Induction of renal senescence marker protein-30 (SMP30) expression by testosterone and its contribution to urinary calcium absorption in male rats

Po-Han Lin¹, Cai-Yun Jian¹, Jou-Chun Chou², Chien-Wei Chen¹, Chih-Chieh Chen¹, Christina Soong³, Sindy Hu^{4,5}, Fu-Kong Lieu³*, Paulus S. Wang^{1,2,6,7}*, Shyi-Wu Wang^{4,8}*

¹Institute and Department of Physiology, School of Medicine, National Yang-Ming University, Taipei 11221;

²Medical Center of Aging Research, China Medical University Hospital, Taichung 40402;

³Department of Rehabilitation, Cheng Hsin General Hospital, Taipei 11212;

⁴Aesthetic Medical Center, Department of Dermatology, Chang Gung Memorial Hospital, Taoyuan 33378;

⁵Department of Medicine, College of Medicine, Chang Gung University, Taoyuan 33302;

⁶Department of Biotechnology, College of Health Science, Asia University, Taichung 41354;

⁷Department of Medical Research and Education, Taipei Veterans General Hospital, Taipei 11217,

⁸Department of Physiology and Pharmacology, College of Medicine, Chang Gung University, Taoyuan 33302, Taiwan, Republic of China.

*Corresponding authors and persons to whom reprint requests should be addressed:

[1] Paulus S Wang, Ph.D.

Institute and Department of Physiology, School of Medicine, National Yang-Ming University, Taipei 11221, Taiwan, Republic of China.

Tel: +886-2-2826-7082 Fax: +886-2-2826-4049 E-mail: pswang3879@gmail.com

[2] Shyi-Wu Wang, Ph.D.
Department of Physiology and Pharmacology, College of Medicine, Chang-Gung University, Taoyuan 33302, Taiwan, Republic of China.
Tel: +886-3-211-8800 ext.5253
Fax: +886-2-211-8700
E-mail: swwang@mail.cgu.edu.tw

[3] Fu-Kong Lieu, M.D.
Department of Rehabilitation, Cheng Hsin General Hospital, Taipei 11212, Taiwan, Republic of China.
Tel: +886-2-2826-4400 ext.3806
Fax: +886-2-2826-4400 ext.3814
E-mail: ch2276@chgh.org.tw

Supplemental Mertinals and Methods:

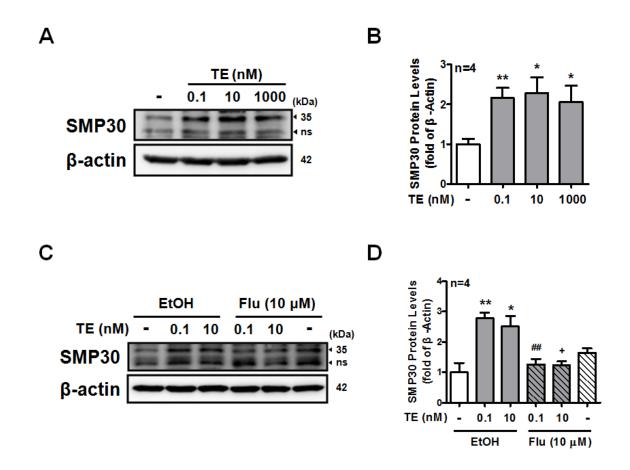
NRK52E cell culture

Normal rat kidney epithelial NRK52E line was purchased from the Bioresource Collection and Research Center (BCRC, Hsinchu, Taiwan). NRK52E cells were maintained in DMEM medium (Gibco Laboratories, Grand Island, NY, USA) supplemented with 5% fetal bovine serum (FBS, Hyclone, *GE* Healthcare Life Sciences), penicillin G (100 IU/ml; Sigma-Aldrich), streptomycin (100 μ g/ml; Sigma-Aldrich), sodium bicarbonate (1.5 g/L; Sigma-Aldrich), glucose (4.5g/L; affymetrix USB, Santa Clara, CA, USA) and L-Glutamine (4 mM; Sigma-Aldrich) in a humidified incubator (37°C, 5% CO₂).

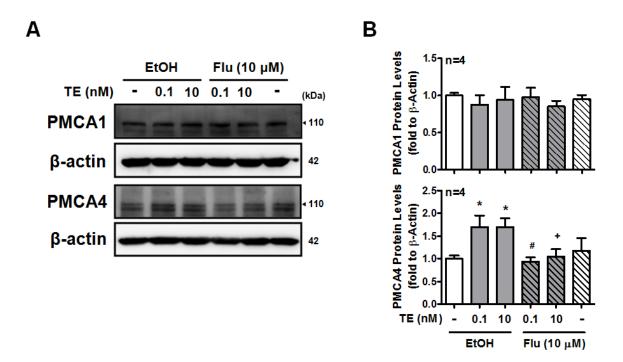
For the expreriments, NRK52E cells had grown to confluence, culture medium was removed and rinsed twice with 1x PBS, following cells were rested by incubation in serum- and phenol red-free DMEM (Gibco) medium for 12 h before the beginning of treatment. The conditional medium was then replaced and incubation for the times indication. The detailed procedures were described in the legends.

Western blot analysis for NRK52E cell

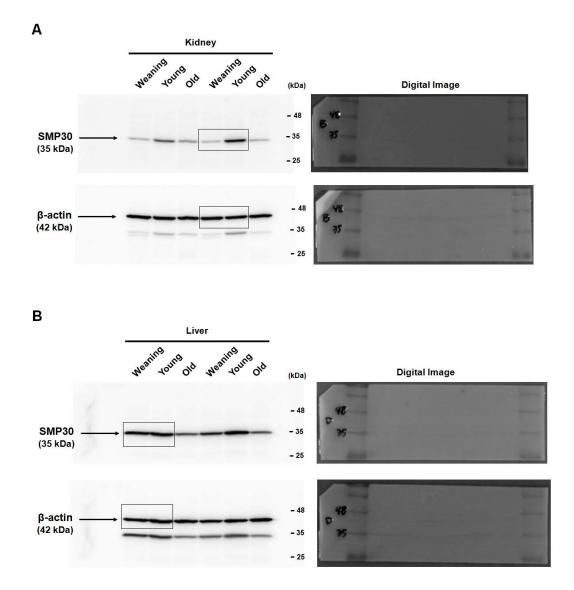
The process of protein lysate was silimar to the tissue samples and SDS-page preparation was described in the methods of manuscript. Primary antibodies used were mouse anti-SMP30 (1:350), rabbit anti-PMCA1 (1:300) and goat anti-PMCA4 (1:300) antibodies (Santa Cruz), and sencondary antibodies were biotinylated goat anti-mouse IgG (1:2000), goat anti-rabbit IgG (1:2000) and rabbit anti-goat IgG (1:2000) (Vector Laboratories, Burlinggame, CA, USA). Binding sensitivity was enhanced by using ABC solution (Standard VECTASTAIN ABC Kit; Vector Laboratories). Subsequently the membranes were developed with enhanced chemiluminescence (PerkinElmer Life Sciences). To quantify the intensity of the protein expression levels, the signal was recorded by Luminescence Imaging System LAS-4000 (*GE* Healthcare Life Sciences).



Supplementary Fig. S1. Involvement of the AR in the testosterone-induced SMP30 expression in NKR52E cells. (A) After resting for 12 h, NRK52E cells were treated with testosterone (0.1, 10, 1000 nM) (Sigma-Aldrich), or absolute alcohol (EtOH) as control vehicle, in serum- and phenol red-free DMEM medium for next 12 hr. At the end of time, cells were collected and the expression of SMP30 protein level was analyed by Western blot, and (B) the quantitative analysis of SMP30 in normalization to β-actin was shown. (C) Cells were pretreated with flutaminde (10 μM) for 1 h and then treated with testosterone (0.1 and 10 nM) or EtOH in serum- and phenol red-free DMEM medium for 12 h. At the end of time, cells were collected and the expression of SMP30 protein level was analyed by the quantitative analysis of SMP30 protein level was analyed by the testosterone (0.1 and 10 nM) or EtOH in serum- and phenol red-free DMEM medium for 12 h. At the end of time, cells were collected and the expression of SMP30 protein level was analyed by Western blot, and (D) the quantitative analysis of SMP30 in normalization to β-actin was shown. Data represent means ± SEM (n=4). **P*<0.05, ***P*<0.01 as compared with the testosterone (0.1 nM) treatment group; **P*<0.01 as compared with the testosterone (0.1 nM) treatment group; **P*<0.01 as compared with the testosterone (0.1 nM) treatment group; **P*<0.01 as compared with the testosterone (0.1 nM) treatment group; **P*<0.01 as compared with the testosterone (10 nM) treatment group. The gels have been run under the same experimental conditions and cropped blots were shown. The entire of membrane pictures of Supplementary Fig. S1 were presented in the Supplementary Fig. S10. TE, testosterone; Flu, flutaminde; ns, non-specific binding.



Supplementary Fig. S2. Effect of TE on the plasma membrane Ca²⁺ ATPase (PMCA) expression in NKR52E cells. (A) After resting for 12 h, NRK52E cells were pretreated with flutamind (10 μ M) for 1 h and then treated with testosteone (0.1 and 10 nM), or EtOH as control vehicle, in serumand phenol red-free DMEM medium for 12 h. At the end of time, cells were collected and the expression of PMCA isoforms, PMCA1 and PMCA4, protein levels were analyed by Western blot, and (B) the quantitative analysis of PMCA1 and PMCA4 in normalization to β -actin was shown, respectively. Data represent means ± SEM (n=4). **P*<0.05 as compared with the testosterone (0.1 nM) treatment group; ⁺*P*<0.01 as compared with the testosterone (10 nM) treatment group. The gels have been run under the same experimental conditions and cropped blots were shown. The entire of membrane pictures of Supplementary Fig. S2 were presented in the Supplementary Fig. S11. TE, testosterone; Flu, flutaminde.

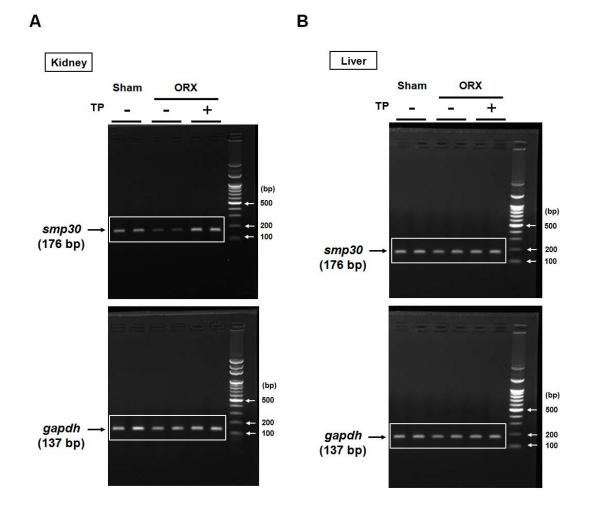


Supplementary Fig. S3. The original blot pictures of Fig. 1. in the text. Two individual rats of each group were shown in the membrane in the kidney and liver, respectively. (A) Corresponding to the section of kidney in Fig. 1B. (B) Corresponding to the section of liver in Fig. 1B. Size distribution of the molecular weight marker was shown in the digital images. The cropping lines and the molecular weight of target proteins were indicated.

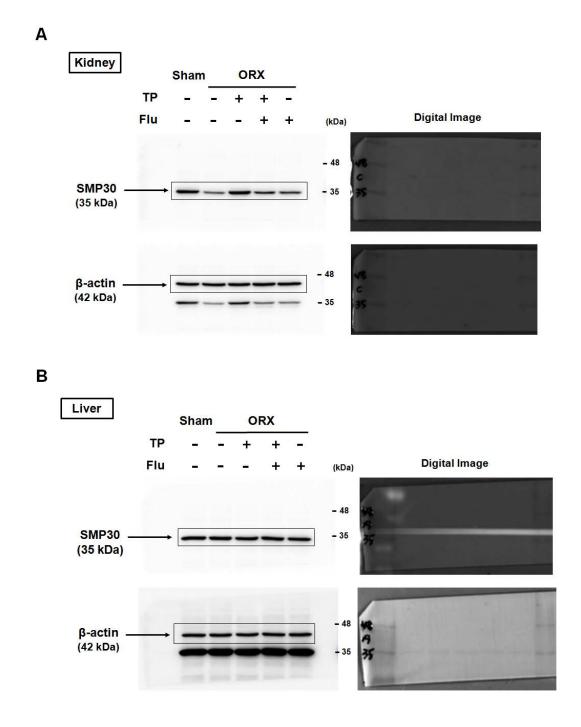
Kidney ORX Sham TP +**Digital Image** (kDa) - 48 SMP30 - 35 (35 kDa) - 25 - 48 β-actin (42 kDa) - 35 - 25 В Liver Sham ORX TP + **Digital Image** (kDa) - 48 SMP30 - 35 (35 kDa) - 25 - 48 β-actin (42 kDa) - 35 - 25

Α

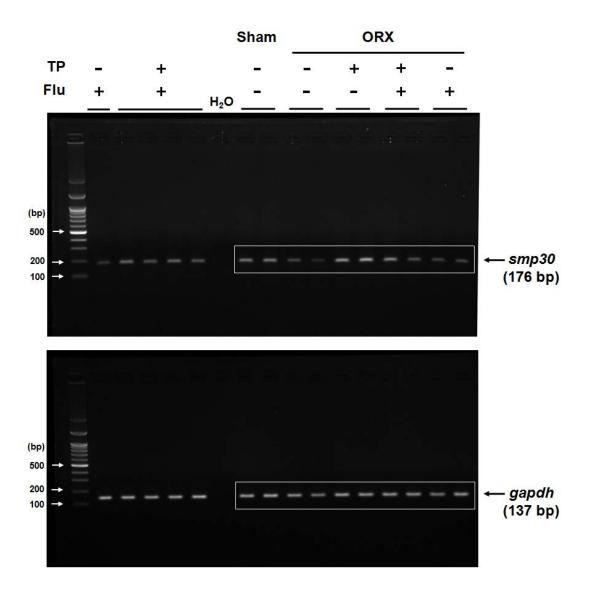
Supplementary Fig. S4. The original blot pictures of Fig. 2. in the text. Two individual rats of each group were shown in the membrane in the kidney and liver, respectively. (A) Corresponding to the section of kidney in Fig. 2A. (B) Corresponding to the section of liver in Fig. 2A. Size distribution of the molecular weight marker was shown in the digital images. The cropping lines and the molecular weight of target proteins were indicated.



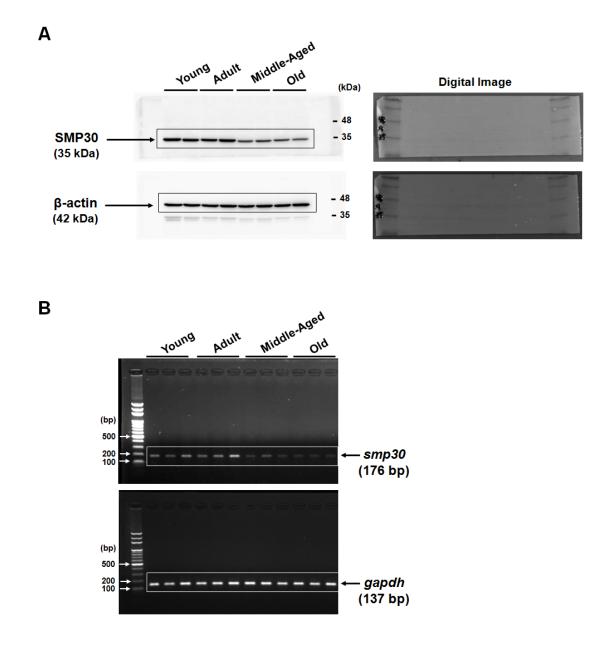
Supplementary Fig. S5. The original gel pictures of Fig. 2. in the text. Two individual rats of each group were shown in the membrane in the kidney and liver, respectively. (A) Corresponding to the section of kidney in Fig. 2D. (B) Corresponding to the section of liver in Fig. 2D. The cropping lines and the molecular weight of target PCR products were indicated.



Supplementary Fig. S6. The original blot pictures of Fig. 3. in the text. (A) Corresponding to Fig. 3B. (B) Corresponding to Fig. 3C. Size distribution of the molecular weight marker was shown in the digital images. The cropping lines and the molecular weight of target proteins were indicated.

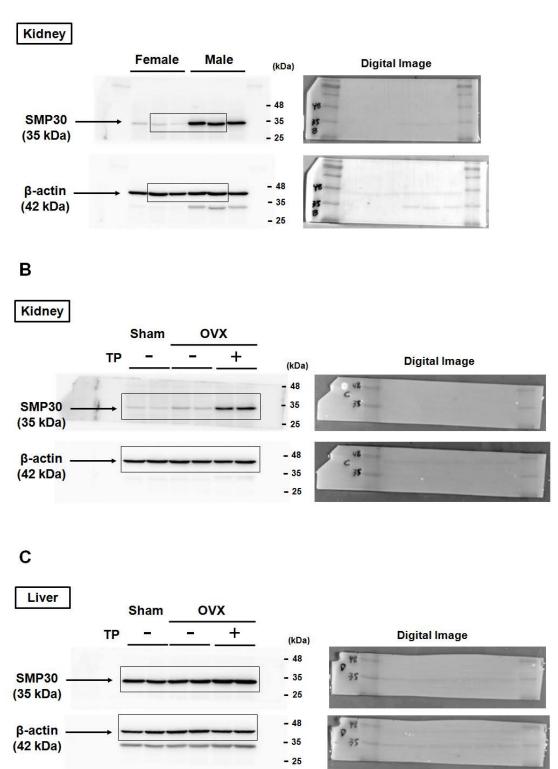


Supplementary Fig. S7. The original gel pictures of Fig. 3. in the text. Corresponding to Fig. 3D. Two individual rats of each group were shown in the gel. The cropping lines and the molecular weight of target PCR products were indicated. H_2O , as a negative control.

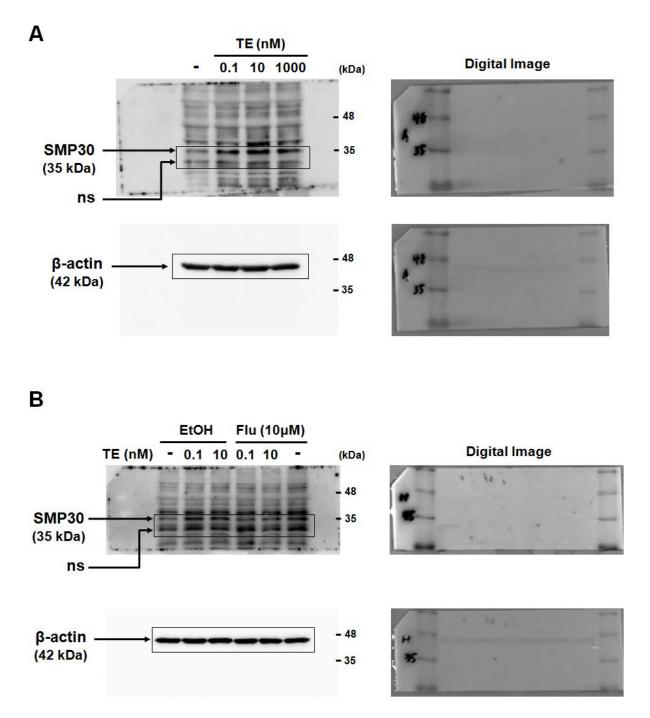


Supplementary Fig. S8. The original blot/gel pictures of Fig. 4. in the text. (A) Corresponding to Fig. 4A. Two individual rats of each group were shown in the membrane. Size distribution of the molecular weight marker was shown in the digital images. (B) Corresponding to Fig. 4B. Three individual rats of each group were shown in the gel. The cropping lines and the molecular weight of target proteins and PCR products were indicated.

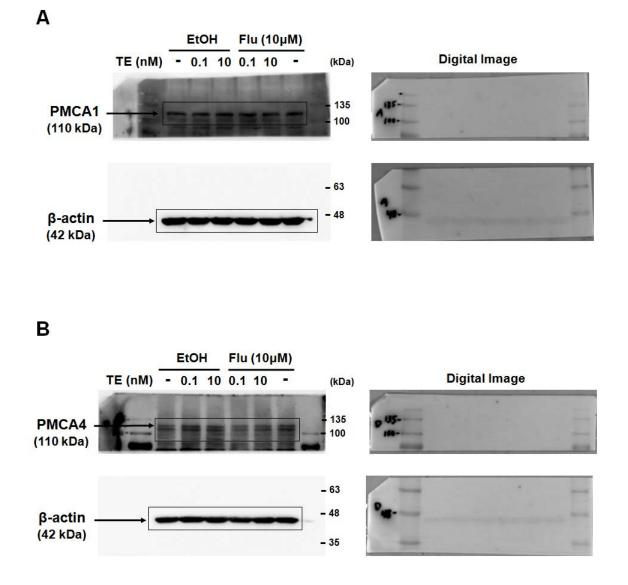
Α



Supplementary Fig. S9. The original blot pictures of Fig. 5. in the text. (A) Corresponding to the section of kidney in Fig. 5A. Three individual rats of each group were shown in the membrane. (B) Corresponding to the section of kidney in Fig. 5B. (C) Corresponding to the section of liver in Fig. 5B. Two individual rats of each group were shown in the membrane in the kidney and liver, respectively. Size distribution of the molecular weight marker was shown in the digital images. The cropping lines and the molecular weight of target proteins were indicated.



Supplementary Fig. S10. The original blot pictures of Supplementary Fig. S1. in the supplementary information. (A) Corresponding to Supplementary Fig. S1A. (B) Corresponding to Supplementary Fig. S1B. Size distribution of the molecular weight marker was shown in the digital images. The cropping lines and the molecular weight of target proteins were indicated. ns, non-specific binding.



Supplementary Fig. S11. The original blot pictures of Supplementary Fig. S2. in the supplementary information. (A) Corresponding to the section of PMCA1 in the top panel of Supplementary Fig. S2A. (B) Corresponding to the section of PMCA4 in the bottom panel of Supplementary Fig. S2A. Size distribution of the molecular weight marker was shown in the digital images. The cropping lines and the molecular weight of target proteins were indicated.

	#Rat number (n)	Sham	ORX	ORX+TP	ORX+TP+Flu	ORX+Flu	One-way ANOVA
Plasma Ca ²⁺ (mg/dl)	5	9.7 ± 0.2	9.6 ± 0.3	9.5 ± 0.4	9.7 ± 0.5	10.3 ± 0.4	<i>P</i> = 0.367
Plasma PTH (ng/ml)	8	3.1 ± 0.3	3.4 ± 0.3	3.5 ± 0.2	3.0 ± 0.3	3.6 ± 0.3	<i>P</i> = 0.497
Urinary Volume (ml/24h)	6	23.0 ± 0.7	23.5 ± 2.0	22.3 ± 2.8	25.2 ± 1.2	23.3 ± 2.0	<i>P</i> = 0.825
Water Intake (ml/24h)	6	41.0 ± 2.3	42.8 ± 2.0	38.5 ± 3.4	42.7 ± 2.0	42.2 ± 4.3	<i>P</i> = 0.775
Food Intake (mg/24h)	6	22.7 ± 1.2	25.2 ± 1.6	21.5 ± 2.4	24.2 ± 0.9	23.6 ± 1.0	<i>P</i> = 0.399

Table S1. Effect of testosterone on plasma Ca²⁺, plasma PTH, urinary volume, water and food intake levels

Table S2. Aging effect on plasma Ca²⁺, plasma PTH, urinary volume, water and food intake levels in male rats

	#Rat number (n)	Young	Adult	Middle-Aged	Old	One-way ANOVA
Plasma Ca ²⁺ (mg/dl)	5	10.1 ± 0.3	-	10.1 ± 0.3	9.8 ± 0.2	<i>P</i> = 0.680
Plasma PTH (ng/ml)	8	3.9 ± 0.3	3.6 ± 0.2	4.0 ± 0.7	3.8 ± 0.4	<i>P</i> = 0.927
Urinary Volume (ml/24h)	5	24.2 ± 2.7	23.4 ± 3.0	24.0 ± 3.6	24.2 ± 3.2	<i>P</i> = 0.997
Water Intake (ml/24h)	5	44.2 ± 4.5	40.4 ± 6.2	40.8 ± 5.4	41.0 ± 6.9	<i>P</i> = 0.951
Food Intake (mg/dl)	5	21.5 ± 0.6	20.4 ± 1.2	21.4 ± 1.5	21.9 ± 2.0	<i>P</i> = 0.845