wang^{1*},
Ning Huang^{1*}

1 Research Unit of Infection and Immunity, Department of Pathophysiology, West
China College of Basic and Forensic Medicine, Sichuan University, Chengdu 610041,
China

* These authors are both Corresponding authors.

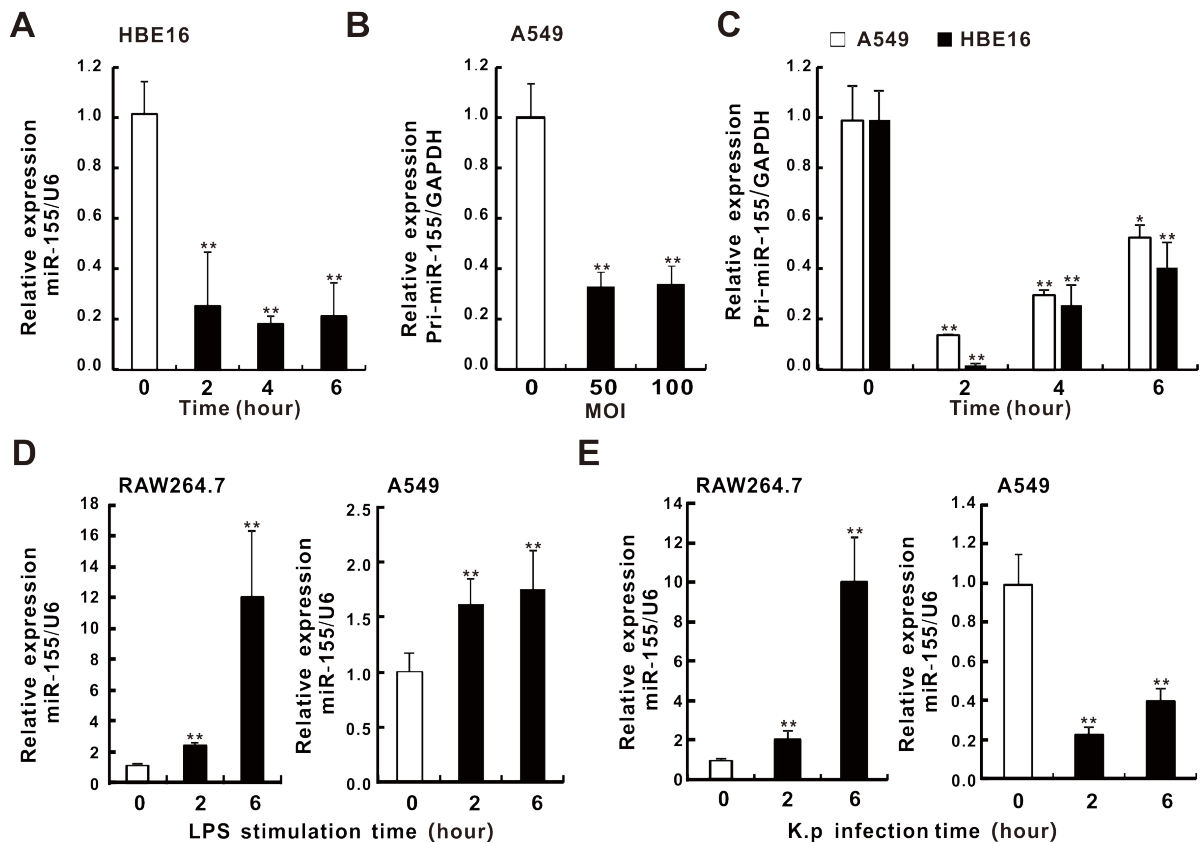
^aThese authors contributed equally to this work.

Tel: +86 02885501243;

E-mail: huangpanxiao@sina.com (Ning Huang). wangyi@stu.scu.edu.cn (Yi Wang)

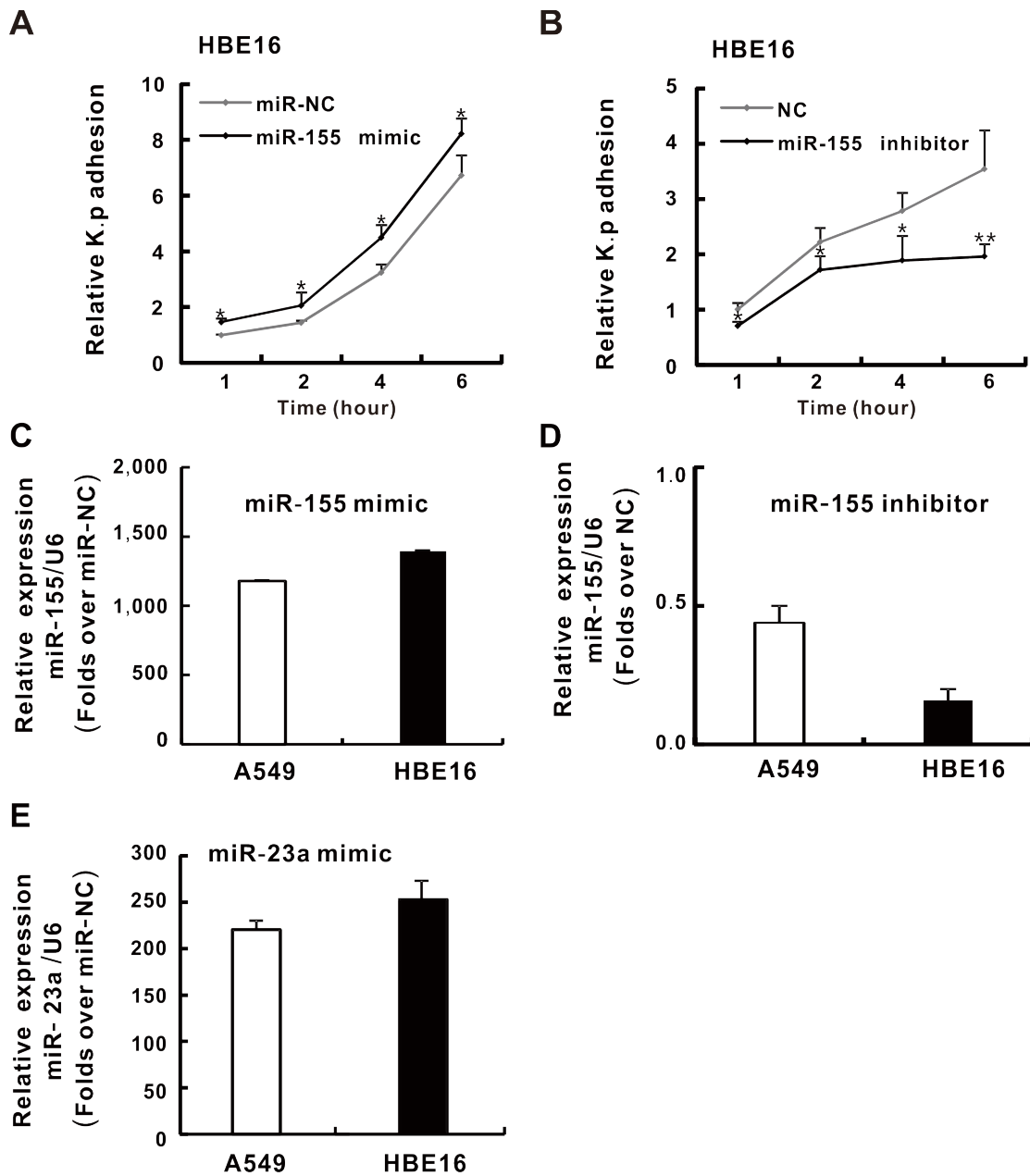
FigS1. Relative expression of miR-155 and pri-miR-155 under *K. pneumoniae* LPS stimulation.

A549 and HBE16 cells were exposed to increasing doses of *K. pneumoniae* (MOI=0, 50, 100) for 2 hours, or fixed amount of bacteria (MOI=100) at indicated time points (0hr, 2hr, 4hr and 6hr). The expression levels of miR-155 (A) and pri-miR-155 (B and C) were examined by RT-qPCR. RAW264.7 and A549 cells were stimulated by LPS (10 ng/mL) or exposed to fixed amount of bacteria (MOI=100) at indicated time points (0hr, 2hr and 6hr), the expression level of miR-155 (D and E) was examined by RT-qPCR. The Relative expression was normalized to U6 and then converted to the fold change over uninfected or unstimulated. (Data are the mean \pm SD and represent three individual experiments. * p <0.05, ** p <0.01 compared with *K. pneumoniae* LPS unstimulated).



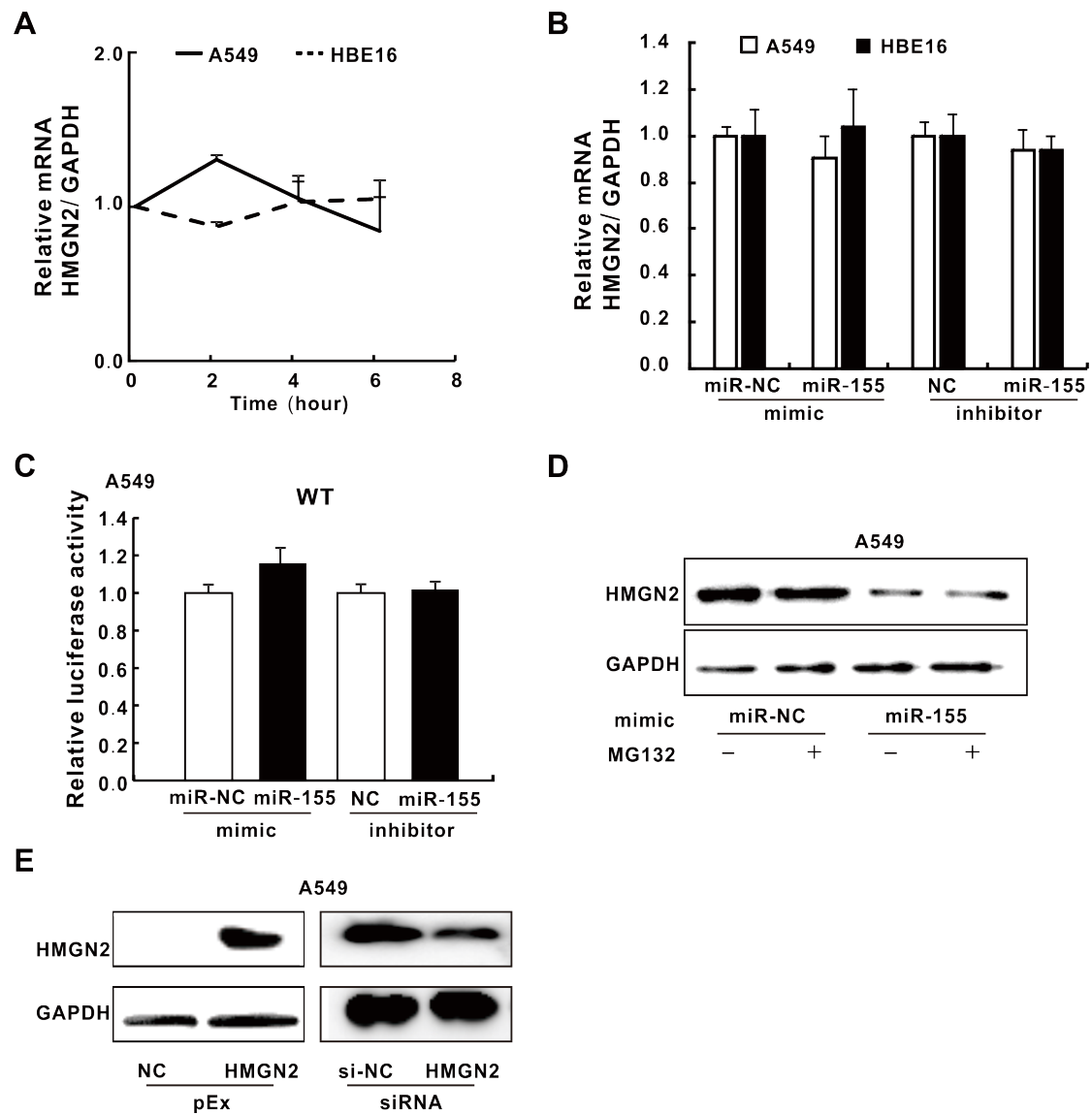
FigS2. MiR-155 regulated the adhesion of *K. pneumoniae*.

HBE16 cells were transfected with miR-155 mimic (A), inhibitor (B) and according negative control (miR-NC or NC) 24 hours prior to 100 MOI of *K. pneumoniae* exposure. The relative *K. pneumoniae* adhesion at indicated time points was determined by colony counts and represented after normalizing to 1 hour bacterial adhesion of miR-NC or NC. Relative expression of indicated miRNA after transfecting miR-155 mimic (C), inhibitor (D) or miR-23a mimic (E) in A549 or HBE16 cells (Folds over miR-NC or NC). (Data are the mean \pm SD and represent three individual experiments. * $p < 0.05$, ** $p < 0.01$).



FigS3. HMGN2 was potential target of miR-155 and miR-23a.

(A) RT-qPCR analysis showing the change of HMGN2 mRNA level in *K. pneumoniae* infected A549 or HBE16 cells at different infection time points (MOI=100). (B) RT-qPCR analysis showing the effect of miR-155 mimic or inhibitor on the mRNA levels of HMGN2 in *K. pneumoniae* infected A549 and/or HBE16 cells (MOI=100 for 2 hours, same as B, C, D, E, and F). (C) A549 cells were co-transfected with miR-155 mimic or inhibitor and HMGN2 3' UTR wild-type (WT) luciferase reporter construct. Luciferase activity was measured 24hr after transfection. (D) A549 cells transfected with miR-155 mimic were treated with proteasome inhibitor (MG132, 20 μ M, 3 hours) the relative HMGN2 protein expression were assessed by Western blot. (E) HMGN2 protein levels of A549 cells transfected with pEx-HMGN2 or si-HMGN2 (Data are the mean \pm SD and represent three individual experiments. ** $p < 0.01$).



FigS4. MiR-155 promoted integrin-Rac1 pathway in *K. pneumoniae*-infected HBE16 cells.

(A) RT-qPCR analysis showing the integrin $\alpha 5$ and $\beta 1$ mRNA expression of HBE16 cells transfected with miR-155 mimic or miR-NC prior to *K. pneumoniae* infection (MOI=100 for 2 hours, same as B, D and E). Western blot analysis showing the levels of integrin $\alpha 5$, integrin $\beta 1$ (B), Rac1-GTP, total Rac1 (D) and F-actin (E) in HBE16 cells treated as (A).(C) FN-cell Adhesion assay was performed to evaluate the effect of miR-155 mimic on HBE16 cells to associate with fibronectin coated plates. (Data are the mean \pm SD and represent three individual experiments. * p <0.05, ** p <0.01 compared with miR-NC).

