

## **Scalable Production of Glioblastoma Tumor-initiating Cells in 3 Dimension Thermoreversible Hydrogels**

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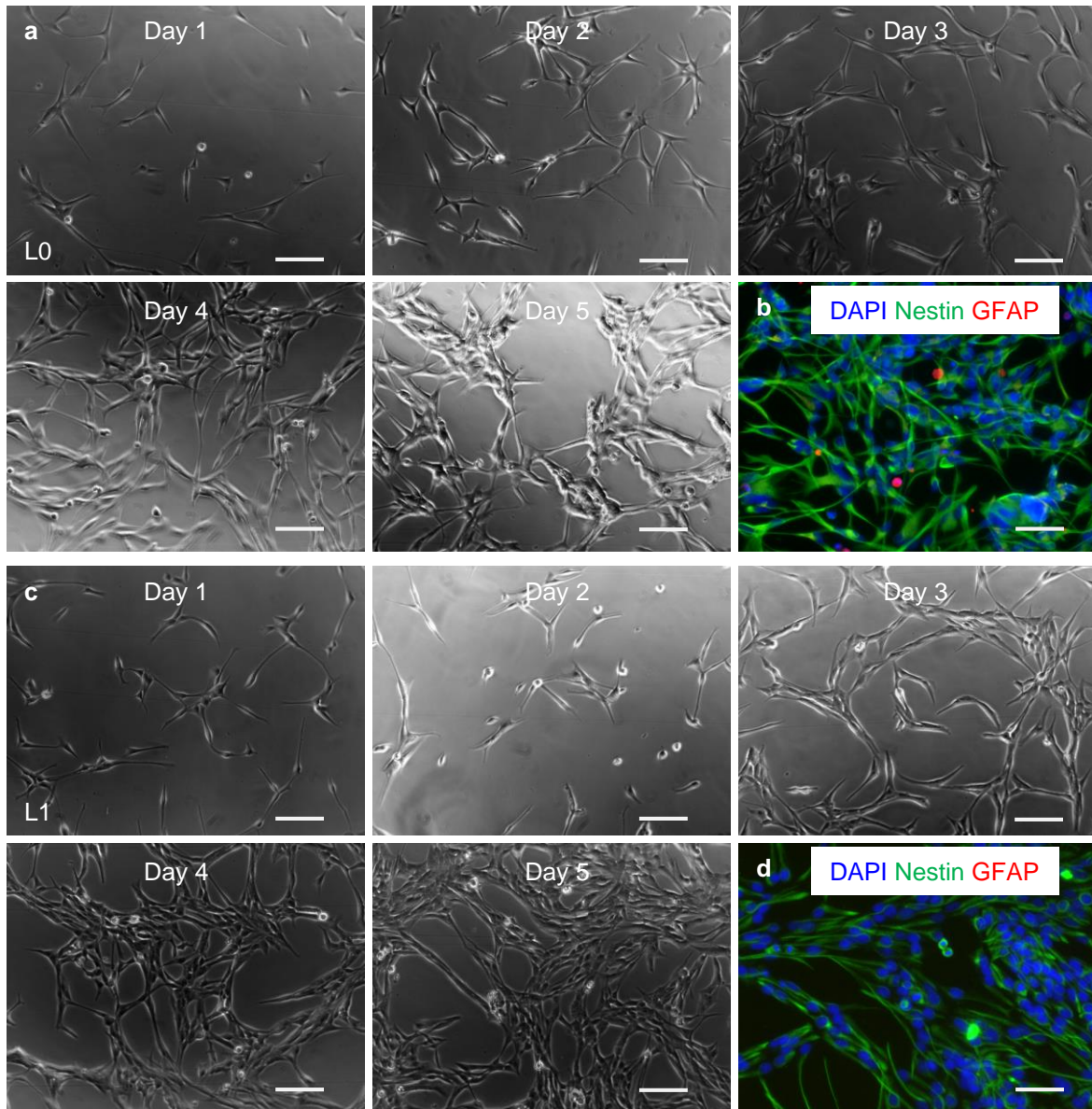
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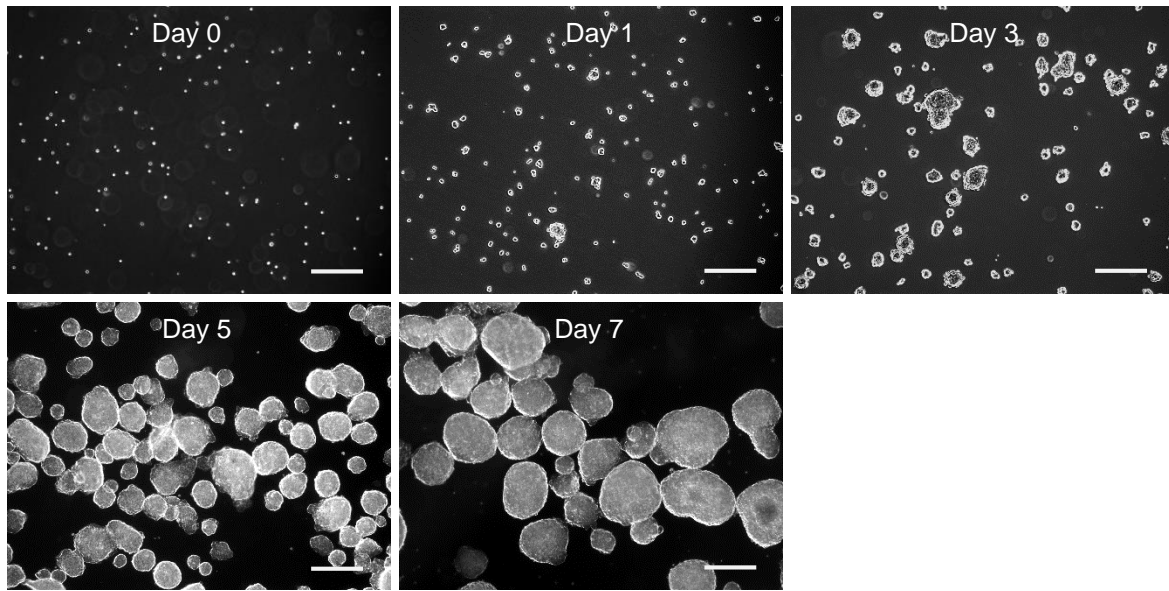
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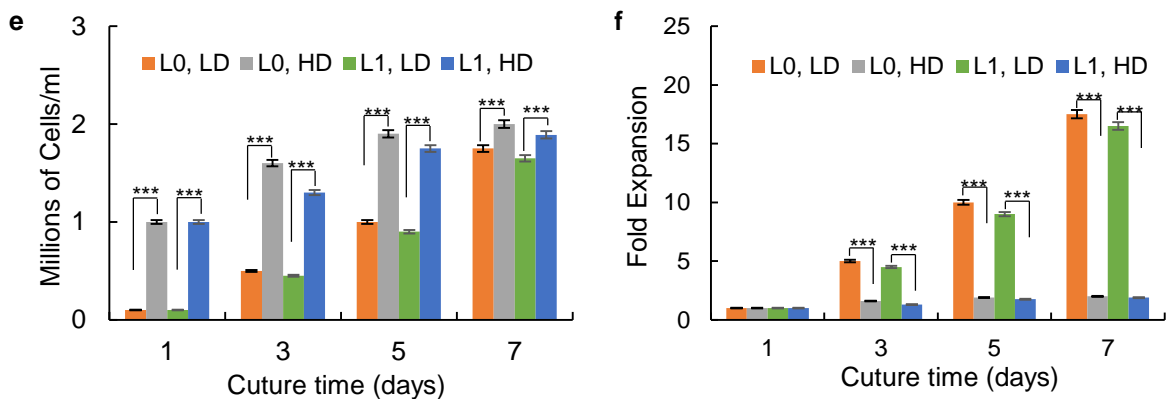
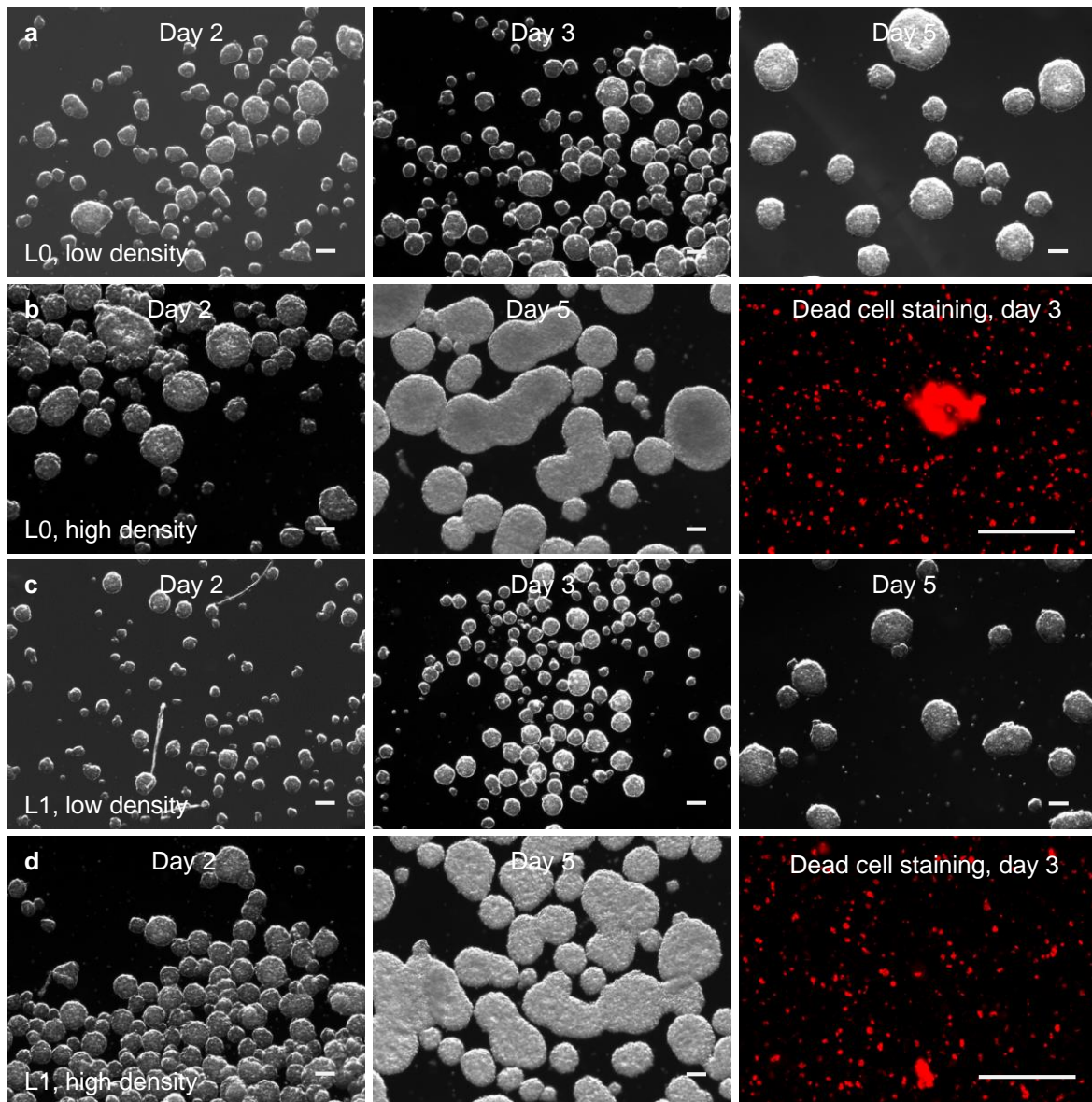
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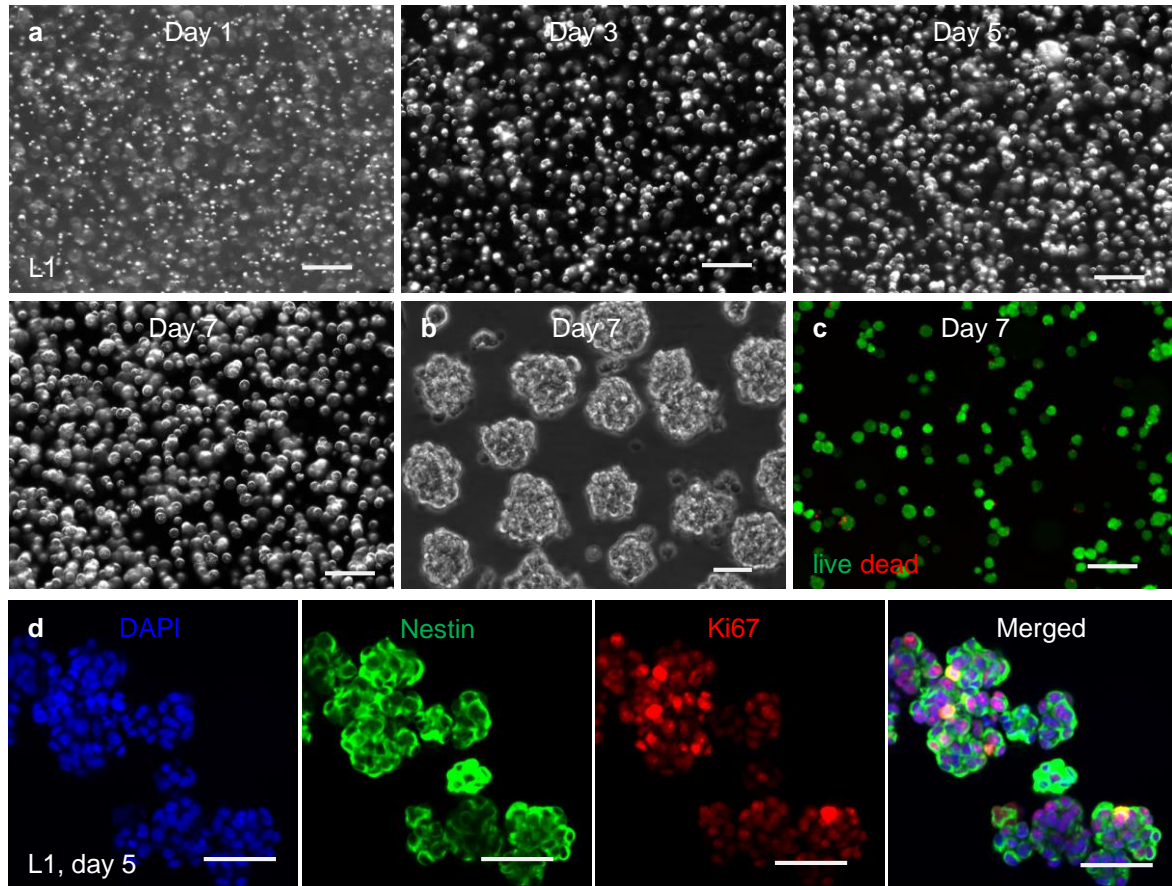
**Figure S1.** Culturing glioblastoma TICs as monolayer in 2D. L0 (a) and L1 (c) TICs were cultured on laminin-coated 6-well plates for 5 days. (b, d) Majority of the cells were Nestin+ stem cells. Very few or no differentiated GFAP+ cells were detected. Scale bar: (a, c) 100  $\mu$ m; (b, d) 50  $\mu$ m.



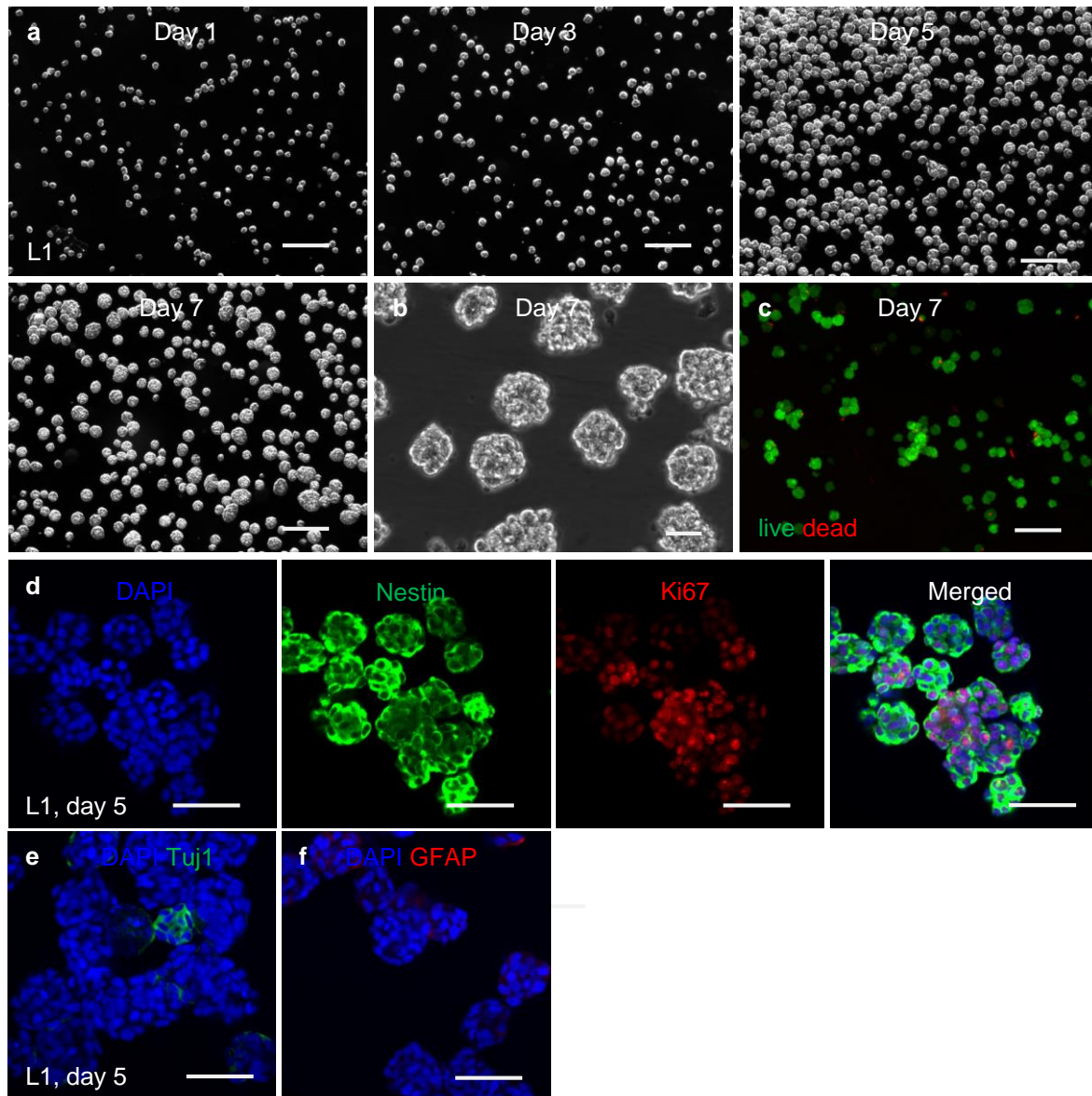
**Figure S2.** Culturing glioblastoma TICs as neurospheres. L0 TICs were suspended in liquid medium statically at  $5 \times 10^4$  cells/ml. Phase pictures of the growing spheroids were shown. Scale bar: 250  $\mu\text{m}$ .



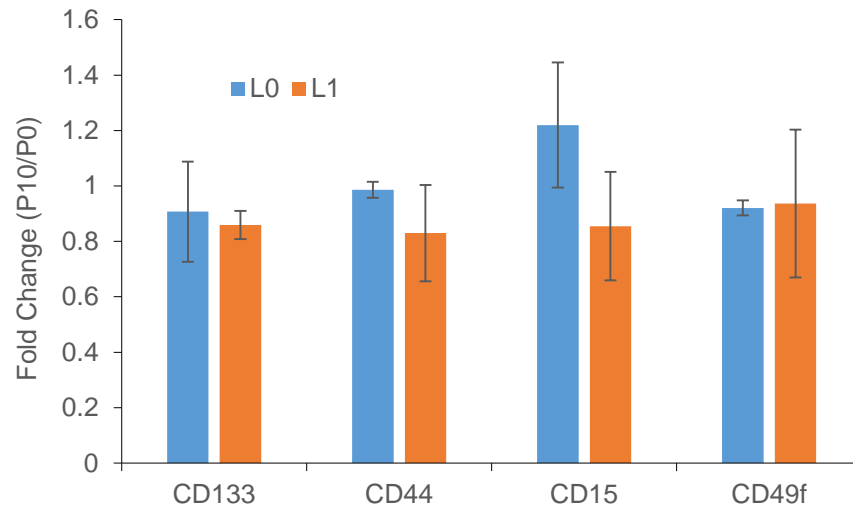
**Figure S3.** Culturing glioblastoma TICs in shaking plates. L0 and L1 TICs were suspended in liquid medium at  $1 \times 10^5$  (low density, LD) and  $1 \times 10^6$  (high density, HD) cells/ml in low adhesion plates on a orbital shaker at 75-90 rpm. Phase pictures and dead cell staining of TICs were shown (a, b, c, d). The cell density and fold of expansion were quantified (e, f). Error bars represent the standard deviation (n=3). \*\*\* indicates statistical significance at a level of  $p < 0.001$ . Scale bar: 100  $\mu\text{m}$ .



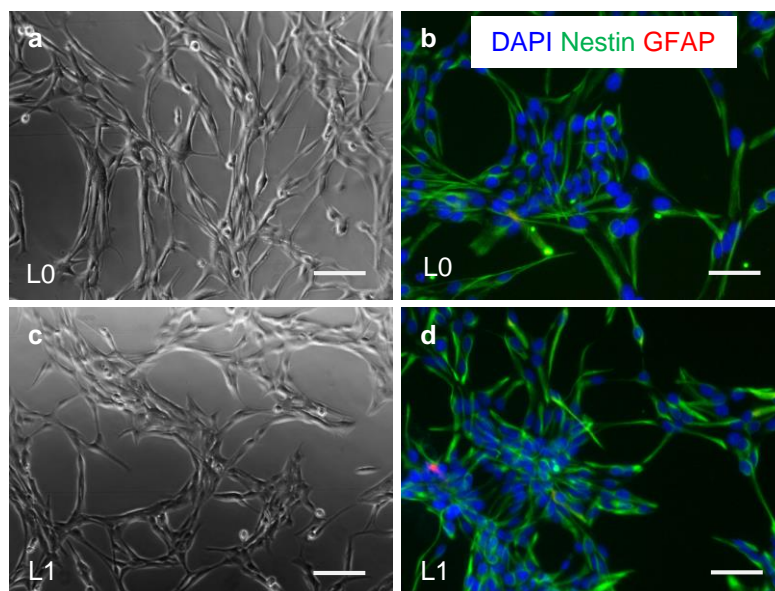
**Figure S4.** Culture glioblastoma TICs in 3D thermoreversible PNIPAAm-PEG hydrogel. **(a)** phase images showing L1 spheroids in the hydrogel on day 1, 3, 5, 7 of the culture. **(b)** Day 7 L1 spheroids released from the hydrogel. **(c)** Live (green) and dead (red) staining of day 7 L1 spheroids. **(d)** Immunostaining of day 5 L0 spheroids. Images were taken after spheroids were released from the hydrogel in **(b, c)**. Scale bar: **(a, c)** 250  $\mu\text{m}$ , **(b, d)** 50  $\mu\text{m}$ .



**Figure S5.** Long-term culture of glioblastoma TICs in 3D thermoreversible PNIPAAm-PEG hydrogel. **(a)** phase images showing L1 spheroids in the hydrogel on day 1, 3, 5, 7 of the culture at passage 10. **(b)** Day 7 L1 spheroids released from the hydrogel at passage 10. **(c)** Live (green) and dead (red) staining of day 7 L1 spheroids at passage 10. **(d, e, f)** Immunostaining of day 5 L0 spheroids at passage 10. Images were taken after the spheroids were released from the hydrogels. Scale bar: **(a, c)** 250 μm, **(b, d, e, f)** 50 μm.

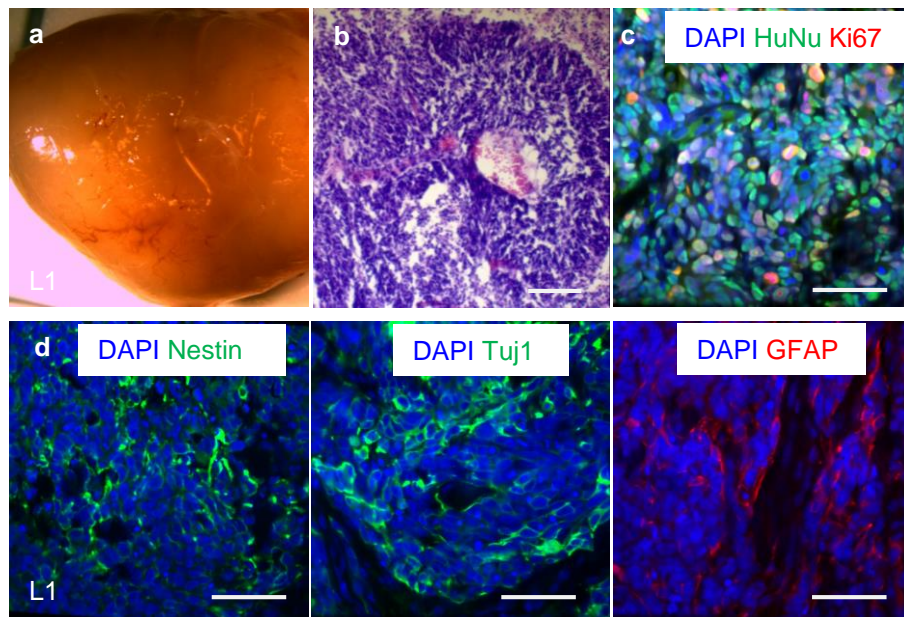


**Figure S6.** qRT-PCR on the mRNA level of CD133, CD44, CD15, CD49f. The ratio of their expression at passage 10 and passage 0 were shown. Error bars represent the standard deviation (n=3).



**Figure S7.** Returning to the 2D culture. After 10 passages in the 3D thermoreversible PNIPAAm-PEG hydrogel, L0 (**a, b**) and L1 (**c, d**) TICs were plated to the laminin-coated 2D surface. Cells on day 5 were shown. Scale bar: (**a, c**) 250  $\mu\text{m}$ , (**b, d**) 50  $\mu\text{m}$ .





**Figure S8.** Xenotransplantation of glioblastoma TICs. After 10 passages in the 3D thermoreversible PNIPAAm-PEG hydrogel, L1 TICs were transplanted subcutaneously to the NOD-SCID mice. (a) Harvested tumor. (b) H&E staining of the tumor section. (c) Majority of the cells in the tumor tissue were human nuclear antigen (HuNu) positive human cells and large percentage of cells were proliferating (Ki67+). (d) Nestin+ TICs, Tuj1+ neurons and GFAP+ glia cells were found in the tumor tissue. Scale bar: 50  $\mu$ m.

**Table S1.** Primer Sequences for qRT-PCR.

Gene Name	Sequences (5'-3')	References
CD133	F: GCGTGATTTCCCAGAAGATA	1
	R: CCCCAGGACACAGCATAGAA	
CD44	F: CAACTCCATCTGTGCAGCAAA	2
	R: GTAACCTCCTGAAGTGCTGCTC	
CD15	F: CTTTGTGCCTTATGGCTACC	3
	R: TTGGCTCAGTTGGTGGTAGT	
CD49F	F: CGAGTGACTGTGTTTCCCTCA	4
	R: GCATCAAGATCCCAGCGAGA	
GAPDH	F: TCGACAGTCAGCCGCATCTTCTTT	5
	R: ACCAAATCCGTTGACTCCGACCTT	

## References

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3. Dewi, Y., Shahib, M., Boesoirie, T. & Achmad, D. Studied of CD14 and CD15 gene expressions for myeloid derived suppressor cell profile at the RNA level as a predictor for progressivity in nasopharyngeal carcinoma. **4**, 6 (2016).
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