

Title: Transcriptome and proteome characterization of surface ectoderm cells differentiated from human iPSCs

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Supplementary Information

Supplementary Material and Methods

Human embryonic stem cell (hESC) culture

hESCs were seeded on feeder-free system using BD Matrigel Matrix and maintained in chemically-defined mTeSR1 medium (Stem Cell Technologies Inc., Vancouver, Canada).

qRT-PCR

Total RNA was extracted using the RNeasy Mini Kit (Qiagen) according to the manufacturer's instructions. Reverse transcription was done using the QuantiTect Reverse Transcription Kit. The qRT-PCR assay was done using an iCycler iQ Real-Time Thermocycler (Bio-Rad Laboratories) as described and GAPDH was used as an internal control. The primers used in this study were listed in Supplementary Table 3. Fold change was calculated according delta delta Ct value. Bar graphs represent mean \pm SD (standard deviation) of three independent experiments.

Chemicals

BMP type I receptor inhibitor LDN-193189, non-selective BMP receptor inhibitor DMH1, TGF β receptor type I/II inhibitor LY2109761, and TGF β receptor type I inhibitor LY2157299 were

purchased from Cayman Chemical (Ann Arbor, MI, USA). The working concentration for each inhibitor was 500 nM, 1 μ M, 2 μ M, and 1 μ M, respectively¹⁻³.

References

- 1 Horbelt, D. *et al.* Small molecules dorsomorphin and LDN-193189 inhibit myostatin/GDF8 signaling and promote functional myoblast differentiation. *J Biol Chem* **290**, 3390-3404, doi:10.1074/jbc.M114.604397 (2015).
- 2 Neely, M. D. *et al.* DMH1, a highly selective small molecule BMP inhibitor promotes neurogenesis of hiPSCs: comparison of PAX6 and SOX1 expression during neural induction. *ACS Chem Neurosci* **3**, 482-491, doi:10.1021/cn300029t (2012).
- 3 Yamasaki, A. *et al.* Identification of the role of bone morphogenetic protein (BMP) and transforming growth factor-beta (TGF-beta) signaling in the trajectory of serotonergic differentiation in a rapid assay in mouse embryonic stem cells in vitro. *J Neurochem* **132**, 418-428, doi:10.1111/jnc.12999 (2015).

Supplementary Figure Legends

Supplementary Figure 1. LifeMap online database analysis of upstream signals and downstream effectors. P63 (**a**) and BMP4 (**b**) are the upstream regulators directing SE differentiation, while Mir450b (**c**) is the upstream regulator inhibiting neural ectoderm differentiation from intraembryonic ectoderm cells.

Supplementary Figure 2. SE differentiation from human embryonic stem cell (hESC) line H9. (a) Western blotting analysis of KRT8, 18, and 19 expression in control hiPSCs and

differentiated SE cells. Actin was used as loading control. **(b)** Cell morphologies of control hESC line H9 and differentiated SE cells. Original magnification $\times 100$. Bars: 100 μm . **(c)** Immunofluorescence staining of SE markers KRT8, KRT18, KRT19, p63, AP-2 α , AP-2 γ , ALDH1A3, p-SMAD1/5/9, CDH1, Desmoglein 3 and FOXG1 endodermal marker AFP, Mesodermal marker Brachyury (T), and neural ectoderm marker OTX2 in control H9 and H9-derived SE cells. DAPI was used to stain nuclei. The secondary antibodies Alexa -488 and Alexa -594 were used to visualize green and red signals, respectively. Original magnification $\times 200$. Bars: 50 μm .

Supplementary Figure 3. Comparison of SE and NE differentiation at different time points.
(a) qRT-PCR results of selected up- and down-regulated genes from cDNA microarray analysis. Relative gene expression levels (SE vs. hiPSCs or SE vs. hESCs, mean \pm SD) are plotted. **(b)** Morphologies of SE induced from iPSCs at 48h and 72h time points are shown. Original magnification $\times 200$. Bars: 100 μm . cDNA microarray analysis was conducted using iPSCs treated with vehicle (control), 48h SE induction (2d-SE), and 72h SE induction (3d-SE). Genes differentially expressed in 2d-SE vs. control and 3d-SE vs. control were identified. IPA was then employed to analyze the correlation of canonical pathways **(c)**, bio-functions **(d)**, and upstream regulators **(e)** between 2d-SE and 3d-SE. **(f)** Morphologies of iPSCs and NE cells are shown. Original magnification $\times 100$. Bars: 100 μm . **(g)** Western blotting of neural and SE marker expression representing SE and neural differentiation. GAPDH was used as loading control. SE: surface ectoderm cells. C: vehicle control (iPSCs). N: NE differentiation.

Supplementary Figure 4. Comparison of SE cells and NE cells using IPA. **(a)** Canonical pathway analysis using IPA shows most significant up- and down-regulated pathways in SE compared to NE cells. Activation z-score (top) and $-\log$ (p-value) (bottom) are shown. **(b)** Bio-

function analysis using IPA shows most significant up- and down-regulated bio-functions in SE compared to NE cells. Activation z-score (top) and $-\log(p\text{-value})$ (bottom) are shown. **(c)** Upstream Regulator Analysis in IPA is a tool that predicts upstream regulators from gene expression data based on the literature and compiled in the Ingenuity Knowledge Base. A Fisher's Exact Test p-value is calculated to assess the significance of enrichment of the gene expression data for the genes downstream of an upstream regulator. Dot plot showed the upstream regulators in SE (X-axis) vs. NE (Y-axis), respectively. Pearson's correlation coefficient was calculated. **(d)** Mechanistic network analysis showed the down-regulation of TGF β superfamily, Wnt/ β -catenin, and NF- κ B signaling pathways in NE cells.

Supplementary Figure 5 Effects of inhibiting TGF β superfamily activity in SE differentiation. **(a)** Cell morphologies in control, SE, and SE with inhibitors. hiPSCs and hESCs were treated with (BMP4+DAPT) or (BMP4+DAPT) and indicated inhibitors for 48h. Representative pictures are shown. Original magnification $\times 100$. Bars: 100 μ m. Left panel: hiPSCs. Right panel: hESCs. **(b)** Western blotting analysis of marker expression in control, SE and SE with inhibitors. Top panel: hiPSCs. Bottom panel: hESCs. Actin was used as loading control.

Supplementary Figure 6. Western blotting images for Fig. 6. The gels have been run under the same experimental conditions.

Supplementary Tables

Supplementary Table 1 SE-associated genes summarized in LifeMap Discovery Database

Species	Gene Symbol	Full Name	Expression	Localization	Method(s)	Validation
m	Foxg1 (Bf1)	forkhead box G1	++	intra		
m	Krt18 (K18)	keratin 18	++	intra	IS	P
m	Krt8	keratin 8	++	intra	IS	P
m	Trp63 (P63)	transformation related protein 63	++	intra	KO	RP
m	Pax6	paired box 6	-	intra		P
m	Aldh1a3 (RALDH3)	aldehyde dehydrogenase family 1, subfamily A3	+	intra	ISH, IS	RP
m	Arhgap31	Rho GTPase activating protein 31	+	intra	ISH, IS	RP
h	BMP2	bone morphogenetic protein 2	+	secreted	ISH	R
h	Bmp7 (OP1)	bone morphogenetic protein 7	+	secreted	ISH, IS	RP
m	BMP7	bone morphogenetic protein 7	+	secreted	ISH	R
ch	DLX5	distal-less homeobox 5	+	intra	IS	P
m	Epcam	epithelial cell adhesion molecule	+	surface		
m	Fgf10	fibroblast growth factor 10	+	secreted	ISH, IS	RP
m	Fgf8	fibroblast growth factor 8	+	secreted	ISH, IS	RP
m	Fgf9	fibroblast growth factor 9	+	secreted	ISH, IS	RP
m	Fgfr2 (Fgfr-2)	fibroblast growth factor receptor 2	+	surface	ISH, IS	RP
m	Fn1 (Fn-1)	fibronectin 1	+	secreted	ISH, KO	RP
m	Gata1	GATA binding protein 1	+	intra		
ch	GATA2	GATA binding protein 2	+	intra	IS	P
h	GRHL2	grainyhead-like 2 (Drosophila)	+	intra		
m	Grhl3	grainyhead-like 3 (Drosophila)	+	intra	ISH, IS	RP
h	GRHL3	grainyhead-like 3 (Drosophila)	+	intra		
m	Krt14 (K14)	keratin 14	+	intra	ISH, IS, Transgenic Mice	RP
m	Ltb	lymphotoxin B	+	surface	Transgenic Mice	RP
m	Mir450b	microRNA 450b	+		ISH	R
m	Msx1 (Hox7)	msh homeobox 1	+	intra	ISH, IS	RP
ch	MSX1	msh homeobox 1	+	intra	IS	P
ch	MSX2 (HOX-8)	msh homeobox 2	+	intra	IS	P
m	Nedd4	neural precursor cell expressed, developmentally down-	+	intra	ISH, IS	RP

regulated 4

m	Notch2	notch 2	+	surface	ISH, IS	RP
m	Otx2	orthodenticle homolog 2 PERP, TP53 apoptosis effector	+	intra	ISH, IS	RP
m	Perp		+	surface	KO	RP
m	Pmel	premelanosome protein	+	surface	ISH, IS	RP
m	Pth1r (Pthr1)	parathyroid hormone 1 receptor	+	surface	ISH, IS	RP
m	Smad1	SMAD family member 1	+	intra		P
m	Smad5	SMAD family member 5	+	intra		
m	Smad9 (Smad8)	SMAD family member 9	+	intra		
m	Sox2	SRY (sex determining region Y)-box 2	+	intra	ISH, IS	RP
m	Tfap2c (AP2gamma)	transcription factor AP-2, gamma	+	intra		
m	Zmiz1	zinc finger, MIZ-type containing 1	+	intra	ISH, IS	RP

Notes: ++, database-selected marker; -, undetected; +, expressed.

Intra: intra cellular; surface: cell surface

IS: immunostaining; ISH: in situ hybridization

P: protein; R: RNA

Supplementary Table 2 Differentially expressed genes in neural and non-neural ectoderm-GXD database analysis

MGI Gene ID	Gene Symbol	Gene Name	Assay Type	Age	Theiler Stage	Structure Detected	PubMed ID
MGI:1914 071	Bcs1l	BCS1-like (yeast) bone morphogenetic protein 4 cadherin, EGF	Immunohistochemistry	E7.0	10	neural ectoderm	Yes 17049929
MGI:8818 0	Bmp4	LAG seven-pass G-type receptor 2	RNA in situ	E7.2 5	10	neural ectoderm	Yes 19393343
MGI:1858 235	Celsr2	chordin-like 1	RNA in situ	E6.7 5	10	neural ectoderm	Yes 11677057
MGI:1933 172	Chrdl1	forkhead box O1	In situ reporter (knock in)	E7.0	10	neural ectoderm	Yes 11118896
MGI:1890 077	Foxo1	general transcription factor II I repeat domain-containing 1	Immunohistochemistry	E7.0	10	neural ectoderm	Yes 21397048
MGI:1861 942	Gtf2ird1	homeobox gene expressed in ES cells	RNA in situ	E7.0	10	neural ectoderm	Yes 12971990
MGI:9607 1	Hesx1	homeobox gene expressed in ES cells	RNA in situ	E7.0	10	neural ectoderm	Yes 10882526
MGI:9607 1	Hesx1	melanoma antigen, family D, 1	Western blot	E7.5	10	neural ectoderm	Yes 8565852
MGI:1930 187	Maged1	paired box 2	RNA in situ	E7.5-8.0	10	neural ectoderm	Yes 12204258
MGI:9748 6	Pax2	sal-like 3 (Drosophila)	RNA in situ	E7.0	10	neural ectoderm	Yes 7577673
MGI:1092 95	Sall3	transcription factor 12	RNA in situ	E7.0	10	neural ectoderm	Yes 8798152
MGI:1018 77	Tcf12	TGFB-induced factor homeobox 1	RNA in situ	E7.0	10	neural ectoderm	Yes 15018808
MGI:1194 497	Tgif1	vang-like 2 (van gogh, Drosophila)	RNA in situ	E7.0-7.5	10	neural ectoderm	Yes 16284942
MGI:2135 272	Vangl2	orthodenticle homolog 2	In situ reporter (knock in)	E7.0	10	non-neural ectoderm	Yes 11431695
MGI:9745 1	Otx2						No 7588062

Supplementary Table 3 Primers used in this study

	Forward	Reverse
TFAP2C	CAGATGGACGAGGTGCAGAATG	GGGGAGGTTCAGAGGGTTCT
TFAP2A	GGAGGTCCCGCATGTAGAAG	CGACCCGGAAGTGAACAGAA
KRT8	GCTGGTGGAGGACTTCAAGA	AGCTCCGGATCTCCTCTTC
KRT18	GATCATCGAGGACCTGAGGG	ATCAATGACCTTGCAGGCC
KRT19	CTACACGACCATCCAGGACC	TCCGTCTCAAACCTGGTCG
OTX2	CCCGGTACCCAGACATCTC	GCGGCACTTAGCTCTCGAT
TUBB3	GAGACAGGTACAGGTCCACG	AGGCACGTACTTGTGAGAAGA
FOXA2	CGTTCCGGGTCTGAACGT	CATGTTGCTCACGGAGGAGT
ACTA1	CACGATGTACCCCTGGGATCG	GCCGATCCACACCGAGTATT
ID1	AATCATGAAAGTCGCCAGTG	ATGTCGTAGAGCAGCACGTTT
ID2	CCGTGAGGTCCGTTAGGAAA	GAGCTTGGAGTAGCAGTCGT
P63	TGGTGCACAAACAAGATTG	ATAGGGACTGGTGGACGAGG
MSX2	TGGATGCAGGAACCCGG	AGGGCTCATATGTCTGGCG
NFKB1	GCTTAGGAGGGAGAGCCCA	TATGGGCCATCTGTTGGCAG

Fig.S1

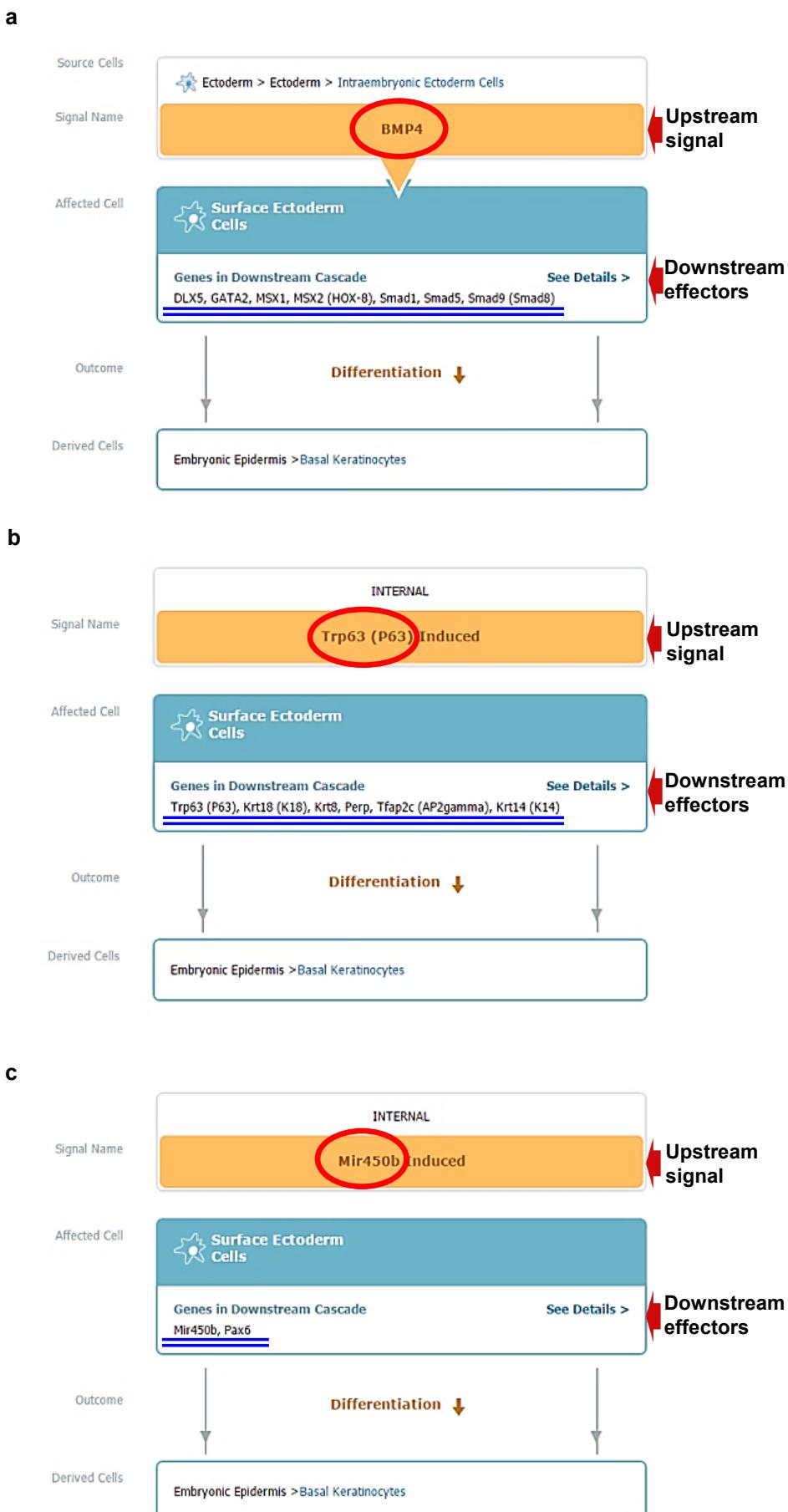


Fig.S2

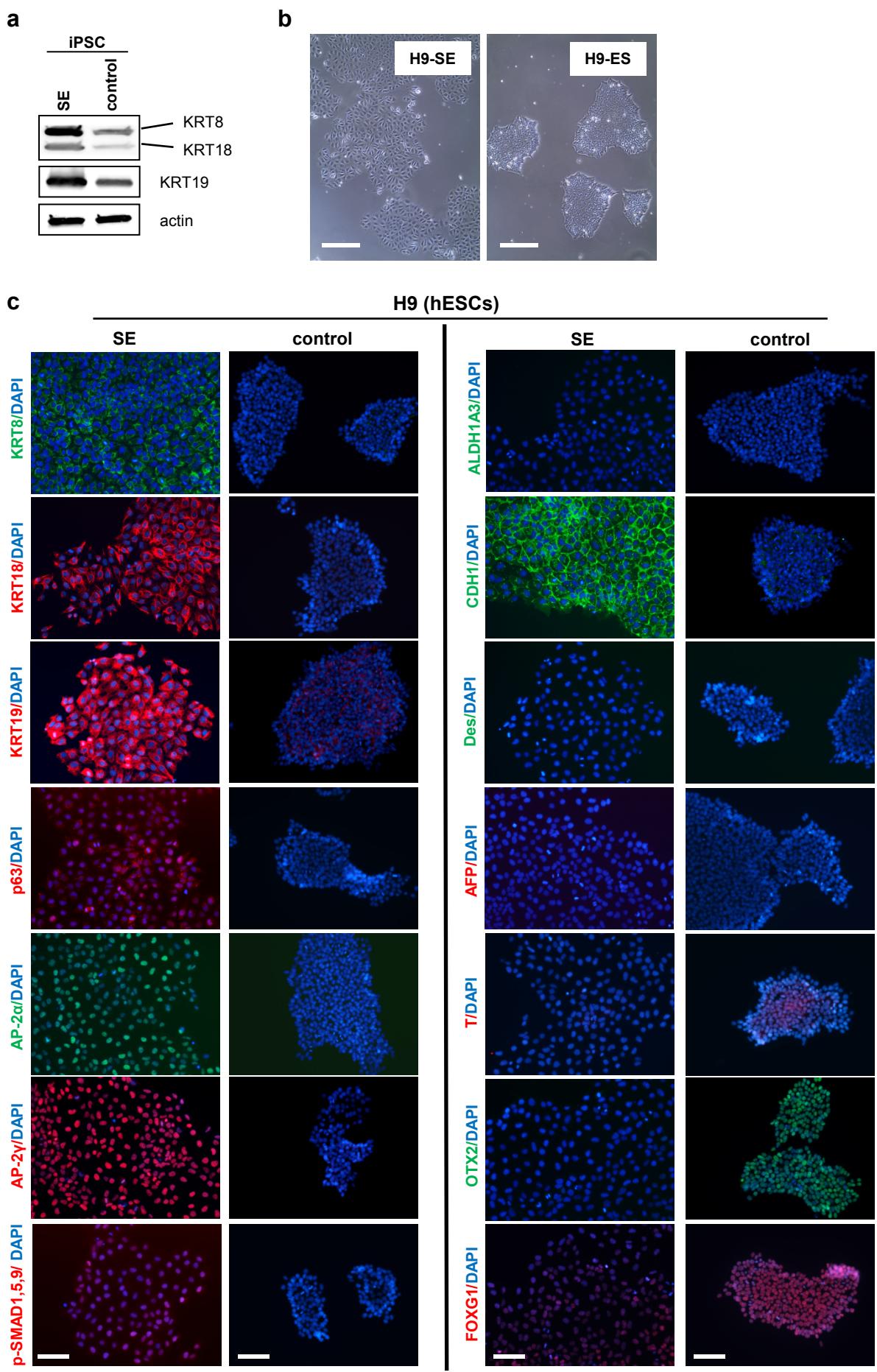
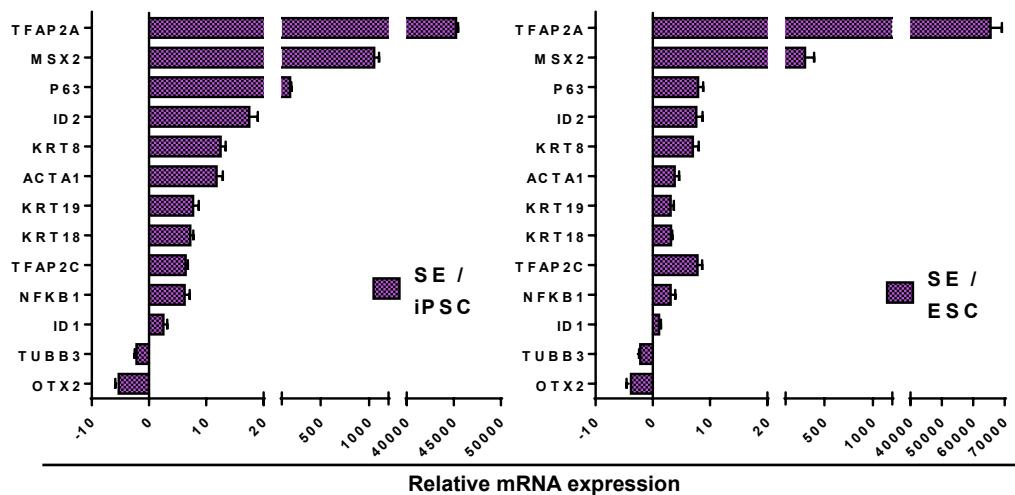
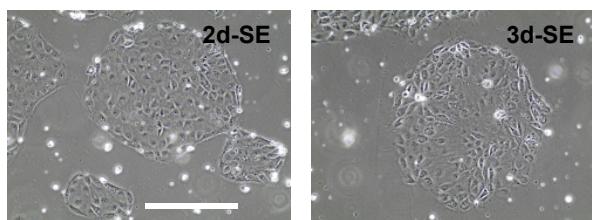


Fig.S3

a



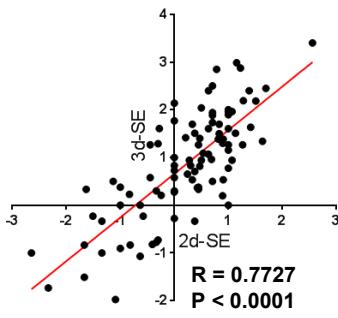
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c

Canonical pathways

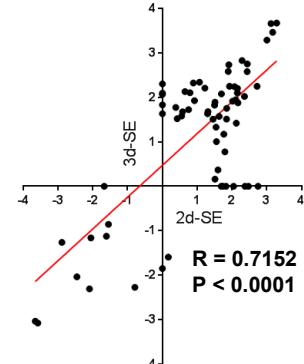
3d-SE vs. 2d-SE



d

Bio functions

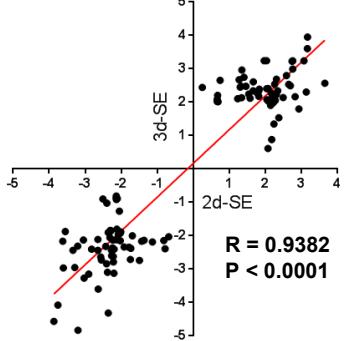
3d-SE vs. 2d-SE



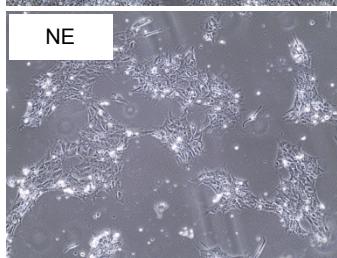
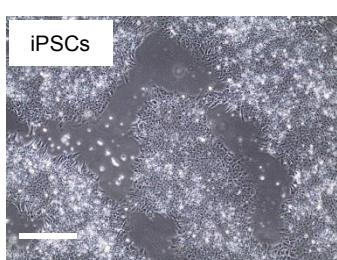
e

Upstream regulators

3d-SE vs. 2d-SE



f



g

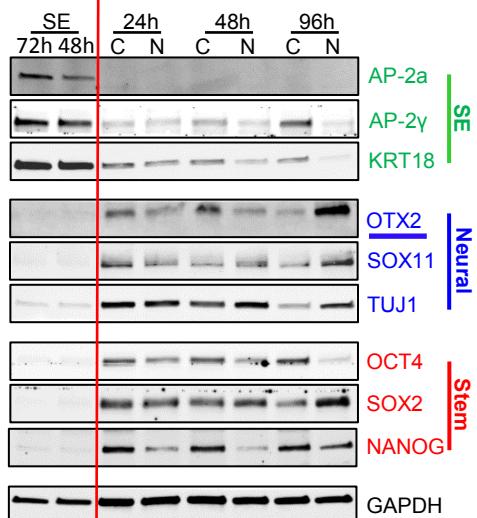


Fig.S4

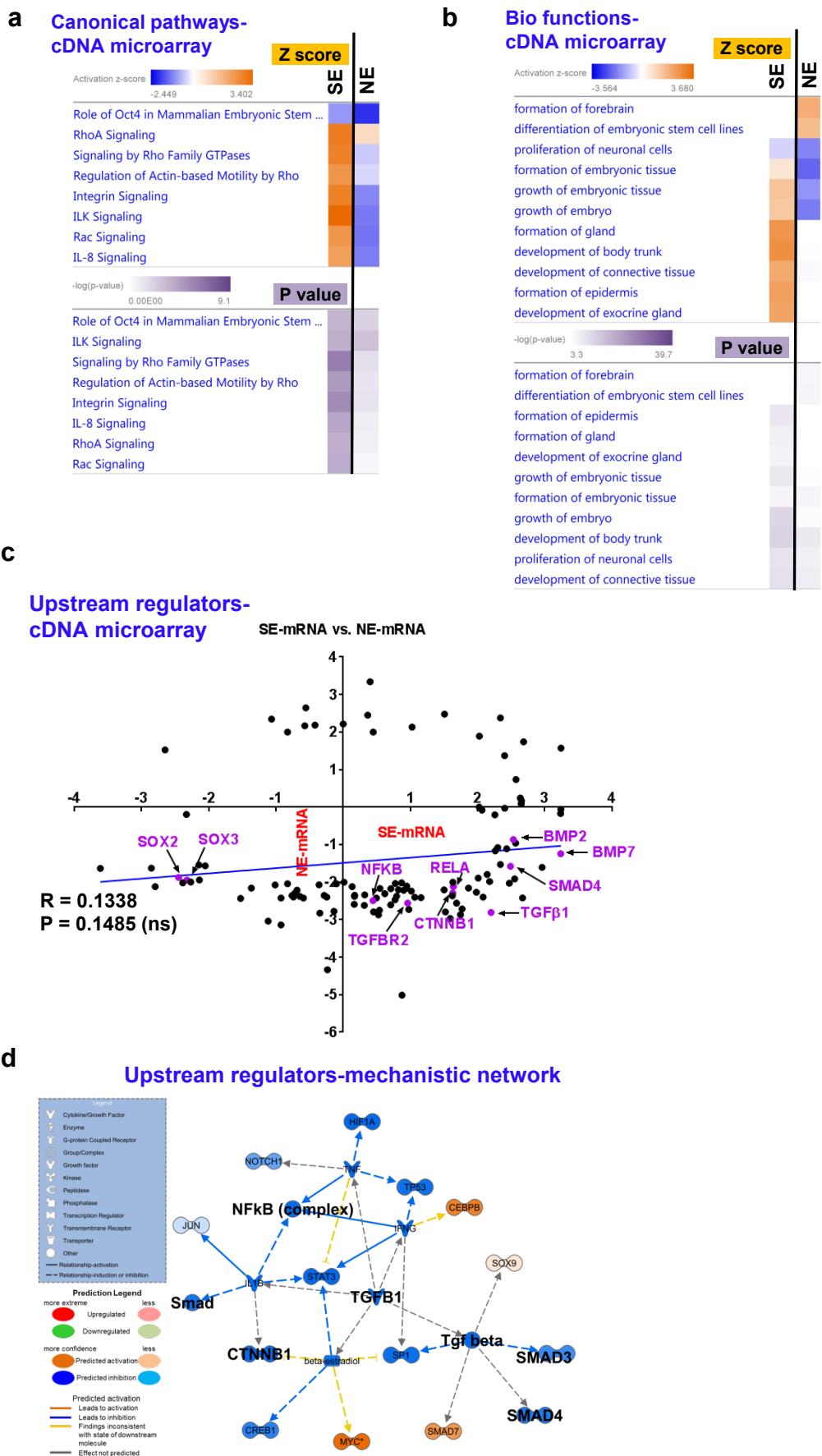
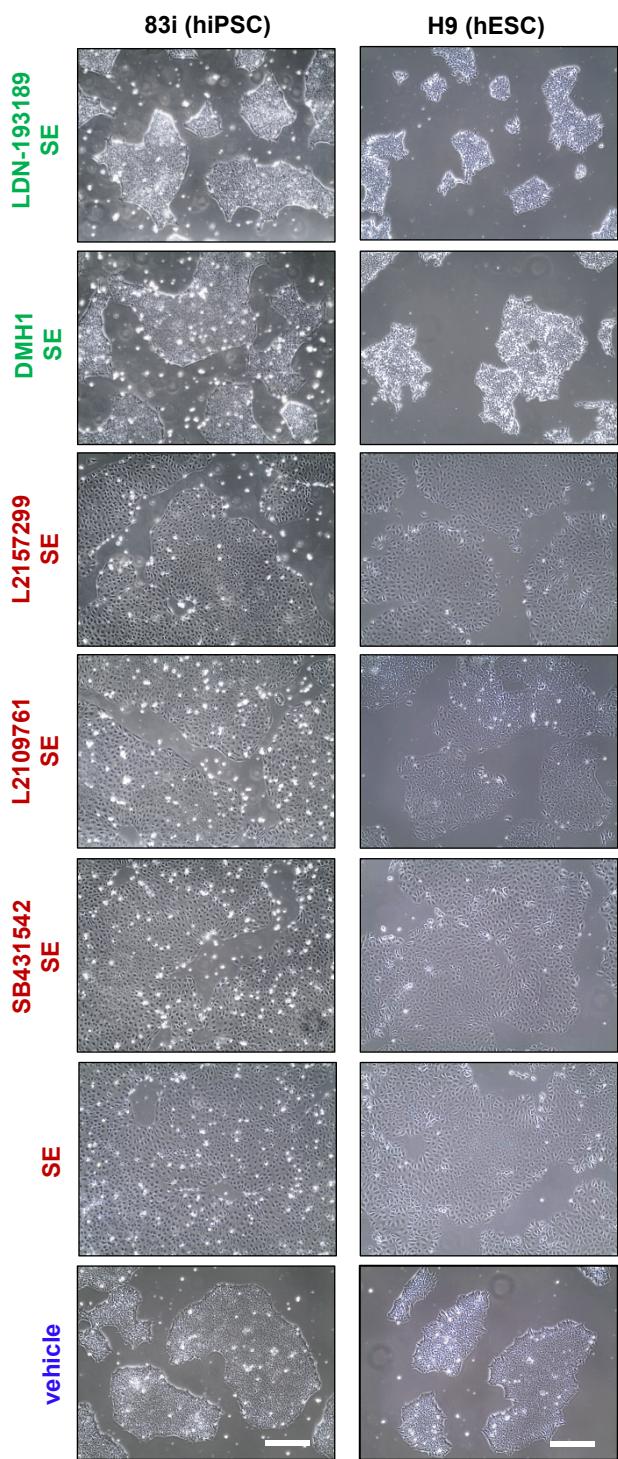


Fig.S5

a



b

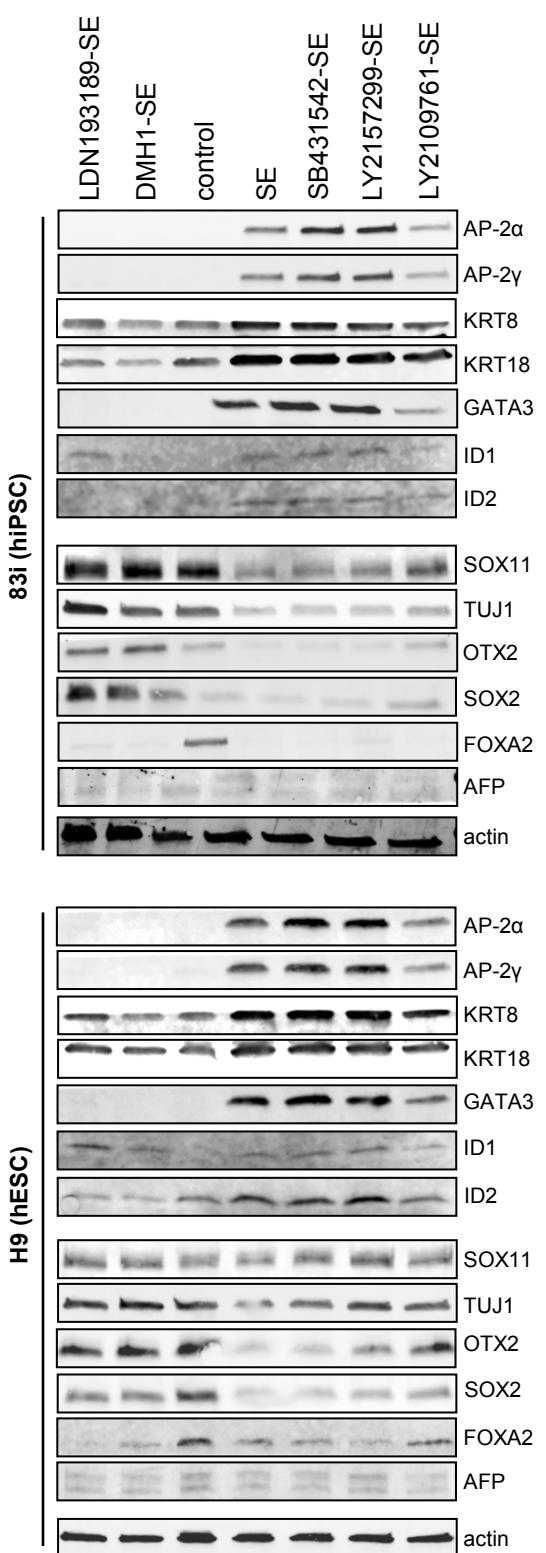


Fig.S6

