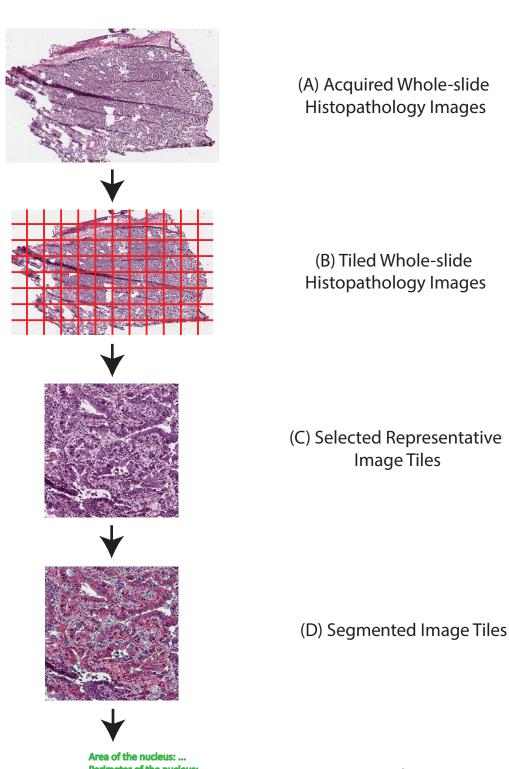
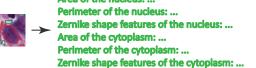
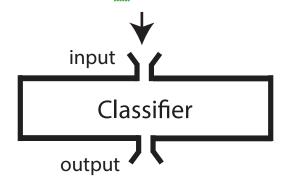
Supplementary Figure 1



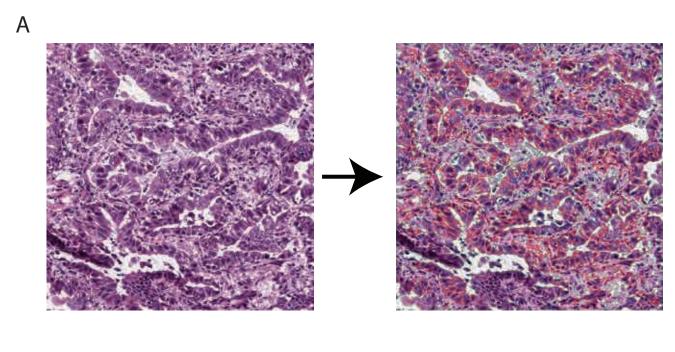


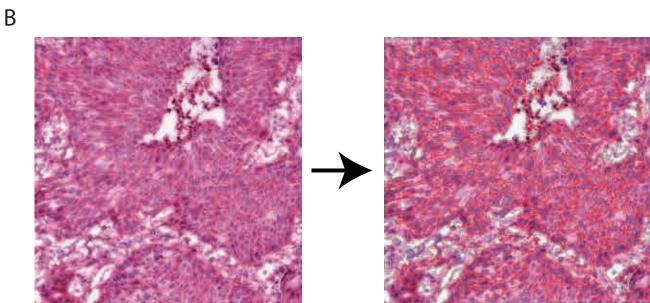
(E) Extracted Quantitative Image Features



(F) Generated Machine Learning Models **Supplementary Figure 1.** (Previous page) Diagram of informatics workflow of histopathology image processing. Digital whole-slide microscopic pathology images were (A) acquired and (B) tiled with open microscopy environment tools. (C) Dense image tiles were selected, as they typically show pathological changes in the lung, such as tumor, lymphocytic infiltration, inflammation, or atelectasis. (D) Tumor nuclei and cytoplasm were segmented with modules in CellProfiler. (E) Quantitative features were extracted from the identified cell nuclei and cytoplasm, (F) and were used for classification tasks.

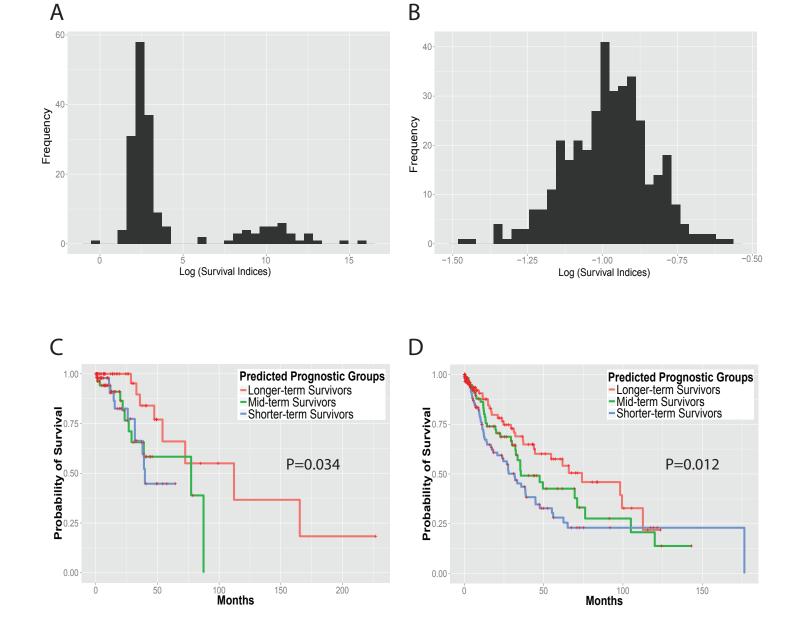
Supplementary Figure 2





Supplementary Figure 2. (Previous page) Fully automated image segmentation algorithm identified most tumor nuclei and tumor cells from the images. (A) Raw histopathology image from a lung adenocarcinoma patient (left) and the segmented image (right). (B) Raw histopathology image from a lung squamous cell carcinoma patient (left) and the segmented image (right). In the segmented image, blue outlines indicated tumor nuclei, whereas red outlines indicated tumor cytoplasm.

Supplementary Figure 3



Supplementary Figure 3. The differences between the survival outcomes of different survival groups are robust for different numbers of groups. (A) Distribution of log-transformed survival indices of TCGA patients with lung adenocarcinoma. (B) Distribution of log-transformed survival indices of TCGA patients with lung squamous cell carcinoma. (C) Sensitivity analysis showed that the survival differences between groups persisted when dividing adenocarcinoma patients into three prognostic groups (P=0.034). (D) The same analysis showed that the survival differences between groups also persisted when dividing lung squamous cell carcinoma patients into three prognostic groups (P=0.012).

Supplementary Table 1. Categories of Features Included in this Study

Feature Category	Feature Category CellProfiler Module Feature Examples			
reature Category	Centromer Module	Feature Examples		
		Focus score, minimum		
Lucia O alta	Manage of Language Control	intensity, maximum		
Image Quality	MeasureImageQuality	intensity		
		Granularity of		
		hematoxylin stained		
		image, granularity of		
		unmixed eosin stained		
Image Granuarity	MeasureGranularity	image		
		Mean intensity, standard		
		deviation of intensity,		
		and lower and upper		
		quartile of intensity of		
		unmixed hematoxylin		
Image Intensity	MeasureImageIntensity	and eosin stained images		
		Area, perimeter, Zernike		
Cell Size and Shape	MeasureObjectSizeShape	shape features		
		Mean intensity, mean		
		edge intensity, lower		
		quartile intensity, upper		
Pixel Intensity of the Cell	MeasureObjectIntensity	quartile intensity		
Radial Distribution of		Mean fractional intensity		
Pixel Intensity of the Cell	MeasureObjectRadialDistribution	at a given radius		
		Haralick texture		
		features, Gabor wavelet		
Cell Texture	MeasureTexture	features		
		Total area occupied by		
		the cells, total perimeter		
Area Occupied by Cells	MeasureImageAreaOccupied	occupied by the cells		
		Correlation coefficients		
		between unmixed		
Pixel Correlation of the		hematoxylin and eosin		
Cells	MeasureCorrelation	stained cell regions		
		The distance to the		
Neighboring Relation of		closest cell, the distance		
the Cells	MeasureObjectNeighbors	to the second closest cell		
		Area, perimeter, Zernike		
Nucleus Size and Shape	MeasureObjectSizeShape	shape features		
		Mean intensity, mean		
		edge intensity, lower		
Pixel Intensity of the		quartile intensity, upper		
Nuclei	MeasureObjectIntensity	quartile intensity		
Radial Distribution of		Mean fractional intensity		
Pixel Intensity of the	MeasureObjectRadialDistribution	at a given radius		

Nuclei		
		Haralick texture
		features, Gabor wavelet
Nucleus Texture	MeasureTexture	features
		Total area occupied by
		the nuclei, total
		perimeter occupied by
Area Occupied by Nuclei	MeasureImageAreaOccupied	the nuclei
		Correlation coefficients
		between hematoxylin
Pixel Correlation of the		and eosin stained
Nuclei	MeasureCorrelation	nucleus regions
		The distance to the
		closest nucleus, the
Neighboring relation of		distance to the second
the Nuclei	MeasureObjectNeighbors	closest nucleus

Supplementary Table 2. Performance of Classifiers in Distinguishing Tumor Parts from Adjacent Dense Normal Tissues

Classifiers	AUCs for Distinguishing Adenocarcinoma from Adjacent Dense Normal Tissues	AUCs for Distinguishing Squamous Cell Carcinoma from Adjacent Dense Normal Tissues
SVM with Gaussian Kernel	0.85	0.88
SVM with Linear Kernel	0.82	0.86
SVM with Polynomial		
Kernel	0.77	0.84
Naïve Bayes Classifier	0.73	0.77
Bagging	0.83	0.87
Random Forest utilizing		
Conditional Inference Trees	0.85	0.87
Breiman's Random Forest	0.85	0.87

^{*}AUC: Area Under Receiver Operating Characteristic Curves

Supplementary Table 3. Performance of Classifiers in Distinguishing Adenocarcinoma from Squamous Cell Carcinoma

Classifiers	AUCs of the TCGA Cohort	AUCs of the TMA Cohort
SVM with Gaussian Kernel	0.75	0.85
SVM with Linear Kernel	0.7	0.82
SVM with Polynomial Kernel	0.74	0.78
Naïve Bayes Classifier	0.63	0.73
Bagging	0.74	0.75
Random Forest utilizing		
Conditional Inference Trees	0.73	0.78
Breiman's Random Forest	0.75	0.76

^{*}AUC: Area Under Receiver Operating Characteristic Curves