

Supplementary Figure 1. PNI increases an extracellular ATP level within the spinal cord 3 days, but not 28 days, after PNI.

Measurement of $[ATP]_e$ content in the ACSF media of ipsilateral and contralateral spinal cord slices taken from wild-type mice (**a**) 3 days and (**b**) 28 days after PNI. Values represent the relative ratio of ATP levels to the contralateral side (**a**, *n*=13; **b**, *n*=8; ****P*=0.0007 vs. Contralateral, Wilcoxon matched-pairs signed rank test). Values are means ± s.e.m.



Supplementary Figure 2. Effect of siRNAs on VNUT expression.

Real-time PCR analysis of *Slc17a9* mRNA in BV2 cells treated with control RNA (10 nM) or VNUT siRNAs (1, 3, 10 nM) for 48 h. Values represent the relative ratio of *Slc17a9* mRNA (normalized to the value for *18s* mRNA) to the control RNA (n=3). Values are means \pm s.e.m.



Supplementary Figure 3. Microglial VNUT is not required for PNI-induced increase in extracellular ATP within the spinal cord and pain hypersensitivity.

(a) Measurement of $[ATP]_e$ content in the ACSF media of spinal cord slices isolated from $Slc17a9^{fl/fl}$ or Cx3cr1- Cre^{ERT2} ; $Slc17a9^{fl/fl}$ mice that had been injected with tamoxifen (TAM), 7 days after PNI (n=14; **P<0.01, one-way ANOVA with post hoc Tukey Multiple Comparison test). (b) PWT of microglia-specific VNUT conditional knockout (Cd11b-Cre; $Slc17a9^{fl/fl}$) mice and their $Slc17a9^{fl/fl}$ littermate controls before and after PNI [$Slc17a9^{fl/fl}$: n=5, Cd11b-Cre; $Slc17a9^{fl/fl}$: n=6; ***P<0.001 vs. Contra ($Slc17a9^{fl/fl}$), two-way ANOVA with post hoc Bonferroni test]. Values are means \pm s.e.m.



Supplementary Figure 4. AAV-mediated Cre recombination in DRG neurons.

(a) Representative images (of three experiments) showing tdTomato fluorescence and Nissl staining in the L4 DRG of ROSA26 (R26)-tdTomato reporter mice intraperitoneally injected with AAV-ESYN-Cre vector. (scale bar, 200 μ m) (b-d) Representative images (of three experiments) showing fluorescent signals of tdTomato with Nissl staining and immunofluorescence labelling of (b) NF200, (c) P2X3R or (d) IB4 in the L4 DRG of R26-tdTomato mice treated with AAV-ESYN-Cre vector. (scale bar, 100 μ m) (e-g) Charts summerize the results of Supplementary Figures 3b-d showing the distribution of tdTomato fluorescence in the L4 spinal cord of R26-tdTomato mice intraperitoneally injected with AAV-ESYN-Cre vector. (scale bar, 100 μ m) (i) Fluorescent signals of tdTomato and immunofluorescence labelling of NeuN in the L4 spinal cord of R26-tdTomato mice treated with AAV-ESYN-Cre vector. (scale bar, 200 μ m) (i) Fluorescent signals of tdTomato and immunofluorescence labelling of NeuN in the L4 spinal cord of R26-tdTomato mice treated with AAV-ESYN-Cre vector. (scale bar, 200 μ m) (i) Fluorescent signals of tdTomato and immunofluorescence labelling of NeuN in the L4 spinal cord of R26-tdTomato signal with NeuN signal. (scale bar, 20 μ m)



Supplementary Figure 5. AAV-mediated Cre recombination in spinal dorsal horn neurons.

(a) Representative images (of three experiments) showing tdTomato fluorescence in the L4 spinal cord of R26-tdTomato mice intraspinally injected with AAV-NSE-Cre vector. (scale bar, 100 μ m) (b) Representative images (of three experiments) showing tdTomato fluorescence with immunofluorescent labelling of Iba1 (microglia), GFAP (astrocyte), APC (oligodendrocyte) or NeuN (neuron) in the L4 spinal cord of R26-tdTomato mice treated with AAV-NSE-Cre vector. Arrowheads indicate colocalization of tdTomato signals with NeuN signals. (scale bar, 20 μ m) (c) Representative images (of three experiments) showing Nissl staining with tdTomato fluorescence in the L4 DRG of R26-tdTomato mice treated with AAV-NSE-Cre vector. (scale bar, 100 μ m)



Supplementary Figure 6. AAV-mediated Cre recombination in spinal dorsal horn neurons.

(a) Representative images (of three experiments) showing tdTomato fluorescence in the L4 spinal cord of R26-tdTomato mice intraspinally injected with AAV-ESYN-Cre vector. (scale bar, 100 μ m) (b) Representative images (of three experiments) showing tdTomato fluorescence with immunofluorescent labelling of Iba1 (microglia), GFAP (astrocyte), APC (oligodendrocyte) or NeuN (neuron) in the L4 spinal cord of R26-tdTomato reporter mice treated with AAV-ESYN-Cre vector. Arrowheads indicate colocalization of tdTomato signals with NeuN signals. (scale bar, 20 μ m) (c) Representative images (of three experiments) showing Nissl staining with tdTomato fluorescence in the L4 DRG of R26-tdTomato mice treated with AAV-ESYN-Cre vector. A small number of Nissl-positive DRG neurons was positive to tdTomato (8.47%: 103 of 1216 Nissl⁺ cells). (scale bar, 100 μ m)



Supplementary Figure 7. VNUT expressed in SDH neurons is involved in increased spinal ATP supply and pain hypersensitivity after PNI.

(a) Schematic diagram of the experimental protocol. (b) PWT of $S/c17a9^{fl/fl}$ mice intraspinally injected with AAV-ESYN-Venus or an AAV-ESYN-Cre viral vectors before and after PNI (AAV-ESYN-Venus; $S/c17a9^{fl/fl}$: n=5, AAV-ESYN-Cre; $S/c17a9^{fl/fl}$: n=6; #P<0.01, ##P<0.001 vs. AAV-ESYN-Venus; $S/c17a9^{fl/fl}$, two-way ANOVA with post hoc Bonferroni test) (c) Measurement of [ATP]_e content in the ACSF media of spinal cord slices isolated from $S/c17a9^{fl/fl}$ mice intraspinally injected with AAV-ESYN-Venus or AAV-ESYN-Cre vectors 7 days after PNI (n=11, Wilcoxon matched-pairs signed rank test).



Supplementary Figure 8. Ectopic expression of VNUT in dorsal horn neurons of *Slc17a9-/-* mice rescues tactile allodynia after PNI.

(a) Real-time PCR analysis of *Slc17a9* mRNA using total RNA extracted from neuronal cell line Neuro2A cells, transduced with pZac-NSE-AcGFP or pZac-NSE-VNUT vectors. Values represent the relative ratio of *Slc17a9* mRNA (normalized to the value for *18s* mRNA) to cells with pZac-NSE-AcGFP (*n*=6). (b) Measurement of [ATP]_e content in the culture media of Neuro2A cells, transduced with pZac-NSE-AcGFP or pZac-NSE-VNUT, with or without treatment of Ca2+ ionopohre ionomycin (5 μ M) for 20 min. Values represent the relative ratio of ATP levels to the control (*n*=5). (c) Real-time PCR analysis of *Slc17a9* mRNA using total RNA extracted from the spinal cord and the DRG of WT mice 28 days after intraspinal viral vector injection (AAV-NSE-AcGFP or AAV-NSE-VNUT). Values represent the relative ratio of *Slc17a9* mRNA (normalized to the value for *18s* mRNA) to the AAV-NSE-AcGFP mice (*n*=6). (d) PWT of *Slc17a9-/-* mice intraspinally injected with AAV-ESYN-AcGFP; *Slc17a9-/-*: *n*=6, AAV-ESYN-VNUT; *Slc17a9-/-*: *n*=7, **P*<0.05, ****P*<0.001 vs. AAV-ESYN-AcGFP; *Slc17a9-/-*; *n*=6, AAV-ESYN-VNUT; with post hoc Bonferroni test). Values are means ± s.e.m.